

# Maximum entropy and population heterogeneity in continuous cell cultures meet experimental data, preliminary results

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## 1 Materials and Method

### 1.1 Model framework

The model framework used is detailed explained in *Cossio1* and *Cossio2*

### 1.2 Experimental data

Experimental data was taken from *Rath*, in this work the author performed 6 continuous cultures, ( $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$ ,  $F$ ), with the cell line AGE1.HN.AA1, which parental line AGE1.HN was established by the company ProBioGen (ProBioGen AG, Berlin, Germany) from a tissue sample of a human brain. All cultures feed mediums was based on the standard 42-Max-UB-medium, which is serum-free and was especially developed for the AGE1.HN cell line *Table1*. The experiments was run under various conditions, differing mainly in the dilution rate ( $D$ ) and the feed medium composition of glucose ( $GLC$ ), glutamine ( $GLN$ ) and galactose ( $GAL$ ) *Table2*.

For each experiment, a steady state condition was reached, ( $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$ ,  $F01$ ), and several observables was reported *Tables3 – 4*. Particularly relevant for this work was the growth rate ( $\mu$ ),  $D$ , the viable cell density ( $Xv$ ) and the medium concentration ( $s$ ) and derived uptake rate ( $u$ ) for a set of metabolites ( $GLC$ , lactose ( $LAC$ ),  $GLN$ , ammonium ( $NH4$ ),  $GAL$ , pyruvate ( $PYR$ ), glutamate ( $GLU$ ), alanine ( $ALA$ ), asparagine ( $ASP$ )). Unit conversion was required for make experimental data and models compatible. For this propose the only external data needed was the cell mass density. It was used  $0.25 \text{ pgDW}/\mu\text{m}^3$  *Niklas*.

### 1.3 Preparing GEMs

In order to preliminary evaluate the capacity of the model to reproduce the experimental data, two Genome-scale Metabolic Models (*GEM*) was prepared.

Substance	Value	Dimension	Analytical method
Pluronic	1.0	g/L	as stated by Xell
NaHCO <sub>3</sub>	2.1	g/L	as stated by Xell
Osmolality	290.0	mOsm/kg	FPDO <sup>a</sup>
pH value	7.4	-	pH meter
GALC	0.5	g/L	AEC <sup>b</sup>
GLC	5.5	g/L	Bioprofile
AMM	0.3	mM	Bioprofile
LAC	0.0	g/L	Bioprofile
PYR	2.9	mM	AEC
GLU	636.9	μM	AEC
ALA	437.1	μM	AEC
ARG	1588.2	μM	AEC
ASN	920.4	μM	AEC
ASP	2197.9	μM	AEC
CYS	963.1	μM	AEC
GLY	1196.0	μM	AEC
HIS	642.7	μM	AEC
ILE	1744.9	μM	AEC
LEU	1893.2	μM	AEC
LYS	1256.0	μM	AEC
MET	601.3	μM	AEC
PHE	1039.4	μM	AEC
PRO	1040.5	μM	AEC
SER	3027.4	μM	AEC
THR	1502.4	μM	AEC
TRP	383.8	μM	AEC
TYR	1109.7	μM	AEC
VAL	1811.9	μM	AEC

a: freezing point depression osmometer (FPDO); b: anion exchanger chromatography (AEC);

Table 1: Measured medium composition of the 42-MAX-UB standard medium. Extracted from *Rath*

Exp. ID	<i>DR</i> (1/h)	Preculture (passage no.)	GLC (mM)	GLN (mM)	GALC (mM)
A	0.0140	7	10	5	3
B	0.0120	7	10	5	3
C	0.0100	5	10	5	3
D	0.0150	8	10	2	3
E	0.0133	4	8	2	3
F01	0.0150	10	10	5	0

Table 2: The dilution rates, preculture ages and the 42-Max-UB-medium modified components concentrations used in *Rath* for the 6 steady states. Table adapted from *Rath*

