

## THE SYNTHESIS OF NEW ADENOSINE A<sub>3</sub> SELECTIVE LIGANDS CONTAINING BIOISOSTERIC ISOXAZOLES

John P. Mogensen, Stanley M. Roberts,<sup>§</sup> Andrew N. Bowler, Christian Thomsen,  
and Lars J. S. Knutsen\*

*Health Care Discovery, Novo Nordisk A/S, Novo Nordisk Park, DK 2760 Måløv, Denmark*

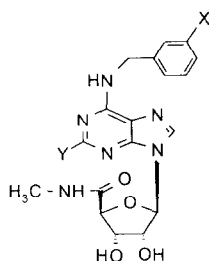
*<sup>§</sup>Department of Chemistry, University of Liverpool, P.O. Box 147, Liverpool, L69 3BX, England*

Received 7 November 1997; accepted 5 June 1998

**Abstract.** The synthesis and purinergic receptor binding of novel adenosine A<sub>3</sub> ligands is described. Many selective A<sub>3</sub> receptor agonists e.g. *N*-(3-iodobenzyl)adenosine-5-methyluronamide (IB-MECA) contain a 4-ribosylalkylamide moiety. We found that this amide and other 4-functional groups could be replaced with an isosteric isoxazole, and the target molecules retained potent binding to the recombinant human A<sub>3</sub> receptor.

© 1998 Elsevier Science Ltd. All rights reserved.

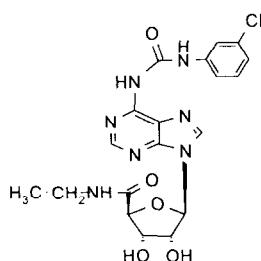
Adenosine is a naturally occurring purine nucleoside that has a variety of well-documented regulatory functions and physiological effects. The pharmacologically distinct adenosine (P<sub>1</sub>) receptor subtypes are known as A<sub>1</sub>, A<sub>2A</sub>, A<sub>2B</sub> (of high and low affinity), and A<sub>3</sub>.<sup>1</sup> Since the discovery of the A<sub>3</sub> receptor early this decade<sup>2,3</sup> an effort has been made to identify ligands at this subtype,<sup>4</sup> but very few selective A<sub>3</sub> agonists have been described to date.



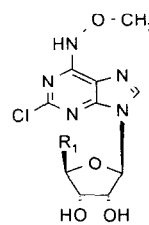
1, X = Y = H, *N*-benzyl MECA

2, X = I, Y = H, IB-MECA

3, X = I, Y = Cl, 2-Cl IB-MECA



4



5, R<sub>1</sub> = Cl-CH<sub>2</sub>-, NNC 21-0113

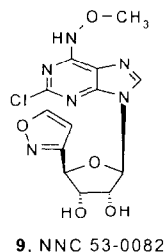
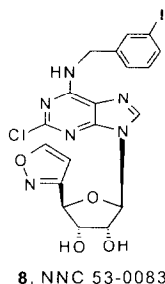
6, R<sub>1</sub> = CH<sub>2</sub>=CH-, NNC 21-0238

7, R<sub>1</sub> = CH<sub>3</sub>OCH<sub>2</sub>-, NNC 53-0055

The first A<sub>3</sub> agonists to be discovered,<sup>5,6</sup> represented by structures 1–3, contain a bulky purine 6-amino substituent (a benzyl group) along with a 4-alkylamide moiety. Potency is increased further by a 3-iodine, as in 2 or 3. This theme of bulky amino substituents was also reflected in a later series<sup>7</sup>, in which compound 4 was the most potent. The recently revealed<sup>8</sup> A<sub>3</sub> ligands 5–7 represent a structural departure compared to 1–4, since they contain both the smaller *N*-methoxy substituent and a new range of ribose 5-modifications. Potential future therapeutic applications of adenosine A<sub>3</sub> ligands appear to be involvement with the control of cytokines, [such as tumor necrosis factor alpha (TNF-α)<sup>9</sup>], as anti-inflammatory agents or in the treatment of asthma.<sup>4</sup>

