

¹ Increasing prevalence of plant-fungal symbiosis across two
² centuries of environmental change

³ Joshua C. Fowler^{1,2*}

Jacob Moutouama¹

Tom E. X. Miller¹

⁴ 1. Rice University, Department of BioSciences, Houston, Texas 77006; 2. University of Miami,
⁵ Department of Biology, Miami, Florida;
⁶ * Corresponding author; e-mail: jcf221@miami.edu.

⁷ *Manuscript elements:* Figure 1 - Figure 5, appendix A (including Figure A1 - Figure A14, Table
⁸ A1, and Supplemental Methods).

⁹ *Keywords:* .

¹⁰ *Manuscript type:* Article.

¹¹ Prepared using the suggested L^AT_EX template for *Am. Nat.*

Abstract

13 Species' distributions and abundances are shifting in response to climate change. Most species
14 harbor microbial symbionts that have the potential to influence these responses. Mutualistic mi-
15 crobial symbionts may provide resilience to environmental change by protecting their hosts from
16 increasing stress. However, environmental change that disrupts these interactions may lead to
17 declines in hosts or symbionts. Microbes preserved within herbarium specimens offer a unique
18 opportunity to quantify changes in microbial symbiosis across broad temporal and spatial scales.
19 We asked how the prevalence of seed-transmitted fungal symbionts of grasses (*Epichloë* endo-
20 phytes), which can protect hosts from abiotic stress, have changed over time in response to
21 climate change, and how these changes vary across host species' ranges. Specifically, we ana-
22 lyzed 2,346 herbarium specimens of three grass host species collected over the last two centuries
23 (1824 – 2019) for the presence or absence of endophyte symbiosis, and evaluated spatial and
24 temporal trends in endophyte prevalence. We found that endophytes increased in prevalence
25 over the last two centuries from ca. 25% prevalence to ca. 75% prevalence, on average, across
26 three host species. We also found that changes in prevalence were associated with observed
27 changes in seasonal climate drivers; notably increasing precipitation corresponding to each host
28 species' peak growing season and changes in off-peak season variability in precipitation. Our
29 analysis performed favorably in an out-of-sample predictive test with contemporary data, how-
30 ever we identified greater local-scale variability in endophyte prevalence in contemporary data
31 compared to historic data, suggesting that model fusion may be an important step moving for-
32 ward. Our results provide novel evidence for a cryptic biological response to climate change that
33 may contribute to the resilience of host-microbe symbiosis through context-dependent benefits
34 that confer a fitness advantage to symbiotic hosts under environmental change.

35 Abstract : 287 words

Introduction

37 Understanding how biotic interactions are altered by global change is a major goal of basic and
38 applied ecological research (Blois et al., 2013; Gilman et al., 2010). Documented responses to
39 environmental change, such as shifts in species' distributions (Aitken et al., 2008) and phenology
40 (Piao et al., 2019), are typically blind to concurrent changes in associated biotic interactions.
41 Empirically evaluating these biotic changes – whether interacting species shift in tandem with
42 their partners or not (HilleRisLambers et al., 2013) – is crucial to predicting the reorganization
43 of Earth's biodiversity under global change. Such evaluations have been limited because few
44 datasets on species interactions extend over sufficiently long time scales of contemporary climate
45 change (Poisot et al., 2021).

46 Natural history specimens, which were originally collected to study and preserve taxonomic
47 diversity, present a unique opportunity to explore long-term changes in ecological interactions
48 across broad spatial and temporal scales (Meineke et al., 2018). Natural history collections, built
49 and maintained by the efforts of thousands of scientists, are invaluable time machines, primarily
50 comprised of physical specimens of organisms along with information about the time and place
51 of their collection. These specimens often preserve physical legacies of ecological processes and
52 species' interactions from dynamically changing environments across time and space. For exam-
53 ple, previous researchers have used plant collections (herbaria) to document shifts in phenology
54 (Berg et al., 2019; Park et al., 2019; Willis et al., 2017), pollination (Duan et al., 2019; Pauw and
55 Hawkins, 2011), and herbivory (Meineke et al., 2019) related to anthropogenic climate change.
56 However, few previous studies have leveraged biological collections to examine climate change-
57 related shifts in a particularly common type of interaction: microbial symbiosis.

58 Microbial symbionts are common to all macroscopic organisms and can have important ef-
59 fects on their hosts' survival, growth and reproduction (McFall-Ngai et al., 2013; Rodriguez et al.,
60 2009). Many microbial symbionts act as mutualists, engaging in reciprocally beneficial interac-
61 tions with their hosts that can ameliorate environmental stress. For example, bacterial symbionts

of insects, such as *Wolbachia*, can improve their hosts' thermal tolerance (Renoz et al., 2019; Truitt et al., 2019), and arbuscular mycorrhizal fungi, documented in 70-90% of families of land plants (Parniske, 2008), allow their hosts to persist through drought conditions by improving water and nutrient uptake (Cheng et al., 2021). On the other hand, changes in the mean and variance of environmental conditions may disrupt microbial mutualisms by changing the costs and benefits of the interaction for each partner, leading the interaction to deteriorate (Aslan et al., 2013; Fowler et al., 2024). Coral bleaching (the loss of symbiotic algae) due to temperature stress (Sully et al., 2019) is perhaps the best known example, but this phenomenon is not unique to corals. Lichens exposed to elevated temperatures experienced loss of photosynthetic function along with changes in the composition of their algal symbiont community (Meyer et al., 2022). How commonly and under what conditions microbial mutualisms deteriorate or strengthen under climate change remain unanswered questions. Previous work suggests that these alternative responses may depend on the intimacy and specialization of the interaction as well as the physiological tolerances of the mutualist partners (Rafferty et al., 2015; Toby Kiers et al., 2010; Warren and Bradford, 2014).

Understanding of how microbial symbioses are affected by climate change is additionally complicated by spatial heterogeneity in the direction and magnitude of environmental change (IPCC, 2021). Beneficial symbionts are likely able to shield their hosts from environmental stress in locations that experience a small degree of change, but symbionts in locations that experience changes of large magnitude may be pushed beyond their physiological limits (Webster et al., 2008). Additionally, symbionts are often unevenly distributed across their hosts' distribution. Facultative symbionts may be absent from portions of the host range (Afkhami et al., 2014), and hosts may engage with a diversity of partners (different symbiont species or locally-adapted strains) across their environments (Frade et al., 2008; Rolshausen et al., 2018). Identifying broader spatial trends in symbiont prevalence is therefore an important step in developing predictions for where to expect changes in the symbiosis in future climates.

Epichloë fungal endophytes are specialized symbionts of cool-season grasses, which have been

89 documented in ~ 30% of cool-season grass species (Leuchtmann, 1992). They are transmitted ver-
90 tically from maternal plants to offspring through seeds. Vertical transmission creates a feedback
91 between the fitness of host and symbiont (Douglas, 1998; Fine, 1975; Rudgers et al., 2009). Over
92 time, endophytes that act as mutualists should rise in prevalence within a host population (Don-
93 ald et al., 2021). *Epichloë* are known to improve their hosts' drought tolerance (Decunta et al.,
94 2021) and protect their hosts against herbivores (Crawford et al., 2010) and pathogens (Xia et al.,
95 2018) likely through the production of a diverse suite of alkaloids and other secondary metabo-
96 lites. The fitness feedback induced by vertical transmission leads to the prediction that endophyte
97 prevalence should be high in populations where these fitness benefits are most important. Pre-
98 vious contemporary survey studies have documented large-scale spatial patterns in endophyte
99 prevalence structured by environmental gradients (Afkhami, 2012; Bazely et al., 2007; Granath
100 et al., 2007; Sneck et al., 2017). We predicted that prevalence should track temporal changes in
101 environmental drivers that elicit these fitness benefits.

102 Early research on *Epichloë* used herbarium specimens to describe the broad taxonomic di-
103 versity of host species that harbor these symbionts (White and Cole, 1985), establishing that
104 endophyte symbiosis could be identified in plant tissue from as early as 1851. However, no
105 subsequent studies, to our knowledge, have used the vast resources of biological collections to
106 quantitatively assess spatio-temporal trends in endophyte prevalence and their environmental
107 correlates. Grasses are commonly collected and identified based on the presence of their re-
108 productive structures, meaning that preserved specimens typically contain seeds, conveniently
109 preserving the fungi along with their host plants on herbarium sheets. This creates the oppor-
110 tunity to leverage the unique spatio-temporal sampling of herbarium collections to examine the
111 response of the symbiosis to historical climate change. Research using historical collections has
112 clearly demonstrated other ecological signatures of a changing climate. However the predictive
113 ability of these historical analyses is rarely tested against contemporary data (Lee et al., 2024).
114 Identifying the ways in which these analyses fall short is a crucial step for the field move from
115 reading signatures in the past to forecasting ecological dynamics into the future.

116 In this study, we assessed the long-term responses of endophyte symbiosis to climate change
117 through the use of herbarium specimens of three North American host grass species (*Agrostis*
118 *hyemalis*, *Agrostis perennans*, and *Elymus virginicus*). We first address questions describing spatial
119 and temporal trends in endophyte prevalence: (i) How has endophyte prevalence changed over
120 the past two centuries? and (ii) How spatially variable are temporal trends in endophyte preva-
121 lence across eastern North America? We then address how climate change may be driving trends
122 in endophyte prevalence by asking: (iii) What is the relationship between variation in temporal
123 trends in endophyte prevalence and changes in climate drivers? We predicted that aggregate en-
124 dophyte prevalence would increase over time in tandem with climate warming, and that hotspots
125 of endophyte change would correspond spatially to hotspots of climate change. Finally, we eval-
126 uated the performance of models built on data from historic specimens with an out-of-sample
127 test, data on endophyte prevalence from contemporary surveys of host populations. To answer
128 these questions we examined a total of 2,346 specimens collected across eastern North America
129 between 1824 and 2019.

130 **Methods**

131 *Focal species*

132 Our surveys focused on three native North American grasses: *Agrostis hyemalis*, *Agrostis peren-*
133 *nans*, and *Elymus virginicus*. Both *Agrostis* species host *Epichloë amarillans* (Craven et al., 2001;
134 Leuchtmann et al., 2014), while *Elymus virginicus* typically hosts *Epichloë elymi* (Clay and Schardl,
135 2002). These C₃ grass species are commonly represented in natural history collections with broad
136 distributions covering much the eastern United States. *A. hyemalis* is a small short-lived perennial
137 species that germinates in the spring and typically flowers between March and July (most com-
138 mon collection month: May). *A. perennans* is of similar stature but is longer lived than *Agrostis*
139 *hyemalis* and flowers in late summer and early autumn (most common collection month: Septem-
140 ber). *A. perennans* is more sparsely distributed, tending to be found in shadier and more moist

¹⁴¹ habitats, while *A. hyemalis* is commonly found in open and recently disturbed ground. Both
¹⁴² *Agrostis* species are recorded from throughout the Eastern US, but *A. perennans* has a slightly
¹⁴³ more northern distribution, whereas *A. hyemalis* is found rarely as far north as Canada and is
¹⁴⁴ listed as a rare plant in Minnesota. *E. virginicus* is a larger and relatively longer-lived species that
¹⁴⁵ is more broadly distributed than the *Agrostis* species. It begins flowering as early as March or
¹⁴⁶ April but continues throughout the summer (most common collection month: July).

¹⁴⁷ *Herbarium surveys*

¹⁴⁸ We visited nine herbaria between 2019 and 2022 (see Table A1 for a summary of specimens in-
¹⁴⁹ cluded from each collection). With permission from herbarium staff, we acquired seed samples
¹⁵⁰ from 1135 *A. hyemalis* specimens collected between 1824 and 2019, 357 *A. perennans* specimens
¹⁵¹ collected between 1863 and 2017, and 854 *E. virginicus* specimens collected between 1839 and
¹⁵² 2019 (Fig. 1, Fig. 2A, Fig. A1). We chose our focal species in part because they are commonly
¹⁵³ represented in herbarium collections, and produce high numbers of seeds, meaning that small
¹⁵⁴ samples would not diminish the value of the specimens for future studies. We collected 5-10
¹⁵⁵ seeds per specimen after examining the herbarium sheet under a dissecting microscope to en-
¹⁵⁶ sure that we collected mature seeds, not florets or unfilled seeds, fit for our purpose of identifying
¹⁵⁷ fungal endophytes with microscopy. We excluded specimens for which information about the
¹⁵⁸ collection location and date were unavailable. Each specimen was assigned geographic coor-
¹⁵⁹ dinates based on collection information recorded on the herbarium sheet using the geocoding
¹⁶⁰ functionality of the ggmap R package (Kahle et al., 2019). Many specimens had digitized collec-
¹⁶¹ tion information readily available, but for those that did not, we transcribed information printed
¹⁶² on the herbarium sheet. Collections were geo-referenced to the nearest county centroid, or near-
¹⁶³ est municipality when that information was available. For a few of the oldest specimens, only
¹⁶⁴ information at the state level was available, and so we used the state centroid.

¹⁶⁵ After collecting seed samples, we quantified the presence or absence of *Epichloë* fungal hy-
¹⁶⁶ phae, which grow intercellularly, in each specimen using microscopy. We first softened seeds

167 with a 10% NaOH solution, then stained the seeds with aniline blue-lactic acid stain and squashed
168 them under a microscope cover slip. We examined the squashed seeds for the presence of fungal
169 hyphae at 200-400X magnification (Bacon and White, 2018). In some cases, the tissues examined
170 during microscopy came from flowers or otherwise non-viable seeds, which were excluded for
171 that specimen. On average we scored 4.7 seeds per specimen of *A. hyemalis*, 4.2 seeds per speci-
172 men of *A. perennans*, and 3.8 seeds per specimen of *E. virginicus*; we scored 10,342 seeds in total. .
173 Due to imperfect vertical transmission (Afkhami and Rudgers, 2008), it is possible that symbiotic
174 host-plants produce a mixture of symbiotic and non-symbiotic seeds. We therefore designated a
175 specimen as endophyte-symbiotic if *Epichloë* hyphae were observed in one or more of its seeds,
176 or non-symbiotic if hyphae were observed in none of its seeds. To capture uncertainty in the
177 endophyte scoring process, we recorded both a "liberal" and a "conservative" endophyte status
178 for each plant specimen. When we identified potential endophytes with unusual morphology,
179 low uptake of stain, or a small amount of fungal hyphae across the scored seeds, we recorded a
180 positive liberal status (more likely to be endophyte-positive) and a negative conservative status
181 (less likely to be endophyte-positive). 89% of scored plants had matching liberal and conservative
182 scores, reflecting high confidence in endophyte status. The following analyses used the liberal
183 status, but we repeated all analyses with the conservative status which yielded qualitatively
184 similar results (Fig. ??)