Variable	Exp. A		Exp. B		Exp. C		Exp. D		Exp. E	
	Average $\pm$ SD	Rel. SD (%)	Average $\pm$ SD	Rel. SD (%)	Average $\pm$ SD	Rel. SD (%)	Average $\pm$ SD	Rel. SD (%)	Average $\pm$ SD	Rel. SD (%)
Setpoints:										
<i>DR</i> (1/h)	0.0140	-	0.0120	-	0.0100	-	0.0150	-	0.0133	-
GLC feed conc. (mM)	10.0	-	10.0	-	10.0	-	10.0	-	8.0	-
GLN feed conc. (mM)	5.0	-	5.0	-	5.0	-	2.0	-	2.0	-
GALC feed conc. (mM)	3.0	-	3.0	-	3.0	-	3.0	-	3.0	-
GLC/GLN ratio (mol/mol)	2	-	2	-	2	-	5	-	4	-
<i>X<sub>V</sub></i> (E6 cells/mL)	2.490 $\pm$ 0.100	4.0	2.745 $\pm$ 0.118	4.3	2.735 $\pm$ 0.056	2.0	1.489 $\pm$ 0.043	2.9	0.998 $\pm$ 0.025	2.5
<i>X<sub>D</sub></i> (E6 cells/mL)	0.210 $\pm$ 0.032	15.2	0.291 $\pm$ 0.021	7.4	0.378 $\pm$ 0.047	12.5	0.086 $\pm$ 0.006	6.5	0.075 $\pm$ 0.014	18.5
<i>CV<sub>V</sub></i> ( $\mu$ mol/L)	6.56 $\pm$ 0.48	7.4	7.19 $\pm$ 0.67	9.4	6.23 $\pm$ 0.21	3.4	4.33 $\pm$ 0.27	4.9	3.40 $\pm$ 0.17	4.9
$\mu$ (1/h)	0.0152 $\pm$ 0.0002	1.2	0.0133 $\pm$ 0.0001	1.1	0.0114 $\pm$ 0.0002	1.4	0.0159 $\pm$ 0.0001	0.6	0.0143 $\pm$ 0.0002	1.4
<i>CD</i> ( $\mu$ m)	17.1 $\pm$ 0.25	1.5	17.2 $\pm$ 0.19	1.1	16.3 $\pm$ 0.24	1.5	17.7 $\pm$ 0.30	1.5	18.7 $\pm$ 0.20	1.1
GLC (mM)	0.0 $\pm$ 0.00	-	0.0 $\pm$ 0.00	-	0.0 $\pm$ 0.00	-	0.0 $\pm$ 0.00	-	0.0 $\pm$ 0.00	-
LAC (mM)	2.1 $\pm$ 1.52	-	0.5 $\pm$ 0.02	-	0.6 $\pm$ 0.04	-	12.5 $\pm$ 0.60	5.0	12.7 $\pm$ 0.40	3.0
GLN (mM)	0.8 $\pm$ 0.08	10.0	5.5 $\pm$ 0.19	3.5	5.9 $\pm$ 0.09	1.5	1.5 $\pm$ 0.10	11.1	0.5 $\pm$ 0.10	13.7
AMM (mM)	3.4 $\pm$ 0.22	6.7	1.9 $\pm$ 0.06	2.9	1.7 $\pm$ 0.02	0.9	2.1 $\pm$ 0.10	2.1	1.6 $\pm$ 0.02	1.6
GALC (mM)	1.9 $\pm$ 0.08	4.5	1.9 $\pm$ 0.06	2.9	1.7 $\pm$ 0.02	0.9	2.1 $\pm$ 0.10	2.5	2.4 $\pm$ 0.10	3.2
PYR (mM)	0.2 $\pm$ 0.00	1.8	0.1 $\pm$ 0.00	3.2	0.1 $\pm$ 0.00	3.2	0.3 $\pm$ 0.04	12.6	1.1 $\pm$ 0.10	5.3
GLU (mM)	1.2 $\pm$ 0.08	6.2	1.3 $\pm$ 0.01	0.5	1.0 $\pm$ 0.05	5.0	0.7 $\pm$ 0.10	12.9	0.8 $\pm$ 0.10	12.8
ALA (mM)	1.4 $\pm$ 0.09	6.4	0.2 $\pm$ 0.03	13.8	0.1 $\pm$ 0.01	14.2	0.3 $\pm$ 0.02	8.9	1.0 $\pm$ 0.05	5.2
ASP (mM)	1.6 $\pm$ 0.06	3.9	1.5 $\pm$ 0.05	3.2	1.3 $\pm$ 0.02	1.2	2.4 $\pm$ 0.17	7.1	2.8 $\pm$ 0.30	10.8
ALAAT (mg/L)	57.9 $\pm$ 3.03	5.2	81.0 $\pm$ 9.21	11.4	72.1 $\pm$ 1.09	1.5	50.9 $\pm$ 1.50	2.9	44.6 $\pm$ 2.80	6.2
