

Stable developmental patterns of gene expression without morphogen gradients

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Abstract

Gene expression patterns are established by cross-regulating target genes that interpret morphogen gradients. However, as development progresses, morphogen activity is reduced, leaving the emergent pattern without stabilizing positional cues. The pattern then can be deteriorated by the intrinsically noisy biochemical processes acting at the cellular level. But remarkably, the established gene expression patterns remain spatially and temporally stable in many biological systems. Here we combine spatial-stochastic simulations with an enhanced sampling method (Non-Stationary Forward Flux Sampling) and a recently developed stability theory to address how spatiotemporal integrity of a gene expression pattern is maintained in developing tissue lacking morphogen gradients. Using a minimal embryo model consisting of spatially coupled biochemical reactor volumes, we study a stripe pattern in which weak cross-repression between nearest neighbor domains alternates with strong repression between next-nearest neighbor domains, inspired by the gap gene system in the *Drosophila* embryo. We find that fine-tuning of the weak repressive interactions to an optimal level can increase temporal stability of the expression patterns by orders of magnitude, allowing for stable patterns over developmentally relevant times in the absence of morphogen gradients. The numerically determined optimal parameter regime closely agrees with the predictions of the stability theory. By analyzing the properties of the reduced phase space defined by pattern asymmetry factors that characterize pattern integrity, we trace back the pattern stability enhancement to the emergence of a metastable basin that protects intact patterns from rapid destruction in the optimal repression regime via restoring forces that counteract pattern perturbations. Altogether our results demonstrate that metastable attractors can emerge as a property of stochastic gene expression patterns even without system-wide positional cues, provided that the gene regulatory interactions shaping the pattern are optimally tuned.

1 Introduction

Maintaining the integrity of spatial gene expression patterns over time is essential in embryonic development. In early embryo development locally expressed morphogenetic proteins spread through the tissue to form gradients of chemical signals [1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11]. Inside developing cells, these chemical signals are interpreted by gene regulatory networks to form remarkably precise and reproducible spatial patterns of gene expression that subsequently give rise to different body parts and organs [12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23]. However, as spatial patterns are established by reading out upstream morphogen gradients, their stability is constantly subject to inherently noisy cellular and extracellular processes [24, 25, 26, 27, 28, 29]. Moreover, the activity of morphogenetic gradients that is interpreted by target cells can decrease over developmental time. This decrease in activity can take different forms, including reduction of the relative signaling range as the embryo grows in size [16, 30], signalling pathway desensitization [31], or complete disappearance of the gradients at later developmental stages [32, 33]. Together, the inherent cellular stochasticity and reduced role of morphogen gradients at later stages raise the question about how stable patterns can be maintained over sufficiently long developmental times.

Focusing on the cellular stochasticity, biological cells are facing two types of noise, namely intrinsic and extrinsic noise, with different notions of robustness against the respective noise types [24, 25, 26, 27, 28, 29]. Intrinsic noise originates from the processes of gene regulation, protein production, and intracellular transport. Thus, robustness of spatial patterns to intrinsic noise amounts to buffering random fluctuations in the copy numbers of patterning proteins. Extrinsic noise, on the other hand, terms the variations originating from different external conditions including cell size variability [34, 35], cell-to-cell variation in ribosome abundance [27] or fluctuations in the cellular environment [36, 37]. Therefore, the robustness of spatial patterns to extrinsic noise refers to the capability of producing precise patterns in spite of imperfect initial conditions, classically termed “canalization” in Waddington’s picture of development [38, 39]. Several gene regulatory strategies providing either type of robustness have been studied [11, 21, 29], but our understanding of how nature orchestrates them in the fully interacting wild-type organism is still incomplete.

Among the regulatory mechanisms that drive developmental pattern formation, the regulatory motif in which two genes mutually repress each other is particularly prevalent [4, 40, 41, 42, 43, 44, 45, 46, 20]. Intriguingly, mutual repression can have a dual role in the establishment of spatial patterns. On the one hand, in systems driven by threshold-dependent activation of patterning genes via morphogen gradients, mutual repression is crucial for shaping out expression domains that are bounded from two sides, thus increasing the positional information carried by the expression pattern [14, 47, 16, 20, 22]. On the other hand, mutual repression can induce bistability leading to stochastic switching between cell fates. Hence, it is *a priori* unclear to which extent mutual repression supports or counter-acts the formation of stable spatial patterns [14, 16, 20]. This issue is particularly relevant to systems that lack external cues for symmetry breaking, such as morphogen or maternal gradients, that could force bistable cells into one of their opposing fates.

Here we ask whether a system of mutually interacting genes can maintain an initially arranged expression pattern in the absence of upstream input gradients. To address this question we study a spatially resolved gene regulatory network, conceptually motivated by the gap gene system in the early embryo of the fruit fly *Drosophila melanogaster* [48, 49, 50, 51, 52, 53]. This system implements a particular regulatory architecture, in which weak and strong mutual repressive interactions between expression domains of different genes alternate depending on whether the domains are adjacent or not. This motif, termed “alternating cushions”, was earlier investigated in terms of stability and robustness against extrinsic noise in initial conditions [41]. That study employed a reaction-diffusion model with step-like activation functions for representing the underlying gene expression dynamics. Using the so-called “moving kink approximation”, the study predicted an extensive basin of pattern stability in the parameter space of the model, where the stability could be attributed to repulsive forces between mutually repressing gene expression domains (“cushions”). More recently, an exact solution was obtained in an analogous model for the dynamics of the contact zones between two gene expression domains and for arbitrary combination of activating or repressing interactions between the involved genes [54]. This work provided exact stability conditions, leading to a better quantitative understanding of the conditions under which stochastically driven gene expression patterns can remain stable. Importantly, it was shown that perfect pattern stability (i.e., a pattern surviving for arbitrarily long time) can only

be achieved for a very specific combination of system parameters; nevertheless, in the vicinity of these states, there exists a continuity of well-defined but slowly travelling gene expression patterns, which can fulfill their biological role for a finite but in many cases sufficiently long period of time. However, it remained unclear whether the derived stability conditions also remain valid under genuinely stochastic conditions.

Here we assess the temporal stability of gene expression patterns interacting via the “alternating cushions” motif by numerical simulations of a minimal spatial-stochastic model that features a full microscopic representation of stochastic gene expression and protein diffusion, thus incorporating the relevant intrinsic noise sources. Using Non-Stationary Forward Flux Sampling (NS-FFS) [55, 56], an enhanced biased sampling technique for stochastic systems changing in time, we quantify for how long patterns shaped and maintained only by mutual repression can self-maintain themselves. Moreover, we expand the stability theory from [54] to the case of multiple interfaces and identify the parameter regime within which boundaries between adjacent gene expression domains are predicted to remain stable.

Our results show that the stability of patterns arranged in the alternating cushions scheme strongly varies with the strength of mutual repression between adjacent gene expression domains. We find that pattern stability time is maximized when spatially adjacent genes repress each other with intermediate strength while the next-nearest neighbor genes repress each other strongly. In this optimal regime, we confirm the existence of robust restoring forces and find signatures of a metastable basin that stabilizes well-ordered patterns (dynamical attractor), in accordance with the previous findings of [41]. Away from the optimum, forces induced by strong nearest neighbor mutual repression destroy the stripe patterns rapidly, while for weaker nearest neighbor repression the forces are imperceptible when compared to stochastic fluctuations. At the same time, the numerically determined optimal parameter regime is situated in the vicinity of the theoretically predicted optimum. While the adapted theory indicates a single optimal choice of parameters ensuring a perfectly stable pattern, the simulations uncover a parameter range over which pattern stability is enhanced but always finite.

Taken together, we demonstrate that forces generated in the alternating cushions scheme can maintain the gene expression pattern subject to stochastic production and diffusion of proteins for extremely long times if the parameters of mutual repression are optimized.

Modelling framework

In order to investigate stability of gene expression patterns without external input gradients, we performed stochastic simulations of a spatial pattern of four mutually repressing genes, using NS-FFS. Here we opted for a minimal spatially resolved stochastic model, shown in schematic Fig. 1, inspired by the the posterior gap gene pattern in *Drosophila* development [48]. The model considers four mutually interacting genes A, B, C and D, arranged in a five-stripe pattern (with order A-B-C-D-A) along a cylindrical spatial lattice. The four genes are analogous to the arrangement of the expression domains of gap genes *hb*, *kr*, *kni* and *gt* in nuclear cycle 14 in the posterior half of the early fly embryo, where *hb* is expressed in two domains, in the first (anteriormost) and last (posteriormost) expression domain [48, 49, 50, 51, 52, 53]. The spatial lattice consists of $N_z \times N_\phi$ equally spaced and well-stirred reaction volumes with periodic boundary conditions in the circumferential (ϕ -) direction motivated by the arrangement of cortical nuclei in the developing fly embryo. Protein diffusion and nuclear exchange are modeled via hopping between neighboring reaction volumes, with a rate proportional to the diffusion coefficient. In each nucleus, proteins of the genes A, B, C and D are produced from their corresponding promoters, dimerize and mutually repress each other by promoter binding. Each gene can repress the promoter of each other gene. Repression is non-competitive, i.e., each promoter has binding sites for each of the three other genes’ dimers and is inactivated when at least one dimer is bound (“OR”-logics). The model combines transcription and translation into one production step, neglecting some features of eukaryotic gene expression such as transcriptional bursts and enhancer dynamics, but previous work has shown that this does not alter the results qualitatively [57, 14].

In the anterior-posterior arrangement A-B-C-D-A, the genes repress each other mutually via the characteristic pattern of strong next-nearest neighbor (NNN) and weaker nearest-neighbor (NN) repression (alternating cushions), as observed in the *Drosophila* embryo [41, 48, 53, 58, 59, 60, 61, 62]. Specifically, there are two pairs of strongly repressing genes, (A,C) and (B,D), and four pairs of genes that repress each other weakly, (A,B), (B,C), (C,D) and (D,A). In our model, the difference in repression strength is tuned via the unbinding rate of the repressors from the repressed promoter. The strong-