185 *Modeling spatial and temporal changes in endophyte prevalence*

186 We assessed spatial and temporal changes in endophyte prevalence across each host distribution,
187 quantifying the "global" temporal trends aggregated across space, and then examining spatial
188 heterogeneity in the direction and magnitude of endophyte change (hotspots and coldspots)
189 across the spatial extent of each host's distribution. To account for the spatial non-independence
190 of geo-referenced occurrences, we used an approximate Bayesian method, Integrated Nested
191 Laplace Approximation (INLA), to construct spatio-temporal models of endophyte prevalence.
192 INLA provides a computationally efficient method of ascertaining parameter posterior distribu-

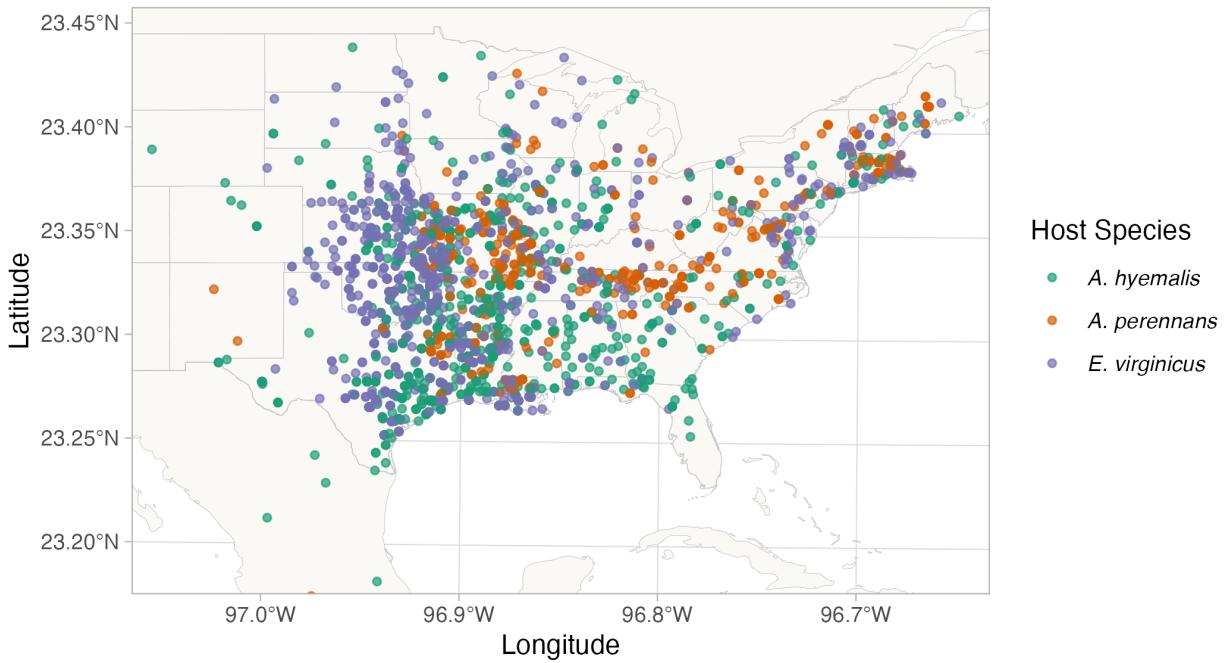


Figure 1: Collection locations of herbarium specimens of three grass host species across eastern North America that were sampled for *Epichloë* endophyte presence or absence.

193 tions for certain models that can be formulated as latent Gaussian Models (Rue et al., 2009). Many
 194 common statistical models, including structured and unstructured mixed-effects models, can be
 195 represented as latent Gaussian Models. We incorporated spatial heterogeneity into this analy-
 196 sis using spatially-structured intercept and slope parameters implemented as stochastic partial
 197 differential equations (SPDE) to approximate a continuous spatial Gaussian process. This SPDE
 198 approach is a flexible method of smoothing across space while explicitly accounting for spatial
 199 dependence between data-points (Bakka et al., 2018; Lindgren et al., 2011). Fitting models with
 200 structured spatial effects is possible with MCMC sampling but can require long computation
 201 times, making INLA an effective alternative, which has been used to model spatial patterns in
 202 flowering phenology (Willems et al., 2022), the abundance of bird species (Meehan et al., 2019)
 203 and butterflies (Crossley et al., 2022), the distribution of temperate trees (Engel et al., 2022) as well
 204 as the population dynamics of endangered amphibians (Knapp et al., 2016) and other ecological

205 processes (Beguin et al., 2012).

206 We estimated global and spatially-varying trends in endophyte prevalence using a joint-
207 likelihood model. For each host species h , endophyte presence/absence of the i^{th} specimen ($P_{[h]i}$)
208 was modeled as a Bernoulli response variable with expected probability of endophyte occurrence
209 $\hat{P}_{[h]i}$. We modeled $\hat{P}_{[h]i}$ as a linear function of intercept $A_{[h]i}$ and slope $T_{[h]}$ defining the global trend
210 in endophyte prevalence specific to each host species as well as with spatially-varying intercepts
211 $\alpha_{[h_1]l[i]}$ and slopes $\tau_{[h_1]l[i]}$ associated with location ($l[i]$, a unique latitude-longitude combination).
212 The joint-model structure allowed us to share variance terms across focal species to account for
213 dependence associated with the collection of specimens and identification of endophytes. Shared
214 variance terms included the spatially-dependent random effect $\delta_{l[i]}$, intended to account for resid-
215 ual spatial variation, and $\chi_{c[i]}$ and $\omega_{s[i]}$ i.i.d.-random effects indexed for each collector identity
216 ($c[i]$), and scorer identity ($s[i]$) of the i^{th} specimen.

$$\text{logit}(\hat{P}_{[h_1]i}) = A_{[h_1]i} + T_{[h_1]} * \text{year}_i + \alpha_{[h_1]l[i]} + \tau_{[h_1]l[i]} * \text{year}_i + \delta_{l[i]} + \chi_{c[i]} + \omega_{s[i]} \quad (1a)$$

$$\text{logit}(\hat{P}_{[h_2]i}) = A_{[h_2]i} + T_{[h_2]} * \text{year}_i + \alpha_{[h_2]l[i]} + \tau_{[h_2]l[i]} * \text{year}_i + \delta_{l[i]} + \chi_{c[i]} + \omega_{s[i]} \quad (1b)$$

$$\text{logit}(\hat{P}_{[h_3]i}) = A_{[h_3]i} + T_{[h_3]} * \text{year}_i + \alpha_{[h_3]l[i]} + \tau_{[h_3]l[i]} * \text{year}_i + \delta_{l[i]} + \chi_{c[i]} + \omega_{s[i]} \quad (1c)$$

217 Previous work suggests that behavior of historical botanists and uneven sampling may intro-
218 duce biases into ecological inferences made from historic collections (Kozlov et al., 2020). Prolific
219 collectors who contribute thousands of specimens may be more or less likely to collect certain
220 species, or specimens with certain traits (Daru et al., 2018). Similarly, the process of scoring seeds
221 for hyphae involved several student researchers who, even with standardized training, may vary
222 in their likelihood of positively identifying *Epichloë* hyphae. By including a random effect for
223 collectors and for scorers, we attempted to account for variance across individual researchers
224 that may bias our predictions of changes in endophyte prevalence.

225 We performed model fitting using the inlabru R package (Bachl et al., 2019). Global intercept
226 and slope parameters A , and T , were given vague priors. Scorer and collector random effects,

²²⁷ χ and ω , were given penalized complexity priors, with distributions approximating a Normal
²²⁸ distribution with standard deviation of 5. Each spatially-structured parameter depended on a
²²⁹ covariance matrix according to the proximity of each collection location (Bakka et al., 2018; Lind-
²³⁰ gren et al., 2011). The covariance matrix was approximated using a Matérn covariance function,
²³¹ with each data point assigned a location according to the nodes of a mesh of non-overlapping
²³² triangles encompassing the study area (Fig. A2). We assessed model fit with visual posterior
²³³ predictive checks (A3) and measurements of AUC (Figs. A4-A5). Priors, termed "range" and
²³⁴ "variance", define the distance of spatial decay described by the Matérn covariance function. Pri-
²³⁵ ors for results presented in the main text reflect a range of 342 kilometers. We found that model
²³⁶ results were sensitive to this choice, and so tested a range of priors (from 68 kilometers to 2160
²³⁷ kilometers) and meshes (Supplemental Material), finding that model results were qualitatively
²³⁸ similar, i.e. the same direction of effects across space, but that the magnitude and uncertainty
²³⁹ varied.

²⁴⁰ *Validating model performance with in-sample and out-of-sample tests*

²⁴¹ We evaluated the predictive ability of the model using both in-sample training data from the
²⁴² herbarium surveys, and with out-of-sample test data from contemporary endophyte surveys, an
²⁴³ important but rarely used strategy in ecological studies (Lee et al., 2024; Tredennick et al., 2021).
²⁴⁴ We used data from contemporary surveys of endophyte prevalence in *A. hyemalis* and *E. virginini-
245 cus* in Texas and the southern US. Surveys of *E. virginicus* were conducted in 2013 as described
²⁴⁶ in Sneck et al. (2017), and surveys of *A. hyemalis* took place between 2015 and 2020. Popula-
²⁴⁷ tion surveys of *A. hyemalis* were initially designed to cover longitudinal variation in endophyte
²⁴⁸ prevalence towards its range edge, while surveys of *E. virginicus* were designed to cover latitu-
²⁴⁹ dinal variation along its range edge. In total, we visited 43 populations of *A. hyemalis* and 20
²⁵⁰ populations of *E. virginicus* across the south-central US, with emphasis on Texas and neighboring
²⁵¹ states (Fig A11). During surveys, we collected seeds from up to 30 individuals per location (av-
²⁵² erage number of plants sampled: 22.9). We quantified the endophyte status of each individual

253 with staining microscopy as described for the herbarium surveys (with 5-10 seeds scored per
254 individual), and calculated the prevalence of endophytes within the population (proportion of
255 symbiotic plants divided by the number of sampled plants). For each population, we compared
256 the observed fraction of endophyte-symbiotic hosts to the predicted probability of endophyte
257 occurrence \hat{P} derived from the model based on location and year. The contemporary survey
258 period (2013-2020) is at the most recent edge of the time period encompassed by the historical
259 observations used for model fitting. We compared the model's prediction for these locations to
260 the observed population prevalence.

261 *Assessing the role of climate drivers*

262 We assessed how the magnitude of climate change may have driven changes in endophyte preva-
263 lence by assessing correlations between changes in climate and changes in endophyte prevalence
264 predicted from our spatial model at evenly spaced pixels across the study area. We first down-
265 loaded monthly temperature and precipitation rasters from the PRISM climate group (Daly and
266 Bryant, 2013) covering the time period between 1895 and 2020 using the 'prism' R package (Hart
267 and Bell, 2015). Prism provides reconstructions of historic climate variables across the United
268 States by spatially-interpolating weather station data (Di Luzio et al., 2008). We calculated 30-
269 year climate normals for seasonal mean temperature and cumulative precipitation for the recent
270 (1990 to 2020) and historic (1895 to 1925) periods. We used three four-month seasons within
271 the year (Spring: January, February, March, April; Summer: May, June, July, August; Autumn:
272 September, October, November, December). This division of seasons allowed us to quantify dif-
273 ferences in climate associated with the two "cool" seasons, when we expected our focal species
274 to be most biologically active (*A. hyemalis* flowering phenology: spring; *E. virginicus*: spring and
275 summer; *A. perennans*: autumn). In addition to mean climate conditions, environmental vari-
276 ability itself can influence population dynamics (Tuljapurkar, 1982) and changes in variability
277 are a key prediction of climate change models (IPCC, 2021; Stocker et al., 2013). Therefore, we
278 calculated the standard deviation for each annual and seasonal climate driver across each 30-

279 year period. We then took the difference between recent and historic periods for the mean and
280 standard deviation for each climate driver (Figs. A12-A14). All together, we assessed twelve
281 potential climate drivers: the mean and standard deviation of spring, summer, and autumn temper-
282 ature, as well as the mean and standard deviation of spring, summer, and autumn cumulative
283 precipitation, cumulative precipitation, and cumulative precipitation.

284 To evaluate whether areas that have experienced the greatest changes in endophyte preva-
285 lence (hotspots of endophyte change) are associated with high degrees of change in climate
286 (hotspots of climate change), we modeled spatially varying slopes of endophyte change through
287 time ($\tau_{[h]l}$) as a linear function of environmental covariates, with a Gaussian error distribution.
288 Data from each host species was analyzed separately. Fitting regressions to many pixels across
289 the study region risks artificially inflating confidence in our results due to large sample sizes, and
290 so we performed this analysis using only a random subsample of 250 pixels across the study
291 region, which provided results qualitatively similar to analysis of the full set of pixels.

292 *Modeling distributions of host species*

293 We modeled the distribution of each host species to generate maps on which we predicted the
294 dynamics of *Epichloë* symbionts. We followed the ODMAP (overview, data, model, assessment,
295 prediction) protocol (Crossley et al., 2022). A full description of the ODMAP can be found in
296 the (Supporting Methods). In short, we used presence-only observations of the host species from
297 Global Biodiversity Information Facility (GBIF) between 1990 to 2020. To reduce the potential
298 influence of sampling bias and spatial autocorrelation, we thinned the occurrences to the spatial
299 scale of our selected climatic predictors. We selected climate variables that aligned with our
300 analysis of climatic influences on trends in endophyte prevalence described above. We calculated
301 the mean and standard deviation of seasonal temperature and precipitation across 1990 to 2020.
302 Among this suite of variables, we chose to include, which were uncorrelated (Variance Inflation
303 Factor > 0.7) and allowed us to predict the occurrence probability of each host species in space
304 and time. These climatic variables are: the mean and standard deviation of spring, summer,

and autumn temperature, as well as the mean and standard deviation of spring, summer, and autumn cumulative precipitation, cumulative precipitation, and cumulative precipitation. We fit maximum entropy (MaxEnt) models using the maxent function in the package dismo (Hijmans et al., 2017). We generated 10,000 pseudo-absences as background points, and split the occurrence data into 75% for model training and 25% for model testing. The performance of models was evaluated with AUC (Jiménez-Valverde, 2012). We found AUC = 0.862, AUC = 0.838, AUC = 0.821 respectively for *Agrostis hyemalis*, *Agrostis perennans*, and *Elymus virginicus*. To convert the continuous predicted probabilities into binary presence - absence maps, we used the training sensitivity (true positive rate) and specificity threshold (true negative rate) (Liu et al., 2005). These binary maps serve as boundaries in presented maps of change in endophyte prevalence, and outline the set of pixels used in our analysis of climate correlates with trends in endophyte prevalence

Results

How has endophyte prevalence changed over time?

We found that endophyte prevalence increased within the examined specimens over the last two centuries for all three host species (Fig. 2). On average, modeling indicated that endophytes of *A. perennans* and *E. virginicus* increased from ~ 40 % to 70% prevalence across the study region, and that of *A. hyemalis* increased from ~ 25% to over 50% prevalence. Our model indicates a high certainty that overall temporal trends are positive across species (99% probability of a positive overall year slope in *A. hyemalis*, 92% probability of a positive overall year slope in *A. perennans*, and 91% probability of a positive overall year slope in *E. virginicus*) (Fig. A6)

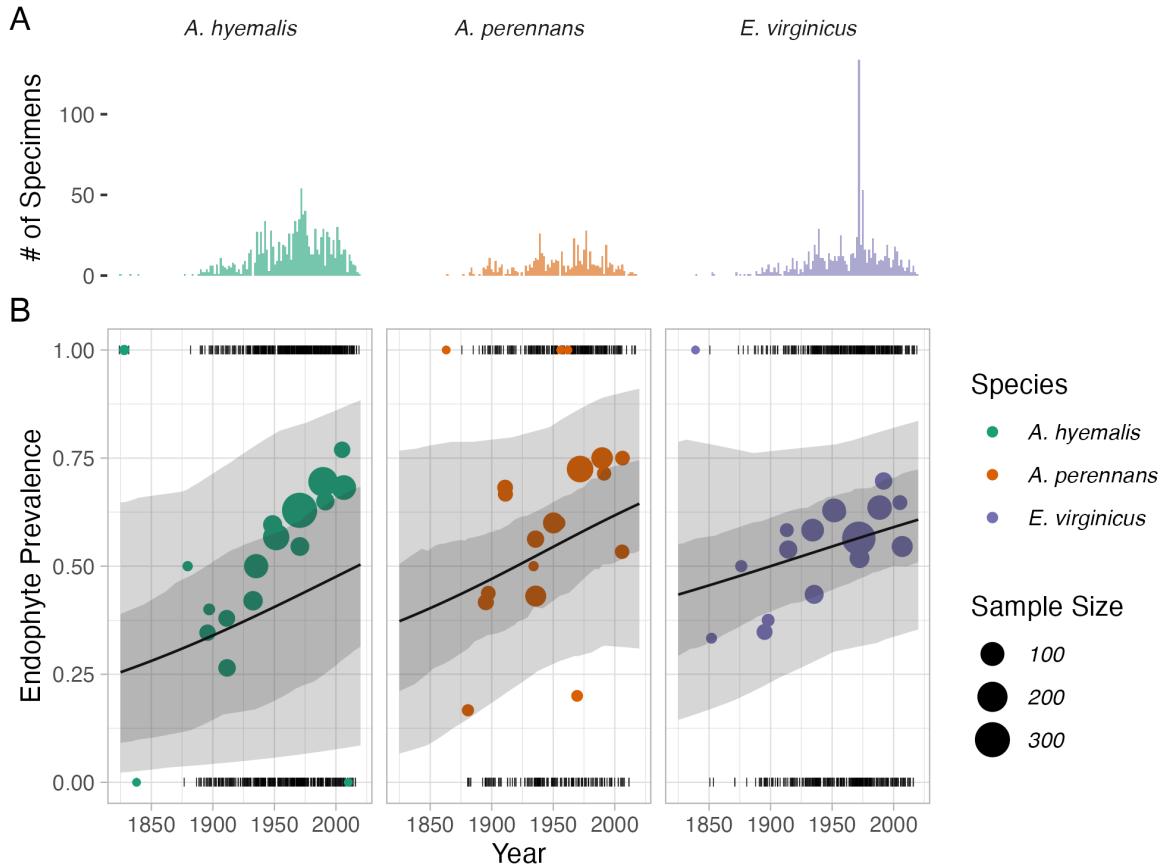


Figure 2: Temporal trends in endophyte prevalence. (A) Histograms show the frequency of scored specimens through time for each host species. (B) Lines show predicted mean endophyte prevalence over the study period along with the 50% and 95% CI bands incorporating uncertainty associated with collector and scorer random effects. Colored points are binned means of the observed endophyte presence/absence data (black dashes). Colors represent each host species and point size represents the number of specimens.