\*Corresponding author. \*Current address: Cerebrus Limited, Oakdene Court, Winnersh, WOKINGHAM, Berkshire, RG41 5UA, England. Tel. (+44) 118 977 3133; fax. (+44) 118 989 9300; E-mail L.Knutsen@Cerebrus.ltd.uk

The use of five-membered heterocycles as isosteric replacements for esters and amides is well established.<sup>10–12</sup> As part of our strategy of incorporating new functionalities in the design of novel adenosine receptor agonists<sup>13,14</sup> we decided to investigate replacement of the methylamide in the selective A<sub>3</sub> agonist 2-chloro-*N*-(3-iodobenzyl)adenosine-5'-methyluronamide (2-Cl-IB-MECA) **3** with an isoxazole moiety. Compound **8** is thereby analogous to compound **3**, and the *N*-alkoxypurine **9** has structural similarities to compounds **5–7**.

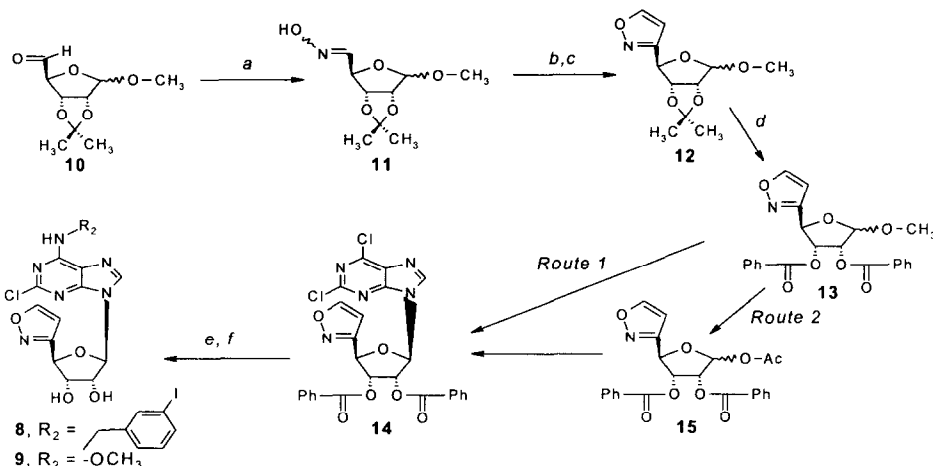


## Chemistry

The synthesis of the novel isoxazole derivatives **8** and **9** is shown in Scheme 1. We opted for a strategy in which the isoxazole was first assembled at the ribose 4-position, and the other heterocycle, a 9*H*-purine, was introduced subsequently. The oxime **11**, derived from the ribose aldehyde **10**,<sup>15</sup> was isolated as an *E/Z* mixture, and from this the isoxazole **12** was built in two steps.<sup>16</sup> First the corresponding nitrile oxide was generated from **11**, and a 1,3-dipolar addition was performed using either TMS-acetylene or vinyl acetate as dipolarophile. The addition products were treated with aqueous sodium hydroxide or DBU to provide the isoxazole **12**. Once the heterocycle was in place, the 1-methylethylidene group was hydrolyzed under acidic conditions and the 2- and 3-hydroxyls were reprotected as the dibenzoate **13**.<sup>17</sup> Attachment of the 2,6-dichloro-9*H*-purine to the ribose moiety was achieved by two different methods. The first method (Route 1, Scheme 1), utilized the 1-*O*-methyl sugar **6** and Vorbruggen's procedure<sup>18,19</sup> with silylated dichloropurine gave 2,6-dichloro-9-[2,3-di-*O*-benzoyl-4-(3-isoxazolyl)-β-*D*-erythrofuranosyl]-9*H*-purine **14** in low yield. The more successful method (Route 2, Scheme 1) involved replacing the ribose 1-*O*-methyl with a 1-*O*-acetyl group, and reacting **15** with 2,6-dichloro-9*H*-purine in an acid-catalyzed process<sup>13</sup> at 140 °C *in vacuo*, provided the key intermediate **14** in good yield. The 6-chloro atom in **14** was displaced with either 3-iodobenzylamine or *O*-methylhydroxylamine, which after de-blocking provided the target purine derivatives **8** and **9** containing the isosteric ribose 4-isoxazole.