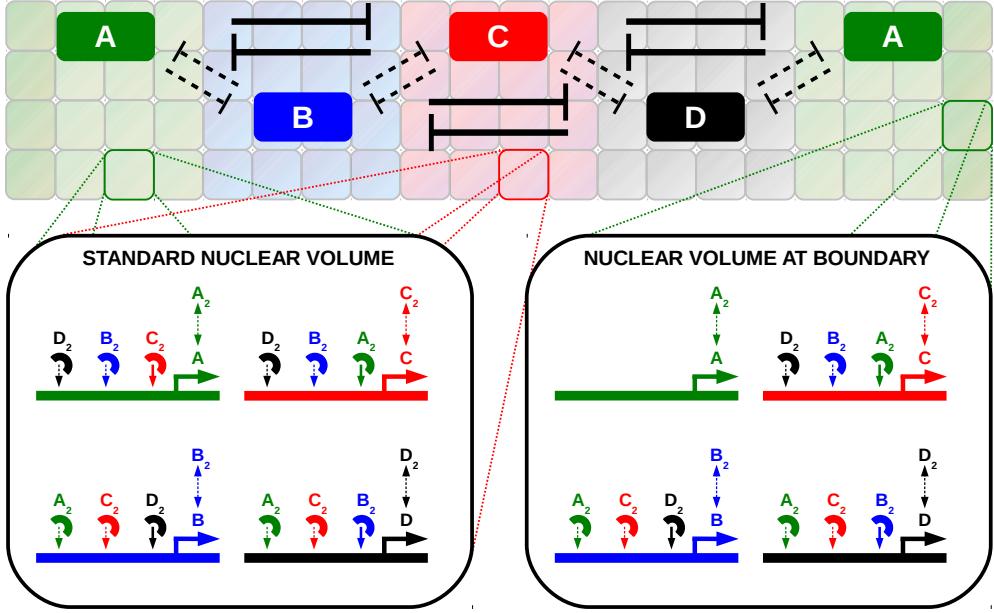


Figure 1: Schematic of the spatial gene-regulatory model. We use a cylindrical lattice of reaction volumes to mimic the arrangement of cortical nuclei in the posterior *Drosophila* embryo at developmental cycle 14. In each nuclear volume (shaded squares) we simulate production, degradation, dimerization and mutual repression of the four genes A, B, C and D via the Gillespie algorithm. Each gene is subject to repression by the protein dimers of the other genes, as indicated by the schematic promoters. Neighboring nuclei can exchange monomers and dimers via diffusive hopping. The system is initialized in a five-stripe pattern of expression domains in the order A–B–C–D–A, corresponding to the experimentally observed order in the fly embryo. The strength of mutual repression varies among gap gene pairs: genes associated with nearest neighbor (NN) domains repress each other weakly (dashed arrows), while next-nearest neighbors (NNN) domains exhibit strong mutual repression (thick arrows). By default, the concentration of A is pinned at the system boundary where the set of modelled reactions differs from the rest of the system by the fact that the A promoter can not be repressed. Details in the Methods, Sec. 4.

repressor unbinding rate k_s^{off} is chosen such that the NNN pairs (A,C) and (B,D) are in the bistable regime, while the weak-repressor unbinding rate k_w^{off} is varied. In this context, bistability corresponds to only one of the two competing genes being expressed at the single-nucleus level. In the absence of cues capable of forcing the bistable systems into a preferred state, stochastic switching is expected to eventually result in one of the domains to dominate over the respective other domain in the NNN pair, causing its elimination and simultaneous expansion of the dominating gene's domain. This partial breakdown of the initial pattern can happen independently for both strongly repressing NNN pairs and thus in random temporal order; however, ultimately one of the strong repression partners is eliminated in each of the NNN pairs and the system settles in a new, effectively irreversible state in which only the remaining two genes are co-expressed.

On the one hand, we expect that the presence of the third expression domain in between the NNN pair domains can impede elimination of (one of) the NNN pair domains when NN repression is present. On the other hand, overly strong NN repression is expected to enhance pattern breakdown because then even the overlapping NN expression domains are brought towards the bistable regime. We therefore study the pattern stability as a function of the *repression strength ratio* κ , defined as

$$\kappa \equiv k_w^{\text{off}} / k_s^{\text{off}}, \quad (1)$$

where k_w^{off} and k_s^{off} are the repressor unbinding rates for weakly repressing NN pairs and strongly repressing NNN pairs, respectively. κ is varied through the weak repression unbinding rate k_w^{off} . For $\kappa = 1$, i.e. $k_w^{\text{off}} = k_s^{\text{off}}$, both the NNN and the NN gene pairs are deeply in the bistable regime, while in the opposite limit $\kappa \rightarrow \infty$ ($k_w^{\text{off}} \rightarrow \infty$) the NN pairs do not affect each other at all.

2 Results

Pattern stability is quantified by asymmetry factors

A typical spatial pattern of gene expression with equally-sized domains is shown in Fig. 2. In order to quantify pattern stability we measure, as a function of κ , the average time until at least one domain is lost. Further, in our system the strong NNN repression effectively prohibits coexistence of the strongly repressing genes at one location. Hence, the increase in size of one domain is always accompanied by a reduction in of the domain with its strong interaction partner. This lead us to introduce the following two order parameters, λ_{AC} and λ_{BD} , here termed *asymmetry factors*, that measure the asymmetry for each of the two strongly antagonistic NNN pairs:

$$\lambda_{AC} \equiv \max([A]_{\text{tot}}, [C]_{\text{tot}})/N, \quad \lambda_{BD} \equiv \max([B]_{\text{tot}}, [D]_{\text{tot}})/N. \quad (2)$$

Here $[P]_{\text{tot}}$ is the *total* copy number of P proteins (counting dimers twice), and $N = [A]_{\text{tot}} + [B]_{\text{tot}} + [C]_{\text{tot}} + [D]_{\text{tot}}$ is the total protein number in the system across all species. In the spatially well-ordered pattern each protein domain occupies roughly the same fraction of the system, such that $\lambda_{AC} \simeq \lambda_{BD} \simeq 0.25$. As expansion of a domain progresses at the expense of its strong antagonist, λ_{AC} (or λ_{BD}) is enlarged and reaches values around 0.5 when the shrinking domain is eventually lost. In order to track progress of complete pattern losing one of its domains, we use sum $\lambda = \lambda_{AC} + \lambda_{BD}$, with values around 0.5 for five-stripe patterns and values above 0.75 indicating pattern breakdown.

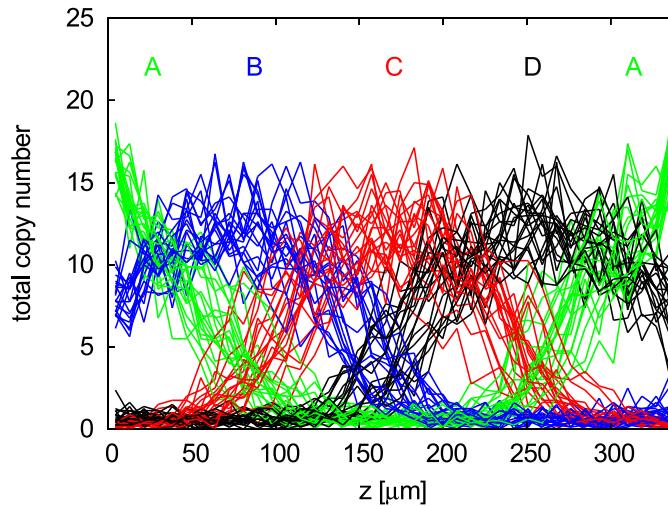


Figure 2: **Spatial pattern of gene expression.** Snapshots of the total copy numbers of all considered patterning proteins as a function of the axial coordinate z of the cylinder, averaged over its circumference. Colors correspond to Fig. 1 (green = A, blue = B, red = C, black = D). Snapshots were taken every 60 min over a total simulated time of 20 h after an initial relaxation phase of 30 min, starting from rectangular domain profiles of equal length. No-flux boundary condition at either end.

Our initial simulations revealed that even for very low protein copy numbers ($\lesssim 20$) the waiting times until one domain is lost are long compared to the duration of the actual breakdown event, and therefore difficult to sample by direct simulation. This lead us to ask, whether the five-stripe pattern is stabilized by a potential barrier that separates it from the pattern with fewer domains. Alternatively, the five-stripe pattern might not be stabilized by a potential barrier, but by a kinetic effect, where progress towards pattern breakdown is a slow directed process. In order to resolve which of these alternative mechanisms is responsible for the stabilization of the expression pattern, we combined our stochastic simulations with Non-Stationary Forward Flux Sampling (NS-FFS), which is particularly suited for enhanced sampling of non-equilibrium rare events. We used λ as the progress coordinate for NS-FFS, which aims at generating a branched and weighted trajectory ensemble that, in the most favorable cases, samples the relevant λ -range uniformly. This allowed us to generate sufficient statistics of rare breakdown events even in the most stable regions of parameter space (see Methods, Sec. 4).

The early simulations also showed that the expression domains of gene A at the boundaries of the system (green expression domains in Fig. 2) are particularly prone to destruction by their opponent domains, because they can only expand into one direction, towards the interior of the system. We therefore decided to keep constant the level of A proteins in the volume at the system boundaries by locally disallowing repression in the outermost nuclei at the boundaries. To assess how this model assumption influences our results, we later compare to simulations in which A could vary at the system boundaries, finding that our main findings also hold in this less restricted system.

