

Tutorial Unix e linha de comandos





24 a 28 JUN 2024

nas instalações do Instituto Ricardo Jorge, em Lisboa



Enquadramento e objetivos

Este curso, de natureza teórico-prática, dá a conhecer as várias etapas envolvidas na análise de variantes de linha germinativa associadas a doença genética, em paralelo com a análise prática de casos reais.

Destinatários: Profissionais de saúde, investigadores e estudantes de mestrado ou doutoramento, que estejam envolvidos em atividades de diagnóstico ou investigação no contexto de estudo de variantes de linha germinativa associadas a doença genética

Formadores: Luís Vieira, José Ferrão, Hugo Martiniano e Daniel Sobral

Coordenação: Luís Vieira



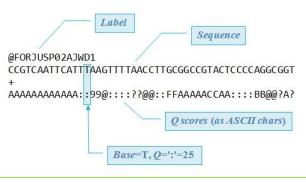
basecalling

mapping

pre variant calling (BQSR, MarkDup)

variant calling

variant annotation/ priorization



.fastq

```
VN:1.0 SO:coordinate
@SQ
      SN:chr20
                    LN:64444167
      ID:TopHat
                    VN:2.0.14
                                 CL:/srv/dna tools/tophat/tophat -N 3 --read-edit-dist 5 --read-rea
lign-edit-dist 2 -i 50 -I 5000 --max-coverage-intron 5000 -M -o out /data/user446/mapping tophat/index/chr
20 /data/user446/mapping tophat/L6 18 GTGAAA L007 R1 001.fastq
HWI-ST1145:74:C101DACXX:7:1102:4284:73714
                                               chr20
                                                    190930 3
      {\tt CCGTGTTTAAAGGTGGATGCGGTCACCTTCCCAGCTAGGCTTAGGGATTCTTAGTTGGCCTAGGAAATCCAGCTAGTCCTGTCTCTCAGTCCCCCCTCT
    XM:i:3 X0:i:0 XG:i:0 MD:Z:55C20C13A9 NM:i:3 NH:i:2 CC:Z:= CP:i:55352714
   AS:i:-15
```

.sam/.bam/.cram

.vcf

```
##FORMAT=<ID=DP, Number=1, Type=Integer, Description="Read Depth">
##FORMAT=<ID=HQ, Number=2, Type=Integer, Description="Haplotype Quality">
#CHROM POS
                                                                                           FORMAT
                                          OUAL FILTER INFO
                                                                                                        NA00001
                                                                                           GT:GQ:DP:HQ 0 0:48:
       14370
                rs6054257 G
                                               PASS
                                                       NS=3; DP=14; AF=0.5; DB; H2
20
       17330
                                                                                           GT:GQ:DP:HQ 0|0:49:
                                                       NS=3; DP=11; AF=0.017
       1110696 rs6040355 A
                                                PASS
                                                       NS=2; DP=10; AF=0.333, 0.667; AA=T; DB GT: G0: DP: H0 1 2: 21:
       1230237 .
                                          47
                                                PASS
                                                       NS=3; DP=13; AA=T
                                                                                           GT:G0:DP:H0 010:54:
       1234567 microsatl GTC
                                  G, GTCT 50
                                                PASS
                                                       NS=3: DP=9: AA=G
                                                                                           GT:G0:DP 0/1:35:
```

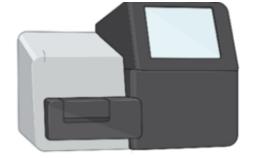
Prioritised Genes

DCAF17 Exomiser Score: 0.986 Phenotype Score: 0.802 Variant Score: 1.000 (p=3.6E-5)AUTOSOMAL RECESSIVE Phenotype Score: 0.802 Exomiser Score: 0.986 Variant Score: 1.000 (p=3.6E-5)Phenotype matches to diseases consistent with this MOI: Phenotypic similarity 0.802 to ORPHA:3464 Woodhouse-Sakati syndrome Phenotypic similarity 0.796 to OMIM:241080 Woodhouse-Sakati syndrome Variants contributing to score: FRAMESHIFT_TRUNCATION DEL 2-171448794-TC-T [1/1:0/1:0/1] rs797045038 Exomiser ACMG: PATHOGENIC [PVS1, PM2, PP4, PP5_Strong] ClinVar: PATHOGENIC (criteria provided, multiple submitters, no conflicts) Variant score: 1.000 CONTRIBUTING VARIANT WHITELIST VARIAN Pathogenicity Data: Frequency Data: Transcripts: No pathogenicity data No frequency data

