

Tutorial Unix e linha de comandos





Análise computacional e bioinformática de variantes em doença genética

10 a 13 Out. 2023

nas instalações do Instituto Ricardo Jorge, em Lisboa

Este curso, de natureza teórico-prática, dá a conhecer as várias etapas envolvidas na análise de variantes de linha germinativa associadas a doença genética, em paralelo com a análise prática de casos reais.





Destinatários: Profissionais de saúde, investigadores e estudantes de mestrado ou doutoramento, que estejam envolvidos em atividades de diagnóstico ou investigação no contexto de estudo de variantes de linha germinativa associadas a doença genética

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Coordenação: Luís Vieira

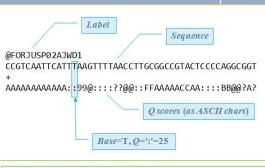


mapping

pre variant calling (BQSR, MarkDup)

variant calling

variant annotation/ priorization



.fastq

```
VN:1.0 SO:coordinate
@50
      SN:chr20
                    LN:64444167
      ID:TopHat
                    VN:2.0.14
                                 CL:/srv/dna tools/tophat/tophat -N 3 --read-edit-dist 5 --read-rea
lign-edit-dist 2 -i 50 -I 5000 --max-coverage-intron 5000 -M -o out /data/user446/mapping tophat/index/chr
20 /data/user446/mapping tophat/L6 18 GTGAAA L007 R1 001.fastg
HWI-ST1145:74:C101DACXX:7:1102:4284:73714
                                               chr20
                                                     190930 3
     {\tt CCGTGTTTAAAGGTGGATGCGGTCACCTTCCCAGCTAGGCTTAGGGATTCTTAGTTGGCCTAGGAAATCCAGCTAGTCCTGTCTCTCAGTCCCCCCTCT}
    AS:1:-15
                XM:i:3 X0:i:0 XG:i:0 MD:Z:55C20C13A9 NM:i:3 NH:i:2 CC:Z:= CP:i:55352714 HI:i:0
```

.sam/.bam/.cram

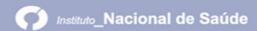
.vcf

www.insa.pt

```
##FORMAT=<ID=DP, Number=1, Type=Integer, Description="Read Depth">
##FORMAT=<ID=HQ,Number=2,Type=Integer,Description="Haplotype Quality">
#CHROM POS
                                                                                           FORMAT
                                                                                                        NA00001
                          REF
                                          QUAL FILTER INFO
       14370
                                                                                           GT: GO: DP: HO 010: 48:
20
               rs6054257 G
                                          29
                                               PASS
                                                       NS=3; DP=14; AF=0.5; DB; H2
                                                                                           GT: GQ: DP: HQ 0 0: 49:
20
       17330
                                          3
                                               q10
                                                       NS=3; DP=11; AF=0.017
20
       1110696 rs6040355 A
                                          67
                                               PASS
                                                       NS=2; DP=10; AF=0.333, 0.667; AA=T; DB GT: GQ: DP: HQ 1 2:21:
20
       1230237 .
                                          47
                                               PASS
                                                       NS=3; DP=13; AA=T
                                                                                           GT:GQ:DP:HQ 0 0:54:
       1234567 microsatl GTC
                                  G, GTCT 50
                                               PASS
                                                       NS=3; DP=9; AA=G
                                                                                           GT:GQ:DP 0/1:35:4
```

Prioritised Genes

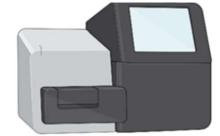
```
DCAF17
                                                                             Phenotype Score: 0.802
                                      Exomiser Score: 0.986
                                                                                                                   Variant Score: 1.000
                                      (p=3.6E-5)
  AUTOSOMAL_RECESSIVE
                                                                            Phenotype Score: 0.802
                                                                                                                Variant Score: 1.000
                                       Exomiser Score: 0.986
  Phenotype matches to diseases consistent with this MOI:
  Phenotypic similarity 0.802 to ORPHA:3464 Woodhouse-Sakati syndrome
  Phenotypic similarity 0.796 to OMIM:241080 Woodhouse-Sakati syndrome
  Variants contributing to score:
   FRAMESHIFT_TRUNCATION DEL 2-171448794-TC-T [1/1:0/1:0/1] rs797045038
  Exomiser ACMG: PATHOGENIC [PVS1, PM2, PP4, PP5_Strong]
                                                                            Pathogenicity Data:
                                                                                                                Frequency Data
   Transcripts:
                                                                            No pathogenicity data
                                                                                                                No frequency data
```



