



3-TRIFLUOROMETHYLCARBACEPHEMS: SYNTHESIS OF BROAD SPECTRUM ANTIBACTERIAL COMPOUNDS

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Abstract. The enhanced stability of the carbacephem nucleus over the corresponding cephalosporin nucleus has allowed the synthesis of 7-arylglycyl-3-trifluoromethyl-carbacephems. These unique carbacephems possess broad spectrum activity and high stability to both plasmid and chromosomally mediated β-lactamases. © 1998 Elsevier Science Ltd. All rights reserved.

Introduction. Carbacephems are structurally related to cephalosporins, differing only by replacement of the sulfur atom with a methylene group. First reported in the early 1970's, the carbacephems maintain the potent antibacterial activity characteristic of the parent cephalosporins. More recently, a cephalosporin vs. carbacephem comparative study demonstrated carbacephem analogs possess enhanced chemical stability. We recognized the carbacephem's equipotent biological activity concomitant with decreased chemical reactivity provided potential medicinal chemistry opportunities. 4-6

D-(-)-arylglycyl side chains attached to the C-7 position of cephalosporin antibiotics allow active transport across the intestinal barrier. Cefaclor 24, the leading oral cephalosporin, is used to treat Gram positive and a limited number of Gram negative organisms. We focused our efforts on obtaining oral carbacephalosporins that possessed a broader antibacterial spectrum and/or enhanced Gram negative activity compared to cefaclor. Considering the carbacephalosporin's enhanced chemical stability, we targeted arylglycylcarbacephems whose cephem counterparts demonstrated low potency in vitro, presumably due to chemical instability. Our target in this study was the 3-trifluoromethyl-7-arylglycylcarbacephems whose cephem analogs were virtually inactive against bacteria at pH 7.9 Lack of in vitro activity of the 3-CF₃-7-arylglycylcephems at physiological pH can probably be attributed to chemical instability of the zwitterion form, since activity greater than cephalexin was reported at pH 5.9 Table I shows a number of representative 3-CF₃ carbacephalosporins acylated with a variety of arylglycyl side chains.

Chemistry. Synthesis of the 7-arylglycyl-3-trifluoromethyl-carbacephems 10 11–23 proceeded according to Scheme 1. We hoped to incorporate the 3-trifluoromethyl group from readily available p-nitrobenzyl (6R,7S)-7-(phenoxyacteamideo)-1-carba-1-dethia-3-[[(trifluoromethyl)sulfonyl]oxy]-3-cephem-4-carboxylate $1.^{11}$ Triflate conversion to vinyl bromide 12 provided a useful substrate for investigating the applicability of known [CF₃M] couplings to aryl halides. 13 Replacement of the p-nitrobenzyl ester with benzhydryl was required due to the difficulty of PNB removal at a later stage. Displacement of the triflates 1 and 2 with LiBr in 2,6-lutidine produced the bromides as a mixture of Δ -2 and Δ -3 isomers that could be isomerized with DBU and crystallized to give the desired Δ -3-vinyl bromides 3 and 4. Pregeneration of Burton's 14 [CF₃Cu] reagent using CF₂Br₂/DMF/zinc/CuBr and subsequent coupling with the bromides (3 and 4) gave the desired trifluoromethyl derivatives 5 and 6. The zwitterion 8, used to obtain compounds 11, 12, 16, and 20–23 was obtained from 5 in three steps: (1) the t-Boc protecting group was first added to the amide of the phenoxyacteyl side chain using

Boc anhydride, (2) treatment with mild base at ice bath temperature removed the phenoxyacteyl group to yield t-Boc amide, ¹⁵ and (3) deprotection with TFA/triethylsilane removed the t-Boc amide and benzhydryl ester. Acylation with a t-Boc phenylglycyl isobutanyl mixed anhydride, t-Boc removal with TFA, and subsequent purification using reverse phase chromatography gave the desired 3-trifluoromethyl-7-arylglycyl carbacephems 11, 12, 16, and 20–23.

