

# PacBio



## Iso-Seq

Scalable  
De Novo  
Isoform Discovery  
from PacBio HiFi Reads



Oxford

**NANOPORE**<sup>TM</sup>  
Technologies

# AN OVERVIEW OF LONG-READ SEQUENCING

Amanda Markee | November 14<sup>th</sup> 2024 | CG2

# OUTLINE

INTRODUCTION

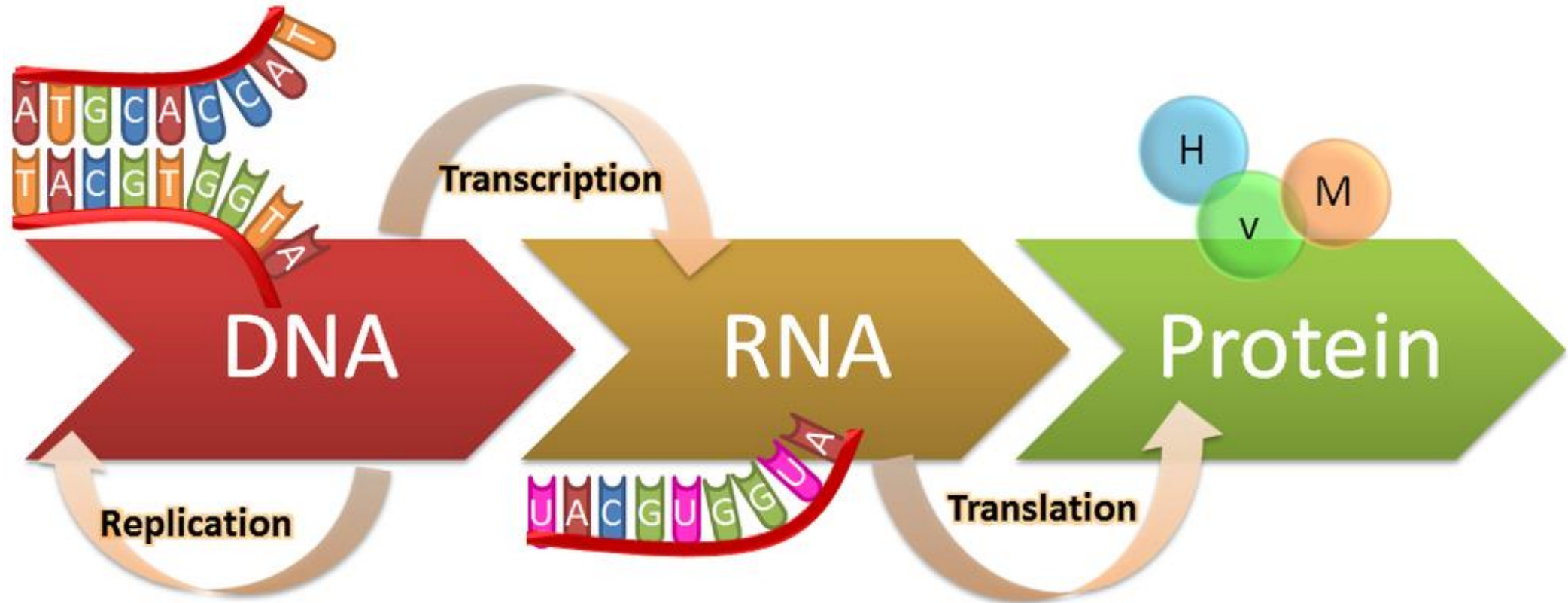
HISTORY OF LONG-READ SEQUENCING

PROS AND CONS

EXAMPLES OF RECENT LITERATURE

CONCLUSIONS

# INTRODUCTION



DNA = GENETIC BLUEPRINT