Clinical exome sequencing

Departamento Genética Humana

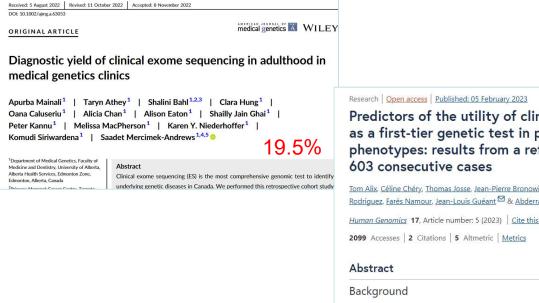






Exome sequencing:

- -diagnosis of genetic disorders
- -discovery of new Mendelian-disease genes
- -Clinical exome sequencing (CES) genes associated to clinical phenotypes



Predictors of the utility of clinical exome sequencing as a first-tier genetic test in patients with Mendelian phenotypes: results from a referral center study on Tom Alix, Céline Chéry, Thomas Josse, Jean-Pierre Bronowicki, François Feillet, Rosa-Maria Guéant-Rodriguez, Farès Namour, Jean-Louis Guéant ≥ & Abderrahim Oussalah Human Genomics 17, Article number: 5 (2023) Cite this article 37.6% Clinical exome sequencing (CES) provides a comprehensive and effective analysis of relevant

disease-associated genes in a cost-effective manner compared to whole exome sequencing

Article Open access | Published: 10 November 2022

Five years' experience of the clinical exome sequencing in a Spanish single center

A. Arteche-López, A. Ávila-Fernández, R. Riveiro Álvarez, B. Almoguera, A. Bustamante Aragonés, I. Martin-Merida, M. A. López Martínez, A. Giménez Pardo, C. Vélez-Monsalve, J. Gallego Merlo, I. García Vara, F. Blanco-Kelly, S. Tahsin Swafiri, I. Lorda Sánchez, M. J. Trujillo Tiebas & C. Ayuso

Scientific Reports 12, Article number: 19209 (2022) | Cite this article

732 Accesses 37 Altmetric Metrics

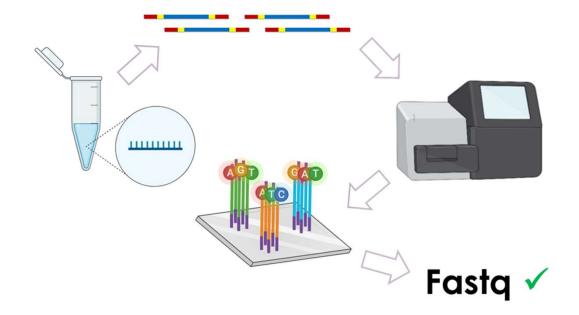
24 62%

Abstract

Nowadays, exome sequencing is a robust and cost-efficient genetic diagnostic tool already implemented in many clinical laboratories. Despite it has undoubtedly improved our diagnostic capacity and has allowed the discovery of many new Mendelian-disease genes, it only provides a molecular diagnosis in up to 25-30% of cases. Here, we comprehensively evaluate the results of a large sample set of 4974 clinical exomes performed in our laboratory

Clinical exome - Experimental procedure

- Library:
 - TruSight One sequencing panel (4 800 genes; ~62 000 targets)
- Sequencing:
 - MiSeq/NextSeq
 - Paired-end, 2x150pb



Variants

.vcf

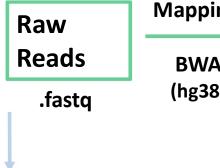
(~8 000)

Exomiser.

