- A Bayesian Model of the DNA Barcode Gap
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- 6 Running Title:

7 Abstract

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₁ 1 Introduction

Since its inception over 20 years ago, DNA barcoding (Hebert et al., 2003a,b) has emerged 12 as a robust method of specimen identification and species delimitation across myriad 13 taxonomic groups which have been sequenced at short, standardized gene regions like 5'-COI 14 for animals. However, the success of the approach depends crucially on two important factors: 15 (1) the availability of high-quality specimen records found in public reference sequence 16 databases such as the Barcode of Life Data Systems (BOLD) Ratnasingham and Hebert 17 (2007), and (2) the establishment of a DNA barcode gap — the idea that the maximum 18 genetic distance observed within species is much smaller than the minimum degree of marker 19 variation found among species (Meyer and Paulay, 2005; Meier et al., 2008). Early work has demonstrated that the presence of a DNA barcode gap hinges strongly on extant levels of 21 species haplotype diversity gauged from comprehensive specimen sampling at wide geographic and ecological scales. Despite this, many taxa lack adequate separation in their pairwise intraspecific and interspecific genetic distances, thereby compromising rapid matching of unknown samples to expertly-validated references.

Recent work has argued that DNA barcoding, in its current form, is lacking in statistical rigor, calling into question the existence of a true species' DNA barcode gap Phillips et al. (2022). To support this notion, novel nonparametric locus-specific metrics based on the multispecies coalescent were recently outlined and shown to hold strong promise when applied to *Agabus* diving beetles Phillips et al. (2024). The metrics quantify the extent of asymmetric directionality of proportional genetic distance distribution overlap/separation for

species within well-sampled genera based on a straightforward distance count. Values of the
metrics close to zero suggest the existence of DNA barcode gaps, whereas values near one lend
credence for the absence of gaps. However, what appears to be missing is an unbiased way
to compute the statistical accuracy of the recommended estimators arising through problems
inherent in frequentist maximum likelihood estimation for discrete probability distributions
having bounded support. To this end, here, a Bayesian model of the DNA barcode gap
coalescent is introduced to rectify such issues. The model allows accurate estimation of
posterior means, posterior standard deviations, posterior quantiles, and credible intervals for
the metrics given datasets of intraspecific and interspecific genetic distances for species of
interest.

⁴² 2 Methods

⁴³ 2.1 DNA Barcode Gap Metrics

Recently, Phillips et al. (2024) proposed novel nonparametric maximum likelihood
estimators (MLEs) of proportional overlap/separation between intraspecific and interspecific
pairwise genetic distance distributions for a given species (x) to aid assessment of the DNA
barcode gap as follows:

$$p_x = \frac{\#\{d_{ij} \ge \min(d_{XY})\}}{\#\{d_{ij}\}} \tag{1}$$

$$q_x = \frac{\#\{d_{XY} \le \max(d_{ij})\}}{\#\{d_{XY}\}} \tag{2}$$

where d_{ij} and d_{XY} are distances within and among species, respectively, and the notation #
reflects a count. Distances are easily computed from a model of DNA sequence evolution,
such as p distance. Similar expressions (denoted p'_x and q'_x) for nearest neighbour species
were also given (see Phillips et al. (2024)), in which d_{XY} included only interspecific distances

between the species of interest and its closest neighbouring species. If a focal species is found to have multiple nearest neighbours, then the species possessing the smallest average 53 pairwise interspecific distance is used. While these schemes differ considerably from the usual definition of the DNA barcode gap laid out by Meyer and Paulay (2005) and Meier et al. (2008), they more accurately account for species' coalescence histories inferred from contemporaneous DNA sequences. such as interspecific hybridization/introgression events (Phillips et al., 2024). Note, distances (and hence the metrics) are constrained to the closed interval [0, 1]. Values of the estimators obtained from equations (1) and (2) close to or equal to zero give evidence for separation between intraspecific and interspecific genetic distance 60 distributions; that is, values suggest the presence of a DNA barcode gap. Conversely, values 61 near or equal to one give evidence for distribution overlap; that is, values likely indicate the 62 absence of a gap.

64 2.2 A Bayesian Implementation

A major criticism of large sample (frequentist) theory is that it relies on asymptotic properties of the MLE (which is assumed to be a fixed but unknown quantity), such as estimator normality and consistency. This problem is especially pronounced in the case of binomial proportions. The estimated Wald SE of the sample proportion, is given by

$$\widehat{SE[\hat{p}]} = \sqrt{\frac{\hat{p}(1-\hat{p})}{n}},\tag{3}$$

where $\hat{p} = \frac{Y}{n}$ is the MLE, Y is the number of successes $(Y = \sum_{i=1}^{n} y)$ and n is the number of trials. However, the above formula is problematic for several reasons. First, Equation (3) makes use of the Central Limit Theorem (CLT); thus, large sample sizes are required for reliable estimation. When few observations are available, SEs will be large and inaccurate, leading to low statistical power. Further, resulting interval estimates could span values less than zero or greater than one, or have zero width, which is practically meaningless. Second, when proprtions are exactly equal to zero or one, resulting SEs will be exactly zero,

