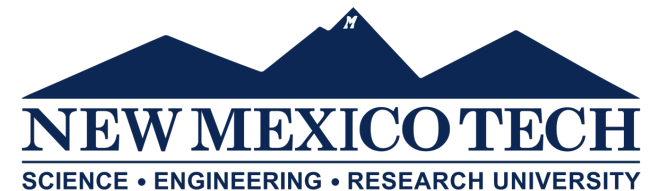


Exploring conservation genomic data in R

Josh Jahner
jpjahner@gmail.com



Outline of today's topics

- 1) Big questions in conservation genetics
- 2) Types of conservation genetic data
- 3) Simulating data with the beta distribution
- 4) Exploring conservation genetic analyses

Genetic diversity and inbreeding

FROM THE COVER

MOLECULAR ECOLOGY
RESOURCES WILEY

Genomics-informed captive breeding can reduce inbreeding depression and the genetic load in zoo populations

Samuel A. Speak^{1,2,3}  | Thomas Birley¹ | Chiara Bortoluzzi^{4,5} | Matthew D. Clark^{2,6} |
Lawrence Percival-Alwyn⁷ | Hernán E. Morales⁸  | Cock van Oosterhout¹ 

Identifying adaptive genetic variation


Received: 30 January 2024 | Revised: 23 April 2024 | Accepted: 30 April 2024

DOI: 10.1111/mec.17383

ORIGINAL ARTICLE

MOLECULAR ECOLOGY WILEY

Natural selection on feralization genes contributed to the invasive spread of wild pigs throughout the United States

Niek W. G. Barmantlo^{1,2}  | Patrick G. Meirmans²  | William H. Stiver³ |
Joseph G. Yarkovich³ | Blake E. McCann⁴ | Antoinette J. Piaggio⁵ | Dominic Wright⁶  |
Timothy J. Smyser⁵  | Mirte Bosse^{1,7} 

Population structure and connectivity



Molecular Ecology

WILEY

MOLECULAR ECOLOGY

ORIGINAL ARTICLE OPEN ACCESS

Shared Dispersal Patterns but Contrasting Levels of Gene Flow in Two Anadromous Salmonids Along a Broad Subarctic Coastal Gradient

Raphaël Bouchard^{1,2}  | Charles Babin^{1,2} | Eric Normandeau³ | Amanda Xuereb^{1,2} | Félix Boulanger⁴ | Angela Coxon⁴ | Sanford Diamond⁵ | Robert Fireman⁵ | John Lameboy⁵ | Natasha Louttit^{4,5} | George Natawapineskum⁵ | Derek Okimaw⁵ | Dante Torio⁶ | Stephanie Varty⁵ | Jean-Sébastien Moore^{1,2} | Dylan Fraser⁷  | Louis Bernatchez^{1,2}

Population size estimation










Evolutionary Applications

WILEY

Evolutionary Applications 




ORIGINAL ARTICLE **OPEN ACCESS**

Estimating the Effective Size of European Wolf Populations

Joachim Mergeay^{1,2}  | Sander Smet² | Sebastian Collet³  | Sabina Nowak⁴  | Ilka Reinhardt⁵  | Gesa Kluth⁵ |
Maciej Szewczyk⁶  | Raquel Godinho⁷  | Carsten Nowak³  | Robert W. Mysłajek⁴  | Gregor Rolshausen³ 

Demographic history reconstruction

Whole Genomes Inform Genetic Rescue Strategy for Montane Red Foxes in North America

Cate B. Quinn ^{1,2,3,*} Sophie Preckler-Quisquater,¹ Michael R. Buchalski ²
Benjamin N. Sacks ^{1,4}

Species delimitation



Current Biology

Report

Comparative species delimitation of a biological conservation icon

Ava Ghezelayagh,^{1,2,13,14,*} Jeffrey W. Simmons,³ Julia E. Wood,¹ Tsunemi Yamashita,⁴ Matthew R. Thomas,⁵
Rebecca E. Blanton,⁶ Oliver D. Orr,¹ Daniel J. MacGuigan,⁷ Daemin Kim,¹ Edgar Benavides,^{8,9} Benjamin P. Keck,¹⁰
Richard C. Harrington,^{1,11} and Thomas J. Near^{1,12}