Scheme 1

(a) LiBr, Lutidine, DMF. (b) DBU, CH₂CL₂. (c) Zn/HCl. (d) Ph₂CN₂, CH₃CN. (e) Zn, CF₂Br₂, DMF, CuBr. (f) Zn/HCl. (g) allylBr, Bu₄NHSO₄. (h) t-Boc anhydride, DMAP. (i) LiOH. (j) TFA, Et₃SiH. (k) t-Boc-arylglycyl mixed anhydride. (l) TFA. (m) PCl₅, i-BuOH. (n) t-Boc-arylglycine, 1-Chloro-3,5-dimethoxytriazine, NMM, CH₂Cl₂. (o) Pd(PPh₃)₄, N-Bu₃SnH.

Compounds 13–15 and 17–19 were made by the following procedure: PNB ester removal from 6 with zinc/HCl and esterification of the tetrabutylammonium salt with allyl bromide yielded 7 whose phenoxyacetyl side chain was cleaved with PCl₅ and butanol to give 9. The amino acid was activated with 1-chloro-3,5-dimethoxytriazine 16 and coupled to the 7-amino group. Deallylation with (Ph₃)₄Pd $^{\circ}$ /tributyltin hydride 17 and reverse phase chromatography provided the desired 3-trifluoromethyl-7-phenylglycyl carbacephems 13–15 and 17–19. Racemic *t*-Boc amino acids were used in the acylation when chiral amino acids were not available (see Table I) and the diastereomers were separated during reverse phase chromatography.

Table 1: 111 yigiyeyi compus										
Compd	X	X'	Route	Yield						
11	Н	Н	A	43%						
12	OH	Н	Α	40%						
13	F	H	В	14%						
14	H	Br	В	9%						
15	NH_2	Н	В	10%						
16*	CH ₃	Н	Α	2%						
17	H	NHSO ₂ Et	В	12%						
18	Н	F	В	14%						
19	H	CF ₃	В	11%						
20*	Н	CH ₃	Α	13%						
21	H	NHSO ₂ CH ₃	Α	39%						
22	OH	C1	Α	41%						
23*	Cl	Cl	Α	3%						

Table I: Arylglycyl Compds

In Vitro Activity. The 3-CF₃ carbacephems' microbiological activity is displayed in Tables 2 and 3 with cefaclor 24 as reference. Overall activity against gram positive organisms (e.g., Staphylococcus sp. and Streptococcus s.p.) is comparable to cefaclor. Methicillin-resistant S. aureus, X400 and S13E, do show a tendency towards less resistance to these compounds than to cefaclor. The 3-CF₃ carbacephems show the same potent activity as cefaclor against Gram negative H. influenza and E. coli strains, but in several other Gram negative organisms the majority of analogs show significantly more activity than cefaclor. Surprisingly lower MIC's for plasmid and chromosomally mediated β-lactamase producing strains (e.g., Klebsiella KAE and Enterobacter cloacae 265A) indicated significant β-lactamase stability that is virtually unseen in comparator compounds. In addition, many Gram negative organisms including Serratia sp., Citrobacter sp., Proteus sp., and Acinetobacter sp. are notably more susceptible to these new carbacephems in vitro. More details regarding these findings will be published later.

Several arylglycyl analogs have potency comparable to or greater than the parent compound (11). 4-OH-Phenylglycine 12 is slightly less potent than the parent 11 against Staphylococcal organisms, but in most other cases, is 1-2 dilutions more potent. Similiarly, 4-OH, 3-Cl-phenylglycine 22 shows 1-2 dilutions lower MIC's than 11 against most organisms. With several exceptions, all compounds show good activity against Gram positive bacteria as well as *H. influenza*, *E. coli*, and *Klebsiella*. Aryl substituents other than hydroxyl result in reduced potency against the Gram negative organisms in Table 3, particularly 3-CF₃-phenylglycine and 3,4-diCl-phenylglycine 19 and 23.

