SimBac: simulation of whole bacterial genomes with homologous recombination

Supplementary Information

Tom Brown¹, Xavier Didelot², Daniel J. Wilson^{3,4,5} and Nicola De Maio^{3,4,*}

- 1 Doctoral Training Centre, University of Oxford, Oxford, United Kingdom
- 2 Department of Infectious Disease Epidemiology, Imperial College, London, United Kingdom
- 3 Institute for Emerging Infections, Oxford Martin School, Oxford, United Kingdom
- 4 Nuffield Department of Medicine, University of Oxford, Oxford, United Kingdom
- 5 Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, United Kingdom
- * E-mail: nicola.demaio@ndm.ox.ac.uk

User Manual

SimBac jointly simulates bacterial genomes with the clonal genealogy under a coalescent model with recombination. Such simulations can be used to test phylogenetic analysis of real data sets.

Running from the command line

SimBac can be run from the command line using a combination of the following options. Passing no arguments to SimBac will display the possible arguments.

- -N NUM Sets the number of isolates (default is 100)
- -T NUM Sets the value of θ , the site-specific mutation rate, between 0 and 1 (default is 0.01)
- -m NUM Sets the lower bound of site-mutation (divergence) in a region of external recombination, between 0 and 1 (default is 0)
- -M NUM Sets the upper bound of site-mutation (divergence) in a region of external recombination, between 0 and 1 (default is 0)
- -R NUM Sets the per-site rate of internal (within species) recombination, R_i , (default is 0.01)
- -r NUM Sets the per-site rate of external (between species) recombination, R_e , (default is 0)
- -D NUM Sets the average length of an internal recombinant interval, δ_i (default is 500)
- -e NUM Sets the average length of an external recombinant interval, δ_e (default is 500)
- -B NUM,...,NUM Sets the number and lengths of fragments of genetic material (default is 10000)
- -G NUM,...,NUM Sets the size of gaps between each fragment, must be the same number of gaps as there are numbers of genetic fragments (default is 0,...,0)
- -s NUM Use given seed to initiate random number generation
- -o FILE Name of file to write generated sequences (FASTA format)
- -c FILE Name of file to write clonal genealogy (Newick format)
- -l FILE Name of file to write local trees (Newick format)
- -b FILE Name of file to write log of internal recombination breaks
- -f FILE Name of file to write log of external recombination breaks
- -d FILE Name of file to export ancestral recombination graph (DOT file)
- -a Include ancestral material in the DOT graph

Output format

SimBac produces the following output files:

- FASTA file of simulated sequences. If more than one fragment of genetic information is specified, the output is in the eXtended Multi-Fasta Alignment (XMFA) format. In this situation the simulated gene fragments are separated with an '=' sign.
- The clonal genealogy in Newick format
- The local trees contained in the simulated data. This file is a list of Newick trees each of which is preceded by the number of sites that share the current local tree.
- A full description of the graph representing the ancestry of the sample in the DOT language (Figure S 2). This can be used in conjunction with the graphviz and the DOT program to produce figures illustrating the ancestry. The examples show the ancestry with and without the ancestral material included at each node. The clonal genealogy is shown in bold and external recombination events are shown in red. In the graph showing the ancestral material, the ancestral material remaining at each node is shown in grey and any external genetic material is shown in red.

Examples

To simulate 100 genomes each 1Mb long with an internal recombination rate of 1% and mutation rate of 1% run:

./SimBac -N 100 -B 1000000 -R 0.01 -T 0.01 -o sequences.fasta -c clonal.nwk -l local.nwk

This produces the simulated sequences and the clonal genealogy in the files 'sequences.fasta' and 'clonal.nwk'. The local trees are written to 'local.nwk'

To simulate 100 genomes with internal and external recombination rate of 1% and average break length of 500bp run:

./SimBac -N 100 -B 1000000 -R 0.01 -D 500 -r 0.01 -e 500 -b internal.log -f external.log

This produces two log files with the start- and end-points of all internal and external recombination events.