## Pharmacological Data

The novel purine derivatives **8** and **9** were evaluated *in vitro* in adenosine A<sub>1</sub> and A<sub>2A</sub> binding assays utilizing published procedures.<sup>6,19</sup> For A<sub>3</sub> receptor binding, the human A<sub>3</sub> receptor was expressed in a human embryonic kidney cell line (HEK 293) and [<sup>125</sup>I]-AB-MECA binding was performed as described below (see reference 19); data for **3** in Table 1 is from ref. 4. The data displayed in Table 1 shows that the isoxazoles **8** and **9** display selectivity in binding towards human A<sub>3</sub> receptors, with relatively weak binding observed to A<sub>1</sub> and A<sub>2A</sub> receptors.

**Scheme 1. Synthesis of adenosine A<sub>3</sub> receptor ligands 8 and 9.**

Reagents and conditions: (a) NH<sub>2</sub>OH.HCl, pyridine; (b) NBS, (CH<sub>3</sub>)<sub>3</sub>Si-acetylene, Et<sub>3</sub>N, 0 °C, 24 h (44%) or NBS, vinyl acetate, Et<sub>3</sub>N, 0 °C, 24 h; (c) 1N NaOH, used in the (CH<sub>3</sub>)<sub>3</sub>Si case (57%) or DBU, THF used in the vinyl acetate case (65% for 2 steps); (d) Dowex 50 H<sup>+</sup>, benzoyl chloride, pyridine, DMAP, 0 °C, 12 h (56% for 2 steps). Route 1: 2,6-dichloro-9H-purine, 1,1,1,3,3,3-hexamethyldisilazane, (CH<sub>3</sub>)<sub>3</sub>Si-triflate, 12 h (12%); Route 2: (1) Ac<sub>2</sub>O, AcOH, H<sub>2</sub>SO<sub>4</sub>, 10 h; (2) 2,6-dichloro-9H-purine, 140 °C, vacuum, 5 h (66%); (e) CH<sub>3</sub>ONH<sub>2</sub>.HCl, Et<sub>3</sub>N, 1,4-dioxan, reflux, 20 h or 3-I-PhCH<sub>2</sub>NH<sub>2</sub>.HCl, Et<sub>3</sub>N, 1,4-dioxan, reflux; (f) 10% NH<sub>3</sub> in CH<sub>3</sub>OH, 4 h (55% in the case of 8 and 34% in the case of 9).

**Table 1. Adenosine receptor binding**

Structure	A <sub>1</sub> receptor, K <sub>i</sub> (nM) (Inhibition of rat brain <sup>3</sup> [H]-R-PIA binding)	A <sub>2A</sub> receptor, K <sub>i</sub> (nM) (Inhibition of rat striatum <sup>3</sup> [H]-CGS 21680 binding)	A <sub>3</sub> receptor, K <sub>i</sub> (nM) (Inhibition of [ <sup>125</sup> I]-AB-MECA binding, human receptor)
1	2400	2100	41
3	54	56	1.1
5	74	4650	26
6	1230	>10000	20
8	1900	1900	7.8
9	620	>10000	31

We conclude from the above data that the isoxazole moiety in 8 is able to act as a bioisostere for methylamides such as 1 and 3<sup>4</sup> and that the same heterocycle is also able to replace the 5'-substituents in A<sub>3</sub> ligands 5 - 7. Functional studies are in progress to establish whether 8 and 9 act as adenosine A<sub>3</sub> agonists or antagonists.