326 *How spatially variable are temporal trends in endophyte prevalence?*

327 Our model revealed hotspots of change in endophyte prevalence. While there was an overall
 328 increase in endophyte prevalence, these changes varied across the host species' ranges (Fig. 3).
 329 In some regions, posterior estimates of spatially varying temporal trends, τ , indicate that *A.*

330 *hyemalis* and *A. perennans* experienced increases in percent prevalence by as much as 2% per
 331 year over the study period, while *E. virginicus* experienced increases up to around 1% per year.
 332 Compared to *E. virginicus*, which had a weaker overall increase in endophytes and less spatial
 333 variability, maps of both *Agrostis* species show areas of strong increase and areas of declining
 334 prevalence. Notably, endophytes increased towards the western range edge of *A. hyemalis* (Fig.
 335 3A) and across the northeastern US for *A. perennans* (Fig. 3B). Posterior estimates of uncertainty
 336 in spatially varying slopes indicate that these hotspots of change may have experienced increases
 337 of up to 5% per year while declines in prevalence may be as great as 4% per year for *A. hyemalis*
 338 and *A. perennans*. For *E. virginicus*, uncertainty ranges between 3.5% increases and 2.5% decreases
 339 (Fig. A7).

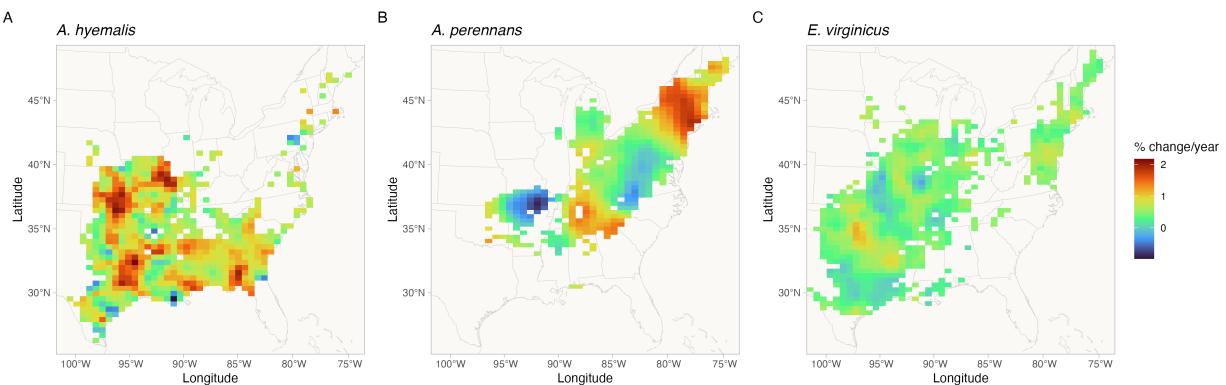


Figure 3: Predicted posterior mean of spatially-varying slopes representing change in endophyte prevalence for each host species. Color indicates the relative change in predicted endophyte prevalence.

340 *What is the relationship between variation in temporal trends in endophyte*
 341 *prevalence and changes in climate drivers?*

342 We found that trends in endophyte prevalence were strongly associated with seasonal climate
 343 change drivers (Fig. 4). For the majority of the study region, the climate has become wetter and

344 cooler over the last century (Fig. A12-A14), a consequence of regional variation in global climate
345 change (IPCC, 2021). Within the study region, spatial variation in climate trends were predic-
346 tive of trends in endophyte prevalence. For example, strong increases in prevalence within *A.*
347 *perennans* were most associated with autumn climate drivers that coincide with its Aug-Sep active
348 growing season. For this species, warmer and wetter autumn climates showed particularly strong
349 relationships, however other seasonal drivers may also contribute to increasing endophyte preva-
350 lence (drier springs and cooler summers). Trends in endophyte prevalence for *A. hyemalis* were
351 most strongly associated with changes in precipitation variability were the strongest predictors.
352 Endophyte prevalence increased the most in regions that experienced greater spring precipita-
353 tion along with increasing variability in summer and autumn precipitation. While this species
354 actively grows and reproduces in the late spring and early summer, climate effects outside of the
355 growing season may indicate that endophytes play a role in persistence during dormant periods
356 through summer droughts or contribute to the ability to successfully germinate. Prevalence of
357 endophytes of *E. virginicus* were least influenced by climate, but decreasing autumn temperature
358 variability and less precipitation in autumn were the strongest predictors.

359 Correlations assessed using all pixels across each species distribution were qualitatively sim-
360 ilar to these results (Fig. A11).

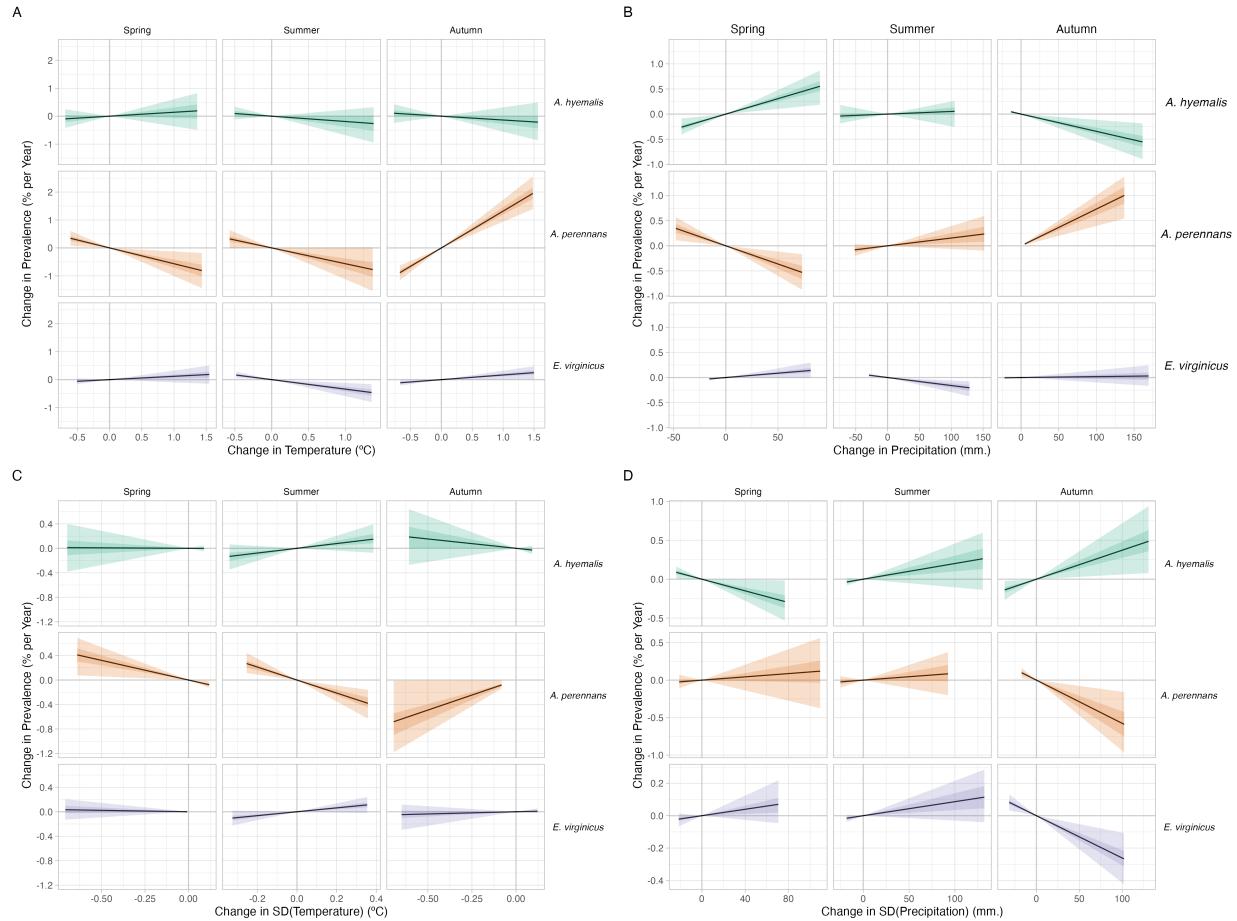


Figure 4: Relationships between changes in seasonal climate drivers and predicted marginal trends in endophyte prevalence. Lines show relationships between seasonal climate (A: mean temperature, B: cumulative precipitation, C: standard deviation in temperature, D: standard deviation in precipitation) and spatially-varying trends in endophyte prevalence for each host species, along with 50 and 95% CI.

361 *Performance on test data*

362 We found that model performance, as judged by AUC, was similar between historic herbarium
 363 specimens used as training data and the out-of-sample test data from contemporary surveys (0.79
 364 and 0.77 respectively; Fig. A5-A4). The model successfully captured broader regional trends
 365 in endophyte prevalence present in the contemporary survey data, such as decline endophyte

³⁶⁶ prevalence towards western longitudes in *A. hyemalis* (Fig. 5A). However, the contemporary
³⁶⁷ data contains additional variability at smaller scales not captured by our sampling of herbarium
³⁶⁸ specimens. We interpreted this to mean that the model captured regional spatial dynamics, but
³⁶⁹ underpredicts local scale dynamics. We discuss potential model improvements in the Discussion.

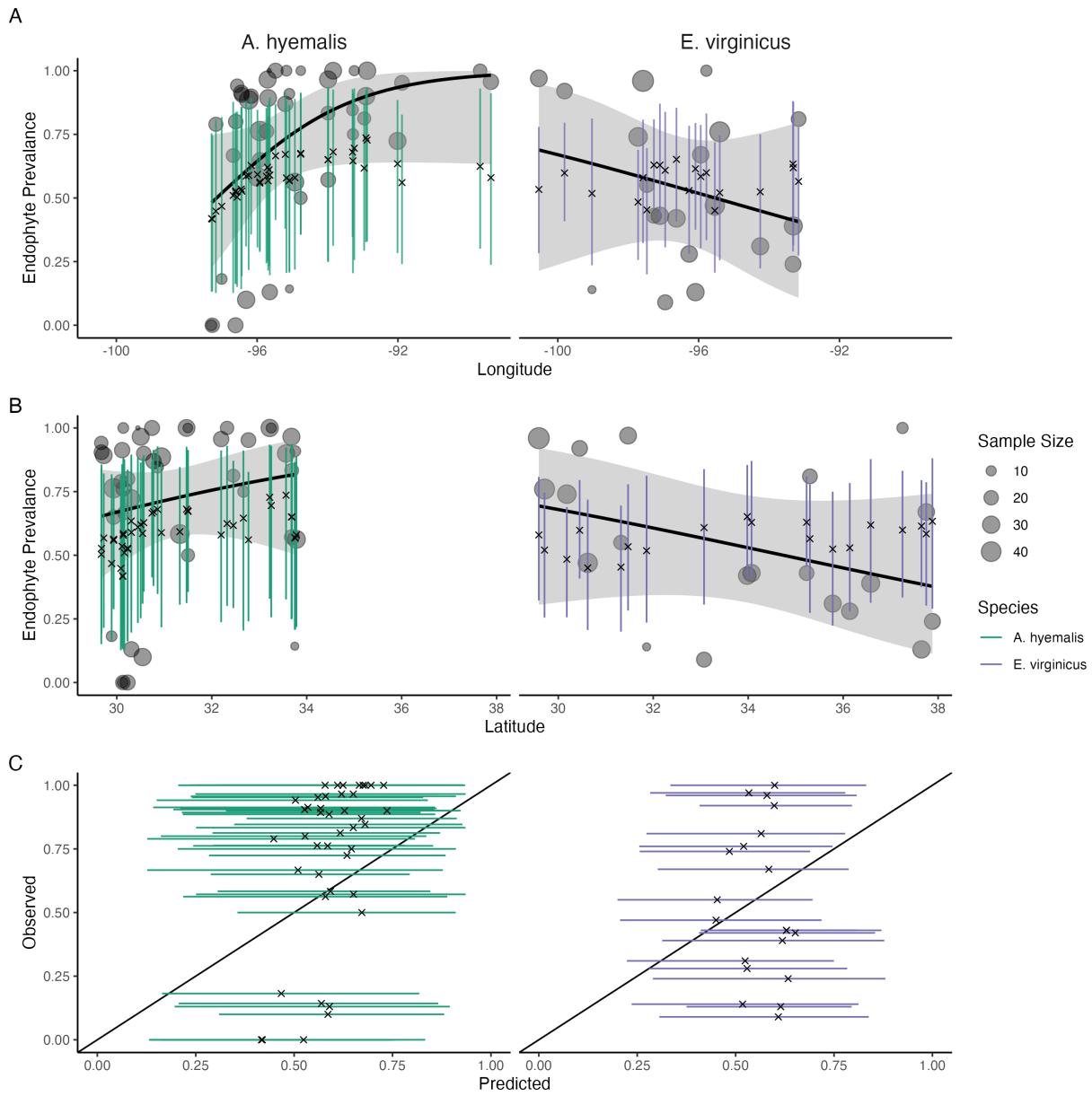


Figure 5: Predicted vs observed endophyte prevalence for contemporary test data. (A) The model, trained on historic herbarium collection data, performed modestly at predicting contemporary endophyte prevalence in *A. hyemalis*, as indicated by some overlap of predicted 95% CI with the 1:1 line, however contemporary test data generally had more variance between populations than model predictions. The model did recapitulate broader regional trends across (B) longitude and (C) latitude. Point size in panels B and C reflect sample sizes of contemporary endophyte population surveys.

370 *Assessing collector and scorer influences on predicted endophyte prevalence*

371 Our modeling effort quantified temporal and spatial trends in endophyte prevalence while ac-
372 counting for potential biases introduced by collectors and by individual scorers who quantified
373 endophyte presence/absence within specimens with the use of random effects. We found no
374 evidence that collector biases influenced our results. Collector random effects were consistently
375 small (Fig. A9), and models fit with and without this random effect provide qualitatively simi-
376 lar results. The identity of individual scorers did contribute to observed patterns in endophyte
377 prevalence. For example, 3 of the 25 scorers were more consistently likely than average to assign
378 positive endophyte status, as indicated by 95% credible intervals greater than zero) (Fig. A10).
379 This may have been driven by differences in scorers biases during the seed scoring process or
380 by unintended spatial clustering of the specimens scored by each scorer (Clayton et al., 1993;
381 Urdangarin et al., 2023). Interpreting our models with the inclusion of variance associated with
382 the scorer effect thus provides conservative estimates of the absolute magnitude of changes in
383 endophyte prevalence.

384 **Discussion**

385 Our examination of historic plant specimens revealed a cryptic biotic reponse to climate change.
386 For the three host species we examined, there have been clear increases in fungal endophyte
387 prevalence over the last two centuries. Increases in prevalence of *Epichloë*, which are vertically
388 transmitted, can potentially be interpreted as adaptive changes that improve the fitness of their
389 hosts under stressful conditions. This interpretation is in line with theory predicting that the
390 positive fitness feedback caused by vertical transmission leads beneficial symbionts to rise in
391 prevalence within a population (Fine, 1975). We found that trends in endophyte prevalence var-
392 ied across the distribution of each species in association with observed changes in climate drivers,
393 suggesting that the endophytes have contributed to host resilience under environmental change.
394 Taken together, this suggests a strengthening of the mutualism over the last two centuries.