VEP



(Automation, Reproducibility)



Mapping

BWA
(hg38)

Mapped
Reads
.bam

Variant calling/filtering

GATK

OLC) Patient's phonetyp

Patient's phenotype

HPO terms

Variant

annotation/priorization

QC - FastQC:

- Q30
- GC
- % reads id

QC - samtools/qualimap:

- Base mean qual
- % reads mapped
- % reads on target
- % target coveraged
- % targets low coverage

QC - vcftools:

- Transit/Transv ratio
- Het/Hom ratio

Priorized-Annotated Variants

.vcf

.html

(~150-200 var)

Virtual gene panel

Phen2Gene → HPO → 150 genes

vs

Variants

Analysis/Interpretation:

- Exomiser Top-10 Variants
- Visualization IGV
- Validation VEP
- DB's (dbSNP, clinVar, HGMD, Uniprot, Decipher, ...)





Graphical user interface (GUI)



Interação meios visuais

vs Command-line interface (CLI)



Interação comandos de texto

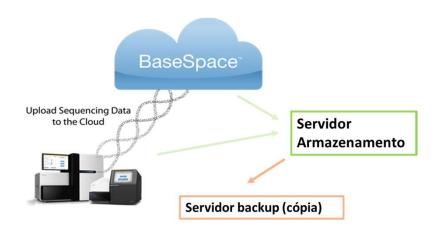
Windows

vs Unix

Ferramentas específicas Grandes datasets/Rec. Inform. Servidores/clusters Automatização Reprodutibilidade

Automatização de procedimentos

Gestão automatizada armazenamento dados em bruto NGS



Automatização controlo de qualidade NGS (InterOp, FastQC)





Gestão automatizada armazenamento dados em bruto NGS

- · Centenas Gigabytes dados por semana
- Gestão automatizada/programada semanal
- Transfere ficheiros corridas NGS para servidor armazenamento dados
- Guarda pasta com designação/formato específico
- · Envia alertas por email



Upload Sequencing Data to the Cloud



Servidor Armazenamento

Servidor backup (cópia)

```
for full_run_dir in $run_output_dir/*

do

full_run_dir=$( basename $full_run_dir )

instrument_type=$( bs run get -i $full_run_dir --retry | grep InstrumentType | sed 's/ //g' | cut -d "|" -f3 )

experiment_name=$( bs run get -i $full_run_dir --retry | grep ExperimentName | cut -d "|" -f3 | sed 's/ //g' )

#run_number=$( bs run get -i $full_run_dir --retry | grep -w Number | cut -d "|" -f3 | sed 's/ //g' )

run_ID_name=$( bs run get -i $full_run_dir --retry | grep "[0-9]* Name" | cut -d "|" -f3 | sed 's/ //g' )

year_start="20"

year_end=$( bs run get -i $full_run_dir --retry | grep "[0-9]* Name" | cut -d "|" -f3 | sed 's/ //g' | cut -c1-2 )

year_complete="${year_start}${year_end}"

if [[ "$instrument_type" == "NextSeq" ]]; then
```



Automatização controlo de qualidade NGS

- Corre os programas de QC Illumina: interop summary e index-summary
- Corre o programa FastQC; Corre o MultiQC para gerar relatório
- Envia por email o relatório MultiQC (*.html)