<i>Y<sub>CVV/glc</sub></i> ( $\mu$ mol/mol)	711 $\pm$ 47	6.6	827 $\pm$ 77	9.4	753 $\pm$ 17	2.3	482 $\pm$ 30	6.3	518 $\pm$ 18	3.5
<i>Y<sub>CVV/gln</sub></i> ( $\mu$ mol/mol)	1679 $\pm$ 140	8.3	1770 $\pm$ 145	8.2	1601 $\pm$ 52	3.3	3617 $\pm$ 215	6.0	2837 $\pm$ 201	7.1
<i>Y<sub>inc/glc</sub></i> (mol/mol)	0.2 $\pm$ 0.15	-	0.0 $\pm$ 0.01	-	0.0 $\pm$ 0.00	-	1.2 $\pm$ 0.10	5.3	1.8 $\pm$ 0.10	3.2
<i>Y<sub>am/gln</sub></i> (mol/mol)	0.8 $\pm$ 0.05	6.6	1.2 $\pm$ 0.04	3.6	1.3 $\pm$ 0.03	1.9	1.0 $\pm$ 0.05	4.9	1.1 $\pm$ 0.10	4.8
<i>Y<sub>ala/gln</sub></i> (mol/mol)	0.2 $\pm$ 0.02	9.5	0.0 $\pm$ 0.00	-	0.0 $\pm$ 0.00	-	0.0 $\pm$ 0.00	-	0.6 $\pm$ 0.04	7.9
<i>q<sub>GLC</sub></i> (mmol/gL h) <sup>a</sup>	21.43 $\pm$ 1.56	7.3	16.17 $\pm$ 1.76	10.9	15.13 $\pm$ 0.51	3.4	33.05 $\pm$ 2.17	6.6	27.64 $\pm$ 1.33	4.8
<i>q<sub>LAC</sub></i> (mmol/gL h) <sup>a</sup>	-4.43 $\pm$ 3.05	-	0.00 $\pm$ 0.00	-	0.00 $\pm$ 0.00	-	-41.23 $\pm$ 4.13	10.0	-49.68 $\pm$ 1.30	2.6
<i>q<sub>GLN</sub></i> (mmol/gL h) <sup>a</sup>	9.10 $\pm$ 0.82	9.0	7.55 $\pm$ 0.72	9.5	7.12 $\pm$ 0.31	4.3	4.40 $\pm$ 0.28	6.4	5.07 $\pm$ 0.41	8.1
<i>q<sub>AMM</sub></i> (mmol/gL h) <sup>a</sup>	-7.19 $\pm$ 0.67	9.3	-9.21 $\pm$ 0.97	10.5	-9.55 $\pm$ 0.30	3.2	-4.48 $\pm$ 0.35	7.8	-5.66 $\pm$ 0.33	5.9
<i>q<sub>GALAC</sub></i> (mmol/gL h) <sup>a</sup>	1.94 $\pm$ 0.21	10.8	1.55 $\pm$ 0.22	14.3	2.46 $\pm$ 1.26	51.1	1.92 $\pm$ 0.24	12.3	1.61 $\pm$ 0.37	23.3
<i>q<sub>PYR</sub></i> (mmol/gL h) <sup>a</sup>	5.82 $\pm$ 0.52	8.9	4.70 $\pm$ 0.57	12.1	4.48 $\pm$ 0.11	2.4	9.10 $\pm$ 0.64	7.1	7.04 $\pm$ 0.55	7.8
<i>q<sub>GLU</sub></i> (mmol/gL h) <sup>a</sup>	-1.31 $\pm$ 0.18	14.0	-1.05 $\pm$ 0.10	9.4	-0.52 $\pm$ 0.09	17.9	-0.71 $\pm$ 0.31	43.5	-0.88 $\pm$ 0.38	43.8
<i>q<sub>ALA</sub></i> (mmol/gL h) <sup>a</sup>	-2.14 $\pm$ 0.27	12.8	0.39 $\pm$ 0.03	6.7	0.54 $\pm$ 0.03	5.9	0.10 $\pm$ 0.12	127.8	-2.81 $\pm$ 0.26	9.4
<i>q<sub>ASP</sub></i> (mmol/gL h) <sup>a</sup>	1.19 $\pm$ 0.19	15.9	1.20 $\pm$ 0.10	8.1	1.49 $\pm$ 0.05	3.5	-0.41 $\pm$ 0.62	150.7	-2.16 $\pm$ 1.15	53.1
<i>q<sub>ALAAT</sub></i> (pM/cell d) <sup>a</sup>	-7.87 $\pm$ 0.19	2.4	-8.54 $\pm$ 1.34	15.7	-6.33 $\pm$ 0.19	3.0	-12.31 $\pm$ 0.60	4.9	-14.29 $\pm$ 1.08	7.5