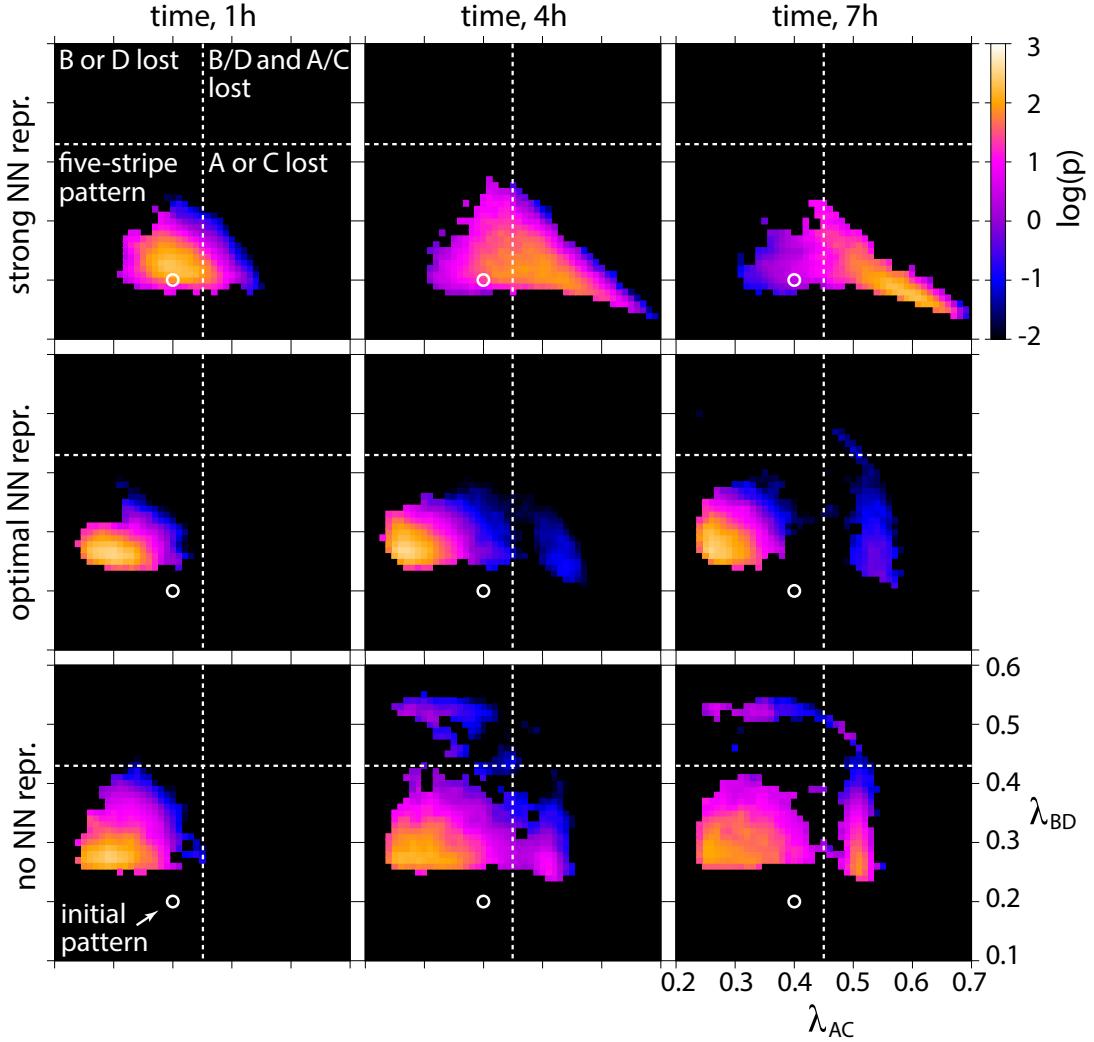


Figure 3: Pattern breakdown in the phase space spanned by asymmetry factors. Probability density snapshots of the phase space spanned by asymmetry factors λ_{AC} and λ_{BD} , defined in (2), at different times t for varied repression strength ratio κ . The conditions are following: strong NN repression, $\kappa = 3.16$ (top row), optimal NN repression for pattern stability, $\kappa = 31.6$ (middle row), and lack of NN repression, $\kappa = \infty$ (bottom row). The simulation was started with the initial rectangular five-stripe A-B-C-D-A pattern ($\lambda_{AC}, \lambda_{BD} = (0.4, 0.2)$) (white circle) in the pinned system. All snapshots are normalized histograms of reweighted $(\lambda_{AC}, \lambda_{BD})$ -points within $t \pm 5$ min. In the middle and bottom rows we identify three densely populated regions: a broad region centered around $(0.30, 0.30)$, R_S , which contains five-stripe patterns, and two smaller regions close to $(0.55, 0.30)$, R_{AC}^\dagger , and $(0.30, 0.55)$, R_{BD}^\dagger , representing patterns with one domain lost (region boundaries (dashed white), details in Methods, Sec. 4). Ultimately trajectories will converge towards region centered around $(0.55, 0.55)$, R^\ddagger , where two domains are lost.

Long-term pattern stability requires optimal repression strengths

In order to see how varied repression strength affects pattern stability, we reweighted histograms of simulated trajectories over the reduced phase space spanned by order parameters λ_{AC} and λ_{BD} at different times, for different values of κ ranging from strong NN repression ($\kappa \simeq 3$) to the limit of non-interacting nearest neighbors ($\kappa = \infty$), see Fig. 3. We found that there exists a region of stable expression patterns in phase space which is populated rapidly and then remains quasi-stationary, indicating that the system can remain in a metastable state if NN repression is moderate. In particular, the velocity with which the system escapes from the quasi-stationary region strongly depends on κ , with low and very high κ resulting in quick pattern deterioration, and intermediate κ values resulting in the most long-lived quasi-stationary states.

Stochastic fluctuations can lead to two different events corresponding to partial pattern destruction: one in which either the A or C domain is lost first and one in which either the B or D domain is lost first. Motivated by these observations we defined a region of stable patterns in terms of the asymmetry factors as $R_S \equiv \{(\lambda_{AC}, \lambda_{BD}) | \lambda_{AC} \leq 0.45 \text{ and } \lambda_{BD} \leq 0.43\}$, see Fig. 3. States that lie outside of R_S are considered deteriorated patterns, and accordingly we also defined two regions R_{AC}^\dagger and R_{BD}^\dagger and a region R^\ddagger accumulating patterns with one expression domain lost and patterns with two domains lost, respectively. The pattern survival probability $S(t) = \iint_{R_S} p(\lambda_{AC}, \lambda_{BD}, t) d\lambda_{AC} d\lambda_{BD}$ is the probability for the system to remain in the region of stable patterns until time t . We have never observed re-entry into R_S . We found that $S(t)$ is well-described by an exponential decay, $S(t) \propto e^{-k_D t}$, for times t larger than a certain lag-time t_{lag} . k_D then defines a deterioration rate, corresponding to average pattern stability time or alternatively mean time until pattern has lost one of its domains, $\tau_D \equiv 1/k_D$ (see Methods, Sec. 4).

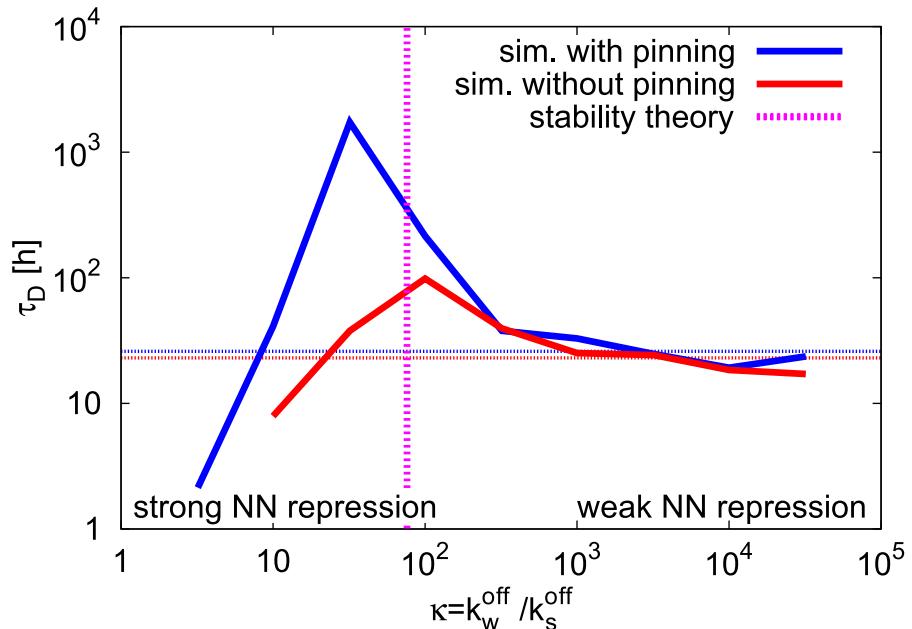


Figure 4: **An optimal strength of nearest neighbor repression maximizes pattern stability.** The mean time until pattern destruction τ_D as a function of κ , the ratio between the weak and strong repressor off-rate, for the system in which expression of gene A is fixed at the boundaries. We observe a pronounced maximum of the stability time when the weak repression is about 30 times weaker than the strong repression ($\kappa_{\text{opt}} = 31.6$) in the system with pinning at the boundaries (blue line). When pinning of the pattern at the system boundaries is relaxed (red line), the maximum of stability time moves to $\kappa = 100$. The dashed horizontal lines indicate the values for the completely uncoupled systems with $\kappa = \infty$. The dashed vertical line (magenta) shows the optimal repression strength ratio predicted analytically by our stability theory, $\kappa_{\text{theor}} \simeq 76$ (see last part of Results section).

By quantifying pattern stability time, we found that τ_D depends strongly on the repression strength ratio, with a maximum of τ_D as a function of κ at $\kappa_{\text{opt}} \simeq 30$, see Fig. 4 (blue curve). For κ values close to κ_{opt} pattern stability is still markedly enhanced. While significantly less stable than in the region around the optimum, patterns with stability time on the order of several hours remain possible in the absence of NN repression ($\kappa \rightarrow \infty$). In contrast, when NN and NNN repression have close to equal strength ($\kappa \rightarrow 1$) patterns collapse almost immediately.

In the maximally stable regime restoring forces reconstitute perturbed patterns

The observation of a phase space region in which system trajectories persist for long times raises the question whether this region constitutes a true metastable basin of attraction. We first addressed this question next by analyzing transient behavior of the perturbed patterns. If enhanced phase space density in certain regions of the $(\lambda_{AC}, \lambda_{BD})$ -space were indeed due to the presence of a metastable basin, perturbations that transiently drive the system away from the stable pattern should be counteracted by restoring forces. To test this hypothesis, we perturbed relaxed five-stripe patterns from the hypothetical basin by artificially enlarging domains in which one gap gene is dominant. Using these perturbed states as initial conditions, we then ran the spatial-stochastic simulator with higher time resolution, and checked whether the perturbed systems relax back into the presumed basin. We investigated two types of asymmetric perturbations: “C expansion”, in which the central C domain is unidirectionally expanded at the expense of the posterior A domain, and the converse “A expansion”, in which the anterior A domain is enlarged at the expense of the C domain. The perturbation experiments are described in detail in the Methods, Sec. 4, and the Supporting Information.

We find that at $\kappa = \kappa_{\text{opt}}$, for both perturbations the perturbed pattern ensembles relax back to their original positions on a timescale $\sim 10 h$ (see Supporting Fig. S1). This demonstrates that for optimal repression strength ratio an effective restoring force counteracts deviations from the five-stripe pattern for varied λ_{AC} . Moreover, this suggests that the probability-enriched region within R_S is a real metastable state confined by an underlying force field. In accordance, the timescale of relaxation is orders of magnitude shorter than the timescale of pattern collapse. Thus, for $\kappa = \kappa_{\text{opt}}$ pattern destruction is a Markovian transition between metastable basins with transition waiting times much longer than the timescales of intra-basin dynamics. In contrast, we could not observe clear restoring behavior in the systems with very weak or no nearest neighbor interaction. Here perturbations of similar strength tend to result in almost immediate pattern destruction.

In summary, for the repression strengths ratio $\kappa_{\text{opt}} \simeq 30$ that maximizes stability, pattern breakdown appears to be an activated process characterized by a restoring force towards the initial state.

Statistical analysis of phase-space dynamics reveals a metastable basin

We further figured that the existence of a true metastable basin should manifest itself also in the statistics of transient dynamics in phase space. Here the local velocities in the $(\lambda_{AC}, \lambda_{BD})$ phase space are particularly informative: forces that drive trajectories back into basins of attraction should translate into local mean phase space velocities with a clear bias towards the bottom of the basin.