PROTEIN = PRODUCT



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PROTEIN = PRODUCT

# INTRODUCTION

**Short-read sequencing (NGS)** produces many short DNA fragments (150bp - 300bp)

**Long-read sequencing** produces longer, contiguous DNA fragments (5,000bp - 30,000 bp)

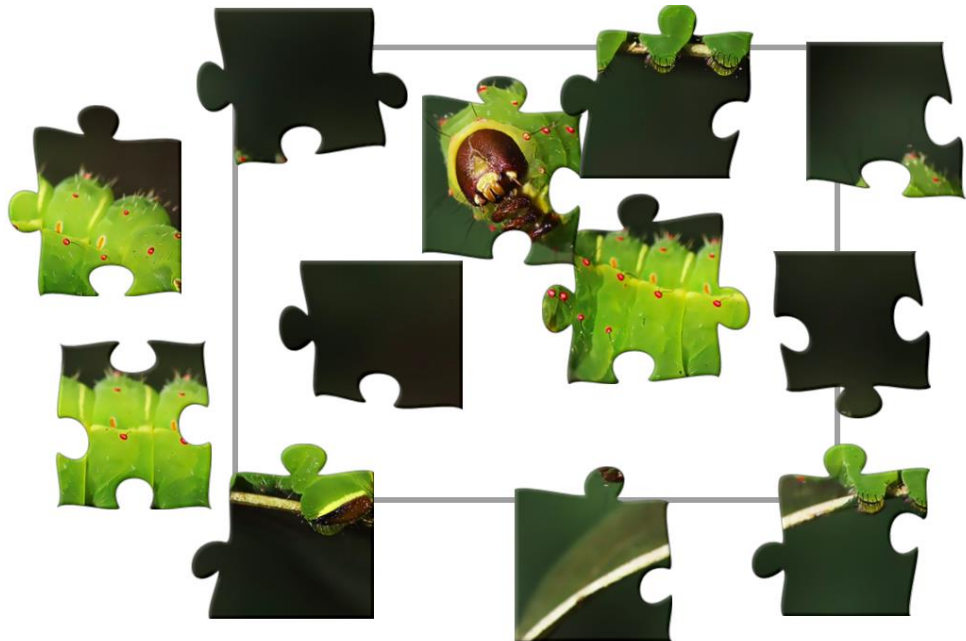




# INTRODUCTION

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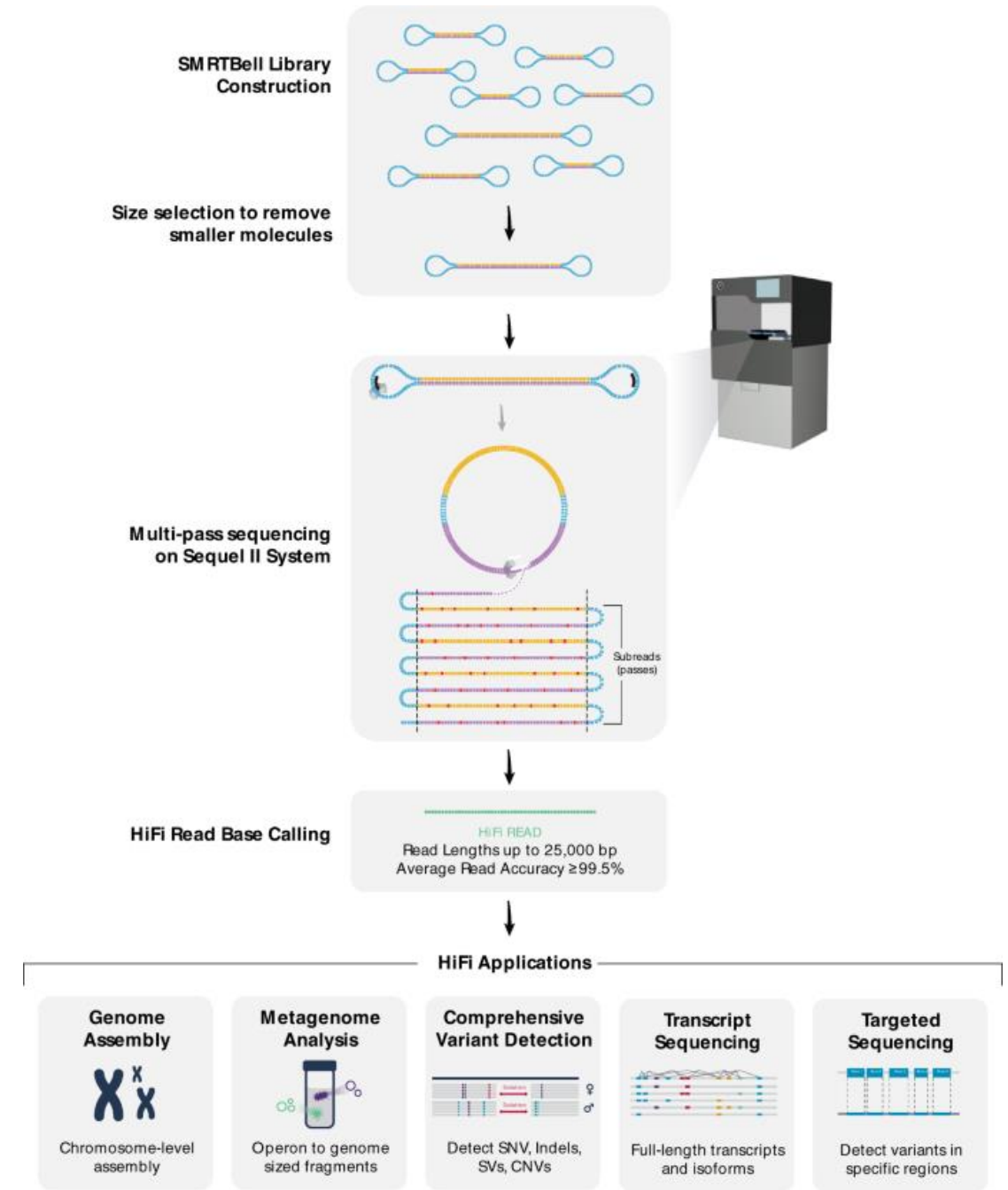
# INTRODUCTION