**a:** Substrate uptake is indicated by a positive rate, whereas a negative value indicates a production rate.

Table 3: Steady-state values reported for *Rath* of different parameters from continuous cultivations with varying GLC and GLN feed concentrations and with 3 mM GAL. Table taken from *Rath*

Variable	Exp. F01	
	Average $\pm$ SD	Rel. SD (%)
Setpoints:		
<i>DR</i> (1/h)	0.0150	-
GLC feed conc. (mM)	10	-
GLN feed conc. (mM)	5	-
GALC feed conc. (mM)	0	-
GLC/GLN ratio (mol/mol)	2	-
$X_V$ (E6 cells/mL)	1.701 $\pm$ 0.023	1.3
$X_D$ (E6 cells/mL)	0.081 $\pm$ 0.004	5.1
$CV_V$ ( $\mu$ L/mL)	4.54 $\pm$ 0.24	5.2
$\mu$ (1/h)	0.0156 $\pm$ 0.0000	0.2
$CD$ ( $\mu$ m)	17.2 $\pm$ 0.33	1.9
GLC (mM)	0.0 $\pm$ 0.00	-
LAC (mM)	7.8 $\pm$ 0.64	8.2
GLN (mM)	1.9 $\pm$ 0.18	9.3
AMM (mM)	3.4 $\pm$ 0.17	5.1
PYR (mM)	0.4 $\pm$ 0.02	6.6
GLU (mM)	1.0 $\pm$ 0.05	5.2
ALA (mM)	0.9 $\pm$ 0.07	8.2
ASP (mM)	1.9 $\pm$ 0.21	10.8
A1AT (mg/L)	44.0 $\pm$ 1.71	3.9
$Y_{CVV/glc}$ ( $\mu$ L/mmol)	626 $\pm$ 25	4.0
$Y_{CVV/gln}$ ( $\mu$ L/mmol)	1865 $\pm$ 201	10.8
$Y_{lac/glc}$ (mol/mol)	0.9 $\pm$ 0.07	8.2
$Y_{amm/gln}$ (mol/mol)	1.3 $\pm$ 0.04	3.1
$Y_{ala/gln}$ (mol/mol)	0.2 $\pm$ 0.02	8.5
$q_{GLC}$ (nmol/ $\mu$ L h) <sup>a</sup>	24.9 $\pm$ 1.08	4.3
$q_{LAC}$ (nmol/ $\mu$ L h) <sup>a</sup>	-22.6 $\pm$ 1.27	5.6
$q_{GLN}$ (nmol/ $\mu$ L h) <sup>a</sup>	8.4 $\pm$ 1.07	12.7
$q_{AMM}$ (nmol/ $\mu$ L h) <sup>a</sup>	-10.9 $\pm$ 1.13	10.3
$q_{PYR}$ (nmol/ $\mu$ L h) <sup>a</sup>	4.5 $\pm$ 0.31	6.9
$q_{GLU}$ (nmol/ $\mu$ L h) <sup>a</sup>	-0.7 $\pm$ 0.15	20.4
$q_{ALA}$ (nmol/ $\mu$ L h) <sup>a</sup>	-1.99 $\pm$ 0.33	16.3
$q_{ASP}$ (nmol/ $\mu$ L h) <sup>a</sup>	1.65 $\pm$ 0.64	39.0
$q_{A1AT}$ (pg/cell d) <sup>a</sup>	-9.2 $\pm$ 0.25	2.7