Hybridization and admixture




Evolution Letters, 2025, 9(1), 1–12

<https://doi.org/10.1093/evlett/qrae057>

Advance access publication 19 October 2024

Letter

Genetic structure and common ancestry expose the dingo-dog hybrid myth

Andrew R. Weeks^{1,2} , Peter Kriesner¹, Nenad Bartonicek^{1,†}, Anthony van Rooyen¹, Kylie M. Cairns³, Collin W. Ahrens¹

Assisted gene flow, genetic rescue, translocations





Evolutionary Applications

WILEY

Evolutionary Applications 

ORIGINAL ARTICLE **OPEN ACCESS**

Genetic Rescue of the Dinaric Lynx Population: Insights for Conservation From Genetic Monitoring and Individual-Based Modelling

Elena Pazhenkova^{1,2}  | Matej Bartol³ | Barbara Boljte^{1,2} | Urša Fležar¹ | Andrea Gazzola⁴ | Tomislav Gomerčič⁵ | Marjeta Konec^{1,2} | Ivan Kos¹ | Miha Krofel¹ | Jakub Kubala⁶ | Ladislav Paule⁶  | Mihai Pop⁴ | Hubert Potočník¹ | Barbara Promberger⁷ | Robin Rigg^{1,8} | Teodora Sin⁴  | Magda Sindičić⁵ | Vedran Slijepčević⁹ | Astrid Vik Stronen^{1,2,10}  | Ira Topličanec⁵ | Tomaž Skrbinšek^{1,2}

Genetic monitoring

Received: 26 September 2023 | Revised: 1 December 2023 | Accepted: 19 January 2024

DOI: 10.1002/ece3.10934

RESEARCH ARTICLE

Ecology and Evolution

Open Access

WILEY

Evaluating genotyping-in-thousands by sequencing as a genetic monitoring tool for a climate sentinel mammal using non-invasive and archival samples

Kate E. Arpin¹ | Danielle A. Schmidt¹ | Bryson M. F. Sjodin¹ | Anthony L. Einfeldt² |
Kurt Galbreath³ | Michael A. Russello¹ 

Outline of today's topics

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Mitochondrial DNA (Sanger sequencing)

Individual 1
ACTTTGAGCTTTCTGA ...
ACTCTGAGCTTTCTGA ...