395 Differences between the responses of each host species underscore that while all of these
396 C_3 grasses share similar broad-scale distributions, each engages in unique biotic interactions
397 and has unique niche requirements. We identified hotspots of change for *A. perennans*, which
398 experienced the strongest absolute changes in endophyte prevalence (Fig. 3). Declines in the
399 southern portion of its range and increases in the north suggest a potential poleward range shift
400 of endophytic plants. Based on previous work demonstrating that endophytes can shield their
401 hosts from drought stress (Decunta et al., 2021), we generally predicted that drought conditions
402 could be a driver of increasing endophyte prevalence. In contrast to this expectation, increasing
403 prevalence for this species was associated with increasing autumn temperature and precipitation
404 (Fig. 4). To our knowledge, the response of the symbiosis in *A. perennans* to drought has not been
405 examined experimentally, but in a greenhouse experiment, endophytes had a positive effect on
406 host reproduction under shaded, low-light conditions (Davitt et al., 2010). Our results also hint
407 that it may be useful to investigate whether lagged climate effects are important predictors of
408 host fitness in this system (Evers et al., 2021). Endophyte prevalence of the spring-flowering *A.*
409 *hyemalis* was most strongly linked to increasing variability in precipitation across summer and
410 autumn. Endophytes could be playing a role helping hosts weather autumn-season droughts
411 while the species is dormant. Previous work has demonstrated drought benefits in a greenhouse
412 manipulation with this species (Davitt et al., 2011), and early life stages may be particularly
413 vulnerable to prolonged droughts. For *E. virginicus*, which experienced the most modest changes
414 in endophyte prevalence overall, we only modest associations with changes in climate drivers.
415 Surveys by Sneck et al. (2017), used as part of the test data in this study, identified a drought index
416 (SPEI) that integrates precipitation with estimated evapotranspiration as an important predictor
417 of endophyte prevalence. *Epichloë* endophytes have also been connected to a suite of non-drought
418 related fitness benefits including herbivore protection (Brem and Leuchtmann, 2001), salinity
419 resistance (Wang et al., 2020), and mediation of the soil microbiome (Roberts and Ferraro, 2015)
420 These effects are potentially mediated by the diverse bioactive alkaloids and other signaling
421 compounds they produce (Saikonen et al., 2013). Increases in symbionts could be explained, at

422 least in part, by these diverse benefits that may help hosts weather a world made increasingly
423 stressful by changes in climate and other anthropogenically introduced stressors. While we show
424 consistent increasing trends in prevalence between the three species, the mechanisms that explain
425 these changes may be diverse and idiosyncratic.

426 Our spatially-explicit model predicted regions of both high and low endophyte prevalence,
427 suggesting that symbiotic and non-symbiotic host plants have overlapping, but non-identical
428 niche requirements. Endophytes fitness benefits potentially explain the spatial distribution of
429 prevalence by allowing their hosts to persist in environments where they otherwise could not
430 (Afkhami et al., 2014; Kazenel et al., 2015). For example, fitness benefits of the symbiosis could
431 explain historically low prevalence in *A. hyemalis* towards its western range edge coinciding with
432 a strong aridity gradient. Previous population surveys for endophytes, which were used as
433 test data for our model, found similar regional trends in prevalence for endophyte host species
434 (Rudgers and Swafford, 2009; Sneck et al., 2017). While the model recreated these large-scale
435 spatial trends, test data contained more population-to-population variability in prevalence. Val-
436 idating our model predictions in this way allows us to evaluate places to improve the model's
437 out-of-sample predictive ability, which will be particularly important for predicting host and
438 symbiont niche-shifts under future climate change. Lack of information on local variability may
439 simply be a feature of data derived from herbarium specimens. They are samples from local pop-
440 ulations, but they are single specimens that are aggregated over in broad-scale model estimates.
441 Poor predictive ability at local scales in this grass-endophyte system is not surprising, as previ-
442 ous studies have found that local variation, even to the scale of hundreds of meters can structure
443 endophyte-host niches (Kazenel et al., 2015). Other studies have found factors including land-
444 use history (Vikuk et al., 2019) and the biotic environment, including herbivory (Rudgers et al.,
445 2016), and host genotype Sneck et al. (2017), to be important predictors of endophyte ecology.
446 Incorporating available climatic and soil layers as covariates is an obvious first step that could
447 improve predictions. Another important step would be integrating data from local and regional
448 scales through modeling to constrain estimates of local and regional variation. These steps will

bridge gaps that often exist between large but broad bioclimatic and biodiversity data and small but local data on biotic interactions, and move towards the goal of predicting the dynamics of microbial symbioses under climate change (Isaac et al., 2020; Miller et al., 2019).

Our analysis advances the use of herbarium specimens in global change biology in two ways. First and foremost, this is the first study to link long-term changes in microbial symbioses to changes in climate using specimens from natural history collections. The responses of microbial symbioses are a rich target for future studies within museum specimens, particularly those that take advantage of advances in sequencing technology. While we used relatively coarse presence/absence data based on fungal morphology, other studies have examined historic plant microbiomes using molecular sequencing and sophisticated bioinformatics techniques, but these studies have so far been limited to relatively few specimens at limited spatial extents (Bieker et al., 2020; Bradshaw et al., 2021; Gross et al., 2021; Heberling and Burke, 2019; Yoshida et al., 2015). Continued advances in capturing historic DNA and in filtering out potential contamination during specimen storage (Bakker et al., 2020; Daru et al., 2019; Raxworthy and Smith, 2021) will be imperative in the effort to scale up these efforts. This scaling up will be essential to be able to quantify changes not just in the prevalence of symbionts, but also in symbionts' intraspecific variation and evolutionary responses to climate change, as well as in changes in the wider microbial community. Answering these questions as well as the unknown questions that future researchers may ask also reiterates the value in capturing meta-information during ongoing digitization efforts at herbaria around the world and during the accession of newly collected specimens (Edwards et al.; Lendemer et al., 2020). Second, we accounted for several potential biases in the data observation process that may be common to many collections-based research questions by using a spatially-explicit random effects model. Spatial autocorrelation (Willems et al., 2022), potential biases introduced by the sampling habits of collectors (Daru et al., 2018), and variation between contemporary researchers during the collection of trait data, if not corrected for could lead to over-confident inference about the strength and direction of historic change. Previous studies that have quantified the effects of collector biases typically find them

476 to be small (Davis et al., 2015; Meineke et al., 2019), and we similarly did not find that collector
477 has a strong effect on the results of our analysis.

478 Ultimately, a central goal of global change biology is to generate predictive insights into the
479 future of natural systems. While this survey of historic endophyte prevalence is necessarily cor-
480 relative, it serves as a foundation to develop better predictive models of the response of microbial
481 symbioses to climate change. Combining the insights from this type of regional-scale survey with
482 field experiments and physiological data could be invaluable. While we found that climate is
483 strongly correlated with endophytes' temporal responses, we do not know why trends in preva-
484 lence were weak in some areas or how endophytes would respond to more extreme changes in
485 climate. For example, transplanting symbiotic and non-symbiotic plants beyond the range edge
486 of *A. hyemalis* could tell us whether persistent low endophyte prevalence in that area is a result of
487 environmental conditions that lead the symbiosis to negative fitness consequences, or is a result
488 of some historical contingency or dispersal limitation that has thus far limited the presence of
489 symbiotic hosts from a region where they would otherwise flourish and provide resilience. While
490 we observed evidence of mutualism resilience, more extreme environmental changes than those
491 observed in our study could potentially push one or both partners beyond their physiological
492 limit, leading to the collapse of the mutualism. Our analysis thus far is agnostic to changes in the
493 distributions of hosts. Mechanistic models could connect the responses of both host and sym-
494 biots to abiotic climate drivers integrating dispersal processes. Beyond host-microbe symbioses,
495 building these types of models would work towards quantitatively attributing biotic responses
496 to anthropogenically driven climate change, similar to methods in climate science and economics
497 (Carleton and Hsiang, 2016; Stott et al., 2010).

498 **Acknowledgments**

499 We thank Dr. Jessica Budke for help in drafting our initial destructive sampling plan, and to the
500 many staff members of herbaria who facilitated our research visits, as well as to the hundreds

501 of collectors who contributed to the natural history collections. Several high schooler and un-
502 dergraduate researchers contributed to data collection, including A. Appio-Riley, P. Bilderback,
503 E. Chong, K. Dickens, L. Dufresne, B. Gutierrez, A. Johnson, S. Linder, E. Scales, B. Scherick,
504 K. Schrader, E. Segal , G. Singla, and M. Tucker. This research was supported by funding from
505 National Science Foundation (grants 1754468 and 2208857) and by funding from the Texas Ecolab
506 Program.

507 **Statement of Authorship**

508 J.C.F. contributed to research conception, data collection, data analysis, and led manuscript draft-
509 ing. J.M. contributed to data analysis and manuscript revisions. T.E.X.M. contributed to research
510 conception, data collection, data analysis, and manuscript revisions.

511 **Data and Code Availability**

512 Data from this publication will be made publically available upon acceptance and before that
513 upon request. Code for analyses can be found through: <https://github.com/joshuacfowler/EndoHerbarium>

Appendix A

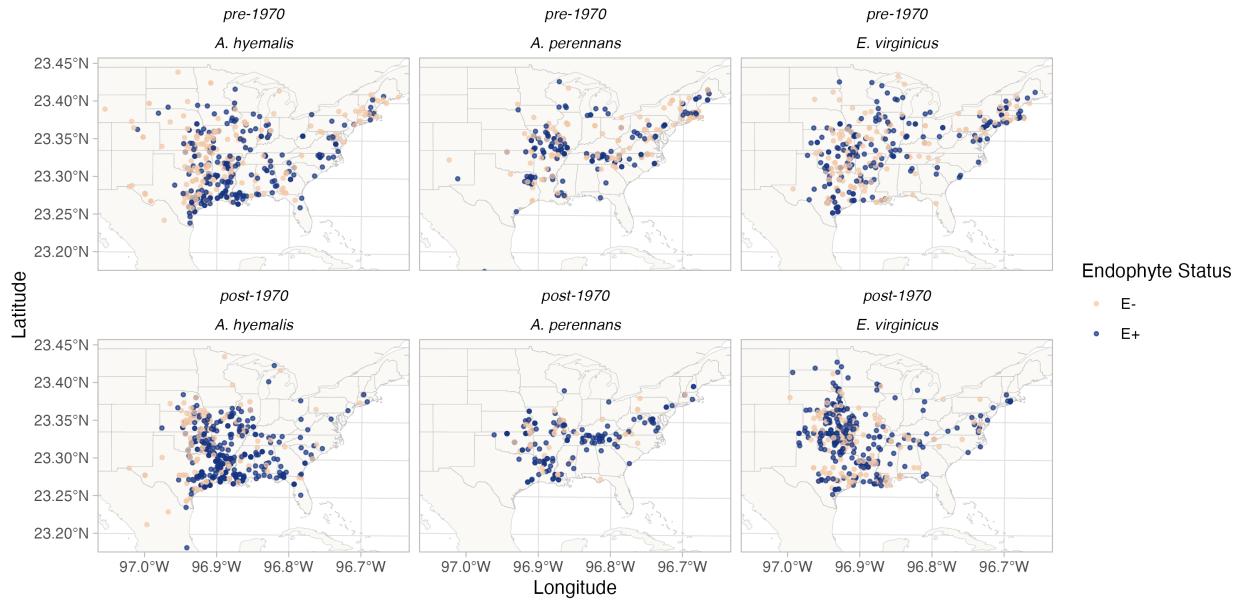


Figure A1: Endophyte presence/absence in specimens of each host species. Points show collection locations colored according to whether the specimen contained endophytes (E+; blue points) or did not contain endophytes (E-, tan points) and are faceted based on collection period.

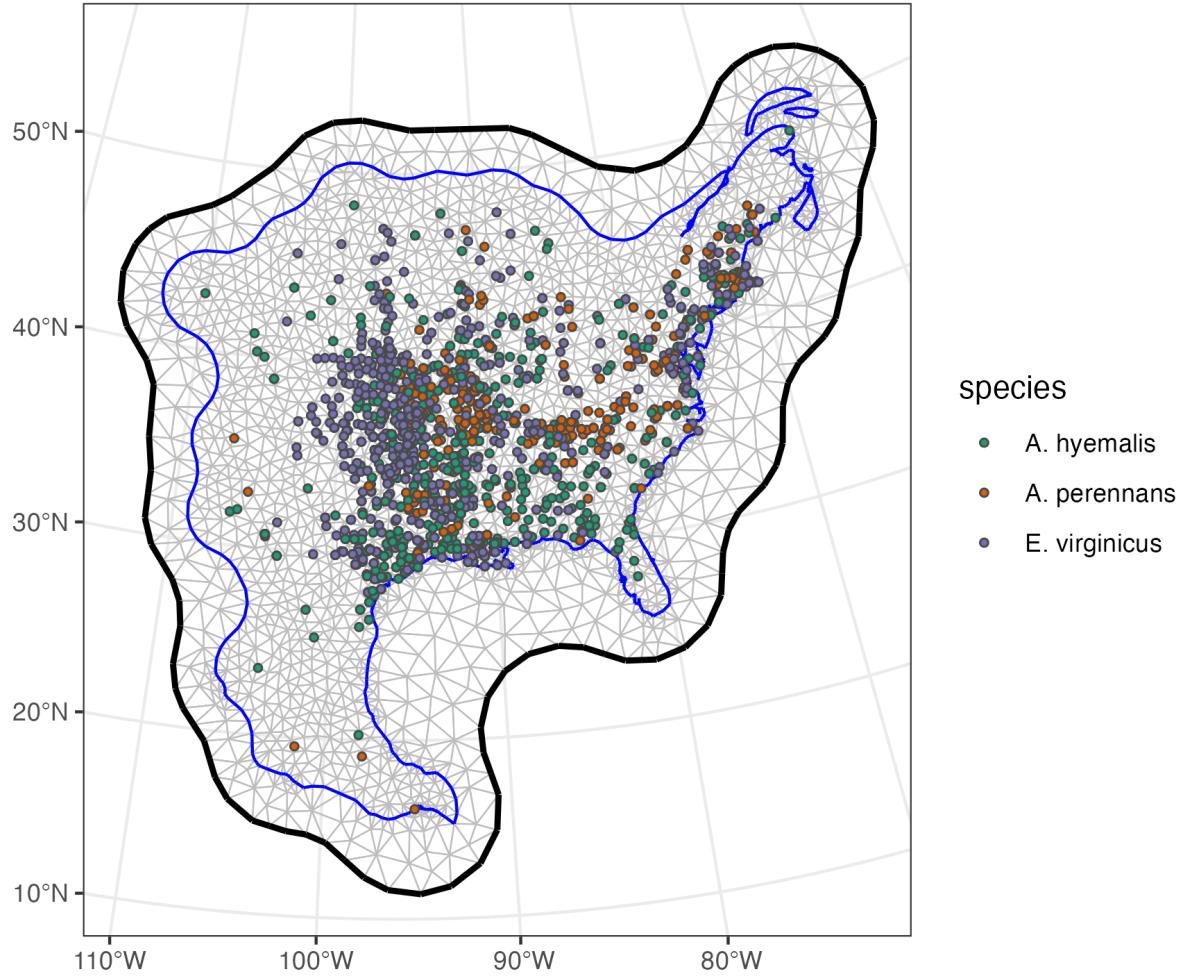


Figure A2: Triangulation mesh used to estimate spatial dependence between data points. Grey lines indicate edges of triangles used to define distances between observations. Colored points indicate locations of sampled herbarium specimens for each host species, and the blue line shows the convex hull and coastline used to define the edge of the mesh around the data points. The thick black line shows the convex hull defining a buffer space around the edge of the mesh to reduce the influence of edge effects on model estimates.