Clinical exome sequencing

Departamento Genética Humana

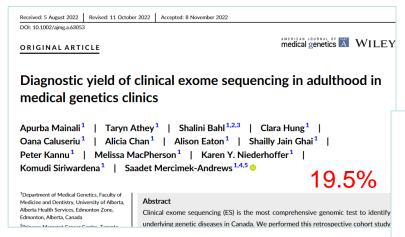






• Exome sequencing:

- -diagnosis of genetic disorders
- -discovery of new Mendelian-disease genes
- -Clinical exome sequencing (CES) genes associated to clinical phenotypes



Research Open access | Published: 05 February 2023

Predictors of the utility of clinical exome sequencing as a first-tier genetic test in patients with Mendelian phenotypes: results from a referral center study on 603 consecutive cases

Tom Alix, Céline Chéry, Thomas Josse, Jean-Pierre Bronowicki, François Feillet, Rosa-Maria Guéant-Rodriguez, Farès Namour, Jean-Louis Guéant [™] & Abderrahim Oussalah [™]

Human Genomics 17, Article number: 5 (2023) | Cite this article

2099 Accesses 2 Citations 5 Altmetric Metrics

37.6%

Abstract

Background

Clinical exome sequencing (CES) provides a comprehensive and effective analysis of relevant disease-associated genes in a cost-effective manner compared to whole exome sequencing.

Article Open access Published: 10 November 2022

Five years' experience of the clinical exome sequencing in a Spanish single center

A. Arteche-López, A. Ávila-Fernández, R. Riveiro Álvarez, B. Almoguera, A. Bustamante Aragonés, I. Martin-Merida, M. A. López Martínez, A. Giménez Pardo, C. Vélez-Monsalve, J. Gallego Merlo, I. García Vara, F. Blanco-Kelly, S. Tahsin Swafiri, I. Lorda Sánchez, M. J. Trujillo Tiebas & C. Ayuso

Scientific Reports 12, Article number: 19209 (2022) Cite this article

732 Accesses 37 Altmetric Metrics

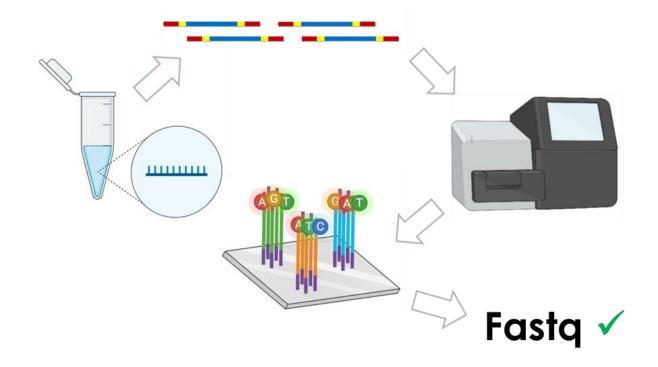
24.62%

Abstract

Nowadays, exome sequencing is a robust and cost-efficient genetic diagnostic tool already implemented in many clinical laboratories. Despite it has undoubtedly improved our diagnostic capacity and has allowed the discovery of many new Mendelian-disease genes, it only provides a molecular diagnosis in up to 25–30% of cases. Here, we comprehensively evaluate the results of a large sample set of 4974 clinical exomes performed in our laboratory

Clinical exome - Experimental procedure

- Library:
 - TruSight One sequencing panel (4 800 genes; ~62 000 targets)
- Sequencing:
 - MiSeq/NextSeq
 - Paired-end, 2x150pb





Clinical exome - Bioinformatics pipeline (SNVs, indels)

(Automation, Reproducibility)



Mapping BWA (hg38)

Mapped Reads .bam

Variant calling/filtering

GATK

Variants

.vcf (~8 000)

Exomiser.