<sup>a</sup>: Substrate uptake is indicated by a positive rate, whereas a negative value indicates a production rate.

Table 4: Steady-state values reported for *Rath* of different parameters from continuous cultivations with varying GLC and GLN feed concentrations and without GAL. Table taken from *Rath*

### 1.3.1 Recon3D

Recon3D represents the most comprehensive human metabolic network model to date *Brunk*. The model, Recon3DModel.301.mat, was downloaded from <http://vmh.life>. The original biomass equation was modified to adjust the biomass demand reported by *Niklas* for the parental line AGE1\_HN. All the others original demands from recon3D were deactivated. An extra demand representing the maintenance demand, not associated with growth, of atp was set according to *Cossio*. All the fluxes representing exchangeable metabolites (external reactions) were set as reversible (lb and ub set to a large number), so the only effective bound constraints (for FBA and EP) are the one produced by the chemostat consideration *Cossio*. Additionally, for including the molecular crowding constraints we map *Shlomi* enzymatic costs, initially defined for recon1, to recon3D.

For reproduce the conditions of the different experiments, the external concentrations of the exchangeable metabolites was set using the data from *Tables1–2*. Additionally external concentrations of salts, oxygen and others metabolites not specified in the medium was set to a large number. Particularly, Recon3D was unable to growth without pe\_hs[e], phosphatidylethanolamine, (or similar) lipid in the feed medium. Later we will discuss the impact of this metabolite in the medium.

### 1.3.2 CHO

Chinese hamster ovary (CHO) host cell-lines GEM was obtained from *Hefzi* and was setup as *Cossio*. The external metabolites concentrations was set to the values indicated in *Table1–2*, similar to Recon3D. Also, a few extra metabolites, taken from *Cossio* was included in the feed medium. An important different from Recon3D was that CHO is able to growth without pe\_hs[e], named pe\_cho\_e in CHO.

## 2 Results

### 2.1 Low concentrations of phosphatidylethanolamine

Because phosphatidylethanolamine isn't a reported component of the 42-Max-UB-medium *Tabla1* and it can be a carbon source for the GEM, its concentration was set first to the lowest value possible. For Recon3D it was fixed to 0.1mM, a hand picked value, and for CHO it wasn't present in the feed medium at all. Flux balance analysis with molecular crowding (FBAwMC) was performed similar to *Cossio*. Plots of  $\mu$  and  $Xv$  as function of  $\xi$  are shown in *Figure1*. As can be appreciated from the comparison of the model prediction (solid lines) with the experimental data (colored circles), FBAwMC have big problem to reproduce both observables. Specially for  $Xv$ , the predictions was really far from the expected, never reaching a high enough value.

