## Part 1 - 8 points

I used Glimmer to predict the ORF and extract sequences from the *Halanaerobium* species sequence file halan.fasta. The species *Halanaerobium pravalens* was used as a reference organism with the file hprev genome.fasta.

1. Below is a screenshot of the .predict file created by running the glimmer commands. I also attached the file halan.predict.

```
>Halanaerobium sp. MDAL1, whole genome shotgun sequence
orf00001
              171
                       350
                            +3
                                  11.68
orf00003
              343
                      1626 +1
                                   8.96
orf00004
                                   6.58
             1629
                      4733 +3
orf00005
                      4971 -3
                                   8.13
             5786
halan.predict (END)
```

2. The glimmer code to produce the output above (halan.predict):

long-orfs hprev\_genome.fasta hprev.longorfs extract -t hprev\_genome.fasta hprev.longorfs > hprev.train build-icm -r hprev.icm < hprev.train glimmer3 -o50 -g110 -t30 halan.fasta hprev.icm halan

And the code to extract the sequences:

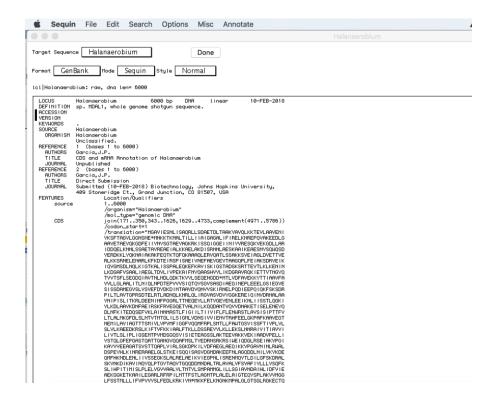
extract halan.fasta halan.predict > halan.glimmer

3. The DNA sequence of the first ORF in FASTA format (I also attached halan.glimmer with all of the ORF sequences):

>orf00001 171 350 len=180

ATGGGGGCAGTAATTGAAAGTAATTTAATTTCGGCTCAGAGATTGTTAAGTGATGCAGAA
ACAGATTTAACTGCTGCAAAATATGCCGTGCAGTTAAAAAAGACAGAAGTTTTGGCTGCA
GTAGAAAATATATATAAGAGCTTTACTGCAGGAGTATTAGGAGGTAATAGTAATGAATAA

4. Below is a screenshot of only the CDS annotation in Sequin (I also attached the generated GeneBank file with both CDS and mRNA annotated):



FGENESB did not contain *Halanaerobium prevalens* to use as a reference organism, so I looked up *Halanaerobium* in the NCBI Taxonomy Browser to find the closest similar organism to use in FGENESB as a comparison. I decided to use *Halobacterium* as my comparison organism. I also tried using Bacteria Generic and got the same result. Below is the result of the FGENESB prediction, which I also attached in a text file.



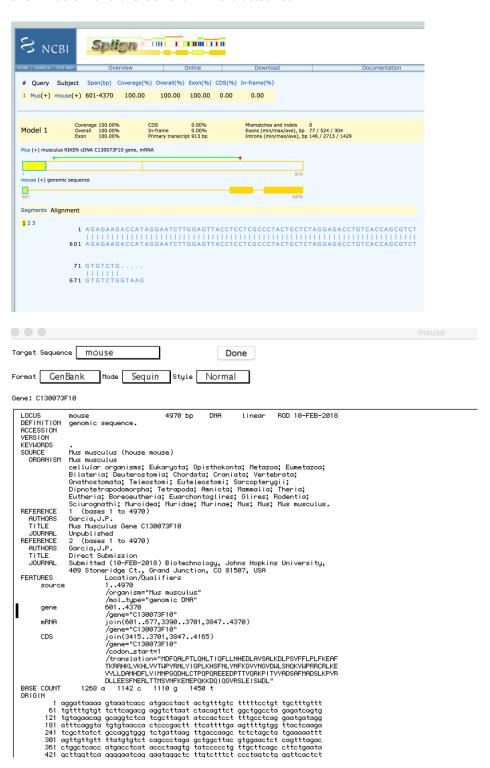
5. Below is a screenshot of the final Sequin file that includes CDS and mRNA annotations.



6. In the screenshots above you can see that FGENESB and Glimmer predicted almost the same coding regions, except for one spot where FGENESB predict the first CDS region to start at 3 and Glimmer predicted it to start at 171.

## Part 2 - 4 points

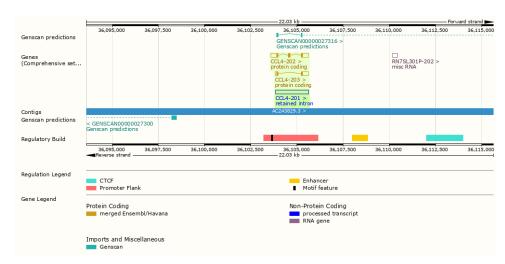
1 & 2. I ran Splign to align the mouse cDNA sequence to the genomic sequence provided. The results of Splign are shown here. One CDS was found with 3 exons. The annotated sequence in GeneBank is also shown below and the GeneBank file is attached.