To extract the velocity field for our system we modeled the coarse-grained pattern dynamics as overdamped diffusive motion in the $\vec{\lambda} \equiv (\lambda_{AC}, \lambda_{BD})$ plane, assuming that these degrees of freedom capture the slowest time scales of the system and making a Markov approximation for the fast dynamics [63, 64]. This technique has been successfully applied in protein folding [65, 66, 67, 68]. The corresponding model equation is

$$\frac{d}{dt} \vec{\lambda} = \langle \vec{v}_\lambda \rangle(\vec{\lambda}) + \sqrt{2D_\lambda(\vec{\lambda})} d\vec{W} \quad (3)$$

where \vec{W} is uncorrelated (2D) white noise with unit covariance. We estimated the local drift $\langle \vec{v}_\lambda \rangle(\vec{\lambda})$ and diffusion coefficient $D_\lambda(\vec{\lambda})$ from our reweighed simulated trajectories by averaging local displacements (see Methods, Sec. 4, and Sec. S1.2 in the Supporting Text). Furthermore, $\langle \vec{v}_\lambda \rangle(\vec{\lambda})$ is proportional to the effective force acting at the reduced phase space point $\vec{\lambda}$ in the overdamped Langevin model. The local mean velocity field $\vec{v}_\lambda(\vec{\lambda})$ is determined by the conditional transition probabilities $\pi(\vec{\lambda}, \vec{\lambda}')$ between states $\vec{\lambda}$ and $\vec{\lambda}'$, and thus can be extracted from our transient simulation data. The resulting

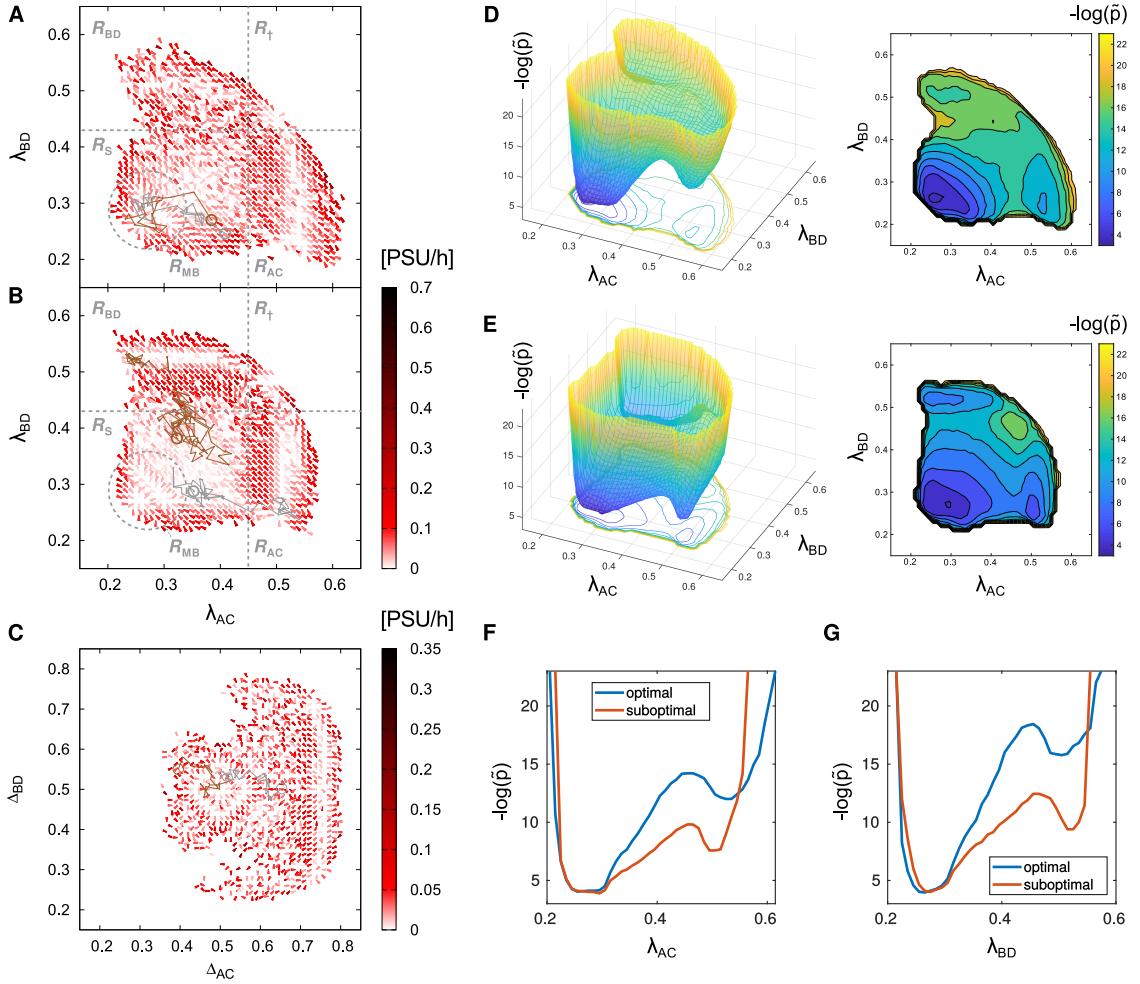


Figure 5: Phase space velocity fields and passage statistics reveal metastable basins. **A, B)** Average phase space velocity fields for the system with optimal ($\kappa = \kappa_{\text{opt}}$, A) and suboptimal ($\kappa = 1000$, B) repression strength ratio, in the phase space spanned by asymmetry factors λ_{AC} and λ_{BD} as defined in (2). The small subregion with concentrically inwards-pointing velocities towards which perturbed trajectories relax, corresponding to metastable basin of five-stripe patterns, is indicated (R_{MB} , dashed circle). Velocity fields were obtained by averaging displacements of all trajectories that exit the local bin (see Methods, Sec. 4). Two examples of trajectories relaxing after perturbations are shown (gray and brown lines) with their starting points (circles). The boundaries of phase space regions (thin dashed lines) are as in Fig. 3. Velocity magnitude is indicated with colors. **C)** The velocity field corresponding to $\kappa = \kappa_{\text{opt}}$ for the alternative asymmetry factors (“differences”) δ_{AC} and δ_{BD} , as in (4). In C the metastable basin R_{MB} is localized around the center $(\delta_{\text{AC}}, \delta_{\text{BD}}) = (\frac{1}{2}, \frac{1}{2})$, corresponding to a perfectly symmetric pattern. The magnitude unit “phase space unit per hour” (PSU/h) is specific to the chosen asymmetry factors. **D, E)** The landscapes of the “pseudopotential” $-\log \tilde{p}$ computed from the total number of phase space trajectories registered in the respective bin of the phase space. The contour plots to the right of the 3D views show a projected view of the same landscapes. **F, G)** Comparison of sections in λ_{AC} and λ_{BD} directions, respectively, at $\lambda_{\perp} = 0.28$ between the optimal and suboptimal choice of the repression strength ratio κ . Here the $-\log \tilde{p}$ profile is almost identical in the metastable basin R_{MB} , but transitions towards the destroyed pattern states face a higher barrier in the system with optimal $\kappa = \kappa_{\text{opt}}$, in both phase space directions.

average velocity field in the reduced phase space of $(\lambda_{\text{AC}}, \lambda_{\text{BD}})$ for the optimal repression strength ratio ($\kappa = 31.6$) is in Fig. 5A, and for suboptimal ($\kappa = 1000$) is shown in Fig. 5B.

Interestingly, in Fig. 5A one can identify two regions of $(\lambda_{AC}, \lambda_{BD})$ -space with low average velocities: one within the region of stable states R_S , the other within the region R_{AC}^\dagger of states in which the C expression domain is lost. The region R_{BD}^\dagger in which either B or D are lost, has no clear boundaries for optimal $\kappa = 31.6$, and only for much larger $\kappa = 1000$ a low-velocity plateau is clearly seen in this region (Fig. 3B). Notably, in the lower-left corner of the R_S plateau we notice a small region in which average velocities are significantly higher and all pointing inwards. We refer to this region as R_{MB} , and identify it as the metastable basin of intact, relaxed five-stripe patterns. In accordance, the two shown exemplary perturbed trajectories relax into R_{MB} after randomly exploring the R_S plateau, and remain confined to the R_{MB} for later times (Fig. 5A). However, if the system drifts far away from R_{MB} , in the direction of R_{AC}^\dagger , the trajectories are quickly absorbed into R_{AC}^\dagger once they reach the edge of R_S characterized by high velocity components towards R_{AC}^\dagger .

In order to further investigate the low-velocity attraction basins and the high-velocity ridges that separate these basins, we use a different representation of pattern asymmetry, defining the shifted difference coordinates

$$\delta_{AC} \equiv \frac{1}{2}([A]_{tot} - [C]_{tot})/N + \frac{1}{2}, \quad \delta_{BD} \equiv \frac{1}{2}([B]_{tot} - [D]_{tot})/N + \frac{1}{2}. \quad (4)$$

These coordinates measure the deviation from a perfectly symmetric pattern $P_0 = (\delta_{AC}, \delta_{BD}) = (\frac{1}{2}, \frac{1}{2})$ in a way that retains information about which of the antagonistic genes becomes dominant. The corresponding average velocities in $(\delta_{AC}, \delta_{BD})$ -space are shown in Fig. 5C. The low-velocity basin, corresponding to R_{MB} , occupies the central part of $(\delta_{AC}, \delta_{BD})$ -space in Fig. 5C. Accordingly, perturbed trajectories relax towards the region enclosed by concentric velocity vectors pointing towards P_0 . This is in line with the Waddington picture of canalization [38, 39], in which developmental stages are seen as successive attractors of the underlying dynamics with the five-stripe pattern representing such an attractor.

In Fig. 5B we show the average velocity field for the case with weaker NN repression ($\kappa = 1000$). Here the velocity fields are even more plateau-like in the region corresponding to weakly asymmetric patterns, and the characteristic concentric velocity pattern indicative of the basin in the optimal case cannot be clearly discerned any more in this case. In accordance, trajectories starting from perturbed patterns do not relax back and progress towards patterns with at least one domain lost.

In addition to the average velocity fields of the registered phase space trajectories, the signatures of the metastable basins are also visible in the local phase space density sampled over many trajectories that explored the phase space during the whole sampled time interval, $p(\vec{\lambda})$. A suitable quantity for visualizing the corresponding phase space “landscape” is the negative logarithm of $p(\vec{\lambda})$; note that in an equilibrated, stationary system this quantity would be proportional to the energy (landscape) defining the stationary probability distribution of the system. Since our system is genuinely non-stationary, this relationship does not hold. Nevertheless we can consider our most stable systems transiently equilibrated in the metastable basins or origin and akin to stationary systems *until* they irreversibly cross the barrier towards one of the basins corresponding to destroyed patterns. Notably, while the depth of these basins grows with the amount of simulated time after the destruction event, the height difference between the metastable basin of intact patterns and the barrier separating it from the destroyed patterns basin is entirely determined by the recurring properties of the destruction process, and not expected to change provided that sufficiently many destruction events have been recorded.

In Fig. 5D and E we plot the “pseudopotential landscape” defined as $-\log(\tilde{p}(\vec{\lambda}))$ for optimal $\kappa = 31.6$ (D) and suboptimal $\kappa = 1000$ (E), where $\tilde{p}(\vec{\lambda})$ is a locally smoothed version of $p(\vec{\lambda})$ which equalizes out small local spikes in $p(\vec{\lambda})$ but preserves the overall structure of the resulting landscape (see Methods for details). The small plots right of the landscape visualizations show sections through the landscapes in direction of the asymmetry factors λ_{AC} and λ_{BD} at chosen constant values of the respective orthogonal factor (see Fig. 5 caption). In both cases we can clearly identify the metastable basin of undestroyed patterns and a barrier separating it from the basins of (half-) destroyed patterns. The basin corresponding to the states in which either the B or D domain is lost is less pronounced for the optimal choice of κ due to its lower accessibility, and—more importantly—separated by a higher barrier. This is best seen in a more detailed explicit comparison of the sections through the landscapes, shown in Fig. 5F. The comparison clearly reveals that the barrier separating the metastable basin of intact patterns from the basin in which the C domain is lost is both higher and wider for the optimal choice of κ , overall leading to a markedly lower rate of pattern destruction.

Taken together, the analysis of both the velocity fields and the empirically sampled phase space density demonstrate that the long-time confinement of phase space trajectories close to the five-stripe pattern at optimal NN repression is due to the existence of a metastable basin which impedes progress towards losing one of the domains by restraining the system from leaving the metastable basin. With decreasing strength of NN repression the basin gradually disappears, thus enhancing the probability of pattern deterioration.