rendering Equation (3) completely uselesss. In the context of the proposed DNA barcode gap metrics, values obtained at the boundaries of their support are often encountered. Therefore, reliable calculation of SEs is not feasible. Given the importance of sufficient sampling of 78 species genetic diversity for DNA barcoding initiatives, a different statistical estimation 79 approach is necessary. Bayesian inference offers a natural path forward in this regard since it allows for direct specification of prior beliefs concerning unknown model parameters and 81 permits the seamless propagation of uncertainty, when data is lacking, through integration with the likelihood function. As a consequence, Bayesian models are much more flexible 83 and generally more easily interpretable compared to frequentist approaches since direct 84 probability statements can be made. 85

$_{\scriptscriptstyle 6}$ 2.3 The Model

Counts, y, of overlapping genetic distances (as expressed in the numerator of Equations 87 (1) and (2)) are treated as binomially distributed with expectation $\mathbb{E}[Y] = k\theta$, where $k = \{N, M, C\}$ are total counts of intraspecific, interspecific, and combined genetic distances for a target species, and $\theta = \{p_x, q_x, p_x^{'}, q_x^{'}\}$. The metrics encompassing θ are presumed to follow a beta (α, β) distribution, which is a natural choice of prior on probabilities. 91 Such a scheme is quite convenient since the beta distribution is conjugate to the binomial distribution. Thus, the posterior distribution is also beta distributed. Parameters were given 93 an uninformative Beta(1, 1) prior, which is equivalent to a standard uniform (Uniform(0, 1)) prior since it places equal probability on all parameter values within its support. As a 95 result, the posterior is Beta(Y+1, n-Y+1), which has expected value $\mathbb{E}[Y] = \frac{Y+1}{n+2}$ and variance $\mathbb{V}[Y] = \frac{(Y+1)(n-Y+1)}{(n+2)^2(n+3)}$. In general however, when possible, it is always advisable to incorporate prior information, even if only weak, rather than simply imposing complete ignorance in the form of a flat prior distribution. The full univariate Bayesian model is thus 99 given by 100

$$\begin{aligned} y_{\text{lwr}} &\sim \text{Binomial}(N, p_{\text{lwr}}) \\ y_{\text{upr}} &\sim \text{Binomial}(M, p_{\text{upr}}) \\ y_{\text{lwr}}' &\sim \text{Binomial}(N, p_{\text{lwr}}') \\ y_{\text{upr}}' &\sim \text{Binomial}(C, p_{\text{upr}}') \\ \end{aligned} \tag{4}$$
$$p_{\text{lwr}}, p_{\text{upr}}, p_{\text{lwr}}', p_{\text{upr}}' &\sim \text{Beta}(1, 1) \end{aligned}$$

given sufficient data, the choice of prior distribution becomes less important since the posterior will be directly proportional to the likelihood.

The model was fitted using the Stan probabilistic programming language (Carpenter et al., 103 2017) framework for Hamiltonian Monte Carlo (HMC) sampling. Four chains were run for 104 2000 iterations each in parallel across four cores with random parameter initializations. 105 Within each chain, a total of 1000 samples was discarded as warmup (i.e., burnin) to reduce 106 dependence on starting conditions. Further, 1000 post-warmup draws were utilized per chain. 107 Each of these reflect default MCMC settings in Stan. Since the DNA barcode gap metrics 108 often attain values either very close to zero or very near one, in addition to more intermediate 109 values, a $\text{Beta}(\frac{1}{2}, \frac{1}{2})$ prior, which is U-shaped symmetric and places greater probability density 110 at the extremes of the distribution, while still allowing for variability in parameter estimates 111 within intermediate values along its domain, was also attempted. However, this resulted in 112 several divergent transitions imposed by complex geometry of the posterior space, despite 113 remedies to resolve them, such as lowering the step size of the HMC sampler. Note that this 114 prior is Jeffreys' prior, which is proportional to the square root of the Fisher information and 115 has several desirable statistical properties, most notably invariance to reparameterization. 116

- 117 3 Results
- 118 4 Discussion
- 5 Conclusion

Supplementary Information

Information accompanying this article can be found in Supplemental Information.pdf.

122 Data Availability Statement

- Raw data, R, and Stan code can be found on GitHub at:
- https://github.com/jphill01/Bayesian-DNA-Barcode-Gap-Coalescent.

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135 Conflict of Interest

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137 Author Contributions

JDP wrote the manuscript, wrote R and Stan code, approved all developed code as well as analysed and interpreted all experimental results.

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