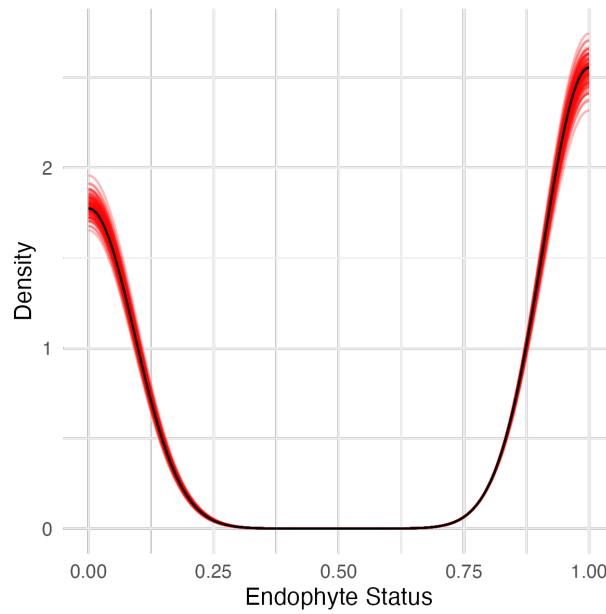


Figure A3: Consistency between real data and simulated values indicate that the fitted model accurately describes the data. Graph shows density curves for the observed data (black) along with 100 simulated datasets (red).

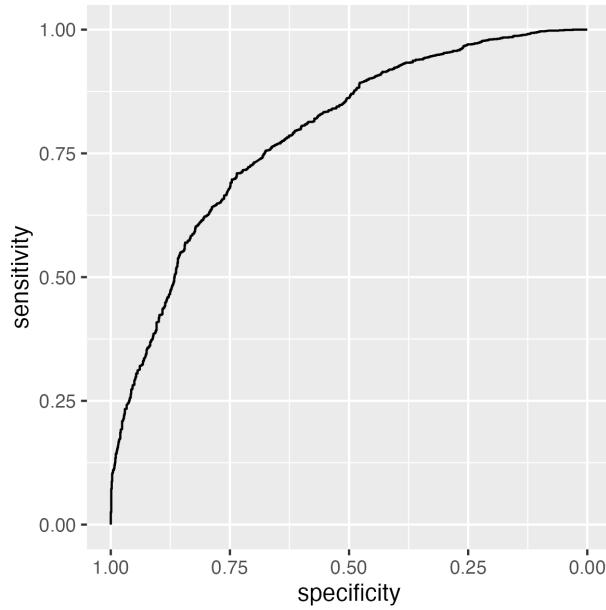


Figure A4: ROC plot showing model performance classifying observations according to endophyte status within the in-sample data. The curves show adequate model performance for observed data. The AUC value is 0.79.

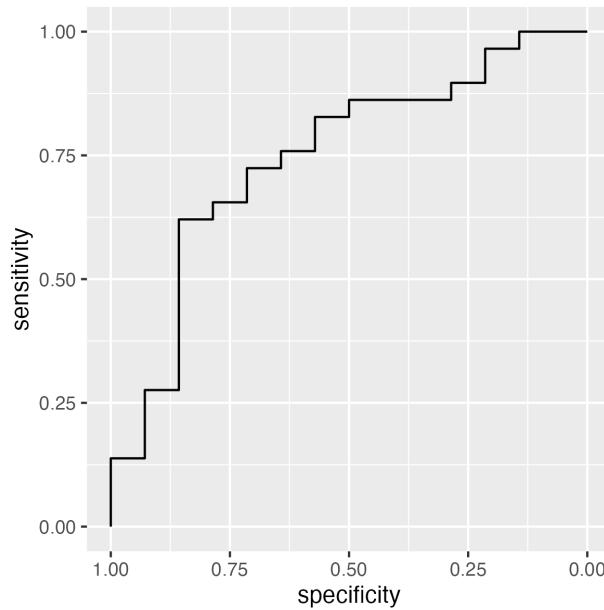


Figure A5: ROC plot showing model performance classifying observations according to endophyte status within the out-of-sample data. The curves show adequate model performance for test data. The AUC value is 0.77.

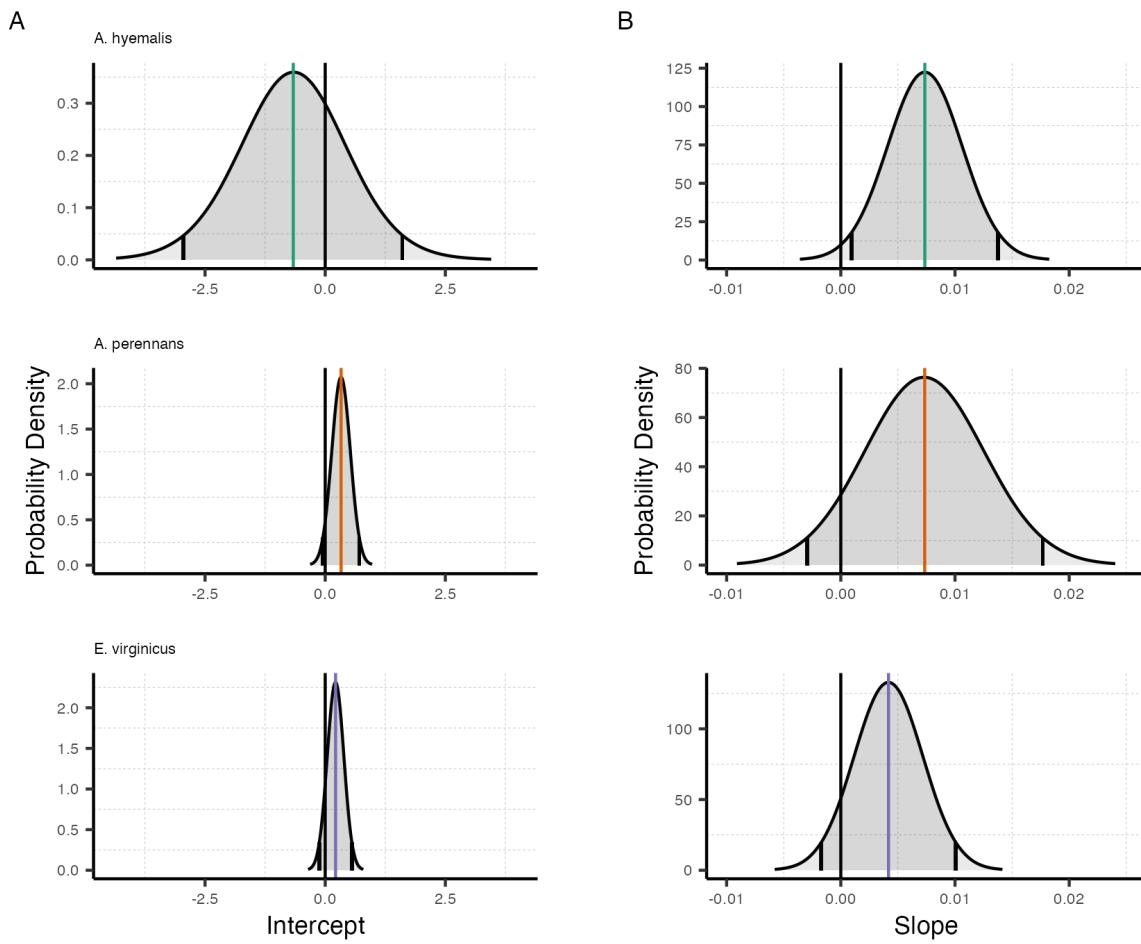


Figure A6: Density curves show the probability density along with mean (colored line) and 95% CI (black lines) for the (A) intercept and (B) slope terms, **A** and **T** respectively. Colors represent each host species

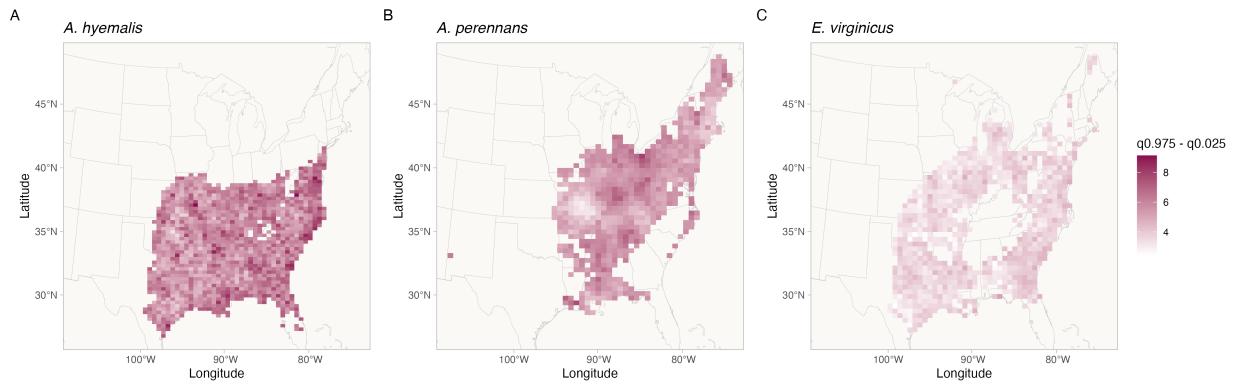


Figure A7: Shading represents the range of the 95% posterior credible interval for spatially varying slopes, τ .

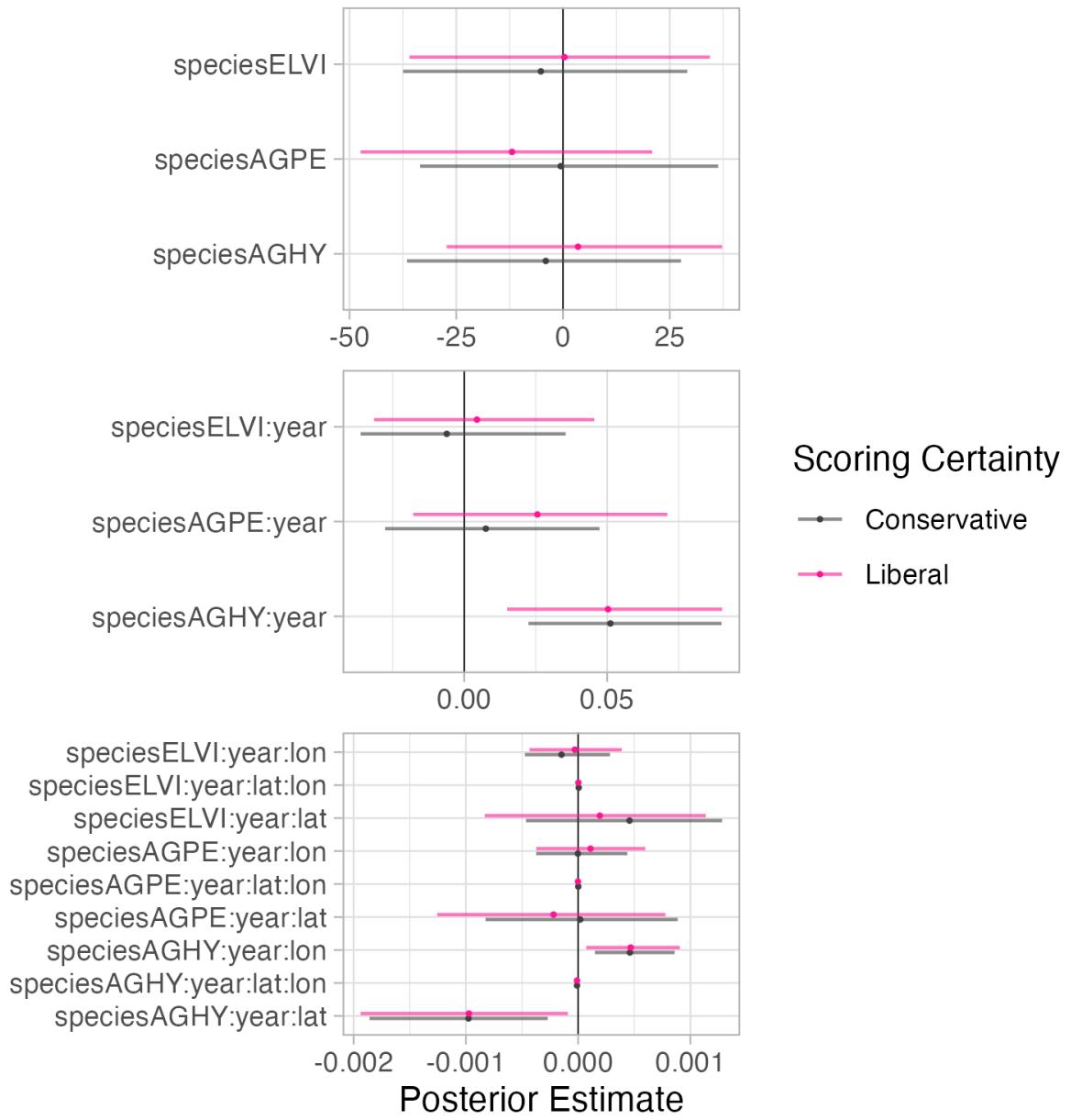


Figure A8: Comparison of posterior estimates of fixed effects when using Liberal or Conservative endophyte scores.

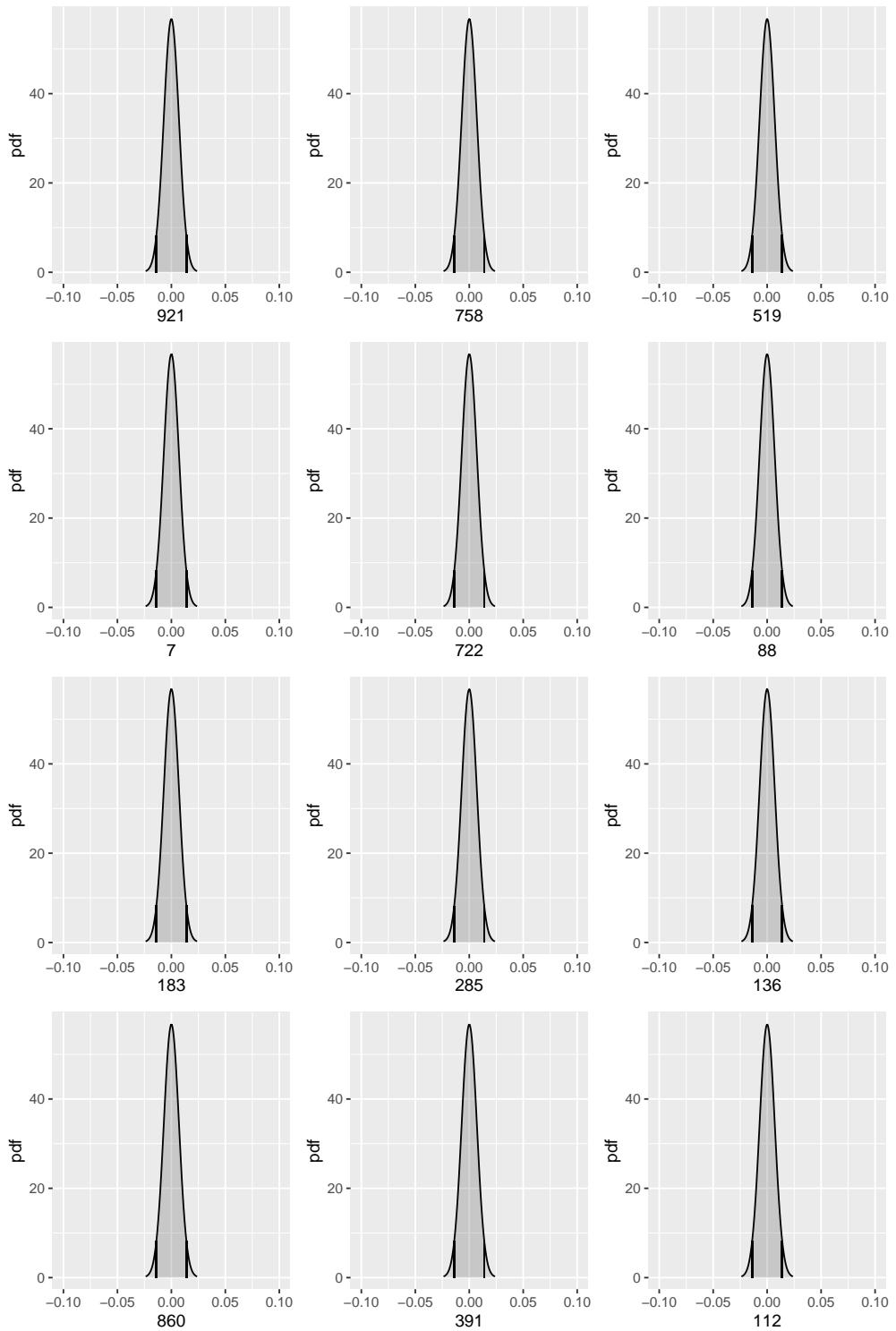


Figure A9: Posterior estimates of collector random effects. Density curves show the posterior estimate along lines indicating the 95% CI for 12 randomly selected collectors.