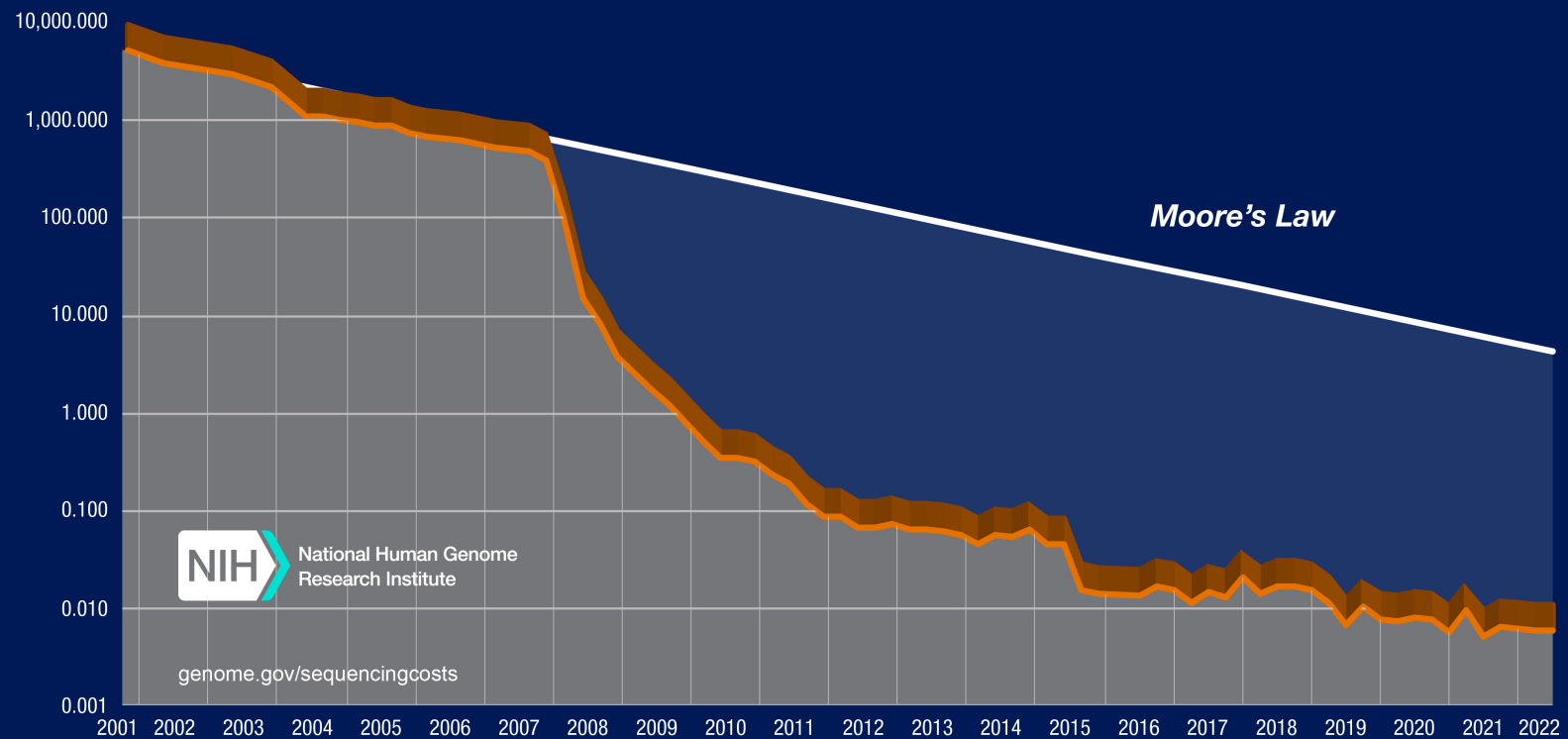
Individual 2
ACTCTGGGCTATCTGA ...
ACTCTGGGCTATCTGA ...

Microsatellites

(aka: simple sequence repeats [SSRs])

Individual 1	AGGAGGAGG AGG AGG AGG AGG AGG AGG AGG	10 repeats
	AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG	12 repeats
Individual 2	AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG	14 repeats
	AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG AGG	14 repeats

Cost per Raw Megabase of DNA Sequence



Reduced representation sequencing

Section of a chromosome



Sequences
from one cut site

CCATGGIGTAATCACCCCTG
CCATGGGGTAATCACCCCTG
CCATGGIGTAATCACCCCTG
CCATGGIGTAATCACCCCTG
CCATGGGGTAATCACCCCTG
CCATGGIGTAATCACCCCTG
CCATGGGGTAATCACCCCTG
CCATGGGGTAATCACCCCTG



Single nucleotide
polymorphism (SNP)