VEP

QC - vcftools:

- **Transit/Transv ratio**
- **Het/Hom ratio**

QC - FastQC:

- **Q30**
- GC
- % reads id

QC - samtools/qualimap:

- Base mean qual
- % reads mapped
- % reads on target
- % target coveraged
- % targets low coverage

Virtual gene panel

Phen2Gene \rightarrow HPO \rightarrow 150 genes

VS

Variants

Analysis/Interpretation:

- Exomiser Top-10 Variants
- Visualization IGV

Patient's phenotype

HPO terms

Variant

annotation/priorization

Priorized-

Annotated

.vcf

.html

(~150-200 var)

Variants

- Validation VEP
- DB's (dbSNP, clinVar, HGMD, Uniprot, Decipher, ...)





Unix



Graphical user interface (GUI) VS



Interação meios visuais

Command-line interface (CLI)



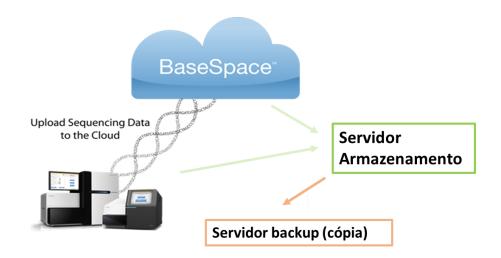
Interação comandos de texto

Windows vs

Ferramentas específicas Grandes datasets/Rec. Inform. Servidores/clusters Automatização Reprodutibilidade

Automatização de procedimentos

Gestão automatizada armazenamento dados em bruto NGS



Automatização controlo de qualidade NGS (InterOp, FastQC)





Gestão automatizada armazenamento dados em bruto NGS

- Centenas Gigabytes dados por semana
- Gestão automatizada/programada semanal
- Transfere ficheiros corridas NGS para servidor armazenamento dados
- Guarda pasta com designação/formato específico
- Envia alertas por email



Servidor Armazenamento

Servidor backup (cópia)

```
for full_run_dir in $run_output_dir/*
do

full_run_dir=$( basename $full_run_dir )
instrument_type=$( bs run get -i $full_run_dir --retry | grep InstrumentType | sed 's/ //g' | cut -d "|" -f3 )
experiment_name=$( bs run get -i $full_run_dir --retry | grep ExperimentName | cut -d "|" -f3 | sed 's/ //g' )

#run_number=$( bs run get -i $full_run_dir --retry | grep "[0-9]* Name" | cut -d "|" -f3 | sed 's/ //g' )

run_ID_name=$( bs run get -i $full_run_dir --retry | grep "[0-9]* Name" | cut -d "|" -f3 | sed 's/ //g' )

year_start="20"

year_complete="${year_start}${year_end}"

if [[ "$instrument_type" == "NextSeq" ]]; then
```



Automatização controlo de qualidade NGS

- Corre os programas de QC Illumina: interop summary e index-summary
- Corre o programa FastQC; Corre o MultiQC para gerar relatório
- Envia por email o relatório MultiQC (*.html)