Análise da qualidade da sequenciação

Dep. de Genética Humana - Unidade de Tecnologia e Inovação A análise primária foi efectuada usando os programas Interop e FastQC

```
printf "\nStep 1/4. Running Illumina interop summary program...\n\n" # prints this message

mkdir qc_tmp_files

interop_summary . --csv=1 > qc_tmp_files/summary.csv # runs the Illumina interop summary program

printf "\nStep 2/4. Running Illumina interop index-summary program...\n\n" # runs the Illumina interop interop_index-summary . --csv=1 > qc_tmp_files/indexing.csv # runs the Illumina interop index-summary printf "\nStep 3/4. Running fastqc program (it may take a while)...\n\n"

fastq_files=($run_output_dir/$run_dir/*/*/*.fastq.gz) # exemplo estrutura pastas retirada do basespace /

#### HGuimaraes_I37546_2022_L004_ds.e3edbc11ee22440c88231ec2669ba356

if fastqc -t 2 -q -f fastq -o qc_tmp_files/ $(ls $fastq_files); then #runs fastqc for all samples (fastq echo "FastQC runned successfully on genome0 (entry node).\n"

else

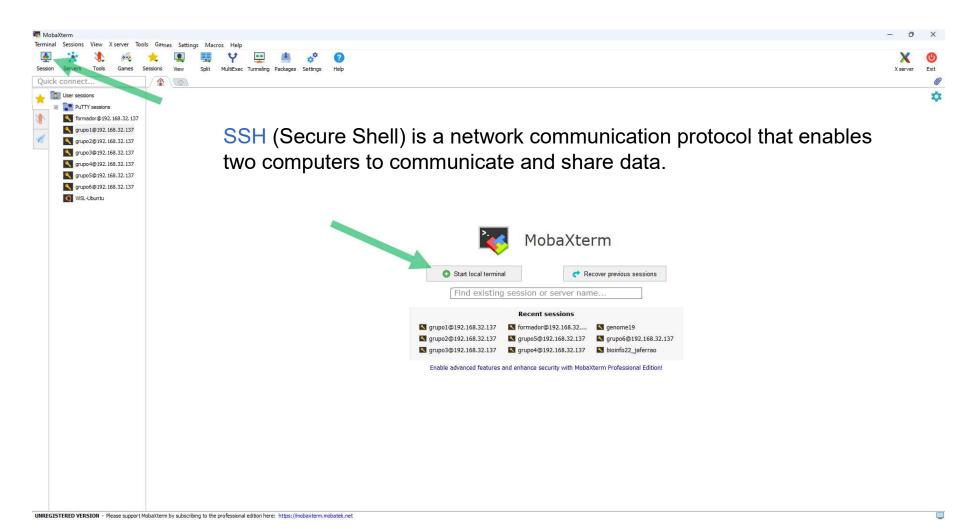
srun -N 1 -n 1 -c 2 --mem-per-cpu=2GB fastqc -t 2 -q -f fastq -o qc_tmp_files/ $(ls $fastq_files))

echo "FastQC runned through Slurm on one of the computation nodes.\n"

fi
```