To simulate sequences undergoing internal and external recombination with a mutation rate, θ of 100 with mutation in an external recombinant interval occurring with probability in the interval [0.5, 1] run:

./SimBac -N 100 -B 100000 -R 0.01 -D 500 -r 0.01 -e 500 -m 0.5 -M 1 -o sequences.fasta -c clonal.nwk This produces the sequences and clonal genealogy.

To produce a DOT file with the ancestral information included in the graph run:

./SimBac -n 10 -B 1000 -R 0.01 -D 50 -r 10 -e 50 -d graph.dot -a

To simulate a linear genome, add a large gap to the end of the genome to prevent any recombinant intervals including both the first and last elements of the genome. For example to simulate a linear genome of length 100kb run:

./SimBac -N 100 -B 100000 -G 1000000 -o sequences.fasta -c clonal.nwk

This places a gap of 1Mb at the end of the genome.

Supplementary Methods

Hereby we will use the notation of [2], except that we will assume that there is a circular genome of length G (with sites 1...G), and that ancestral material of each lineage is a subset of this genome. Ancestral material for a sample (an ARG tip) need not be the whole genome, but might be a subset of the genome made of different loci, for example in the case of MLST data, so that we can simulate both genome data or MLST data. Ancestral material for a node consists of b non-overlapping ordered intervals, $I_1...I_b$, of lengths respectively $L_1...L_b$, and with $I_i = [s_i, e_i]$ (implying $e_i - s_i = L_i - 1$). Also for easiness of presentation (due to genome circularity) we will set $e_0 = e_b - G$, which is intendedly negative. The recombination rate per site per genome will be R/2. It should be noted that the rate of recombination, R, is twice that of r_{ij} in LDhat [3]. Lastly, recombining intervals have a geometric distribution with mean δ .

Effective recombination rate for a lineage

In [2], the recombination rate per site is R/2. We call a the ancestral material of this lineage. If a recombination event happens on the considered lineage, then a recombining interval r is picked at random from the genome, and if $r \cap a \neq \emptyset$ (and $a - r \neq \emptyset$ for lineages not in the clonal frame) then the two new recombining lineages are created, otherwise the recombination event is rejected.

Here we propose to sample recombination events and recombining intervals conditional on $r \cap a \neq \emptyset$, $a - r \neq \emptyset$, or just on $r \cap a \neq \emptyset$ for lineages in the clonal frame, such that no rejection ever occurs while simulating. To do this, we first define a lineage-specific effective recombination rate. This is the rate at which recombination events occur satisfying $r \cap a \neq \emptyset$, $a - r \neq \emptyset$ (or just $r \cap a \neq \emptyset$ for clonal frame lineages). As in [2], we assume that the rate of initiation of a recombination event is the same for each site of the genome. Under these assumptions, and assuming as in [1] a geometric distribution with mean δ for recombination interval lengths, the rate at which a recombination event is started between e_0 and e_1 , and includes e_2 , is:

$$\frac{R_{s_1-e_0}}{2} = \frac{R}{2} \sum_{i=0}^{s_1-e_0-1} (1-\delta^{-1})^i =$$

$$= \frac{R}{2} \left[\sum_{i=0}^{\infty} (1-\delta^{-1})^i - \sum_{i=s_1-e_0}^{\infty} (1-\delta^{-1})^i \right] =$$

$$= \frac{R}{2} \left[\delta - \delta (1-\delta^{-1})^{(s_1-e_0)} \right] =$$

$$= \frac{R}{2} \delta (1 - (1-\delta^{-1})^{(s_1-e_0)}).$$

Where $(1 - \delta^{-1})^i$ is the probability of a recombinant break having length greater than i. Now, let us assume we have a lineage with ancestral material $a = \bigcup_{i=1}^b [s_i, e_i]$ union of non-empty, ordered, disjoint intervals. As mentioned before, $e_0 = e_b - G$. The amount of ancestral material in a lineage is defined as: $L = \sum_{i=1}^b L_i$ The rate of recombination events satisfying $r \cap a \neq \emptyset$ for that lineage is then:

$$\frac{R_a}{2} = \left(\sum_{i=1}^b \frac{R_{s_i - e_{i-1}}}{2}\right) + \frac{R}{2}(L - b).$$