**Long-read** sequencing for de-novo genome assembly, capturing complex genomic regions (structural variants, isoforms, etc.), or highly repetitive regions.

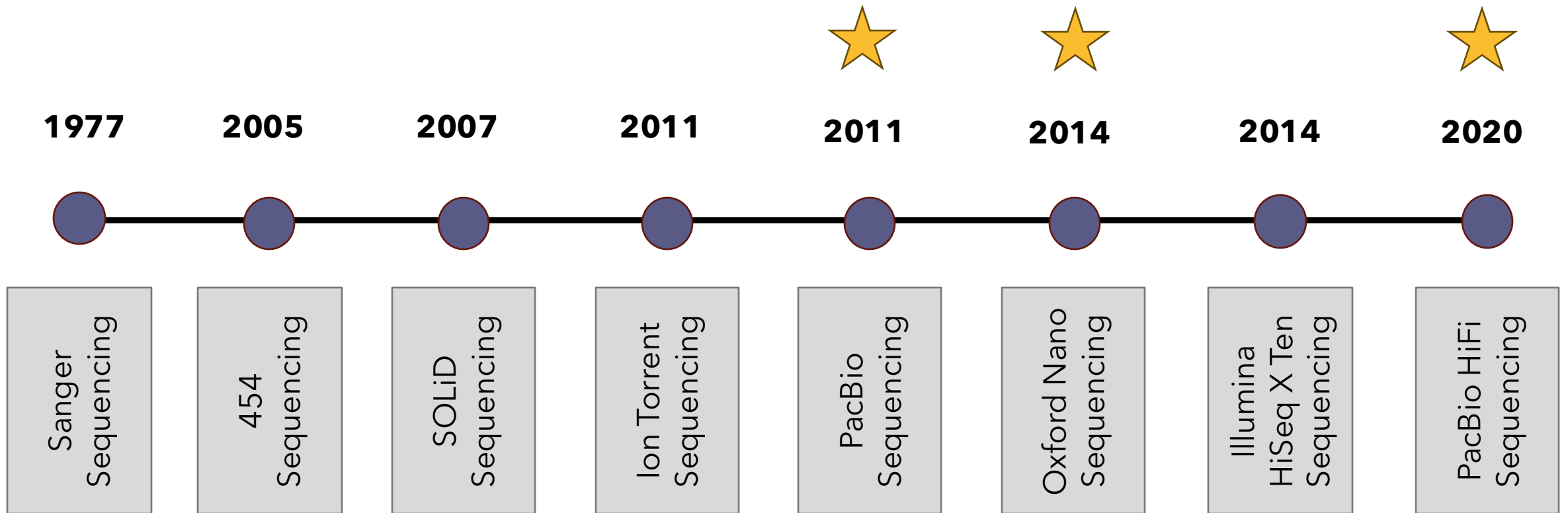
**SMRT cell** (PacBio) sequencing takes place on a chip lined with individual wells

**DNA polymerase** is used to sequence a complementary strand, and the fluorescence is measured to identify the corresponding nucleotides.