Stability enhancement does not require pinning

To assess whether pinning of the A-domains at the system boundaries is necessary for the observed stability enhancement at intermediate NN repression, we repeated our simulations and analysis for a system without pinning. In contrast to the system with pinning, here the promoters of gene A in the nuclei at the system boundaries can be inhibited by the repressors of A. We found that also in the system without pinning, pattern stability is markedly enhanced by the presence of weak interaction partners between two strongly repressing gene domains. In Figure 4 the red curve shows the mean destruction time τ_D against the ratio of the repressor off-rates κ for the system without pinning. We again find the highest pattern stability at an optimal repression strength ratio $\kappa_{\text{opt}}^{(\text{np})} = 100$ (red curve), which is close to the optimum in the system with pinning ($\kappa_{\text{opt}}^{(\text{p})} \approx 30$, blue curve), albeit with about 10 times lower overall stability times; yet, these stability times are still about an order of magnitude larger than without fine-tuning of NN interactions.

Overall this demonstrates that enhancement of pattern stability by at least one order of magnitude is possible both with and without pinning of expression at the system boundaries. However, pinning alters the proportion of destruction pathways that the collapsing patterns pursue; we discuss this effect in more detail in Sec. S1.3 of the Supporting Text.

The optimal pattern stability regime is predicted analytically by a mathematical model of expression domain competition

The problem of pattern stability has been recently addressed analytically in [54], where general and exact stability conditions for a contact zone between two interacting domain boundaries were derived. In this section, we show that these stability conditions can be successfully employed in the current multi-gene system, in order to make a coarse-grained prediction of stabilizing parameters set.

The major finding reported in [54] is that for sufficiently strong interactions between two genes, a contact zone of well-defined width is established between two domains of gene expression. However, in general, this pattern is dynamic, as one domain can still shrink and the other grow in coordinated manner, which preserves the width of the contact zone. This means that the contact zone can undergo a drift. It was found that the velocity of this motion asymptotically tends to a certain constant value, determined by the system parameters. This defines an attractor of the pattern dynamics, as the drift velocity and the width of contact zone are restored if concentrations of gene expression products are perturbed. A special case of this behaviour are (perfectly) stable patterns, for which the drift velocity is exactly zero. In these systems patterns can survive infinitely long. Conversely, increasing deviations of the system parameters from the stabilizing values translate into growing drift velocity, which eventually destroys the pattern. While the systems simulated in this work are highly stochastic, their dynamics also incorporate the deterministic component, which is expected to follow the principles outlined in [54]. Thus, the survival time of the patterns is not only hampered by noise, but also by that deterministic component, unless stability conditions are met. We therefore assessed whether the numerically determined optimal κ_{opt} coincides with the theoretically predicted values of κ_{theor} that ensure pattern stability.

To this end we mapped the microscopic model used in our stochastic simulations onto the effective reaction-diffusion model analysed in [54] (see Methods). With the necessary readjustments, the adapted effective model equation reads

$$\partial_t X_2(x, t) = D \partial_{xx} X_2(x, t) - \gamma X_2(x, t) + H \boldsymbol{\theta} \left(1 - \sum_{Y \neq X} \epsilon_{XY} Y_2(x, t) \right), \quad (5)$$

where $X, Y \in \{A, B, C, D\}$ denotes the particular proteins, $X_2(x, t), Y_2(x, t)$ is the concentration profile

of their respective dimers, D is the diffusion constant, γ the degradation constant, H a production constant, and ϵ_{XY} are gene-gene interaction strengths. $\theta(\dots)$ denotes the Heaviside step function, corresponding to steep Hill kinetics. Note that the derivations in [54] only apply to systems with size $L \gg \lambda$, where $\lambda \equiv \sqrt{D/\gamma}$ is the characteristic length of gene interaction. For the systems studied here, $\lambda \approx 8.62$ nm, which is markedly smaller than the system size $L \simeq 340$ nm, warranting application of the theory.

While the original theory in [54] describes only the contact zone between exactly two domain boundaries, we can adapt it to the four-gene system studied here. Fig. 2 shows that in the alternating cushions system there are only two types of contact zones: (i) between two strongly interacting genes with the third, weakly interacting gene, expressed in the background or (ii) between two weakly interacting genes, with all other genes having close-to-zero expression level. Thus, we will consider stability of both contact zone types separately.

In the type-(i) contact zone, the dynamics of gene expression is described by the effective equations

$$\begin{cases} \partial_t X_2(x, t) = D\partial_{xx}X_2(x, t) - \gamma X_2(x, t) + H\theta\left(1 - K_w^{-1}\frac{H}{\gamma} - K_s^{-1}Y_2(x, t)\right) \\ \partial_t Y_2(x, t) = D\partial_{xx}Y_2(x, t) - \gamma Y_2(x, t) + H\theta\left(1 - K_w^{-1}\frac{H}{\gamma} - K_s^{-1}X_2(x, t)\right) \end{cases} \quad (6)$$

where we approximate that the third “background gene”, has a constant expression level over the contact zone. The equilibrium value of this expression level is H/γ . K_w and K_s are the weak and strong repression constants, respectively and they satisfy (cf. Eq. 1 and Methods, Sec. 4):

$$\kappa = \frac{K_w}{K_s} = \frac{k_w^{\text{off}}}{k_s^{\text{off}}} . \quad (7)$$

Type-(i) contact zones are established between genes A and C (with B or D in the background) as well as between B and D (with C in the background). In the type-(ii) contact zone, the equations take the form:

$$\begin{cases} \partial_t X_2(x, t) = D\partial_{xx}X_2(x, t) - \gamma X_2(x, t) + H\theta\left(1 - K_w^{-1}Y_2(x, t)\right) \\ \partial_t Y_2(x, t) = D\partial_{xx}Y_2(x, t) - \gamma Y_2(x, t) + H\theta\left(1 - K_w^{-1}X_2(x, t)\right) \end{cases} \quad (8)$$

This contact zone emerges between genes A and B, B and C, C and D as well as D and A.

The general stability conditions derived in [54] define a relation between the system parameters that ensures zero drift of the contact zone. Applied to the system studied here these conditions read:

$$(1 - |R_X|)^\lambda = (1 - |R_Y|)^\lambda , \quad -1 \leq R_X \leq 1 , \quad -1 \leq R_Y \leq 1 , \quad \text{sgn } R_X = \text{sgn } R_Y , \quad (9)$$

where

$$R_X = \frac{-2\gamma^2\tilde{C}_X - \epsilon_{XX}\gamma H}{\epsilon_{XY}H} - 1 , \quad R_Y = \frac{-2\gamma^2\tilde{C}_Y - \epsilon_{YY}\gamma H}{\epsilon_{YX}H} - 1 , \quad (10)$$

Equations (9) can be applied separately to the type-(i) and type-(ii) contact zones in order to identify the range of stabilizing parameters in each case. Subsequently, we will investigate whether these ranges overlap, as the entire pattern is only stabilized when both types of contact zones are stable at the same time. We specify the constants in (9) for each contact zone type as follows:

$$(i) \quad \tilde{C}_X = \tilde{C}_Y = 1 - K_w^{-1}\frac{H}{\gamma} , \quad \epsilon_{XY} = \epsilon_{YX} = -K_s^{-1} , \quad \epsilon_{XX} = \epsilon_{YY} = 0 , \quad (11)$$

$$(ii) \quad \tilde{C}_X = \tilde{C}_Y = 1 , \quad \epsilon_{XY} = \epsilon_{YX} = -K_w^{-1} , \quad \epsilon_{XX} = \epsilon_{YY} = 0 . \quad (12)$$

Notice that as we choose D , γ and H to be the same for all dimers, $R_X = R_Y$ for each contact zone, and the first and the last of stability conditions (9) are automatically satisfied. The remaining condition is $-1 \leq R_X \leq 1$, which requires closer examination.

Employing parameters (11) for the type-(i) contact zone and parameters (12) for the type-(ii) contact zone in $-1 \leq R_X \leq 1$, we obtain two sets of inequalities:

$$\begin{aligned} (i) \quad -1 &\leq \frac{-2(1 - K_w^{-1}\frac{H}{\gamma})\gamma}{-K_s^{-1}H} - 1 \leq 1 , \\ (ii) \quad -1 &\leq \frac{-2\gamma}{-K_w^{-1}H} - 1 \leq 1 . \end{aligned} \quad (13)$$

Solving these inequalities results in:

$$(i) \quad K_w \geq \frac{H}{\gamma}, \quad K_s \leq \frac{1}{(\frac{H}{\gamma})^{-1} - K_w^{-1}}, \quad (14)$$

$$(ii) \quad K_w \leq \frac{H}{\gamma}.$$

These conditions show that the addition of weak interactions is instrumental for increasing system stability. On the one hand, the type-(i) contact zone is stable provided that the weak interaction strength K_w^{-1} does not exceed $(H/\gamma)^{-1}$; otherwise it would prevent the expression of strongly interacting genes in this region. On the other hand, for the type-(ii) interface it is necessary that K_w^{-1} is greater than $(H/\gamma)^{-1}$, as this minimal strength of repression is required to prevent co-expression of both weakly interacting genes in the same region. This shows that in order to simultaneously stabilize both types of contact zones, one must negotiate between these two largely opposite goals. This trade-off can be achieved only for the most marginal value in both parameter ranges, $K_w = H/\gamma$. This highlights why in the alternating cushions architecture the weak interactions have to be fine-tuned for pattern stability; in contrast, but in line with the numerical findings, the strong interactions characterized by K_s can be arbitrarily large, $K_s \leq +\infty$. The simulations in this work were performed for $K_s \simeq 0.003 \mu\text{m}^{-3}$ with K_w varied to obtain different κ values, see Methods, Sec. 4. Calculated from these microscopic parameters, $H/\gamma \simeq 0.23 \mu\text{m}^{-3}$. The resulting theoretical value of κ ensuring stability is $\kappa_{\text{theor}} \simeq 76$. This number is of the same order of magnitude as the optimal κ in the simulated stochastic systems, showing better agreement with the no pinning case ($\kappa_{\text{opt}}^{(\text{np})} \simeq 100$) than with the case with pinning ($\kappa_{\text{opt}}^{(\text{p})} \simeq 30$), see Fig. 4.

3 Discussion

In many developing organisms, morphogen gradients provide a long-range positioning system by activating downstream patterning genes in a concentration-dependent manner. Prominent examples are the gap gene system in *Drosophila*, whose main maternal regulators are the morphogen gradients of *Bcd* and *Cad* spreading along the embryo axis [52, 48, 69, 70, 71, 72, 73, 74, 75], and the vertebrate neural tube with *Shh* and *BMP/Wnt* secreted from the opposite sides of the neural tube [4, 7, 8, 18, 20, 23]. For the *Drosophila* embryo, multiple studies have shown that mutual interactions between gap genes play a crucial role in abdominal segmentation [14, 48, 50, 51, 52, 47, 76, 77, 78], leading to the formation of stable domains with slow effective dynamics [41]. However, it remains unclear how such a system could be robust given the stochastic nature of gene expression and regulation. Moreover, it is observed that maternal regulators such as the *Bcd* gradient disappear while the expression patterns invoked downstream persist [79, 80].