^{*} Racemic amino acid starting material

	Staphylococcus			Streptococcus sp.			H. influ		E. coli		Klebsiella	
	aureus		Gp. A pneu. D		Amp-S B-lact		sens B-lact		pneumoniae			
Compd	X1.1	X400	S13E	C203	PARK	2041	C.L.	76	EC14	TEM	X26	KAE
11	0.5	32	8	0.03	0.25	16	1	1	0.5	1	0.25	4
12	0.5	64	32	0.01	0.03	4	0.5	0.5	0.25	0.5	0.06	1
13	1	64	32	0.06	0.25	32	0.5	1	1	1	0.5	8
14	1	16	8	0.13	0.13	8	0.5	1	4	4	0.25	16
15	2	64	64	NG	NG	16	1	1	0.5	1	0.5	16
16	1	4	4	0.06	0.13	8	2	4	2	4	0.5	8
17	4	32	32	0.06	0.13	32	1	4	1	1	0.25	32
18	2	64	32	0.06	0.25	32	2	2	4	2	2	16
19	2	16	16	NG	0.13	16	1	1	8	16	1	64
20	1	4	4	0.03	0.25	8	1	2	2	4	1	16
21	2	16	16	0.06	0.13	16	2	2	0.5	1	0.5	16
22	1	8	8	0.03	0.06	4	0.5	0.5	0.5	1	0.13	4
23	0.5	64	8	0.02	0.13	16	2	2	32	16	1	64
24	0.5	128	128	0.06	0.5	32	2	1	1	2	0.25	128+

Table 2: Microbiological Activity I*

MIC's (minimum inhibitory concentrations) were determined by a standardized agar-dilution method (ref 18) using Mueller-Hinton agar (BBL Microbiology System, Cockeysville, MD) and an inoculum of approximately 10,000 CFU/spot.

Table 3: Microbiological Activity II*

	Enterobacter				Salm.	Serratia			Providencia		Citro	Acin
	aerogenes		cloacae					morg.	stu.	rett.		
Compd	C32	EB17	EB5	265a	X514	X99	SE3	PR15	PR33	C24	CF17	AC12
11	4	2	4	8	0.25	8	16	32	4	8	4	32
12	0.25	0.5	1	1	0.06	2	16	16	1	4	4	16
13	8	8	8	8	0.5	16	32	16	8	8	8	64
14	16	32	64	64	4	128	128	16	16	16	32	16
15	8	4	16	32	0.25	16	32	8	32	16	16	64
16	8	8	16	32	4	16	64	8	16	16	16	16
17	16	16	32	32	1	64	128	16	16	32	32	64
18	16	8	16	32	2	32	64	32	16	16	16	32
19	128	128	4	128	32	128	128+	32	32	64	128	32
20	32	4	16	32	8	16	32	8	8	16	16	32
21	2	4	16	32	2	16	32	8	8	16	16	64
22	2	1	8	16	0.125	4	16	4	4	4	4	32
23	128	128	128	128	32	128+	128+	32	32	64	128	128
24	32	8	32	128+	0.5	64	128+	128+	64	64	128+	128

^{*} See ref 19 for organism descriptions.

Summary. A new series of compounds, the 3-trifluoromethyl-7-arylglycyl-carbacephems, has been synthesized by 3-vinyl bromide coupling with in situ generated [CF₃Cu]. Microbiological potency of this series is high at pH 7 with incubation at 37°C and, in most cases, is comparable to cefaclor. These compounds show surprising activity against strains with plasmid and chromosomally mediated β-lactamases. In vivo activity of compounds 11–12 will be reported in due course.