**CCS reads** (PacBio) ensure high accuracy



# HISTORY OF LONG-READ SEQUENCING





# PROS

- Accuracy and Resolution:** CCS reads allow for equally as accurate reads as NGS with high accuracy up to 99.5%
- Improved Assembly:** Long reads allow for a more complete and contiguous assembly, less gaps than short reads, especially useful for *de-novo* genome assembly.
- Applications in Challenging Genomes:** Useful in sequencing large repetitive genomes such as plant or squamate genomes. Easily resolves repetitive regions

# CONS

- Cost:** If going for high-quality, can be more costly (1 SMRT Cell ~ \$3000; 1 NGS lane ~ \$800)
- Error Rate:** Long read technologies tend to have higher raw error rates than short-read sequencing, though error correction methods can mitigate this (CCS)
- Technical Limitations:** sample preparation more specialized (HMW DNA, needs little degradation, etc. harder for in-field work, or degraded DNA samples like museum specimens)

# Rapid molecular diversification and homogenization of clustered major ampullate silk genes in Argiope garden spiders

Richard H Baker <sup>1</sup>, André Corvelo <sup>2</sup>, Cheryl Y Hayashi <sup>1</sup>

Affiliations + expand

PMID: 36508456 PMCID: PMC9779670 DOI: 10.1371/journal.pgen.1010537

## Evolution of Opsin Genes in Caddisflies (Insecta: Trichoptera)

Ashlyn Powell <sup>1,\*</sup>, Jacqueline Heckenhauer <sup>2,3</sup>, Steffen U. Pauls <sup>2,3</sup>, Blanca Ríos-Touma <sup>4</sup>, Ryoichi B. Kuranishi <sup>5,6</sup>, Ralph W. Holzenthal <sup>7</sup>, Ernesto Razuri-Gonzales <sup>3</sup>, Seth Bybee <sup>8</sup>, Paul B. Frandsen <sup>1,\*</sup>

## Long Reads Are Revolutionizing 20 Years of Insect Genome Sequencing

Scott Hotaling <sup>1,\*</sup>, John S. Sproul <sup>2</sup>, Jacqueline Heckenhauer <sup>3,4</sup>, Ashlyn Powell <sup>5</sup>, Amanda M. Larracuent <sup>2</sup>, Steffen U. Pauls <sup>3,4,6</sup>, Joanna L. Kelley <sup>1</sup>, and Paul B. Frandsen <sup>3,5,7,\*</sup>

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## Long-read human genome sequencing and its applications

Glennis A. Logsdon, Mitchell R. Vollger & Evan E. Eichler 

*Nature Reviews Genetics* **21**, 597–614 (2020) | [Cite this article](#)

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Darwin  
**TREE**  
of  
**LIFE**

Sequencing the genomes of 70,000  
plants, fungi, animals and protists  
in Britain and Ireland



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# Darwin Tree of Life

The Darwin Tree of Life project aims to sequence the genomes of 70,000 species of eukaryotic organisms in Britain and Ireland.

It is a collaboration between biodiversity, genomics and analysis partners that is transforming the way we do biology, conservation and biotechnology.



# EXAMPLE PAPER

JOURNAL ARTICLE

## De Novo Long-Read Genome Assembly and Annotation of the Luna Moth (*Actias luna*) Fully Resolves Repeat-Rich Silk Genes

Amanda Markee, Rebekah Keating Godfrey, Paul B Frandsen, Yi-Ming Weng, Deborah A Triant, Akito Y Kawahara