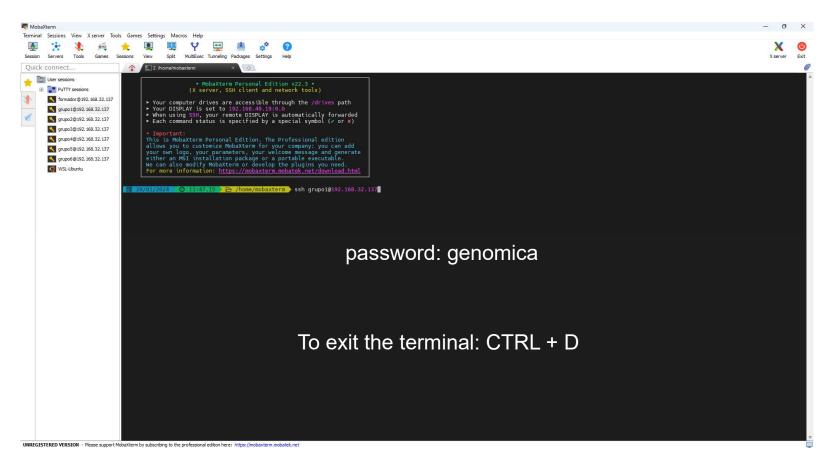


Connection to the Unix server

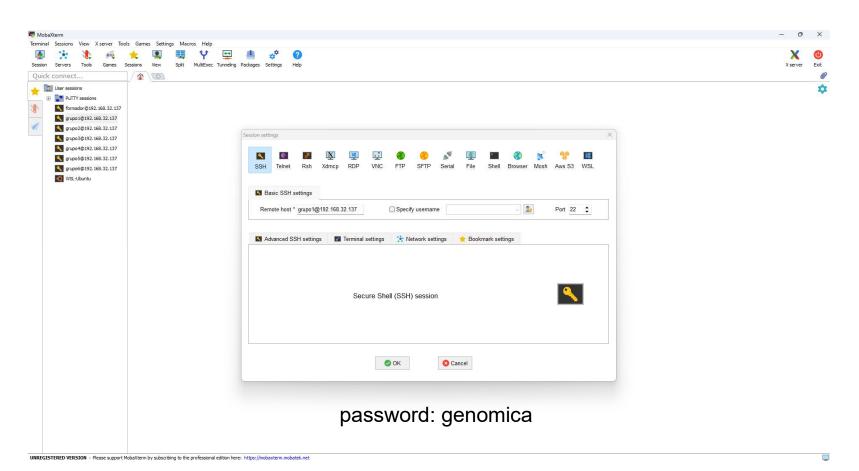




Connection from local terminal: ssh grupo1@192.168.32.137 [ENTER]

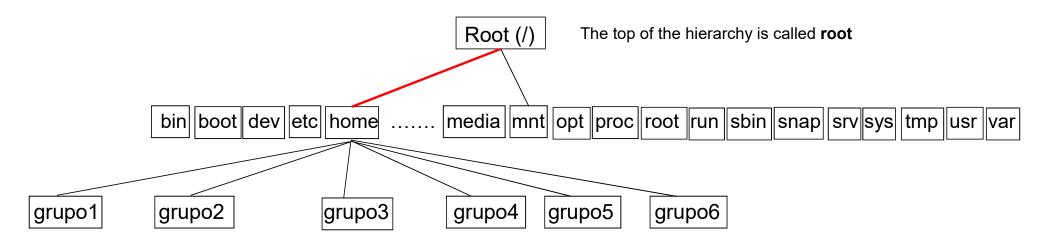


Connection using a session

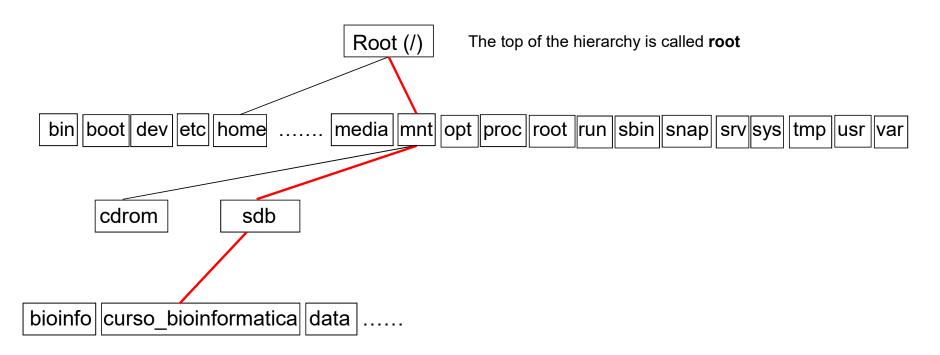




File-system on the Unix server



File-system on the Unix server







Git and GitHub



- Version control:
 - helps developers track and manage changes to code
- Colaboration
- CLI

- user-friendly interface (GUI)
- public code repository for free
- popular open-source projects

Tutorial linha de comandos Unix

https://github.com/krother/bash_tutorial (clone this repository on home dir)

cd ~
git clone https://github.com/krother/bash_tutorial.git

• Extra: https://ubuntu.com/tutorials/command-line-for-beginners#1-overview

Conda



- Conda provides package, dependency, and environment management for any language.
- Conda allows users to install different versions of <u>binary</u> software packages and any required libraries appropriate for their <u>computing platform</u>. Also, it allows users to switch between package versions and download and install updates from a <u>software repository</u>.
- A popular Conda channel for <u>bioinformatics software</u> is *Bioconda*, which provides multiple software distributions for computational biology.

Conda



- conda env list
- conda activate curso_amb
- conda list
- conda deactivate
- conda activate curso_amb_vep