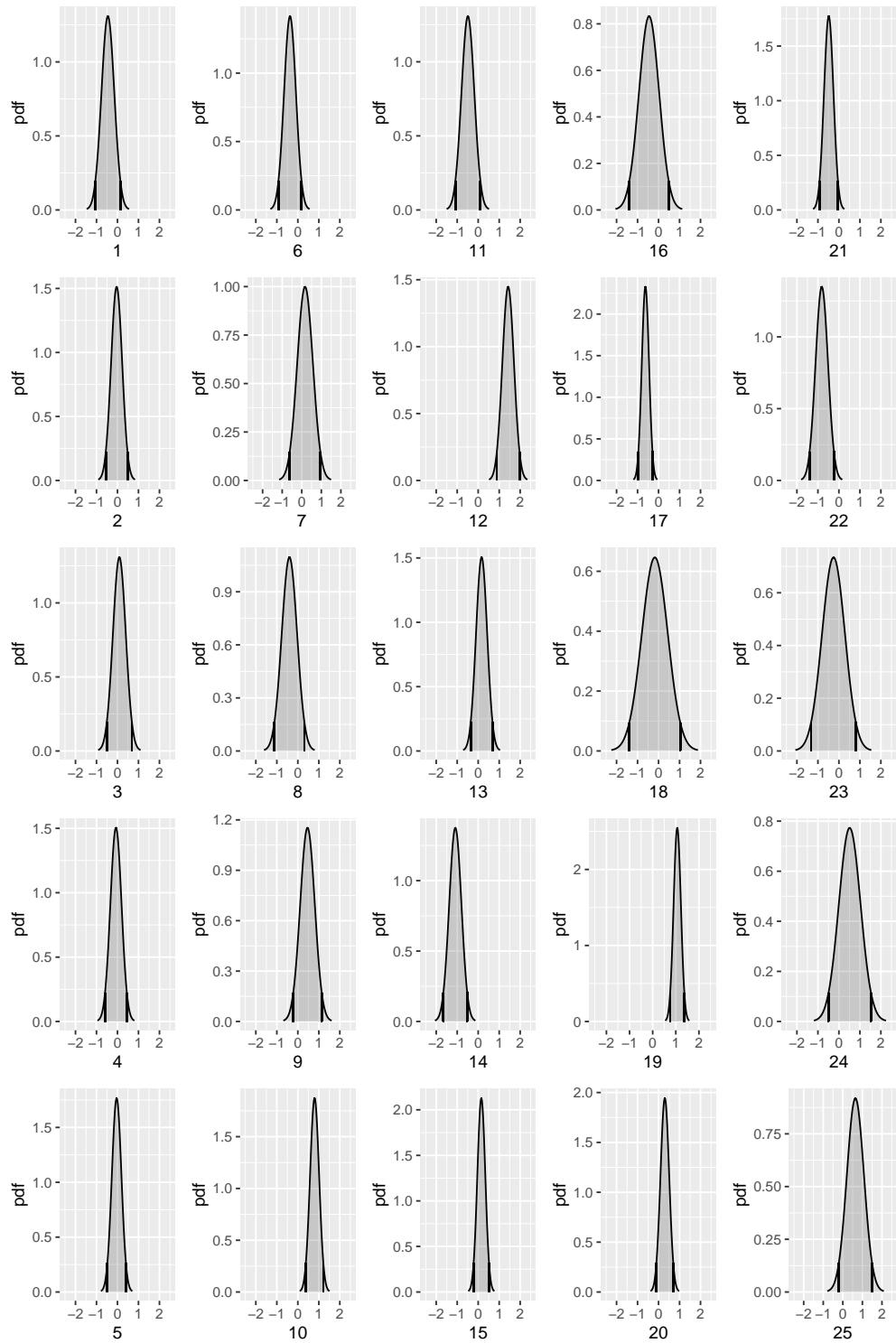


Figure A10: Posterior estimates of scorer random effects. Density curves show the posterior estimate along lines indicating the 95% CI for 25 scorers.

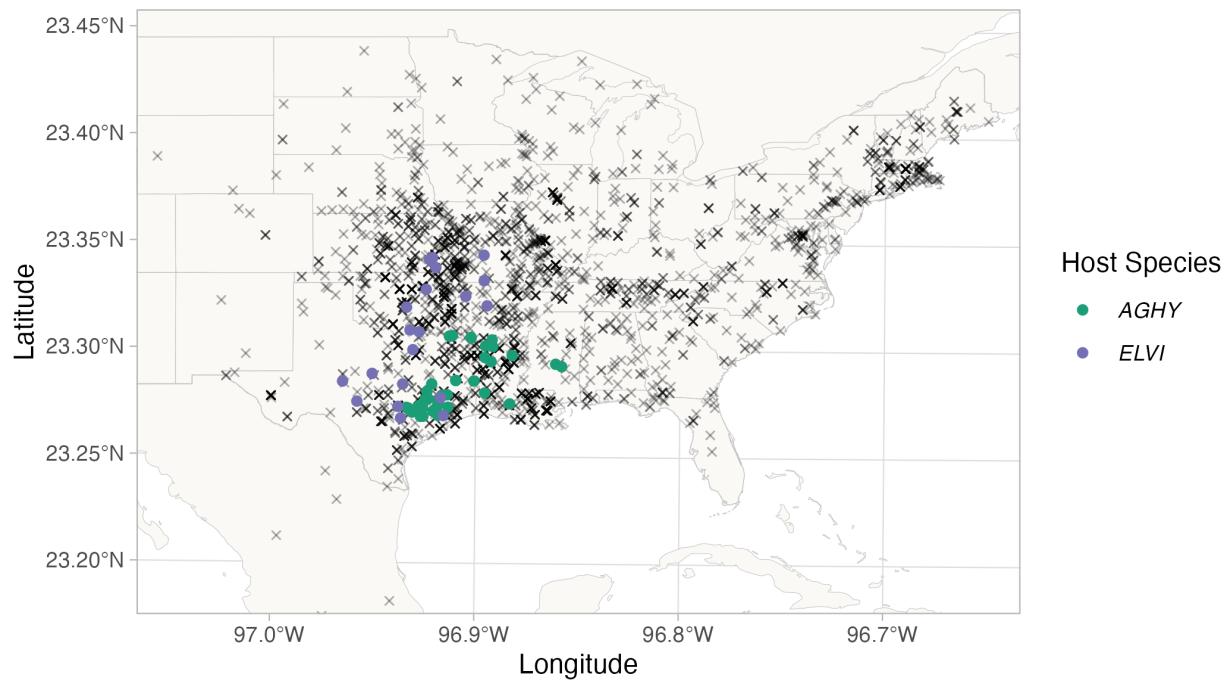


Figure A11: Locations of contemporary surveys of endophytes in *A. hyemalis* used as "test" data (red points), relative to the historical collection data (black crosses).

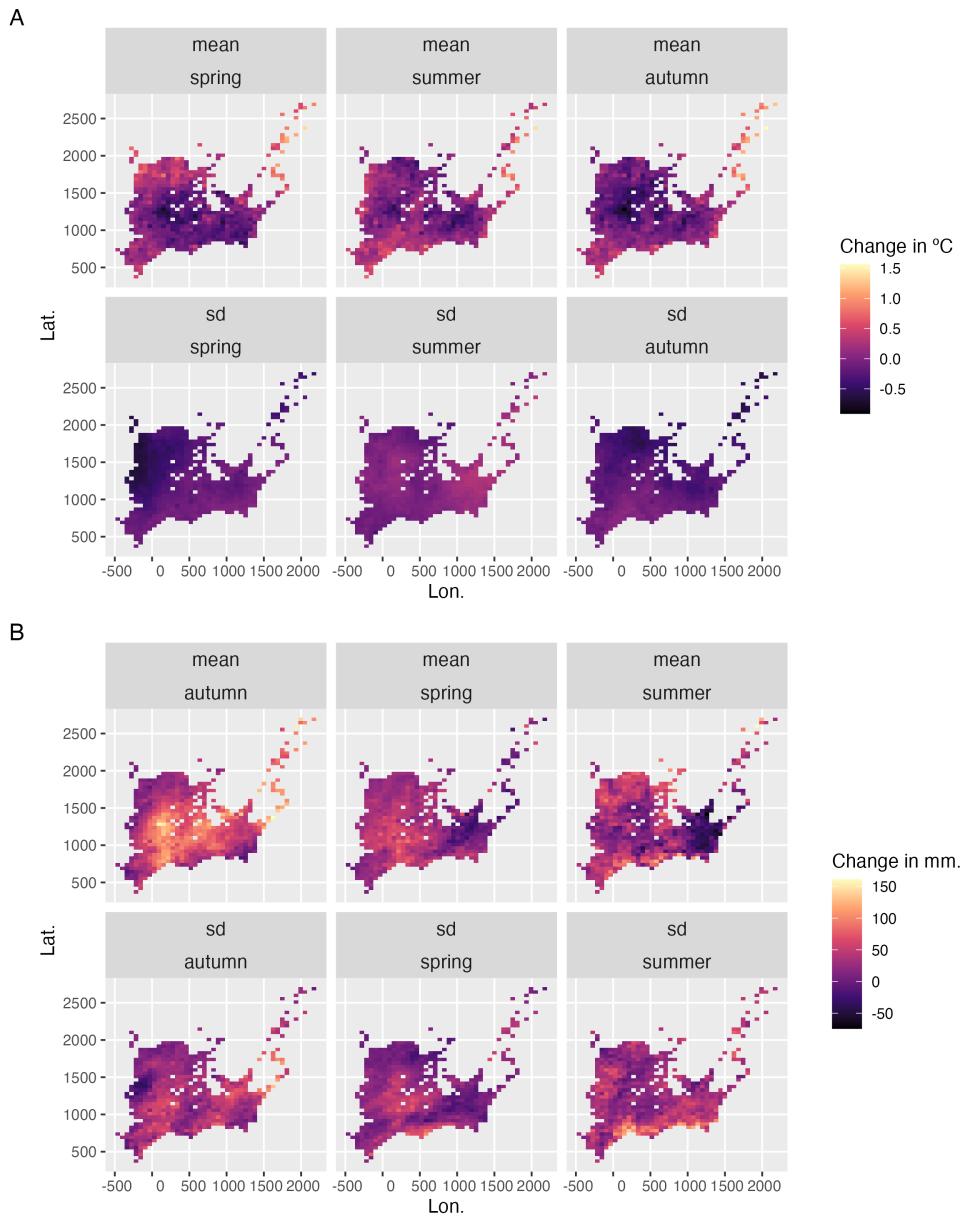


Figure A12: Change in seasonal climate variables between the periods 1895-1925 and 1990-2020. Color represents change in (A) seasonal temperature and (B) seasonal precipitation. Maps show pixels covering the modeled distribution of *A. hyemalis* used in post-hoc climate correlation analysis.

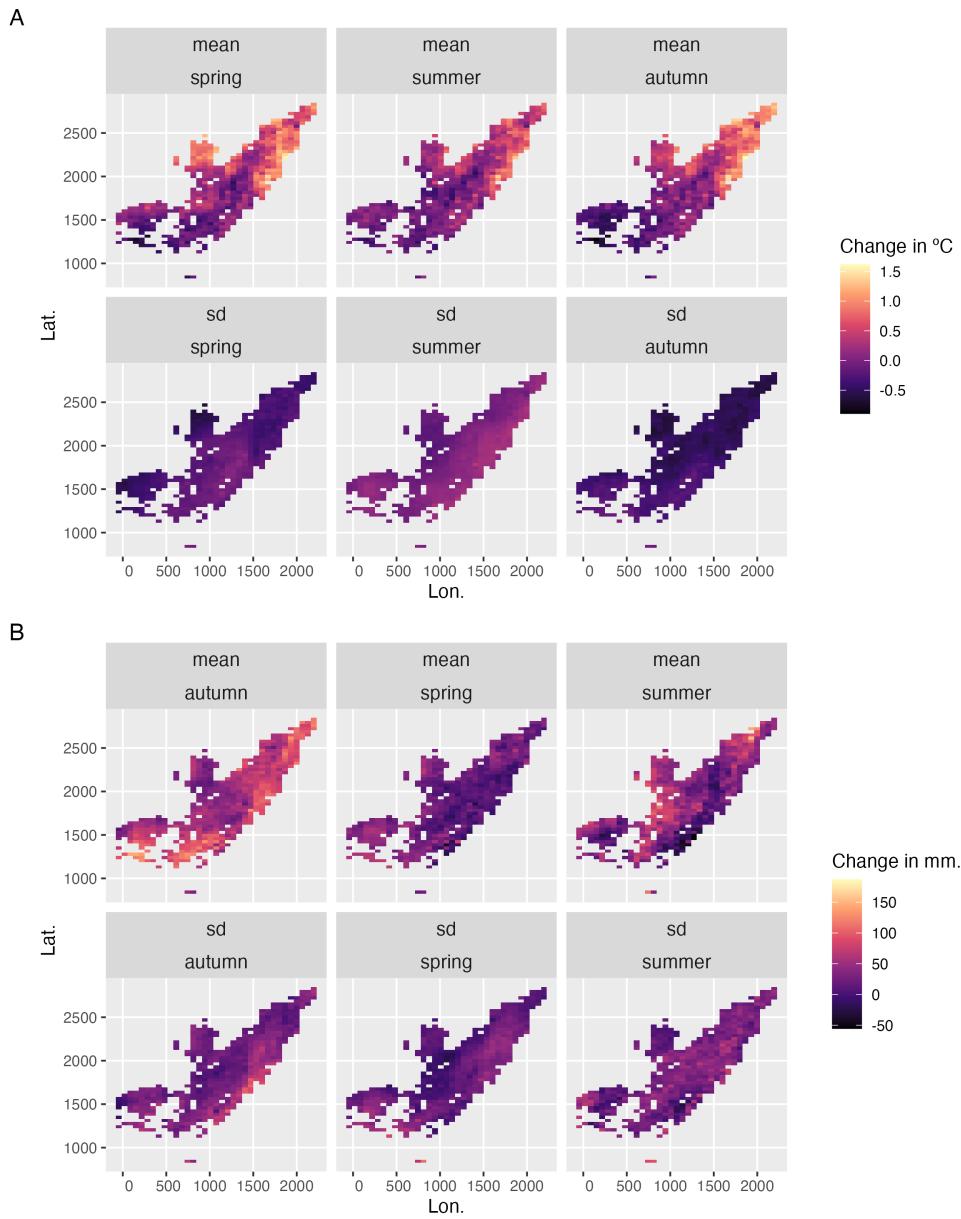


Figure A13: Change in seasonal climate variables between the periods 1895-1925 and 1990-2020.

Color represents change in (A) seasonal temperature and (B) seasonal precipitation. Maps show pixels covering the modeled distribution of *A. perennans* used in post-hoc climate correlation analysis.