The correlations of the experimental and the modeled uptakes, *Figure2*, show

better results for CHO. This could be, maybe, explained because Recon3D is a general GEM, allowing access to all the possible genome repertory present in humans, fact that is not accurate in a *in vivo* scenario.

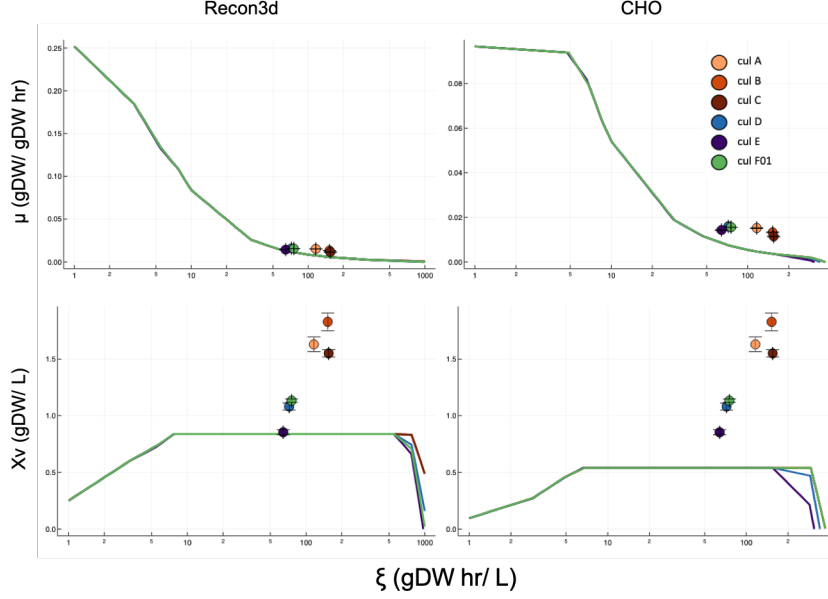


Figure 1: FBAwMC results showing the growth rate  $\mu$  and the viable cell density  $X_v$  dependence of  $\xi$  for the six culture conditions. The solid lines represents the model predictions and the color points shows the experimental results. The model data was obtained for feed mediums with low (Recon3D) or zero (CHO) concentrations of phosphatidylethanolamine

## 2.2 High concentrations of phosphatidylethanolamine

## 3 Bibliography



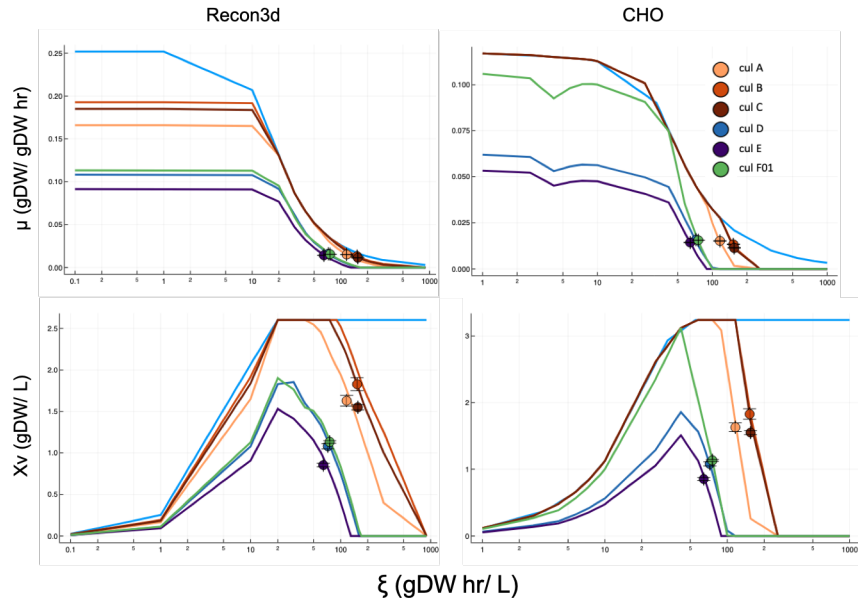


Figure 3: EP and FBA results showing the growth rate and the viable cell density dependence of  $\xi$  for the six culture conditions. The solid lines represents the model predictions and the color points shows the experimental results. FBA results are shown as the solid light blue line.