Here we asked whether a system of mutually repressing developmental patterning genes arranged in successive expression domains can indeed be stable over developmentally relevant time intervals without upstream morphogen gradients while facing unavoidable fluctuations in the expressed gene products. Such copy number fluctuations can induce bistable switching at the domain boundaries, resulting in stochastic movement of the boundary which ultimately can lead to destruction of one of the gene expression domains. We quantified the mean stability time of a five-stripe expression pattern formed by four interacting genes in a stochastic model conceptually inspired by the posterior *Drosophila* embryo in cycle 14 as a function of the repression strength between neighboring stripes. To be able to simulate the breakdown of very stable patterns we employed Non-Stationary Forward Flux Sampling (NS-FFS), an enhanced sampling scheme for simulating rare events in non-stationary systems with transient dynamics [56]. We find that for an optimal value of the repression strength between adjacent expression domains the stability of the pattern is increased by about an order of magnitude. This stability optimum can be traced back to the fact that bistable switching at the boundary between domains of strongly mutually repressing genes is inhibited by an intervening cushion domain of a gene that weakly represses both strong partners. This stabilizing mechanism works best if the spacer gene represses its nearest neighbors (NN) with moderate strength: very weak NN repression has no effect while strong NN repression globally destabilizes overlapping domains. At the optimal repression strength ($\kappa = \kappa_{\text{opt}}$) the cushion thus slows down the random motion of the domain boundary and subsequent pattern destruction.

Stability is enhanced even more, by one more order of magnitude, if expression of the outermost gene is pinned at the system boundaries, which effectively anchors the whole expression pattern. Such a situation may emerge when the outermost gene remains under control of maternal cues, such as maternally deposited mRNA, while the other gene stripes form only by zygotic interactions. In that case we find that five-stripe patterns form a metastable attractor of the dynamics with a restoring force that counteracts perturbations, such as non-perfect initial conditions. In the optimal stability regime, our observations are consistent with the Waddington picture [38, 39] of development as canalization into successive metastable states, with the ordered initial gap gene pattern representing one of the metastable states in this succession. Earlier work already demonstrated that developmental attractors may emerge as an intrinsic property of the gene expression pattern established through mutual interactions [51, 78]. Here, we demonstrate that also without morphogen gradients metastable basins can arise and protect expression patterns against stochastic fluctuations.

Further insight comes from the application of the stability theory derived in [54] to the model of four genes interacting in the alternating cushions scheme. In agreement with the simulations, these analytical calculations reveal that the presence of weak interactions is necessary for stabilizing the system and establishing long-surviving patterns. More specifically, theoretical analysis shows that requirements for stability of type-(i) contact zones (i.e. two strongly interacting genes with the third weakly interacting in the background) and type-(ii) contact zones (i.e. two weakly interacting genes with other genes at very low expression level) are to certain degree incompatible, and the agreement can be achieved only for the most marginal value of $\kappa = \kappa_{\text{theor}}$ in the respective stability range for each type. As a consequence, simultaneously ensuring perfect stability of both contact zone types requires the fine-tuning of weak repression strength, quantified by the corresponding dissociation constant K_w . This analytical prediction of one optimal value of κ is in qualitative agreement with the numerical simulations, which show a very sharp rise in the survival time of expression pattern near one particular value of $\kappa = \kappa_{\text{opt}}$, see Fig. 4.

Quantitatively, the numerical κ_{opt} and theoretical κ_{theor} agree particularly well in the no-pinning case (for pinning: $|\kappa_{\text{opt}}^{(\text{p})} - \kappa_{\text{theor}}|/\kappa_{\text{opt}}^{(\text{p})} = 139\%$, for no-pinning: $|\kappa_{\text{opt}}^{(\text{np})} - \kappa_{\text{theor}}|/\kappa_{\text{opt}}^{(\text{np})} = 24\%$). This is in line with the assumptions of [54], in which pinning was also neglected. Differences between κ_{theor} and κ_{num} are expected due to the nature of approximations employed in the mapping of microscopic model on its effective representation (5). It is plausible that this discrepancy could be resolved by constructing an even higher-level stability theory that takes into account the spatial variability of all four genes in each contact zone.

The simulations carried out in this work are both highly stochastic and microscopically detailed. Yet, the approximate agreement between κ_{opt} and κ_{theor} suggests that the deterministic dynamics of the generic model (5), described in [54], is still important for this system. This deterministic theory suggests that, unless the system parameters are chosen to satisfy the stability conditions perfectly, the contact zones between the domains undergo drift, which would be able to eventually destroy the pattern on its own. This behaviour is overshadowed by strong stochastic fluctuations in the numerical simulations, but the significant increase in survival time in the vicinity of κ_{theor} suggests that nearly perfectly stable systems are significantly more robust against noise. However, the presence of noise seems to prevent perfect stability, as the increase of pattern survival time near κ_{theor} , though remarkable, was finite in all simulations. Nevertheless, taken together, these results demonstrate that the stability theory might provide useful information also in the context of gene expression models with substantial levels of noise.

The observed stability times appear sufficient for early fly embryogenesis ($\simeq 2h$ until cycle 14) for all NN repression strengths weaker than the optimal value, with or without pinning, even for the reduced system size considered here for computational feasibility. In a more realistic description, stability times will be likely modified due to the influence of system size. We expect that as expression domains grow in spatial extension or maximal copy numbers stability will be further enhanced. In the first case, stability is enhanced because more local cell fates need to be switched while the noise level remains the same. In the second case, the relative noise level is reduced in the first place. Nevertheless, based on the theoretical and numerical evidence we believe that the stability enhancing mechanisms uncovered in this study will also apply to biologically relevant system sizes. Other factors potentially affecting stability are autoactivation interactions and interactions with other genes not included in the simplified regulatory network studied here, which will likely affect the dynamics of the gene expression pattern. Note, however, that the adapted stability theory clearly identifies the weak nearest-neighbor repression strength as the key parameter for enhancing stability, while the strong repressive interactions are found not to affect stability as long as they are chosen strong enough.

In developmental perspective, the present work leads to the interesting question of how a properly ordered stripe pattern of mutually repressing genes is established initially. While pinning at the system boundaries can break the pattern symmetry between the genes forming the innermost and outermost stripes, additional symmetry breaking mechanisms would be required to position the remaining two genes. It is conceivable that the timing of the onset of gene expression for the different gap genes may determine their initial arrangement, which then gets stabilized by mutual interactions.

The stabilizing mechanism arising from fine-tuning nearest-neighbor interactions in the alternating cushions scheme can be also considered in the broader class of regulatory mechanisms providing pattern stability against intrinsic and extrinsic noise [11, 21, 29]. In future studies, it may be instrumental to further numerically and analytically explore the proposed model by including other biologically relevant features. Possible extensions include growth of the tissue by cell divisions, self-correcting mechanisms through cell-to-cell communication other than diffusive exchange of proteins, or inclusion of more specific noise types. These extensions could further test the validity of our stability under more realistic biological conditions. However, due to the remarkable agreement between our adapted stability theory and the numerical simulations of the minimal model studied in this work, we believe that more realistic variants of it will result in quantitative but not qualitative changes in our predictions.

4 Methods

Details of the model

Our model is inspired by arguably the most paradigmatic developmental system in which development of distinct cell fates is determined by local protein expression patterns driven by external morphogen gradients, the early embryo of the fruit fly *Drosophila melanogaster*. We model the egg-shaped embryo with its cortical layer of nuclei as a cylindrical array of reaction volumes coupled by diffusion of proteins. Every volume (nucleus) contains four individual promoters for each of the genes A, B, C and D. Each promoter can be repressed by the products of the three others with different affinities; this system of four mutually inhibiting genes represents the gap gene system in the early fly embryo, formed by the four genes *hb*, *kr*, *kni* and *gt*, and comprises its essential regulatory interactions. For combined repressive interactions, we employ OR-logic, i.e. whenever one of the three repressor sites is occupied expression of the gene is completely blocked. There is no competition for repressor sites on the promoters. In the unrepresed state the promoters exhibit constitutive protein production, i.e. no external activator signal is required. This deliberately mimics a situation in which activation of the genes is not provided by external morphogen gradients but by either an omnipresent master activator or auto-activation with a low activation threshold. Consequently, our model explicitly does not include morphogen gradients. As a simplifying assumption, we treat the whole production process, i.e. transcription, elongation and translation, as one step governed by a single rate β . Proteins however can form (homo)dimers and dedimerize again [81, 82], and only in their dimeric form they act as repressors. This is to ensure that antagonistic genes form bistable pairs for sufficiently strong mutual repression. Initially, all simulations are set up in a stripe pattern similar to the experimentally observed order in the embryo posterior, i.e. A-B-C-D-A [52, 48, 53]. This implies a fixed definition of “gene neighborhood” to which we refer throughout this paper: by nearest neighbors (NN) we mean the pairs (A, B), (B, C), etc., while the pairs (A, C) and (B, D) are considered next-nearest neighbors (NNN). A key ingredient of our model is that nearest-neighbor repression is weaker than repression between next-nearest neighbor domains (see “Parameter choice” in Methods). By default we pin the expression of A at the system boundaries, i.e. in nuclei on the two outermost rings of the cylinder the A promoter is irrepressible, and therefore constitutively produces A proteins. This is motivated by the fact that in the real *Drosophila* embryo the gene *Hb* is under strict control by the maternal morphogen *Bcd* throughout the anterior half [83], while in the posterior a second enhancer exposes *Hb* to positive regulation by the maternal terminal system [84, 85, 86]. We compare this system to a system in which there is no pinning and all nuclei are identical.

Simulations

To perform rare-event sampling of the spatially resolved system we integrate our “Gap Gene Gillespie” (GGG) simulator used in previous work [14, 57] with the NS-FFS scheme [56]. NS-FFS is used to monitor and process a progress coordinate written out by GGG at regular simulation interrupts, at which GGG trajectories are cloned and restarted in a way that sampling is enhanced in the direction of increased progress coordinate, i.e. towards pattern destruction.

Spatially resolved stochastic simulations (GGG)

In GGG, the model is implemented via the Stochastic Simulation Algorithm by Gillespie [87, 88] on a cylindrical 2D lattice of reaction volumes at constant distance $l = 8.5 \mu\text{m}$, with periodic boundary conditions in the circumferential direction of the array. An abstract graph of the reaction network that displays the set of reactions for any of the simulated promoters is shown in Figure S8 in the Supporting Text. Diffusive chemical species (patterning gene proteins and their dimers) hop between neighboring volumes via the next-subvolume method [89] which integrates diffusion into the Gillespie algorithm by annihilation of a species copy in the volume of origin and instantaneous insertion of that copy in a randomly chosen neighboring volume with a rate $k_{\text{diff}} = 4D_P/l^2$, where D_P corresponds to the protein diffusion coefficient. The source code of GGG with examples can be downloaded from <https://github.com/YetAnotherTomek/GGG>.

Forward flux sampling

We employ the recently developed non-stationary forward flux sampling (NS-FFS) method [56, 55, 90] to enhance stochastic sampling of system realizations that increase a (reaction) progress coordinate λ while retaining correct statistical weight. NS-FFS achieves this by branching off multiple child trajectories upon crossing predefined interfaces in undersampled regions of (λ, t) -space and pruning trajectories that cross interfaces in oversampled regions. The NS-FFS scheme aims at equilizing the flux of simulated trajectories in the reaction coordinate direction among the time bins. The rate of branching and pruning is calculated from the temporal trajectory crossing statistics collected during runtime. To that purpose the time domain is subdivided into equidistant time intervals. For a detailed account of the reweighting procedure we refer to [56].