**Acknowledgements:** We wish to thank Prof. Mikael Begtrup of the Royal School of Pharmacy, Denmark; Dr. John Bondo Hansen (Novo Nordisk) for useful discussions, and V. Sik of the University of Exeter, U.K. for NMR analyses. Financial support by Novo Nordisk for studies at Exeter University is gratefully acknowledged.

**Reference and Notes**

1. Fredholm, B. B.; Abbracchio, M. P.; Burnstock, G.; Daly, J. W.; Harden, T. K.; Jacobson, K. A.; Williams, M. *Pharmacol. Rev.* **1994**, *46*, 143-156.
2. Zhou, Q. Y.; Li, C. Y.; Olah, M. E.; Johnson, R. A.; Stiles, G. L.; Civelli, O. *Proc. Natl. Acad. Sci. U.S.A.*, **1992**, *89*, 7432-7436.
3. Salvatore, C. A.; Jacobson, M. A.; Taylor, H. E.; Linden, J.; Johnson, R. G. *Proc. Natl. Acad. Sci. U.S.A.*, **1993**, *90*, 10365-10369.

4. Linden, J. *TIPS* **1994**, 15, 298-306; Jacobson, K. A.; Kim, H. A.; Siddiqi, S. M.; Olah, M. E.; Stiles, G. L.; von Lubitz, D. K. J. E. *Drugs of the Future* **1995**, 20, 689-699; Collis, M. G.; Hourani, S. M. O. *TIPS* **1993**, 14, 360-366.
5. Gallo-Rodriguez, C.; Ji, X. D.; Melman, N.; Siegman, B. D.; Sanders, L. H.; Orlina, J.; Pu, Q. L.; Olah, M. E.; van Galen, P. J. M.; Stiles, G. L.; Jacobson, K. A. *J. Med. Chem.* **1994**, 37, 636-646.
6. Kim, H. O.; Ji, X. D.; Siddiqi, S. M.; Olah, M. E.; Stiles, G. L.; Jacobson, K. A. *J. Med. Chem.* **1994**, 37, 3614-3621; Siddiqi, S. M.; Jacobson, K. A.; Esker, J. L.; Olah, M. E.; Li, X.-d.; Melman, N.; Tiwari, K. N.; Secrist III, J. A.; Schneller, S.; Cristalli, G.; Stiles, G. L.; Johnson, C. R.; IJzerman, A. P. *J. Med. Chem.* **1995**, 38, 1174-1178.
7. Baraldi, P. G.; Caciari, B.; Spalluto, G.; Ji, X. D.; Olah, M. E.; Stiles, G. L.; Dionisotti, S.; Zocchi, C.; Ongini, E.; Jacobson, K. A. *J. Med. Chem.* **1996**, 39, 802-806.
8. Bowler, A. N.; Olsen, U. B.; Thomsen, C.; Knutsen, L. J. *S. Drug Dev. Res.* **1996**, 173. See also Knutsen, L. J. S.; Olsen, U. B.; Bowler, A. N. WO 97/33591A1 (18.09.1997).
9. Sajjadi, F. G.; Takabayashi, K.; Foster, A. C.; Domingo, R. C.; Firestein, G. S. *J. Immunol.* **1996**, 156, 3435-3442.
10. King, F. D. In *Medicinal Chemistry: Principles and Practice*; King, F. D., Ed., The Royal Society of Chemistry: Cambridge; 1994, pp 206-225.
11. Andersen, K. E.; Lundt, B. F.; Jørgensen, A. S.; Bræstrup, C. *Eur. J. Med. Chem.* **1996**, 32, 417-425.
12. Williams, M.; Nadzan, A. M. In *Textbook of Drug Design and Development*; Krosgaard-Larsen, P.; Bundgaard, H., Eds.; Harwood Academic: Chur, 1991; pp 1-25.