Análise da qualidade da sequenciação

Dep. de Genética Humana - Unidade de Tecnologia e Inovação

A análise primária foi efectuada usando os programas Interop e FastQC

```
printf "\nStep 1/4. Running Illumina interop summary program...\n\n" # prints this message

mkdir qc_tmp_files

interop_summary . --csv=1 > qc_tmp_files/summary.csv # runs the Illumina interop summary program

printf "\nStep 2/4. Running Illumina interop index-summary program...\n\n" # runs the Illumina interop i

interop_index-summary . --csv=1 > qc_tmp_files/indexing.csv # runs the Illumina interop index-summary pr

printf "\nStep 3/4. Running fastqc program (it may take a while)...\n\n"

fastq_files=($run_output_dir/$run_dir/*/*/*.fastq.gz) # exemplo estrutura pastas retirada do basespace /

HGuimaraes_I37546_2022_L004_ds.e3edbc11ee22440c88231ec2669ba356

if fastqc -t 2 -q -f fastq -o qc_tmp_files/ $(ls $fastq_files); then #runs fastqc for all samples (fastq

echo "FastQC runned successfully on genome0 (entry node).\n"

else

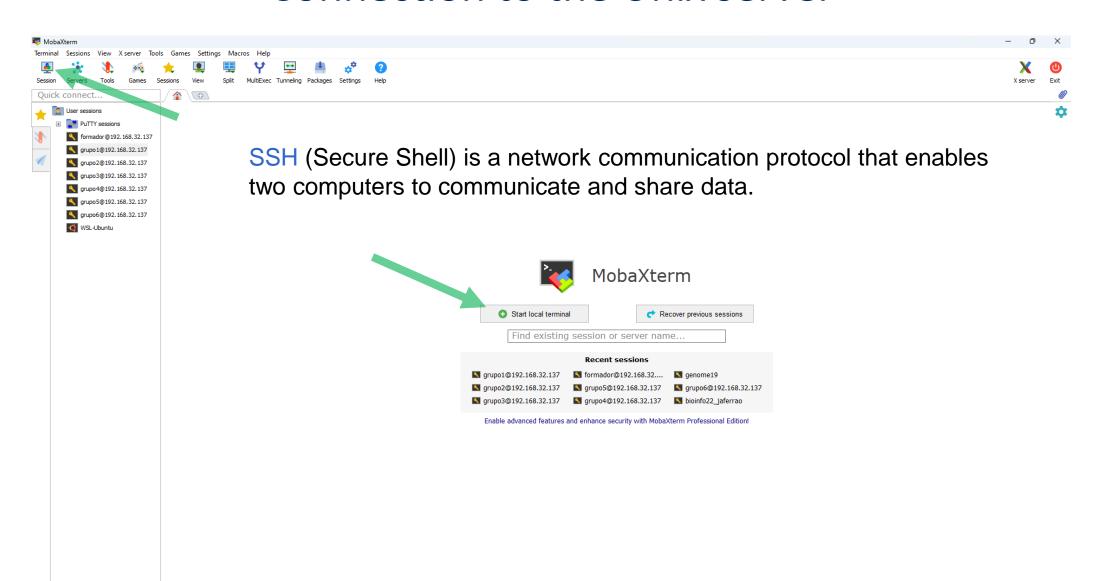
srun -N 1 -n 1 -c 2 --mem-per-cpu=2GB fastqc -t 2 -q -f fastq -o qc_tmp_files/ $(ls $fastq_files))

echo "FastQC runned through Slurm on one of the computation nodes.\n"

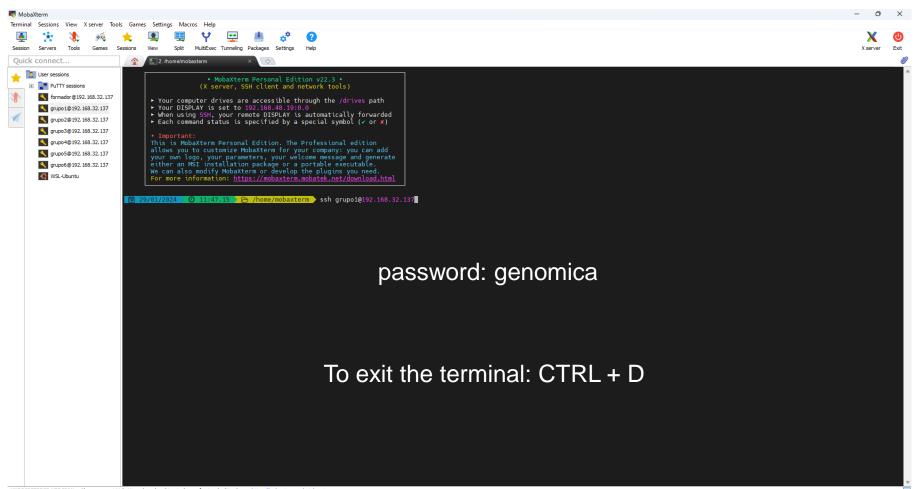
fi
```