## Part 3 - 12 points

1. In the human CCL4, the transcript described on NCBI gene contains all three exons, the first including the 5' UTR, the second being only a coding sequence, and the third containing the 3' UTR. This primary transcript encodes a protein 92aa long, while the second transcript on Ensembl skips the middle exon, only includes the first and third exons and encodes a shorter protein of 52aa. The third transcript is a retained intron transcript and does not encode a protein.

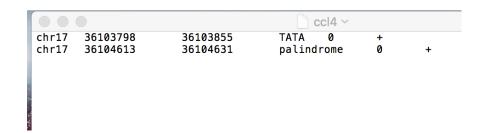
2.

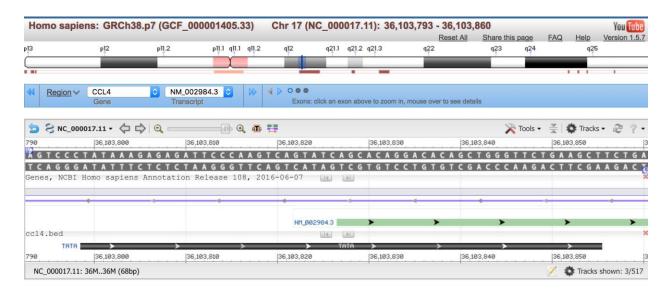


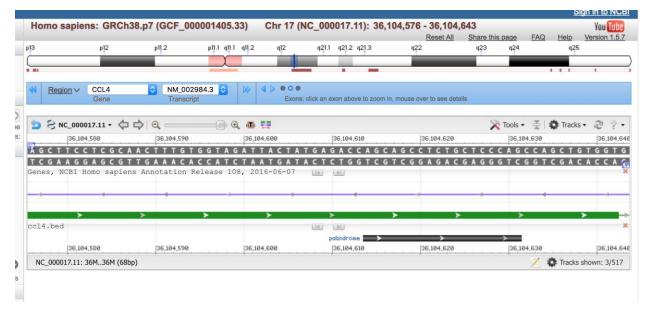
3. The gene encodes a protein that contains a Chemokine interleukin-8-like domain at position 33-88aa as can be seen in the image from Ensembl below. It is found in the primary transcript of CCL4-202, but not in either of the alternative transcripts CCL4-201 and CCL4-203.



4. I created a BED file containing the TATA box and the DNA location of the palindromic sequence. Below is a screenshot and I attached the file to my submission. Below are two screenshots of the BED file loaded into Variation Viewer, one with the TATA Box location and one with the palindrome location.







## Part 4 - 10 points

Search OMIM.org for "huntington's disease". The first five entires all have this or a similar phrase in the title. Record the five identifiers (six-digit numbers) of those five records. The corresponding biomaRt filter name for these identifiers is "mim\_morbid". Use biomaRt to retrieve two tables with the following attributes, limiting to the five MIM values you found:

```
huntingtons <- c("143100", "613004", "604802", "606483", "603218")
huntingtons_genes <- getBM(attributes=c("entrezgene", "hgnc_symbol", "ensembl_gene_id"),
            filters="mim morbid accession", values=huntingtons, mart=ensembl)
huntingtons_genes
  entrezgene hgnc_symbol ensembl_gene_id
1
2
         9096
                       TBX18 ENSG00000112837
                         HTT ENSG00000197386
         3064
3
         5621
                        PRNP ENSG00000171867
huntingtons transcripts <- getBM(attributes=c("hgnc symbol", "ensembl gene id",
"ensembl transcript id"),
             filters="mim_morbid_accession", values=huntingtons, mart=ensembl)
huntingtons transcripts
```

hgnc\_symbol ensembl\_gene\_id ensembl\_transcript\_id 1 TBX18 ENSG00000112837 ENST00000330469 TBX18 ENSG00000112837 ENST00000606784 2 3 4 ENST00000369663 TBX18 ENSG00000112837 TBX18 ENSG00000112837 ENST00000607343 5 TBX18 ENSG00000112837 ENST00000606521 6 7 8 9 TBX18 ENSG00000112837 ENST00000606325 ENST00000606621 TBX18 ENSG00000112837 HTT ENSG00000197386 ENST00000355072 HTT ENSG00000197386 ENST00000506137 10 HTT ENSG00000197386 ENST00000512909 HTT ENSG00000197386 11 ENST00000510626 12 HTT ENSG00000197386 ENST00000509618 13 HTT ENSG00000197386 ENST00000513639 14 HTT ENSG00000197386 ENST00000513326 <u>1</u>5 ENST00000509043 HTT ENSG00000197386 ENST00000502820 16 HTT ENSG00000197386 17 HTT ENSG00000197386 ENST00000509751 18 HTT ENSG00000197386 ENST00000512068  $\overline{19}$ HTT ENSG00000197386 ENST00000513806 20 HTT ENSG00000197386 ENST00000508321 21 22 23 PRNP ENSG00000171867 ENST00000379440 PRNP ENSG0000171867 ENST00000430350 PRNP ENSG0000171867 ENST00000424424

PRNP ENSG00000171867

Because we included the Ensembl Transcript Id in the second query, the table lists each gene id and each transcript for each gene. For example, gene ENSG00000112837 has 7 transcripts with the transcript ids shown in the third column.

ENST00000457586