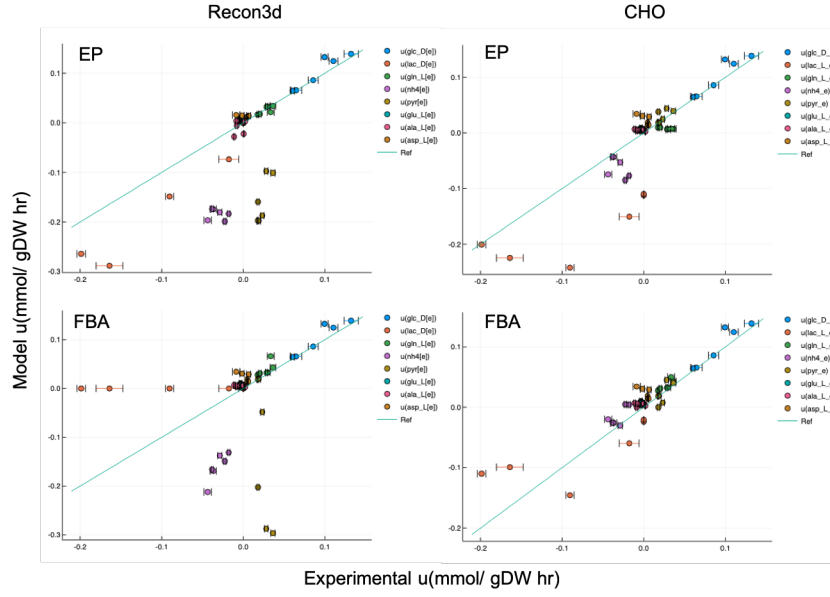


Figure 4: Correlations of all the experimental uptakes compared with the predicted value from EP and FBA.

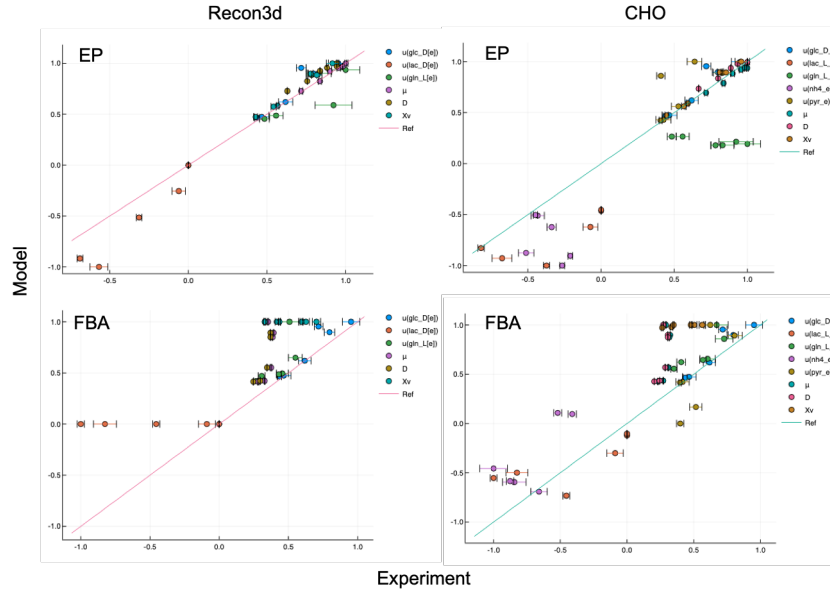


Figure 5: Normalized correlations of some experimental uptakes, growth rate, dilution rate and viable cell density compared with the predicted value from EP and FBA