Progress coordinates

The choice of a suitable progress coordinate is a critical step of the FFS technique. Here, we seek to enhance progress of the simulated patterns towards their destroyed state. The destruction events are in particular characterized by the disappearance of one of the partners within each of the strongly repressing gene pairs. Progress towards destruction thus is accompanied by increasing pair asymmetry, which can be quantified for each pair separately by the following two asymmetry factors:

$$\lambda_{AC} \equiv \max([A]_{tot}, [C]_{tot})/N \quad (15)$$

$$\lambda_{BD} \equiv \max([B]_{tot}, [D]_{tot})/N \quad (16)$$

where $N = [A]_{tot} + [B]_{tot} + [C]_{tot} + [D]_{tot}$ is the number of all proteins in the system. Based on this we define our progress coordinate, which increases whenever asymmetry among any of the pairs is augmented, via

$$\lambda \equiv \lambda_{AC} + \lambda_{BD} = [\max([A]_{tot}, [B]_{tot}) + \max([B]_{tot}, [D]_{tot})]/N. \quad (17)$$

Since NS-FFS features multi-dimensional reaction coordinates we compared our standard choice to a setup in which the two components λ_{AC} , λ_{BD} of the reaction coordinate λ are treated as two separate reaction coordinates with an own set of interfaces each. While an orthogonal pair of reaction coordinates captures the principal reaction paths in our system more accurately, the acquisition of crossing statistics is prolonged because of the increased number of bins in these simulations, and we did not find any substantial advantage of this choice in terms of branching behavior. We therefore preferred the standard definition.

Combination of simulation methods

In order to wrap NS-FFS around the GGG simulator we run GGG for a predefined simulation time $t_{GGG} = 60$ s. At the end of the simulation the reaction coordinates are calculated and passed on to the NS-FFS module, and the end state of the simulation is recorded. The NS-FFS module then determines whether an interface crossing has occurred and, if so, decides on whether the trajectory shall be branched or pruned. In case of branching NS-FFS will prompt $n_B \geq 1$ restarts of the GGG simulator with the recorded end state as initial condition, different random seeds and with new statistical weights. At each crossing and at measuring times spaced by a regular interval Δt the time, branch weight and reaction coordinate values are stored in a tree-like data structure that facilitates later analysis.

Trajectory trees are started from a standartized, regular-stripe initial condition passed to the first call of GGG. Propagation of the tree stops when all child branches have either reached the end of the time histogram or have been pruned. Subsequently a new tree is started with a different random seed. NS-FFS monitors the cumulative simulated time T_{cum} and terminates simulation when T_{cum} exceeds a predefined maximal simulation time T_{max} and the last trajectory tree has been propagated towards the end. Typically, $T_{max} = 3 - 7$ h and $T_{cum} = 2 - 5 \cdot 10^7$ s, which usually results in several thousand independent starts from the initial condition.

By default we start from an artificial pattern consisting of five non-overlapping stripes with rectangular profiles occupying an equal part of the total system length $L/5$ each and equal number of monomers (no dimers) in each nucleus close to the expected total copy numbers. We find that these initial patterns quickly relax towards typical metastable patterns, i.e. into the metastable main basin of attraction, which justifies our approach a posteriori.

Parameter choice

Repression

We are mainly concerned about the importance of distinct repression strength of nearest-neighbor (NN) as compared to next-nearest neighbor (NNN) interaction. We assume repressor binding-rates to be diffusion-limited via $k_{\text{on}}^{\text{R}} = 4\pi\sigma_{\text{R}}D_{\text{N}}$, where D_{N} is the *intracellular* diffusion constant and σ_{R} an effective target radius. Repression strength therefore is varied by changing the unbinding rates of the repressing dimers. The main parameter in our simulations is $\kappa = k_{\text{w}}^{\text{off}}/k_{\text{s}}^{\text{off}}$, the ratio between NN and NNN repressor off-rate. In this work only $k_{\text{w}}^{\text{off}}$ is varied, while $k_{\text{s}}^{\text{off}}$ is chosen sufficiently low to guarantee bistability between next-nearest neighbor genes, which is a precondition for the formation of individual stripe domains in the first place, see Table S1 in the Supporting Text. For $\kappa = 1$ NN and NNN repressive interactions are equally strong, while for large κ values NN repression is much weaker than NNN repression. In the “uncoupled limit” $\kappa \rightarrow \infty$ the two bistable pairs coexist without sensing each other. We do not consider cases with $\kappa < 1$.

Dimerization

We set the dimerization forward rate k_{on}^{D} to be equal to two times the diffusion-limited repressor binding rate, which is accounting for the fact that both reaction partners are diffusing. The dimerization backward rate is set via $k_{\text{off}}^{\text{D}} = k_{\text{on}}^{\text{D}}/V_{\text{N}}$ (V_{N} = nuclear volume) as in [91, 92, 93, 14] to ensure that at any moment most of the proteins are dimerized.

Production and degradation

In our model both monomers and dimers are degraded. This leads to a nontrivial dependence of the total copy number on production, degradation and (de)dimerization rates, as we discuss with more detail in [14]. Since we did not find any experimental reports of gap protein lifetimes, we chose equal monomeric (μ_{M}) and equal dimeric degradation rate (μ_{D}) for all genes and set these quantities to values that lead to a reasonable effective lifetime of the corresponding proteins of $t_{\text{eff}} \simeq 100$ s. The steady-state copy number is tuned via the production rate β . By default, we consider copy numbers as low as possible ($\simeq 15$) to minimize computational effort. The effect of increasing the average copy number is discussed in the “Discussion” section.

Geometry and internuclear transport

The choice of our geometric parameters, in particular of the lattice constant, is inspired by experimental measurements in the *Drosophila* embryo by Gregor et al. [1]. Information on the diffusion constants of proteins involved in early *Drosophila* patterning is scarce. The diffusion constant of the morphogen *Bcd* has been measured by several groups, yet its true value is still under debate [12, 94]. In our model we therefore set for all patterning proteins an effective internuclear diffusion constant $D_{\text{P}} = 1 \mu\text{m}^2/\text{s}$, which comprises both protein import/export and actual diffusion. This value is a reasonable cytoplasmic diffusion coefficient and well within the bounds reported for *Bcd*.

The simulated lattice is 40 nuclei long so that the total system length L roughly corresponds to the posterior 2/3 of the *Drosophila* embryo in cycle 14. To reduce computation effort we simulate a system with smaller circumference (8 nuclei) as compared to the living embryo. This is justified by the fact that for our standard diffusion constant D_{P} and effective protein lifetime μ_{eff} the diffusive correlation length $l_{\text{corr}} = \sqrt{D_{\text{P}}/\mu_{\text{eff}}}$ is ≤ 2 nuclei. A larger circumference therefore is not expected to introduce new features into the system, but might alter the timescales of expression boundary movement and domain desintegration. We discuss the effect of reduced system size on measured stability times in the “Discussion” section.

A complete overview of the specific numerical values of our model parameters is found in Table S1 of the Supporting Text.

Data analysis

Quantification of pattern stability

In order to analyse pattern stability we represent each simulated pattern as a point in $(\lambda_{AC}, \lambda_{BD})$ phase space. For every pattern simulation from time $t = 0$ until time $t = t_{end}$ the temporal sequence of these points corresponds to a trajectory in the $(\lambda_{AC}, \lambda_{BD})$ space. For each parameter choice and pinning scenario, we restarted the simulations with 6000 trajectories started from the relaxed initial patterns at $t = 0$; the trajectories ensemble is then further enriched by the branching process at the NS-FFS interfaces. Next, the trajectories are binned with the statistical weight assigned by NS-FFS, and then the histograms are normalized. As a result, we can identify a few distinct regions that accumulate probability. In order to formally define these regions we define rectangular boundaries that enclose accumulated probability regions corresponding to different types of patterns:

- the metastable main basin with five-stripe pattern:
 $R_S \equiv \{(\lambda_{AC}, \lambda_{BD}) | \lambda_{AC} \leq 0.45 \wedge \lambda_{BD} \leq 0.43\}$
- the basin in which either the A or C protein domain was lost:
 $R_{AC}^\dagger \equiv \{(\lambda_{AC}, \lambda_{BD}) | \lambda_{AC} > 0.45 \wedge \lambda_{BD} \leq 0.43\}$
- the basin in which either the B or D protein domain was lost:
 $R_{BD}^\dagger \equiv \{(\lambda_{AC}, \lambda_{BD}) | \lambda_{AC} \leq 0.45 \wedge \lambda_{BD} > 0.43\}$
- the basin in which either A or C and one of B or D were lost:
 $R^\ddagger \equiv \{(\lambda_{AC}, \lambda_{BD}) | \lambda_{AC} > 0.45 \wedge \lambda_{BD} > 0.43\}$

Note that the location of the regions slightly changes for different values of κ . We found that the above boundary definitions constitute a good compromise. For each basin we compute the fraction of total probability as a function of time by integrating the weights of trajectories that are within the basin at time t . We define the *pattern survival probability* to be the integrated probability in R_S at time t after initialization: $S(t) = \iint_{R_S} p(t) d\lambda_{AC} d\lambda_{BD}$. As expected, $S(t)$ displays roughly exponential decay behavior after a certain lag phase that can be attributed to initial relaxation. To obtain the *pattern destruction rate* k_D we fit a function $f(x) \equiv \exp(-k_D(t - t_{lag}))$ to $S(t)$. This only yields satisfactory results if the fitting range is adapted accordingly, i.e. only $S(t)$ values for $t > t_{lag}$ are taken into account. Since t_{lag} itself is a fitting parameter we adopted the following protocol: Starting from a value of t_{start} that is clearly in the relaxation regime we perform the fit on the interval $[t_{start}, t_{end}]$ where t_{end} is the largest time recorded. We then choose the fitted values k_D and t_{lag} for which $|t_{lag} - t_{start}|$ is minimal. From this we compute the pattern stability time (average time until pattern has lost one of the domains) via $\tau_D \equiv 1/k_D$. In most considered cases the patterns are very stable, i.e. k_D very small, and we can expand $S(t) \simeq 1 - k_D(t - t_{lag})$. As a control, we therefore also fitted $g(t) \equiv k_D(t - t_{lag})$ to $1 - S(t)$ for a fixed t_{lag} clearly in the exponential regime and obtained almost identical results.

Computation of average probability fluxes

To quantify which destruction pathways are dominant we computed the average fluxes J_{avg} into the regions of (partly) destroyed patterns. Here the average flux is defined as the average rate of increase in time of the fractional probability in the region and obtained by fitting a linear function $h(t) \equiv J_{avg}t + P_0$ to $P_R(t) \equiv \iint_R p(t) d\lambda_{AC} d\lambda_{BD}$ for $R \in \{R_{AC}^\dagger, R_{BD}^\dagger, R^\ddagger\}$ over the interval $[t_{start}, t_{end}]$ with t_{start} chosen such that $\partial_t P_R(t) \neq 0$ for $t > t_{start}$. P_0 depends on the particular choice of t_{start} and is discarded.