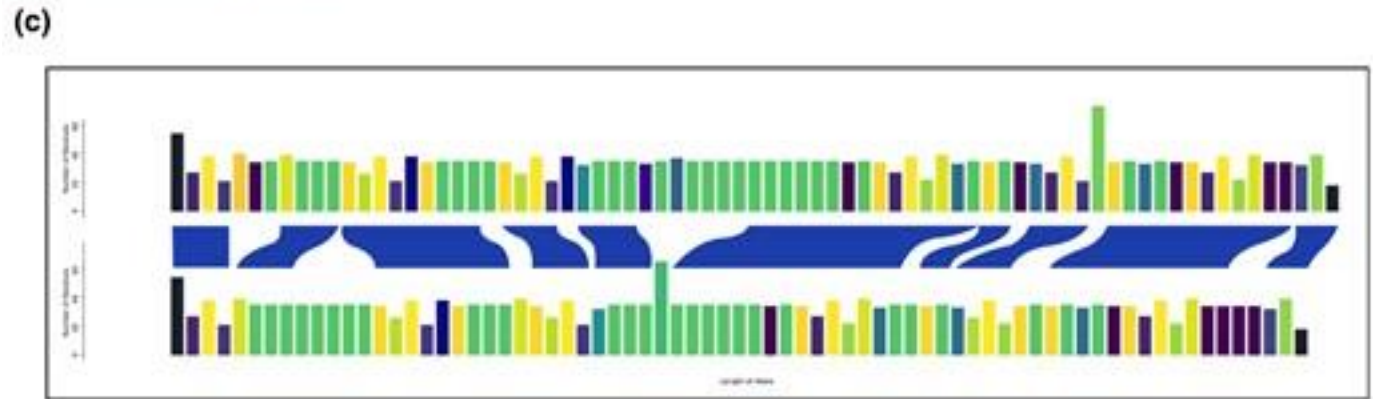
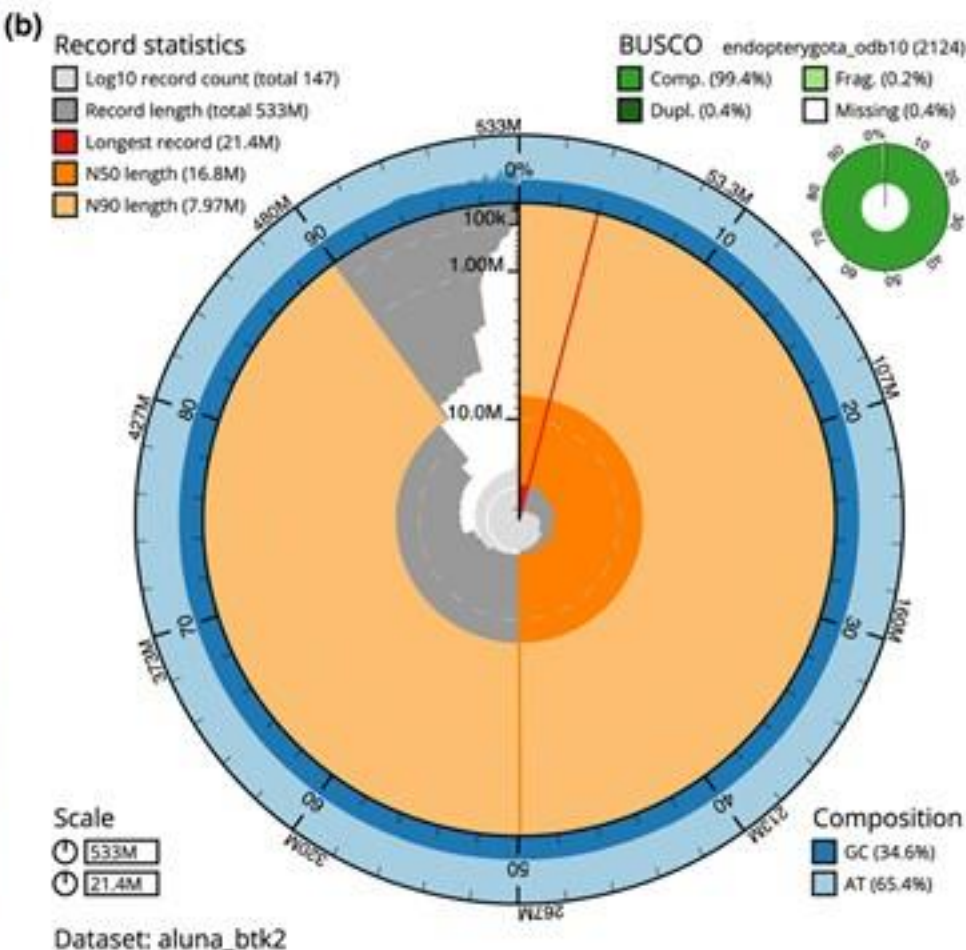
Genome Biology and Evolution, Volume 16, Issue 7, July 2024, evae148, <https://doi.org/10.1093/gbe/evae148>

Published: 03 July 2024 Article history

**Table 2** A comparison of genome completeness between the only other existing *A. luna* genome assembly available to date

	BUSCO complete (%)	Single Copy (%)	Duplicated (%)	Fragmented (%)	Missing (%)	Ref.
<i>Actias luna</i> <sup>a</sup>	99.4	99.0	0.4	0.2	0.4	Authors
<i>Actias luna</i>	71.4	63.9	7.5	15.4	13.2	GCA_010014465.3

<sup>a</sup>Genomes produced with long-read sequencing platforms, e.g. PacBio or Oxford Nanopore.