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References and Notes

- 1. Bruwin, D. M.; Lowe, G. Parker; J. J. Chem. Commun. 1971, 865, Bender, D. R.; Bjeldanes, L. F.; Knapp, D. R.; McKean, D. R.; Rapoport, H. J. Org. Chem. 1973, 38, 3439.
- 2. Guthikonda, R. N.; Cama, L. D.; Christensen, B. G. J. Am. Chem. Soc. 1974, 96, 7584.
- 3. Blaszczak, L. C.; Brown, R. F.; Cook, G. C., Hornback, W. J.; Hoying, R. C.; Indelicato, J. M.; Jordan, C. L.; Katner, A. S.; Kinnick, M. D.; McDonald, III, J. H.; Morin, J. M.; Munroe, J. E.; Pasini, C. E. J. Org. Chem. 1990, 33. 1656.
- 4. Cook, G. K.; Hornback, W. J.; Jordan, C. L.; McDonald, III, J. H.; Munroe, J. E. J. Org. Chem. 1989, 54 5828.
- Crowell, T. A.; Halliday, B. D.; McDonald, III, J. H.; Indelicato, J. M.; Pasini, C. E.; Wu, E. C. Y. J. Med. Chem. 1989, 32, 2436.
- Cook, G. K.; McDonald, III, J. H.; Alborn, Jr., W. Boyd, D. B.; Eudaly, J. A.; Indelicato, J. M.; Johnson, R.; Kasher, J. S.; Pasisi, C. E.; Preston, D. A.; Wu, E. C. Y. J. Med. Chem. 1989, 32, 2442.
- 7. Dantzig, A. H.; Duckworth, D. C.; Tabas, L. B. Biochim. Biophys. Acta 1994, 1191, 7.
- 8. Tsuji, E.; Nakashima, E.; Kagami, I.; Yamana, T. J. Pharm. Sci. 1981, 70, 768.
- 9. Kawano, Y.; Watanabe, T.; Sakai, J.; Nagano, M.; Nishimura, T.; Miyadera, T. *Chem. Pharm. Bull.* 1980, 28, 70.
- 10. Some details of this chemistry were disclosed in the following publication: Fisher, J. W; Dunigan, J. M. Hatfield, L. D.; Hoying, R. C.; Ray, J. E.; Thomas, K. L. *Terahedron Lett.* **1993**, *34*, 4755.
- 11. Evans, D. A.; Sjorgen, E. B.; *Tetrahedron. Lett.* 1985, 26, 3787. Bodurow, C. C.; Boyer, B. D.; Brennen, J.; Bunnell, C. A.; Burks, J. E.; Carr, M. A.; Doecke, C. W.; Eckrich, T. M.; Fisher, J. W.; Gardner, J. P.; Graves, B. J.; Hines, P.; Hoying, R. C.; Jackson, B. G.; Kinnick, M. D.; Kochert, C. D.;

- Lewis, J. S.; Luke, W. D.; Moore, L. L.; Morin, J. M.; Nist, R. L.; Prather, D. E.; Sparks, D. L.; Vladuchick, W. C. *Tetrahedron Lett.* **1989**, *30*, 2321.
- 12. Procedure according to U.S. Patent #5142039.
- 13. Kobayashi, Y.; Yamamoto, K.; Kumadaki, I.; Tetrahedron Lett. 1979, 42, 4071.
- 14. Burton, D. J. J. Amer. Chem. Soc. 1985, 107, 5014.
- 15. Spry, D. O.; Snyder, N. J.; Bhala, A. R.; Pasini, C. E.; Indelicato, J. M. Heterocycles, 1987, 26, 2911.
- 16. Kaminski, Z. Tetrahedron .Lett. 1985, 26, 2901.
- 17. Guide, F.; Dangles, O.; Balavoine, G.; Lavielle, S.; Marquet, A. J. Org. Chem. 1987, 52, 4984.
- National Committee for Clinical Laboratory standards, 1990; approved standard M7-A2. Methods for dilution antimicrobial susceptibility testing for bacteria that grow aerobically, 2nd ed. National Committee for Clinical Laboratory Standards, Villanova, Pa.
- 19. The bacterial strains used in this study are a collection of Eli Lilly and Co. isolates (see refs Hornback, W. J.; Munroe, J. E.; Counter, F. T. J. Antibiot. 1994, 1052. Counter, F. T.; Ensminger, P. W.; Preston, D. A.; Wu, C. Y. E.; Greene, J. M.; Felty-Duckworth, A. M.; Paschal, J. W.; Kirst, H. A. Antimicrob. Agents Chemother. 1991, 35, 1116. The strains used were X1.1 S. aureus pencillin sensitive; X400 S. aureus methicillin resistant and macrolide resistant; S13E S. aureus methicillin resistant penicillin resistant macrolide sensitive (PBP2a producer); C203 Streptococcus pyogenes Gp. A; Park S. pneumoniae; 2041 Enterococcus faecalis Gp. D streptococcus penicillin susceptible; C. L. Haemophilus influenzae ampicillin sensitive; 76 H. influenzae b-lactamase producing; EC14 Escherichia coli sensitive; TEM E. coli b-lactamase producing; X26 Klebsiella pneumoniae; KAE K. pneumoniae; C32 Enterobacter aerogenes; EB17 E. aerogenes; EB5 E. cloacae; 265a E. cloacae b-lactamase producing; X514 Salmonella; X99 Serratia; SE3 Serratia; PR15 Morganella morganii; PR33 Providencia stuartii; C24 Providencia rettgeri; CF17 Citrobacter; AC12 Acinetobacter.