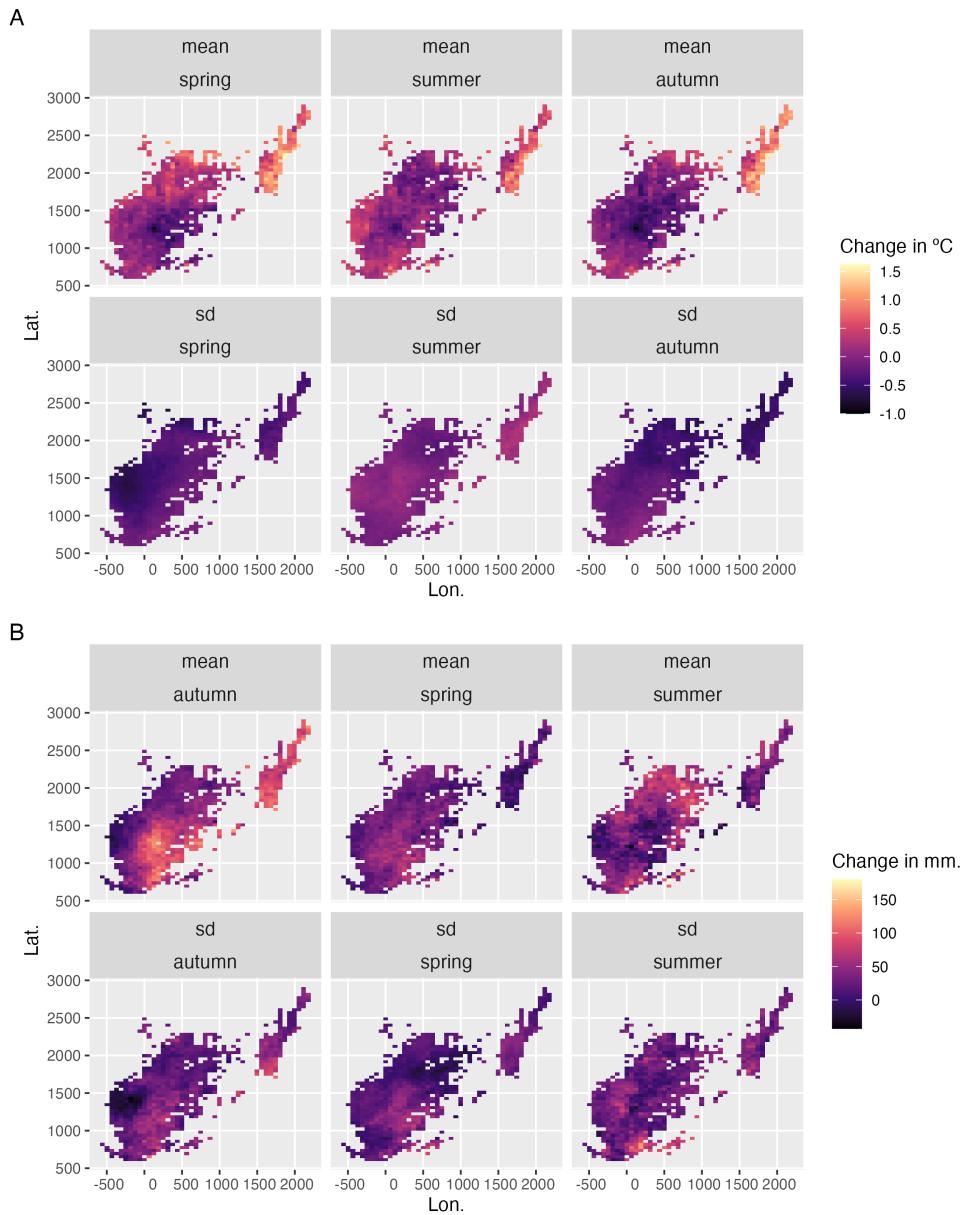


Figure A14: Change in seasonal climate variables between the periods 1895-1925 and 1990-2020. Color represents change in (A) seasonal temperature and (B) seasonal precipitation. Maps show pixels covering the modeled distribution of *E. virginicus* used in post-hoc climate correlation analysis.

Table A1: Summary of herbarium samples across collections

Herbarium Collection	AGHY	AGPE	ELVI
Botanical Research Institute of Texas	350	190	198
Louisiana State University	72	38	62
Mercer Botanic Garden	3	–	6
Missouri Botanic Garden	210	205	122
Texas A&M	100	–	72
University of Kansas	134	34	197
University of Oklahoma	85	34	95
University of Texas & Lundell	183	91	102
Oklahoma State University	51	10	74

515

Supporting Methods

516

ODMAP Protocol

517 **Overview**

518 **Model purpose:** Mapping current distribution of *Epichloë* host species.

519 **Target species:** *Agrostis hyemalis*, *Agrostis perennans*, and *Elymus virginicus*.

520 **Study area:** Eastern North America

521 **Spatial extent:** -125.0208, -66.47917, 24.0625, 49.9375 (xmin, xmax, ymin, ymax).

522 **Spatial resolution:** 0.04166667, 0.04166667 (x, y).

523 **Temporal extent:** 1990 to 2020.

524 **Boundary:** Natural.

525 **Data**

526 **Observation type:** Occurrence records from Global Biodiversity Information Facility and herbar-

527 ium collection across eastern North America. We used 713 occurrences records for *Agrostis hye-*

528 *malis*, 656 occurrence records for *Agrostis perennans* and 2338 for *Elymus virginicus*.

529 **Response data type:** occurrence record, presence-only.

530 **Coordinate reference system:** WGS84 coordinate reference system (EPSG:4326 code)

531 **Climatic data:** raster data extracted from PRISM

532 **Model**

533 **Model assumption:** We assumed that the target species are at equilibrium with their environment.

535 **Algorithms:** Maximum entropy (maxent)

536 **Workflow:** We described the workflow in the method section of the manuscript.

537 **Software:** All statistics were performed using Maxent 3.3.4 and R4.3.1 with packages terra, usdm, spThin and dismo.

539 **Code availability:** Available through this link: <https://github.com/joshuacfowler/EndoHerbarium>

540 **Data availability:** Will be available upon acceptance

541 **Assessment**

542 We used AUC to test model performance.

543 **Prediction**

544 We predicted the probability of presence of the host species as a binary maps (presence or absence)

Literature Cited

- 547 Michelle E Afkhami. Fungal endophyte–grass symbioses are rare in the California floristic
548 province and other regions with mediterranean-influenced climates. *Fungal ecology*, 5(3):345–
549 352, 2012.
- 550 Michelle E Afkhami and Jennifer A Rudgers. Symbiosis lost: imperfect vertical transmission of
551 fungal endophytes in grasses. *The American Naturalist*, 172(3):405–416, 2008.
- 552 Michelle E Afkhami, Patrick J McIntyre, and Sharon Y Strauss. Mutualist-mediated effects on
553 species' range limits across large geographic scales. *Ecology letters*, 17(10):1265–1273, 2014.
- 554 Sally N Aitken, Sam Yeaman, Jason A Holliday, Tongli Wang, and Sierra Curtis-McLane. Adap-
555 tation, migration or extirpation: climate change outcomes for tree populations. *Evolutionary*
556 *applications*, 1(1):95–111, 2008.
- 557 Clare E Aslan, Erika S Zavaleta, Bernie Tershy, and Donald Croll. Mutualism disruption threatens
558 global plant biodiversity: a systematic review. *PLoS one*, 8(6):e66993, 2013.
- 559 Fabian E Bachl, Finn Lindgren, David L Borchers, and Janine B Illian. inlabru: an R package for
560 bayesian spatial modelling from ecological survey data. *Methods in Ecology and Evolution*, 10(6):
561 760–766, 2019.
- 562 Charles W Bacon and James F White. Stains, media, and procedures for analyzing endophytes.
563 In *Biotechnology of endophytic fungi of grasses*, pages 47–56. CRC Press, 2018.
- 564 Haakon Bakka, Håvard Rue, Geir-Arne Fuglstad, Andrea Riebler, David Bolin, Janine Illian, Elias
565 Krainski, Daniel Simpson, and Finn Lindgren. Spatial modeling with r-inla: A review. *Wiley*
566 *Interdisciplinary Reviews: Computational Statistics*, 10(6):e1443, 2018.
- 567 Freek T Bakker, Vanessa C Bieker, and Michael D Martin. Herbarium collection-based plant
568 evolutionary genetics and genomics, 2020.

- 569 Dawn R Bazely, John P Ball, Mark Vicari, Andrew J Tanentzap, Myrtille Bérenger, Tomo Rakoc
570 cevic, and Saewan Koh. Broad-scale geographic patterns in the distribution of vertically-
571 transmitted, asexual endophytes in four naturally-occurring grasses in sweden. *Ecography*,
572 30(3):367–374, 2007.
- 573 Julien Beguin, Sara Martino, Håvard Rue, and Steven G Cumming. Hierarchical analysis of
574 spatially autocorrelated ecological data using integrated nested laplace approximation. *Methods*
575 in *Ecology and Evolution*, 3(5):921–929, 2012.
- 576 Colette S Berg, Jason L Brown, and Jennifer J Weber. An examination of climate-driven flowering-
577 time shifts at large spatial scales over 153 years in a common weedy annual. *American Journal*
578 of *Botany*, 106(11):1435–1443, 2019.
- 579 Vanessa C Bieker, Fátima Sánchez Barreiro, Jacob A Rasmussen, Marie Brunier, Nathan Wales,
580 and Michael D Martin. Metagenomic analysis of historical herbarium specimens reveals a
581 postmortem microbial community. *Molecular ecology resources*, 20(5):1206–1219, 2020.
- 582 Jessica L Blois, Phoebe L Zarnetske, Matthew C Fitzpatrick, and Seth Finnegan. Climate change
583 and the past, present, and future of biotic interactions. *Science*, 341(6145):499–504, 2013.
- 584 Michael Bradshaw, Uwe Braun, Marianne Elliott, Julia Kruse, Shu-Yan Liu, Guanxiu Guan, and
585 Patrick Tobin. A global genetic analysis of herbarium specimens reveals the invasion dynamics
586 of an introduced plant pathogen. *Fungal Biology*, 125(8):585–595, 2021.
- 587 D Brem and A Leuchtmann. Epichloë grass endophytes increase herbivore resistance in the
588 woodland grass *brachypodium sylvaticum*. *Oecologia*, 126(4):522–530, 2001.
- 589 Tamara A Carleton and Solomon M Hsiang. Social and economic impacts of climate. *Science*, 353
590 (6304):aad9837, 2016.
- 591 Shen Cheng, Ying-Ning Zou, Kamil Kuča, Abeer Hashem, Elsayed Fathi Abd_Allah, and Qiang-

- 592 Sheng Wu. Elucidating the mechanisms underlying enhanced drought tolerance in plants
593 mediated by arbuscular mycorrhizal fungi. *Frontiers in Microbiology*, 12:4029, 2021.
- 594 Keith Clay and Christopher Schardl. Evolutionary origins and ecological consequences of endo-
595 phyte symbiosis with grasses. *the american naturalist*, 160(S4):S99–S127, 2002.
- 596 David G Clayton, Luisa Bernardinelli, and Cristina Montomoli. Spatial correlation in ecological
597 analysis. *International journal of epidemiology*, 22(6):1193–1202, 1993.
- 598 KD Craven, PTW Hsiau, A Leuchtmann, W Hollin, and CL Schardl. Multigene phylogeny of
599 epichloë species, fungal symbionts of grasses. *Annals of the Missouri Botanical Garden*, pages
600 14–34, 2001.
- 601 Kerri M Crawford, John M Land, and Jennifer A Rudgers. Fungal endophytes of native grasses
602 decrease insect herbivore preference and performance. *Oecologia*, 164:431–444, 2010.
- 603 Michael S Crossley, Timothy D Meehan, Matthew D Moran, Jeffrey Glassberg, William E Snyder,
604 and Andrew K Davis. Opposing global change drivers counterbalance trends in breeding north
605 american monarch butterflies. *Global change biology*, 28(15):4726–4735, 2022.
- 606 Christopher Daly and Kirk Bryant. The prism climate and weather system—an introduction.
607 *Corvallis, OR: PRISM climate group*, 2, 2013.
- 608 Barnabas H Daru, Daniel S Park, Richard B Primack, Charles G Willis, David S Barrington,
609 Timothy JS Whitfeld, Tristram G Seidler, Patrick W Sweeney, David R Foster, Aaron M Ellison,
610 et al. Widespread sampling biases in herbaria revealed from large-scale digitization. *New
611 Phytologist*, 217(2):939–955, 2018.
- 612 Barnabas H Daru, Elizabeth A Bowman, Donald H Pfister, and A Elizabeth Arnold. A novel proof
613 of concept for capturing the diversity of endophytic fungi preserved in herbarium specimens.
614 *Philosophical Transactions of the Royal Society B*, 374(1763):20170395, 2019.

- 615 Charles C Davis, Charles G Willis, Bryan Connolly, Courtland Kelly, and Aaron M Ellison.
616 Herbarium records are reliable sources of phenological change driven by climate and pro-
617 vide novel insights into species' phenological cueing mechanisms. *American journal of botany*,
618 102(10):1599–1609, 2015.
- 619 Andrew J Davitt, Marcus Stansberry, and Jennifer A Rudgers. Do the costs and benefits of fungal
620 endophyte symbiosis vary with light availability? *New Phytologist*, 188(3):824–834, 2010.
- 621 Andrew J Davitt, Chris Chen, and Jennifer A Rudgers. Understanding context-dependency in
622 plant–microbe symbiosis: the influence of abiotic and biotic contexts on host fitness and the
623 rate of symbiont transmission. *Environmental and Experimental Botany*, 71(2):137–145, 2011.
- 624 Facundo A Decunta, Luis I Pérez, Dariusz P Malinowski, Marco A Molina-Montenegro, and
625 Pedro E Gundel. A systematic review on the effects of epichloë fungal endophytes on drought
626 tolerance in cool-season grasses. *Frontiers in plant science*, 12:644731, 2021.
- 627 Mauro Di Luzio, Gregory L Johnson, Christopher Daly, Jon K Eischeid, and Jeffrey G Arnold.
628 Constructing retrospective gridded daily precipitation and temperature datasets for the con-
629 terminous united states. *Journal of Applied Meteorology and Climatology*, 47(2):475–497, 2008.
- 630 Marion L Donald, Teresa F Bohner, Kory M Kolis, R Alan Shadow, Jennifer A Rudgers, and
631 Tom EX Miller. Context-dependent variability in the population prevalence and individual
632 fitness effects of plant–fungal symbiosis. *Journal of Ecology*, 109(2):847–859, 2021.
- 633 AE Douglas. Host benefit and the evolution of specialization in symbiosis. *Heredity*, 81(6):599–
634 603, 1998.
- 635 Yuan-Wen Duan, Haibao Ren, Tao Li, Lin-Lin Wang, Zhi-Qiang Zhang, Yan-Li Tu, and Yong-Ping
636 Yang. A century of pollination success revealed by herbarium specimens of seed pods. *New
637 Phytologist*, 224(4):1512–1517, 2019.