13. Knutsen, L. J. S.; Lau, J.; Sheardown, M. J.; Thomsen, C. *Bioorg. Med. Chem. Lett.* **1993**, 3, 2661-2666. See also Imai, K.; Nohara, A.; Honjo, M. *Chem Pharm. Bull.* **1966**, 14, 1377-1381.
14. Knutsen, L. J. S.; Lau, J.; Eskesen, K.; Sheardown, M. J.; Thomsen, C.; Weis, J. U.; Judge, M. E.; Klitgaard, H. In *Adenosine and Adenine Nucleotides: From Molecular Biology to Integrative Physiology*; Belardinelli, L. and Pelleg, A., Eds.; Kluwer Academic: Norwell, 1995; pp 479-487.
15. Ranganathan, R. S.; Jones, G. H.; Moffatt, J. G. *J. Org. Chem.*, 1974, 39, 290-298.
16. Humphrey, G. R.; Wright, S. H. B. *J. Het. Chem.* **1989**, 26, 23-24.
17. Physical data for selected compounds: 2,3-Dibenzoyl-1-*O*-methyl- $\beta$ -D-erythrofuranos-4-yl)-isoxazole **13**,  $m/z$  409.11570 ( $M^+$ ),  $C_{21}H_{18}NO_7$  requires 409.11615;  $\delta_H$  (400 MHz,  $CDCl_3$ ) 3.52 (3H, s,  $CH_3O$ ), 5.22 (1H, d,  $J = 1$  Hz, H-1), 5.59 (1H, dd,  $J = 1$  and 7.5 Hz, H-2), 5.74 (1H, d,  $J = 5$  Hz, H-4), 5.95 (1H, dd,  $J = 5$  and 7.5 Hz, H-3), 6.50 (1H, d,  $J = 2$  Hz, H-4 in isoxazole), 7.50 (6H, m, Ph H-3, H-4 and H-5), 7.90 (2H, dd,  $J = 1$  and 7 Hz, Ph H-2 and H-6), 8.02 (2H, dd,  $J = 1$  and 7 Hz, Ph H-2 and H-6), 8.45 (1H, d,  $J = 2$  Hz, isoxazole H-4);  $\delta_C$  (100 MHz,  $CDCl_3$ ) 55.29, 74.57, 74.77, 74.98, 102.33, 106.45, 127.88, 127.98, 128.05, 128.28, 128.33, 128.59, 129.32, 129.35, 129.66, 132.98, 133.11, 158.75, 161.79, 164.74. 2,6-Dichloro-9-[2,3-di-*O*-benzoyl-4-(3-isoxazolyl)- $\beta$ -D-erythrofuranosyl]-9H-purine **14**,  $m/z$  565.05813 ( $M^+$ ),  $C_{26}H_{17}N_5O_6Cl_2$  requires 565.05559;  $\delta_H$  (400 MHz,  $CDCl_3$ ) 5.71 (1H, d,  $J = 3.5$  Hz, H-1'), 6.25 (2H, m, H-2' and H-3'), 6.21 (1H, d,  $J = 5$  Hz, H-4'), 6.70 (1H, d,  $J = 1.5$  Hz, isoxazole H-4), 7.5 (6H, m, H-3, 2 x Ph H-4 and H-5), 7.90 (2H, dd,  $J = 1$  and 7.5 Hz, Ph H-2 and H-6), 8.10 (2H, dd,  $J = 1$  and 7.5 Hz, Ph H-2 and H-6), 8.52 (1H, d,  $J = 1.5$  Hz, isoxazole H-4), 8.60 (1H, s, H-8);  $\delta_C$  (100 MHz,  $CDCl_3$ ) 73.80, 74.01, 77.23, 87.12, 102.76, 128.01, 128.10, 128.13, 128.21, 129.39, 129.44, 129.80, 130.01, 133.52, 133.58, 144.31, 151.87, 152.33, 152.36, 152.80, 159.45, 159.54, 164.67, 164.77. 2-Chloro-9-[4-(3-isoxazolyl)- $\beta$ -D-erythrofuranosyl]-*N*-methoxyadenine **9**,  $m/z$  368.06289 ( $M^+$ ),  $C_{11}H_{13}N_5O_4Cl$  requires 368.06360.  $\delta_H$  (400 MHz,  $DMSO-d_6$ ) 3.72 (3H, s,  $CH_3O$ ), 4.51 (1H, m, H-3'), 4.72 (1H, m, H-2'), 5.05 (1H, dd,  $J = 5$ , H-4'), 5.74 (1H, d,  $J = 6$ , 3'-OH), 5.78 (1H, d,  $J = 6$ , 2'-OH), 6.02 (1H, d,  $J = 5$ , H-1'), 6.81 (1H, s,  $J = 2$ , isoxazole H-4), 8.41 (1H, s, N-H), 8.96 (1H, s,  $J = 2$ , isoxazole H-5); 8.50 (1H, s, H-8);  $\delta_C$  (100 MHz,  $DMSO-d_6$ ) 56.42, 74.71, 76.11, 76.90, 84.75, 101.89, 146.01, 150.33, 151.83, 152.78, 153.65, 160.34, 161.24. 2-Chloro-*N*-3-iodobenzyl-9-[4-(3-isoxazolyl)- $\beta$ -D-erythrofuranosyl]adenine **8**,  $m/z$  553.99893 ( $M^+$ ),  $C_{19}H_{16}N_5O_4ICl$  requires 553.99663;  $\delta_H$  (400 MHz,  $CDCl_3$ ) 4.85 (5H, m, *N*-CH<sub>2</sub>, H-2', H-3' and 3'-OH), 5.43 (1H, d,  $J = 4$  Hz, H-4'), 6.05 (1H, d,  $J = 5.5$  Hz, H-1'), 6.49 (1H, d,  $J = 2$  Hz, isoxazole H-4), 7.09 (1H, dd,  $J = 8$  Hz, Ph H-5), 7.35 (1H, d,  $J = 8$  Hz, Ph H-6), 7.63 (1H, d,  $J = 8$  Hz, Ph H-4), 7.72 (1H, s, Ph H-2), 7.91 (1H, s, H-8), 8.50 (1H, d,  $J = 2$  Hz, isoxazole H-5). For full experimental details, see Knutsen, L. J. S.; Olsen, U. B.; Roberts, S. M.; Varley, D. R.; Bowler, A. N. WO 98/01459A1 (15.01.1998).
18. Vorbruggen, H.; Hofle, G. *Chem. Ber.* **1981**, 114, 1256 - 1268; Vorbruggen, H.; Krolkiewicz, K.; Bennua, B. *Chem. Ber.* **1981**, 114, 1234-1253.
19. Klitgaard, H.; Knutsen, L. J. S.; Thomsen, C. *Eur. J. Pharmacol.* **1993**, 224, 221-228. Adenosine A<sub>1</sub> receptor binding was performed on whole cells using a Krebs-Henseleit assay buffer supplemented with 2 U/ml adenosine deaminase. Cells were incubated for 45 min at 37 °C with test compounds and 0.2 nM [<sup>125</sup>I]-AB-MECA and washed twice with ice-cold assay buffer. The cells were solubilized with NaOH (2 M) and bound [<sup>125</sup>I]-AB-MECA was quantitated in a  $\gamma$ -counter. IC<sub>50</sub> values (mean of 2-3 experiments) from competition curves were calculated by a non-linear regression analysis and K<sub>d</sub> values were calculated using the equation  $K_d = IC_{50}/(1 - [I]/K_d)$ , where [I] is the concentration of radioligand and K<sub>d</sub> the dissociation constant. These parameters were: ([I] ~ 1 nM, K<sub>d</sub> = 1.2 nM) for [<sup>3</sup>H] *R*-PIA binding, ([I] ~ 2 nM, K<sub>d</sub> = 7 nM) for [<sup>3</sup>H] CGS 21680 binding and ([I] ~ 0.2 nM, K<sub>d</sub> = 0.6 nM) for [<sup>125</sup>I]-AB-MECA binding.