Computation of average flux velocities

The average local drift velocity and diffusion constant of the trajectories in the $(\lambda_{AC}, \lambda_{BD})$ phase space are computed by averaging displacements $\Delta\lambda_{AC(BD)} \equiv \lambda_{AC(BD)}(t + \Delta t) - \lambda_{AC(BD)}(t)$ and squared displacements $\Delta\lambda^2 \equiv \Delta\lambda_{AC}^2 + \Delta\lambda_{BD}^2$ on a two-dimensional lattice of bins covering the whole phase space. Displacements $\Delta\lambda_{AC(BD)}$ are assigned to the bin at $\vec{\lambda} \equiv (\lambda_{AC}, \lambda_{BD})$, i.e. we are averaging outgoing displacements and the averaged vector $\langle \vec{\Delta\lambda} \rangle(\vec{\lambda})$ therefore will represent the average velocity with which trajectories leave this bin. The local phase space diffusion constant is calculated as $D_\lambda(\vec{\lambda}) \equiv \frac{1}{4\Delta t} \left[\langle \Delta\lambda^2 \rangle(\vec{\lambda}) - \left(\langle \Delta\lambda_{AC} \rangle^2(\vec{\lambda}) + \langle \Delta\lambda_{BD} \rangle^2(\vec{\lambda}) \right) \right]$. This is done in the same way for other

combinations of phase space coordinates. The diffusion-drift decomposition is explained in more detail in the Supporting Text.

Computation of “pseudopotential” landscapes

The trajectory binning procedure used for computing the average flux velocities as described above was at the same time used for computing the “pseudopotential” $-\log(\tilde{p}(\vec{\lambda}))$. Herein $\tilde{p}(\vec{\lambda})$ is the local density calculated from the reweighed number of trajectories *leaving* the bin at $\vec{\lambda} = (\lambda_{AC}, \lambda_{BD})$, and smoothed afterwards by 2D median filtering over n_{filt} neighboring bins. For the 2D median filtering we used the `medfilt2` function from the MATLAB Image Processing Toolbox. We empirically chose $n_{\text{filt}} = 4$ as we found that this choice efficiently removes local spikes in $p(\vec{\lambda})$ without changing the overall shape of the landscape.

Perturbation experiments

Simulations starting from perturbed initial conditions were performed directly via the GGG simulator. First the systems were relaxed to representative states within the metastable basin for a simulated time of $t_{\text{relax}} = 30$ min. The final states of these runs then were post-modified according to the following two protocols:

1. “C expansion”: starting from mid-embryo the central C protein domain was expanded as follows: the configurations in the nuclei just posterior to mid-embryo were copied and used to overwrite configurations in the subsequent Δ rows in the axial (z -) direction of the cylinder. The original configurations were stored and for each nucleus at row $z_i > N_z/2 + \Delta$ (counting from the anterior) the configuration was overwritten by the original configuration at $z_i - \Delta$. The posterior-most nucleus was exempted from overwriting to preserve pinning.
2. “A expansion”: here the anterior A protein domain was enlarged at the expense of the C protein domain. To this purpose we applied the same copy-paste procedure as above starting from $z_i = 5$, however only nuclei up to mid-embryo ($z_i \leq N_z/2$) were overwritten by the original configurations at $z_i - \Delta$.

Δ quantifies the severity of perturbation. We found that $\Delta < 4$ results in changes to the pattern that were hard to distinguish from noise, while for $\Delta > 12$ perturbations were large enough to induce immediate pattern destruction with high probability. We therefore limited systematic tests to perturbations with $\Delta \in \{4, 8, 12\}$. Starting from the perturbed initial conditions simulations were continued for $t_{\text{sim}} = 20$ h and snapshots of the current configurations in all nuclei were written out with an acquisition interval of 10 min (simulated time). 10 samples starting from 10 different perturbed initial conditions were produced for each set of parameters.

In order to overcome the difficulties of boundary detection we quantified the motion of protein domains by tracking their center of mass (CoM) along the z -axis of the cylinder. For each considered gene G we define the CoM z_G as

$$z_G \equiv \frac{\int_z \int_r z G_{\text{tot}}(r, z) dr dz}{\int_z \int_r G_{\text{tot}}(r, z) dr dz} \quad (18)$$

where $G_{\text{tot}} = [G] + 2[G_2]$ is the total copy number. Since our system features two A domains we calculate z_A separately for the anterior (A_{ant}) and the posterior (A_{post}) part of the embryo by restricting z -integration adequately. While the CoM remains unchanged upon symmetric changes of the domain boundaries or global copy number increase, it is well-suited to indicate relaxations from the asymmetric perturbations that we apply. To find general trends in the time-evolution of the domains CoM trajectories were averaged over the 10 samples.

Effective reaction-diffusion dynamics of dimer expression

In this section we map the fully microscopic model defined in Fig. 1 onto the effective model from [54]. First, we postulate that the stochastic dynamics of gene expression studied in this paper corresponds

to the following effective dynamical equations:

$$\begin{aligned}\partial_t X_1(x, t) &= -\mu_M X_1(x, t) - k_{\text{on}}^D X_1^2(x, t) + k_{\text{off}}^D X_2(x, t) + \beta f(\{Y_2(x, t)\}_{Y \neq X}) , \\ \partial_t X_2(x, t) &= D_0 \partial_{xx} X_2(x, t) - (k_{\text{off}}^D + \mu_D) X_2(x, t) + k_{\text{on}}^D X_1^2(x, t) ,\end{aligned}\quad (19)$$

where $X, Y \in \{A, B, C, D\}$ denotes the expressed protein species, X_1, Y_1 are the concentrations of its monomer, and X_2, Y_2 are concentrations of its dimers. The synthesis and decay of dimers is described by rates k_{on} and k_{off} . Both, monomers and dimers degrade with rates μ_D and μ_M . In this system, only dimers are allowed to diffuse (with diffusivity D_0) and only monomers are primarily synthesized, with maximal production rate β and production kinetics described by function $f(\{Y_2(x, t)\}_{Y \neq X})$, which we specify later. However, we assume that in the absence of other dimers $Y_2 \neq X_2$ the production is active by default, so $f(\{0\}) = 1$. Since the system has cylindrical symmetry, we will treat the axis x as distinguished and treat the system as effectively one-dimensional.

The fact that the model defined by eqs. (19) involves monomers and dimers complicates its mapping onto the model in [54]. We therefore translate it into a simplified model, tracking the effective dynamics of dimers only. To this end, we will first determine the ratio between stationary concentrations \tilde{X}_1 and \tilde{X}_2 in the absence of other dimers ($Y_2(x, t) = 0$) and assuming system homogeneity. In this case, equations (19) turn into:

$$\begin{aligned}0 &= -\mu_M \tilde{X}_1 - k_{\text{on}}^D \tilde{X}_1^2 + k_{\text{off}}^D \tilde{X}_2 + \beta , \\ 0 &= -(k_{\text{off}}^D + \mu_D) \tilde{X}_2 + k_{\text{on}}^D \tilde{X}_1^2 .\end{aligned}\quad (20)$$

Solving for \tilde{X}_1 and \tilde{X}_2 , we obtain:

$$\begin{aligned}\tilde{X}_1 &= \frac{1}{2k_{\text{on}}^D} \left(-\frac{k_{\text{off}}^D + \mu_D}{\mu_D} + \sqrt{\frac{(k_{\text{off}}^D + \mu_D)^2}{\mu_D^2} \mu_M^2 + 4k_{\text{on}}^D \beta \frac{k_{\text{off}}^D + \mu_D}{\mu_D}} \right) , \\ \tilde{X}_2 &= \frac{\beta}{\mu_D} - \frac{\mu_M}{\mu_D} \tilde{X}_1 .\end{aligned}\quad (21)$$

We will now sum both equations in (19) to obtain

$$\partial_t (X_1(x, t) + X_2(x, t)) = D \partial_{xx} X_2(x, t) - \mu_D X_2(x, t) - \mu_M X_1(x, t) + \beta f(\{Y_2\}_{Y \neq X}) , \quad (22)$$

and approximate

$$X_1(x, t) \approx \frac{\tilde{X}_1}{\tilde{X}_2} X_2(x, t) . \quad (23)$$

In other words, we assume that $X_1(x, t)$ follows strictly $X_2(x, t)$. The advantage of this approximation is that it becomes exact in the stationary state. This procedure results in the following effective equation for $X_2(x, t)$:

$$\partial_t X_2(x, t) = D_X \partial_{xx} X_2(x, t) - \gamma_X X_2(x, t) + H_X f(\{Y_2\}_{Y \neq X}) , \quad (24)$$

where the rescaled constants are:

$$D_X = \frac{D_0}{1 + \frac{\tilde{X}_1}{\tilde{X}_2}} , \quad \gamma_X = \frac{\mu_D + \mu_M \frac{\tilde{X}_1}{\tilde{X}_2}}{1 + \frac{\tilde{X}_1}{\tilde{X}_2}} , \quad H_X = \frac{\beta}{1 + \frac{\tilde{X}_1}{\tilde{X}_2}} . \quad (25)$$

We can now specify the kinetics function. The microscopic dynamics is such that each gene X is produced, unless it is blocked by the biding of any other dimer to its repressor site on the promoter. In the averaged-out description, we expect that a sufficiently high concentration of free repressor particles effectively shuts down the production of X . Similarly to [54], we will assume that this transition is steep, so we can choose the functional form of the regulatory Hill function in (24) to have the overall shape of Heaviside step function:

$$f(\{Y_2\}_{Y \neq X}) = \theta \left(\sum_{Y \neq X} \epsilon_{XY} Y_2(x, t) - C_X \right) . \quad (26)$$

Finally, we relate the effective gene interaction constants ϵ_{XY} to microscopic parameters by the following reasoning: In the microscopic simulations, the attachment of Y_2 to the repressor site is described by the constant k_{on}^R and detachment by k_s^{off} or k_w^{off} . Assuming that the repressor production speed can be approximated by Michealis-Menten kinetics, with the repressor site acting like a “catalyst”, we know that

$$K_Y = k_{w,s}^{\text{off}}/k_{\text{on}}^R \quad (27)$$

where $k_{w,s}^{\text{off}}$ stands for either k_w^{off} or k_s^{off} , is the concentration of repressor dimers Y_2 at which the velocity of production of Y_1 is at the half of its maximal value. We postulate that at this point Y_2 effectively switches off the production of X , and we equate this point with reaching the threshold for production in (26). Hence, the following is satisfied:

$$\epsilon_{XY} K_Y - C_X = 0 . \quad (28)$$

Solving for ϵ_{XY} we obtain:

$$\epsilon_{XY} = \frac{C_X}{K_Y} . \quad (29)$$

We choose $C_X < 0$ to ensure that the production of X is active by default, in the absence of repressive dimers ($Y_2(x, t) = 0$). Since C_X is now present in every term in (26), we can factor it out and neglect. Taken together, and assuming that diffusion, degradation and production constants are the same for all genes, that is: $D_X = D$, $\gamma_X = \gamma$ and $H_X = H$ for all $X \in \{A, B, C, D\}$; the microscopic dynamics of gene expression mapped onto the effective model results in eq. (5).

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