9/12

- prepare 500 mL each of cathode & anode buffers
  - Anode: 50 mM Bis Tris

$$X\ (1000\ mM) = (500\ mL)(50\ mM)$$

$$X = 25\ mL\ \text{Bis-Tris} + 475\ mL\ H_2O$$

  - Cathode: 15 mM Bis Tris, 50 mM Tricine

$$X\ (1000\ mM) = (500\ mL)(15\ mM)$$

$$X = 7.5\ mL\ \text{Bis Tris}$$

$$X\ (1000\ mM) = (500\ mL)(50\ mM)$$

$$X = 25\ mL\ \text{Tricine}$$

$$+ 467.5\ mL\ H_2O$$

B4 5/19/17
0.75 mg/mL
19.65 μM

- need 3X dilution of Hao's "Blue Native Gel Dye" in all samples except ladder
  - ↳ 6.6 μL each sample, 3.3 μL dye
- prep 15 μL of 1 mg/mL A2, then use 2-fold dilutions to decrease to 0.5 mg/mL and 0.25 mg/mL
  - ↳ $X\ (9.62\ mg/mL) = (15\ \mu L)(1\ mg/mL)$

$$X = 1.56\ \mu L + 13.44\ \mu L\ \cancel{H_2O}\ \text{PBS}$$

A2 7/2/18
9.62 mg/mL
209 μM

- start w/ 0.75 mg/mL B4, then dilute to 0.5, 0.25
  - ↳ $X\ (0.75\ mg/mL) = (15\ \mu L)(0.5\ mg/mL)$

$$X = 10\ \mu L + 5\ \mu L\ \cancel{H_2O}\ \text{PBS}$$

- load 8 μL / sample
- lanes:

1. ladder
2. A2 1 mg/mL
3. A2 0.5 mg/mL
4. A2 0.25 mg/mL
5. ladder
6. B4 0.75 mg/mL
7. B4 0.5 mg/mL
8. B4 0.25 mg/mL
9. ladder

- start gel 11 AM, 150 V

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- after ~10 min, no sample is visually detectable in any wells
  - ↳ perhaps the gel was oriented backwards
  - ↳ repeat in opposite orientation
- this time load 9.5 μL each
- also used newer "Native PAGE" gel, instead of "NuPAGE"
- new gel start 11:45 AM
  - ↳ added 25 min to run