- 638 Erika J Edwards, Brent D Mishler, and Charles D Davis. University herbaria are uniquely impor-
639 tant. *Trends in Plant Science*.
- 640 Markus Engel, Tobias Mette, and Wolfgang Falk. Spatial species distribution models: Using
641 bayes inference with inla and spde to improve the tree species choice for important european
642 tree species. *Forest Ecology and Management*, 507:119983, 2022.
- 643 Sanne M Evers, Tiffany M Knight, David W Inouye, Tom EX Miller, Roberto Salguero-Gómez,
644 Amy M Iler, and Aldo Compagnoni. Lagged and dormant season climate better predict plant
645 vital rates than climate during the growing season. *Global Change Biology*, 27(9):1927–1941,
646 2021.
- 647 Paul EM Fine. Vectors and vertical transmission: an epidemiologic perspective. *Annals of the New
648 York Academy of Sciences*, 266(1):173–194, 1975.
- 649 Joshua C Fowler, Shaun Ziegler, Kenneth D Whitney, Jennifer A Rudgers, and Tom EX Miller.
650 Microbial symbionts buffer hosts from the demographic costs of environmental stochasticity.
651 *Ecology Letters*, 27(5):e14438, 2024.
- 652 PR Fraude, F De Jongh, F Vermeulen, J Van Bleijswijk, and RPM Bak. Variation in symbiont
653 distribution between closely related coral species over large depth ranges. *Molecular Ecology*,
654 17(2):691–703, 2008.
- 655 Sarah E Gilman, Mark C Urban, Joshua Tewksbury, George W Gilchrist, and Robert D Holt. A
656 framework for community interactions under climate change. *Trends in ecology & evolution*, 25
657 (6):325–331, 2010.
- 658 Gustaf Granath, Mark Vicari, Dawn R Bazely, John P Ball, Adriana Puentes, and Tomo Rakoce-
659 vic. Variation in the abundance of fungal endophytes in fescue grasses along altitudinal and
660 grazing gradients. *Ecography*, 30(3):422–430, 2007.
- 661 Andrin Gross, Célia Petitcollin, Cyril Dutech, Bayo Ly, Marie Massot, Julie Faivre d'Arcier, Laure

- 662 Dubois, Gilles Saint-Jean, and Marie-Laure Desprez-Loustau. Hidden invasion and niche con-
663 traction revealed by herbaria specimens in the fungal complex causing oak powdery mildew
664 in europe. *Biological Invasions*, 23:885–901, 2021.
- 665 Edmund M. Hart and Kendon Bell. prism: Download data from the oregon prism project. 2015.
666 doi: 10.5281/zenodo.33663. URL <https://github.com/ropensci/prism>. R package version
667 0.0.6.
- 668 J Mason Heberling and David J Burke. Utilizing herbarium specimens to quantify historical
669 mycorrhizal communities. *Applications in plant sciences*, 7(4):e01223, 2019.
- 670 Robert J Hijmans, Steven Phillips, John Leathwick, Jane Elith, and Maintainer Robert J Hijmans.
671 Package ‘dismo’. *Circles*, 9(1):1–68, 2017.
- 672 Janneke HilleRisLambers, Melanie A Harsch, Ailene K Ettinger, Kevin R Ford, and Elinore J
673 Theobald. How will biotic interactions influence climate change-induced range shifts? *Annals*
674 *of the New York Academy of Sciences*, 1297(1):112–125, 2013.
- 675 IPCC. Climate change 2021: The physical science basis, 2021. URL
676 <https://www.ipcc.ch/report/ar6/wg1/>.
- 677 Nick JB Isaac, Marta A Jarzyna, Petr Keil, Lea I Dambly, Philipp H Boersch-Supan, Ella Browning,
678 Stephen N Freeman, Nick Golding, Gurutzeta Guillera-Arroita, Peter A Henrys, et al. Data
679 integration for large-scale models of species distributions. *Trends in ecology & evolution*, 35(1):
680 56–67, 2020.
- 681 Alberto Jiménez-Valverde. Insights into the area under the receiver operating characteristic curve
682 (auc) as a discrimination measure in species distribution modelling. *Global Ecology and Biogeog-*
683 *raphy*, 21(4):498–507, 2012.
- 684 David Kahle, Hadley Wickham, and Maintainer David Kahle. Package ‘ggmap’. *Retrieved Septem-*
685 *ber*, 5:2021, 2019.

- 686 Melanie R Kazenel, Catherine L Debban, Luciana Ranelli, Will Q Hendricks, Y Anny Chung,
687 Thomas H Pendergast IV, Nikki D Charlton, Carolyn A Young, and Jennifer A Rudgers. A
688 mutualistic endophyte alters the niche dimensions of its host plant. *AoB plants*, 7:plv005, 2015.
- 689 Roland A Knapp, Gary M Fellers, Patrick M Kleeman, David AW Miller, Vance T Vredenburg,
690 Erica Bree Rosenblum, and Cheryl J Briggs. Large-scale recovery of an endangered amphibian
691 despite ongoing exposure to multiple stressors. *Proceedings of the National Academy of Sciences*,
692 113(42):11889–11894, 2016.
- 693 Mikhail V Kozlov, Irina V Sokolova, Vitali Zverev, Alexander A Egorov, Mikhail Y Goncharov,
694 and Elena L Zvereva. Biases in estimation of insect herbivory from herbarium specimens.
695 *Scientific Reports*, 10(1):12298, 2020.
- 696 Benjamin R Lee, Evelyn F Alecrim, Tara K Miller, Jessica RK Forrest, J Mason Heberling,
697 Richard B Primack, and Risa D Sargent. Phenological mismatch between trees and wildflow-
698 ers: Reconciling divergent findings in two recent analyses. *Journal of Ecology*, 112(6):1184–1199,
699 2024.
- 700 James Lendemer, Barbara Thiers, Anna K Monfils, Jennifer Zaspel, Elizabeth R Ellwood, Andrew
701 Bentley, Katherine LeVan, John Bates, David Jennings, Dori Contreras, et al. The extended
702 specimen network: A strategy to enhance us biodiversity collections, promote research and
703 education. *BioScience*, 70(1):23–30, 2020.
- 704 A Leuchtmann. Systematics, distribution, and host specificity of grass endophytes. *Natural Toxins*,
705 1(3):150–162, 1992.
- 706 Adrian Leuchtmann, Charles W Bacon, Christopher L Schardl, James F White Jr, and Mariusz
707 Tadych. Nomenclatural realignment of neotyphodium species with genus epichloë. *Mycologia*,
708 106(2):202–215, 2014.
- 709 Finn Lindgren, Håvard Rue, and Johan Lindström. An explicit link between gaussian fields and

- 710 gaussian markov random fields: the stochastic partial differential equation approach. *Journal*
711 *of the Royal Statistical Society: Series B (Statistical Methodology)*, 73(4):423–498, 2011.
- 712 Canran Liu, Pam M Berry, Terence P Dawson, and Richard G Pearson. Selecting thresholds of
713 occurrence in the prediction of species distributions. *Ecography*, 28(3):385–393, 2005.
- 714 Margaret McFall-Ngai, Michael G Hadfield, Thomas CG Bosch, Hannah V Carey, Tomislav
715 Domazet-Lošo, Angela E Douglas, Nicole Dubilier, Gerard Eberl, Tadashi Fukami, Scott F
716 Gilbert, et al. Animals in a bacterial world, a new imperative for the life sciences. *Proceedings*
717 *of the National Academy of Sciences*, 110(9):3229–3236, 2013.
- 718 Timothy D Meehan, Nicole L Michel, and Håvard Rue. Spatial modeling of audubon christmas
719 bird counts reveals fine-scale patterns and drivers of relative abundance trends. *Ecosphere*, 10
720 (4):e02707, 2019.
- 721 Emily K Meineke, Charles C Davis, and T Jonathan Davies. The unrealized potential of herbaria
722 for global change biology. *Ecological Monographs*, 88(4):505–525, 2018.
- 723 Emily K Meineke, Aimée T Classen, Nathan J Sanders, and T Jonathan Davies. Herbarium
724 specimens reveal increasing herbivory over the past century. *Journal of Ecology*, 107(1):105–117,
725 2019.
- 726 Abigail R Meyer, Maria Valentin, Laima Liulevicius, Tami R McDonald, Matthew P Nelsen, Jean
727 Pengra, Robert J Smith, and Daniel Stanton. Climate warming causes photobiont degradation
728 and c starvation in a boreal climate sentinel lichen. *American Journal of Botany*, 2022.
- 729 David AW Miller, Krishna Pacifici, Jamie S Sanderlin, and Brian J Reich. The recent past and
730 promising future for data integration methods to estimate species' distributions. *Methods in*
731 *Ecology and Evolution*, 10(1):22–37, 2019.
- 732 Daniel S Park, Ian Breckheimer, Alex C Williams, Edith Law, Aaron M Ellison, and Charles C
733 Davis. Herbarium specimens reveal substantial and unexpected variation in phenological sen-

- 734 sitivity across the eastern united states. *Philosophical Transactions of the Royal Society B*, 374
735 (1763):20170394, 2019.
- 736 Martin Parniske. Arbuscular mycorrhiza: the mother of plant root endosymbioses. *Nature Reviews
737 Microbiology*, 6(10):763–775, 2008.
- 738 Anton Pauw and Julie A Hawkins. Reconstruction of historical pollination rates reveals linked
739 declines of pollinators and plants. *Oikos*, 120(3):344–349, 2011.
- 740 Shilong Piao, Qiang Liu, Anping Chen, Ivan A Janssens, Yongshuo Fu, Junhu Dai, Lingli Liu,
741 XU Lian, Miaogen Shen, and Xiaolin Zhu. Plant phenology and global climate change: Current
742 progresses and challenges. *Global change biology*, 25(6):1922–1940, 2019.
- 743 Timothée Poisot, Gabriel Bergeron, Kevin Cazelles, Tad Dallas, Dominique Gravel, Andrew Mac-
744 Donald, Benjamin Mercier, Clément Violet, and Steve Vissault. Global knowledge gaps in
745 species interaction networks data. *Journal of Biogeography*, 48(7):1552–1563, 2021.
- 746 Nicole E Rafferty, Paul J CaraDonna, and Judith L Bronstein. Phenological shifts and the fate of
747 mutualisms. *Oikos*, 124(1):14–21, 2015.
- 748 Christopher J Raxworthy and Brian Tilston Smith. Mining museums for historical dna: advances
749 and challenges in museomics. *Trends in Ecology & Evolution*, 36(11):1049–1060, 2021.
- 750 François Renoz, Inès Pons, and Thierry Hance. Evolutionary responses of mutualistic insect-
751 bacterial symbioses in a world of fluctuating temperatures. *Current opinion in insect science*, 35:
752 20–26, 2019.
- 753 Elizabeth Lewis Roberts and Aileen Ferraro. Rhizosphere microbiome selection by epichloë en-
754 dophytes of festuca arundinacea. *Plant and soil*, 396:229–239, 2015.
- 755 RJ Rodriguez, JF White Jr, Anne E Arnold, and a RS and Redman. Fungal endophytes: diversity
756 and functional roles. *New phytologist*, 182(2):314–330, 2009.

- 757 Gregor Rolshausen, Francesco Dal Grande, Anna D Sadowska-Deś, Jürgen Otte, and Imke
758 Schmitt. Quantifying the climatic niche of symbiont partners in a lichen symbiosis indicates
759 mutualist-mediated niche expansions. *Ecography*, 41(8):1380–1392, 2018.
- 760 Jennifer A Rudgers and Angela L Swafford. Benefits of a fungal endophyte in *elymus virginicus*
761 decline under drought stress. *Basic and Applied Ecology*, 10(1):43–51, 2009.
- 762 Jennifer A Rudgers, Michelle E Afkhami, Megan A Rúa, Andrew J Davitt, Samantha Hammer,
763 and Valérie M Huguet. A fungus among us: broad patterns of endophyte distribution in the
764 grasses. *Ecology*, 90(6):1531–1539, 2009.
- 765 Jennifer A Rudgers, Rebecca A Fletcher, Eric Olivas, Carolyn A Young, Nikki D Charlton, Dean E
766 Pearson, and John L Maron. Long-term ungulate exclusion reduces fungal symbiont prevalence
767 in native grasslands. *Oecologia*, 181:1151–1161, 2016.
- 768 Håvard Rue, Sara Martino, and Nicolas Chopin. Approximate bayesian inference for latent gaus-
769 sian models by using integrated nested laplace approximations. *Journal of the royal statistical
770 society: Series b (statistical methodology)*, 71(2):319–392, 2009.
- 771 Kari Saikkonen, Pedro E Gundel, and Marjo Helander. Chemical ecology mediated by fungal
772 endophytes in grasses. *Journal of chemical ecology*, 39:962–968, 2013.
- 773 Michelle E Sneck, Jennifer A Rudgers, Carolyn A Young, and Tom EX Miller. Variation in the
774 prevalence and transmission of heritable symbionts across host populations in heterogeneous
775 environments. *Microbial Ecology*, 74:640–653, 2017.
- 776 Thomas F Stocker, Dahe Qin, G-K Plattner, Lisa V Alexander, Simon K Allen, Nathaniel L Bindoff,
777 F-M Bréon, John A Church, Ulrich Cubasch, Seita Emori, et al. Technical summary. In *Climate
778 change 2013: the physical science basis. Contribution of Working Group I to the Fifth Assessment
779 Report of the Intergovernmental Panel on Climate Change*, pages 33–115. Cambridge University
780 Press, 2013.

- 781 Peter A Stott, Nathan P Gillett, Gabriele C Hegerl, David J Karoly, Dáithí A Stone, Xuebin Zhang,
782 and Francis Zwiers. Detection and attribution of climate change: a regional perspective. *Wiley*
783 *interdisciplinary reviews: climate change*, 1(2):192–211, 2010.
- 784 S Sully, DE Burkepile, MK Donovan, G Hodgson, and R Van Woesik. A global analysis of coral
785 bleaching over the past two decades. *Nature communications*, 10(1):1–5, 2019.
- 786 E Toby Kiers, Todd M Palmer, Anthony R Ives, John F Bruno, and Judith L Bronstein. Mutualisms
787 in a changing world: an evolutionary perspective. *Ecology letters*, 13(12):1459–1474, 2010.
- 788 Andrew T Tredennick, Giles Hooker, Stephen P Ellner, and Peter B Adler. A practical guide to
789 selecting models for exploration, inference, and prediction in ecology. *Ecology*, 102(6):e03336,
790 2021.
- 791 Amy M Truitt, Martin Kapun, Rupinder Kaur, and Wolfgang J Miller. Wolbachia modifies thermal
792 preference in *drosophila melanogaster*. *Environmental microbiology*, 21(9):3259–3268, 2019.
- 793 Shripad D. Tuljapurkar. Population dynamics in variable environments. III. Evo-
794 lutionary dynamics of r-selection. *Theoretical Population Biology*, 21(1):141–165,
795 February 1982. ISSN 0040-5809. doi: 10.1016/0040-5809(82)90010-7. URL
796 <http://www.sciencedirect.com/science/article/pii/0040580982900107>.
- 797 Arantxa Urdangarin, Tomás Goicoa, and María Dolores Ugarte. Evaluating recent methods to
798 overcome spatial confounding. *Revista Matemática Complutense*, 36(2):333–360, 2023.
- 799 Veronika Vikuk, Carolyn A Young, Stephen T Lee, Padmaja Nagabhyru, Markus Krischke, Mar-
800 tin J Mueller, and Jochen Krauss. Infection rates and alkaloid patterns of different grass species
801 with systemic epichloë endophytes. *Applied and Environmental Microbiology*, 85(17):e00465–19,
802 2019.
- 803 Zhengfeng Wang, Chunjie Li, and James White. Effects of epichloë endophyte infection on

- 804 growth, physiological properties and seed germination of wild barley under saline conditions.
805 *Journal of Agronomy and Crop Science*, 206(1):43–51, 2020.
- 806 Robert J Warren and Mark A Bradford. Mutualism fails when climate response differs between
807 interacting species. *Global Change Biology*, 20(2):466–474, 2014.
- 808 Nicole S Webster, Rose E Cobb, and Andrew P Negri. Temperature thresholds for bacterial
809 symbiosis with a sponge. *The ISME journal*, 2(8):830–842, 2008.
- 810 James F White and Garry T Cole. Endophyte-host associations in forage grasses. i. distribution
811 of fungal endophytes in some species of lolium and festuca. *Mycologia*, 77(2):323–327, 1985.
- 812 Franziska M Willems, JF Scheepens, and Oliver Bossdorf. Forest wildflowers bloom earlier as
813 europe warms: lessons from herbaria and spatial modelling. *New Phytologist*, 235(1):52–65,
814 2022.
- 815 Charles G Willis, Elizabeth R Ellwood, Richard B Primack, Charles C Davis, Katelin D Pearson,
816 Amanda S Gallinat, Jenn M Yost, Gil Nelson, Susan J Mazer, Natalie L Rossington, et al. Old
817 plants, new tricks: Phenological research using herbarium specimens. *Trends in ecology &*
818 *evolution*, 32(7):531–546, 2017.
- 819 Chao Xia, Nana Li, Yawen Zhang, Chunjie Li, Xingxu Zhang, and Zhibiao Nan. Role of epichloë
820 endophytes in defense responses of cool-season grasses to pathogens: A review. *Plant disease*,
821 102(11):2061–2073, 2018.
- 822 Kentaro Yoshida, Eriko Sasaki, and Sophien Kamoun. Computational analyses of ancient
823 pathogen dna from herbarium samples: challenges and prospects. *Frontiers in plant science*,
824 6:771, 2015.