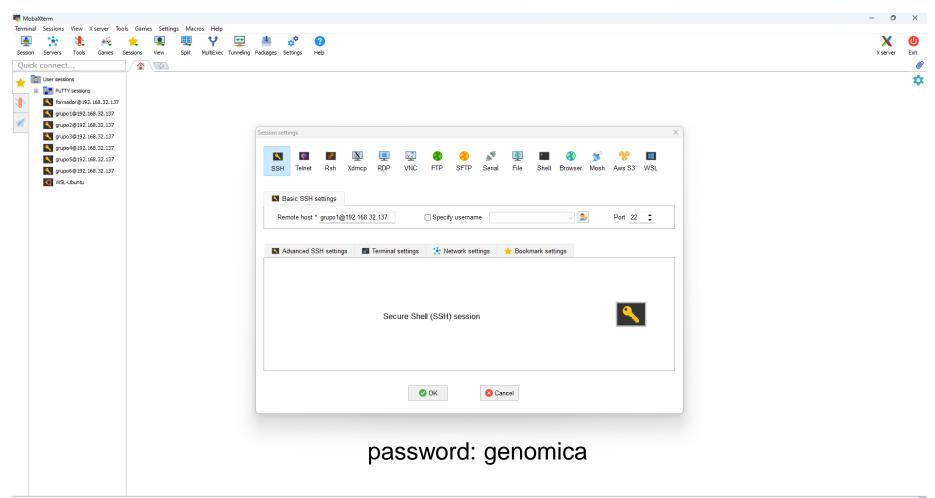
Connection to the Unix server



Connection from local terminal: ssh grupo1@192.168.32.137 [ENTER]

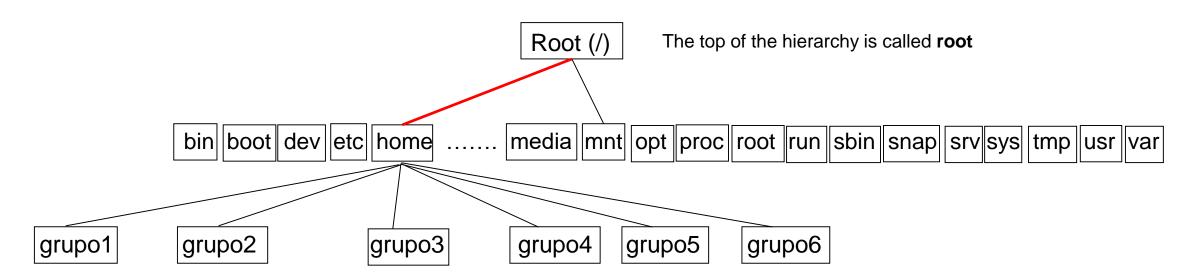


Connection using a session

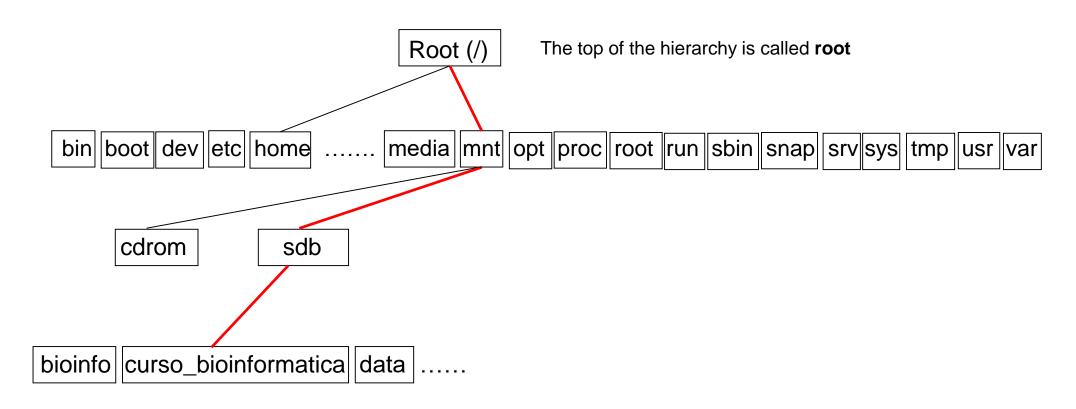


www.insa.pt

File-system on the Unix server



File-system on the Unix server







Git and GitHub



- Version control:
 - helps developers track and manage changes to code
- Colaboration
- CLI

- user-friendly interface (GUI)
- public code repository for free
- popular open-source projects

Tutorial linha de comandos Unix

https://github.com/krother/bash_tutorial (clone this repository on home dir)

cd ~
git clone https://github.com/krother/bash_tutorial.git

• Extra: https://ubuntu.com/tutorials/command-line-for-beginners#1-overview



Conda



- Conda provides package, dependency, and environment management for any language.
- Conda allows users to install different versions of <u>binary</u> software packages and any required libraries appropriate for their computing platform. Also, it allows users to switch between package versions and download and install updates from a software repository.
- A popular Conda channel for <u>bioinformatics software</u> is *Bioconda*, which provides multiple software distributions for computational biology.

Conda



- conda env list
- conda activate curso_amb
- conda list
- conda deactivate
- conda activate curso_amb_vep