Finally, the lineage-specific recombination rate satisfying $r \cap a \neq \emptyset$, and $a - r \neq \emptyset$ is:

$$\frac{R'_a}{2} = \frac{R_a}{2} - \sum_{i=1}^b \frac{R_{s_i - e_{i-1}}}{2} (1 - \delta^{-1})^{G - (s_i - e_{i-1})} - \frac{R}{2} (1 - \delta^{-1})^{G - 1} (L - b).$$

Additionally, for a clonal lineage without ancestral material the recombination rates will be 0.

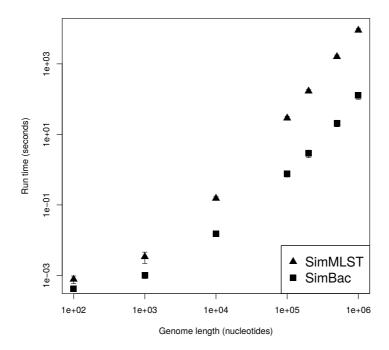


Fig. S 1: Comparison of run-time between SimMLST [2] and SimBac. The recombination rate was fixed at R = 0.01 and the length of the genome increased from 100b to 1Mb. 100 simulations were performed for each genome length and error bars represent ± 2 SD.

Probability of recombination initiating sites

Conditional on an effective recombination event on a non clonal frame lineage (that is, satisfying $r \cap a \neq \emptyset$, $a - r \neq \emptyset$) occurring on ancestral material a, the probability that the first ancestral site affected by r is s_i is:

$$P'_{s_i} = \frac{R_{s_i - e_{i-1}} (1 - (1 - \delta^{-1})^{G - (s_i - e_{i-1})})}{R'_{s_i}},$$

and the probability that it is any other site in a is

$$\frac{R(1-(1-\delta^{-1})^{G-1})}{R'_a}.$$

If the considered recombining lineage is in the clonal frame instead (with recombination satisfying only $r \cap a \neq \emptyset$), the probabilities are

 $P_{s_i} = \frac{R_{s_i - e_{i-1}}}{R_a},$

and

 $\frac{R}{R_a}$

respectively.

After the starting site of $r \cap a$ has been picked, the ending site of r is chosen according to a geometric distribution with mean δ for a lineage in the clonal frame. In a non-clonal lineage, the ending site of r is chosen according to the same geometric distribution, but conditional on $|r| \leq G - (s_i - e_{i-1})$ if the starting site of r is s_i , or $|r| \leq G - 1$ otherwise.

External recombination events

Simulation of external recombination events follows the same protocol as that of a clonal recombination event, with R_e and δ_e replacing R and δ , respectively. As we are only interested in the imported fragment from the external species, the recombinant interval need only satisfy the condition $r \cap a \neq \emptyset$, as in a clonal lineage.

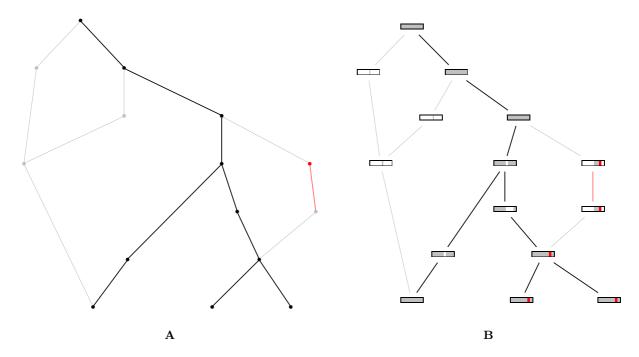


Fig. S 2: Examples of Ancestral Recombination Graphs (ARGs). The Nodes represent lineages and the events that make up the ARG. A shows the clonal frame in black, with the non-clonal lineage in grey and a recombinant event involving an external species in red. B shows the ancestral material in each lineage of the ARG. The ancestral material is shown at each node in grey. Material shown in red represents genetic material imported from an external species.