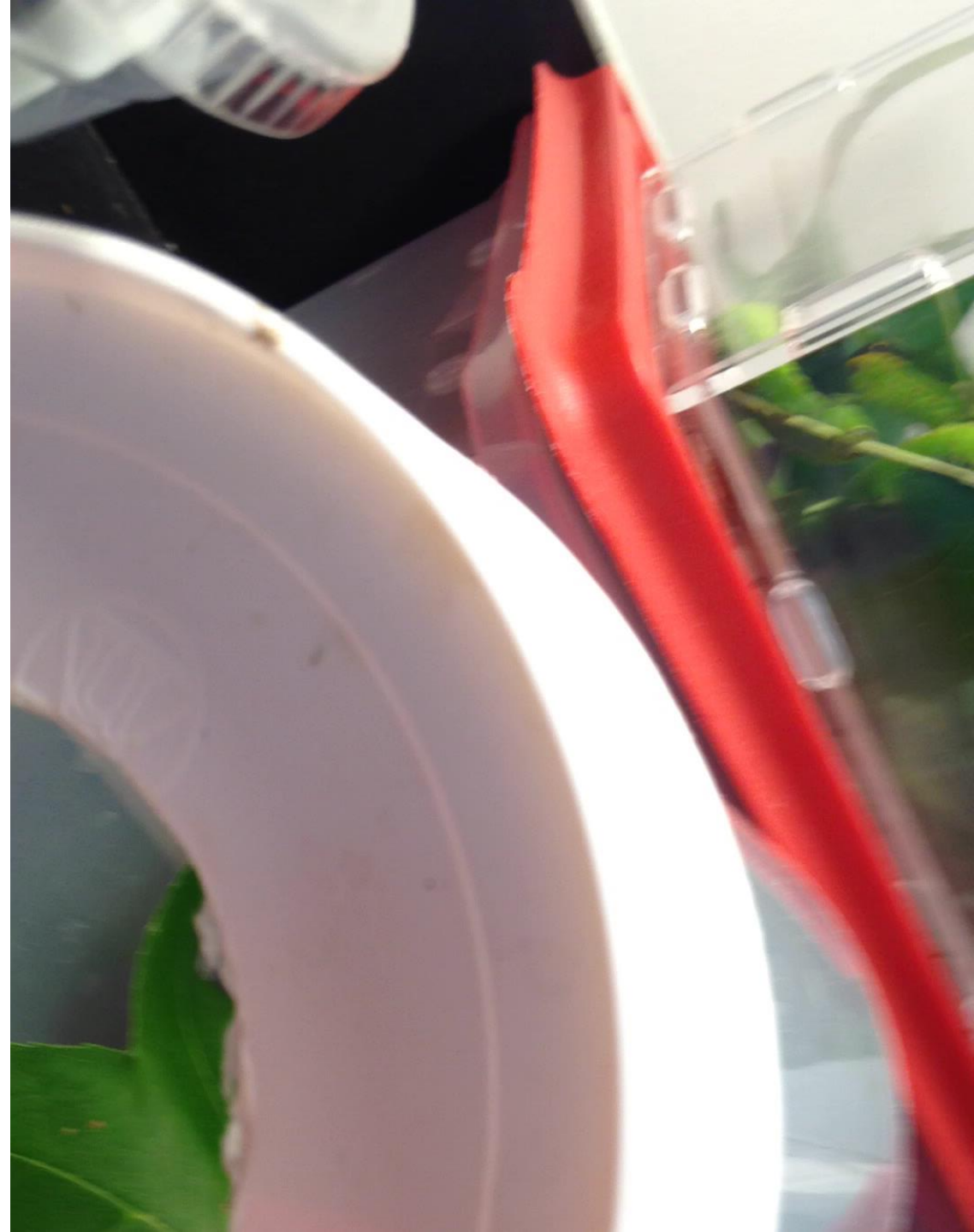


# ASSEMBLY STATS EXAMPLE

Parameter	<i>Actias luna</i> (long-read)	<i>Actias luna</i> (short-read)
Reference	This study	2016 genome
Platform	PacBio Sequel IIe	Illumina MiSeq/HiSeq
Genome completeness	99.4%	71.4%
Number of contigs	155	541,894
Contig N50	16,802,800	2,189
Contig L50	14	64,346
GC content	34.65%	35.50%
Shortest contig (bp)	6,149	1
Longest contig (bp)	21,422,706	136,946
Mean contig (bp)	3,441,632	1055

# CONCLUSIONS

- **Long read sequencing** has provided the opportunity for an influx of high-quality genomic resources for both model and non-model systems
- **Previous concerns** of lower accuracy continue to be addressed and built upon, making long-read sequencing arguably the most high-quality option for new genomes
- **Comparatively** long reads are **costly** and hopefully in time, will become more affordable and accessible across disciplines.



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