Technical description of SimBac

end while

Here, n denotes the number of isolates for which the data is to be simulated. For a genome of length G, simulate the ARG with internal recombination rate $R_i/2$ and external recombination rate $R_e/2$. The average length of internal and external recombining segments are given by δ_i and δ_e , respectively.

```
Input: n, R_i/2, R_e/2, \delta_i, \delta e, G
 Output: Simulated ancestral recombination graph with clonal genealogy
Set number of lineages k = n
for x = 1 \dots n do
   Calculate internal and external recombination rates, \frac{R_{i,x}}{2}, \frac{R_{e,x}}{2}
    Ancestral material a_x is given by [0, G]
   Lineage x is clonal
end for
while k > 1 do
    Calculate the rates of internal and external recombination given by:
    \frac{R_i}{2} = \sum_{x=1}^k \frac{R_{i,x}}{2} and \frac{R_e}{2} = \sum_{x=1}^k z \frac{R_{e,x}}{2}
   Increment current time by an amount distributed exponentially with parameter \binom{k}{2} + \frac{R_i}{2} + \frac{R_e}{2}
   Let u \sim U(0,1)
   if u < k(k-1)/(k(k-1) + R_i + R_e) then
       Coalescent event
       Choose two lineages x and y at random from the k remaining nodes and replace with the new lineage z
       The lineage z is clonal if x or y is clonal
       The ancestral material of z is a_z = a_x \cup a_y
       Update total number of lineages containing each element of the genome
       for all Nucleotides do
           if Number of lineages containing given nucleotide is one then
                Remove nucleotide from a_z
           end if
       end for
       Calculate the internal and external rate of recombination for the new lineage, \frac{R_{i,z}}{2} and \frac{R_{e,z}}{2}
       The number of lineages, k is decreased by one
   else if u < (k(k-1) + R_i)/(k(k-1) + R_i + R_e) then
       Internal recombination event
       Choose one lineage x weighted by \frac{R_{i,x}}{2}
Determine a recombining interval, r, distributed geometrically with parameter \delta_i.
       if x is clonal then
           Choose r such that r \cap a_x \neq \emptyset
            Choose r such that r \cap a_x \neq \emptyset and a - r \neq \emptyset
       end if
       Create two new lineages, y and z.
       Lineage z is clonal if x is clonal
       a_y = a_x \cap r and a_z = a_x - r
       Lineage y is not clonal
       Calculate the new recombination rates for lineages y and z, \frac{R_{i,y}}{2}, \frac{R_{e,y}}{2}, \frac{R_{i,z}}{2} and \frac{R_{e,z}}{2}
       The number of lineages, k is increased by one
   else
       External recombination event
       Choose one lineage, x weighted by \frac{R_{e,x}}{2}
       Choose a recombinant interval, r, distributed geometrically with parameter \delta_e conditioned on r \cap a_x \neq \emptyset
       The material satisfying a_x \cap r will be simulated as genetic material from an external species
   end if
```

References

- [1] Didelot, X., Falush, D. (2006) Inference of Bacterial Microevolution Using Multilocus Sequence Data. Genetics, 175, 1251-1266.
- [2] Didelot, X. Lawson, D, Falush, D. (2009) SimMLST: simulation of multi-locus sequence typing data under a neutral model *Bioinformatics*, **25**(11): 1442–1444.
- [3] McVean, G. Awadalla, P. Fearnhead, P. (2002) A Coalescent-Based Method for Detecting and Estimating Recombination From Gene Sequences *Genetics*, **160**: 1231–1241.