Reduced representation sequencing

Section of a chromosome

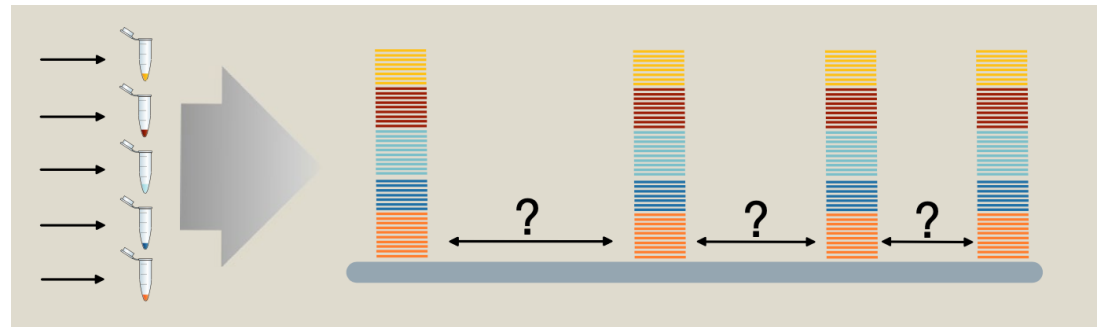


Sequences
from one cut site

CCATGGTGTAAATCACCCCTG
CCATGGGTAATCACCCCTG
CCATGGTGTAAATCACCCCTG
CCATGGTGTAAATCACCCCTG
CCATGGGTAATCACCCCTG
CCATGGTGTAAATCACCCCTG
CCATGGGTAATCACCCCTG
CCATGGGTAATCACCCCTG
CCATGGGTAATCACCCCTG

↑
Single nucleotide
polymorphism (SNP)

RADseq approaches



Reduced representation sequencing

Section of a chromosome

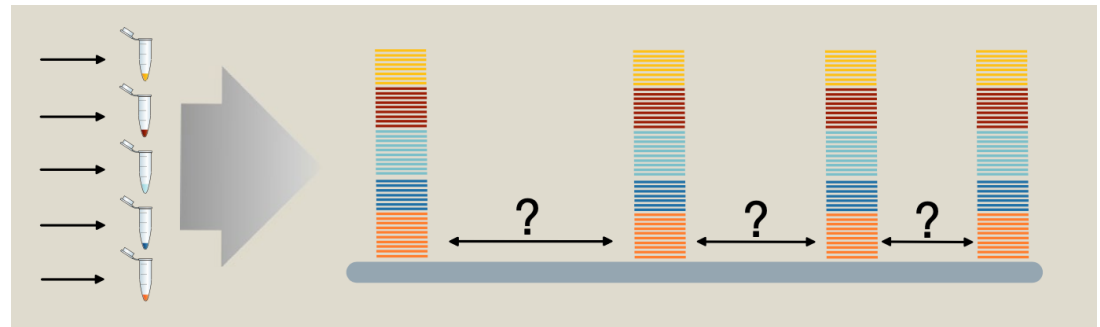


Sequences
from one cut site

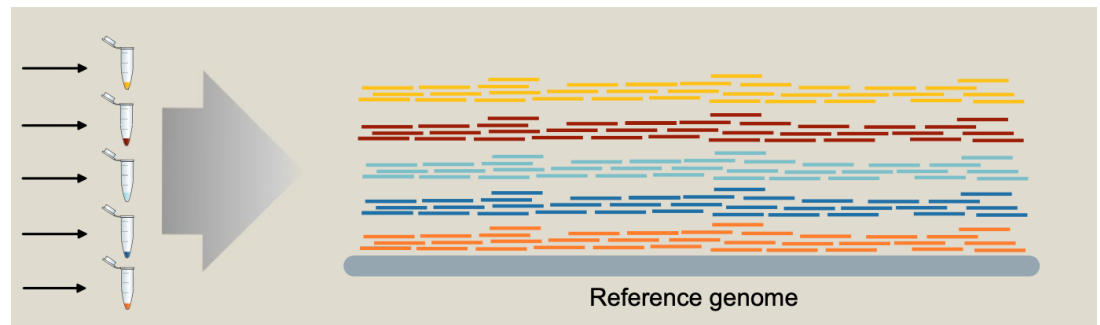
CCATGGT**I**GTAATCACCCCTG
CCATGG**G**GTAATCACCCCTG
CCATGGT**I**GTAATCACCCCTG
CCATGGT**I**GTAATCACCCCTG
CCATGG**G**GTAATCACCCCTG
CCATGGT**I**GTAATCACCCCTG
CCATGG**G**GTAATCACCCCTG
CCATGG**G**GTAATCACCCCTG

↑
Single nucleotide
polymorphism (SNP)

RADseq approaches



Whole genome resequencing



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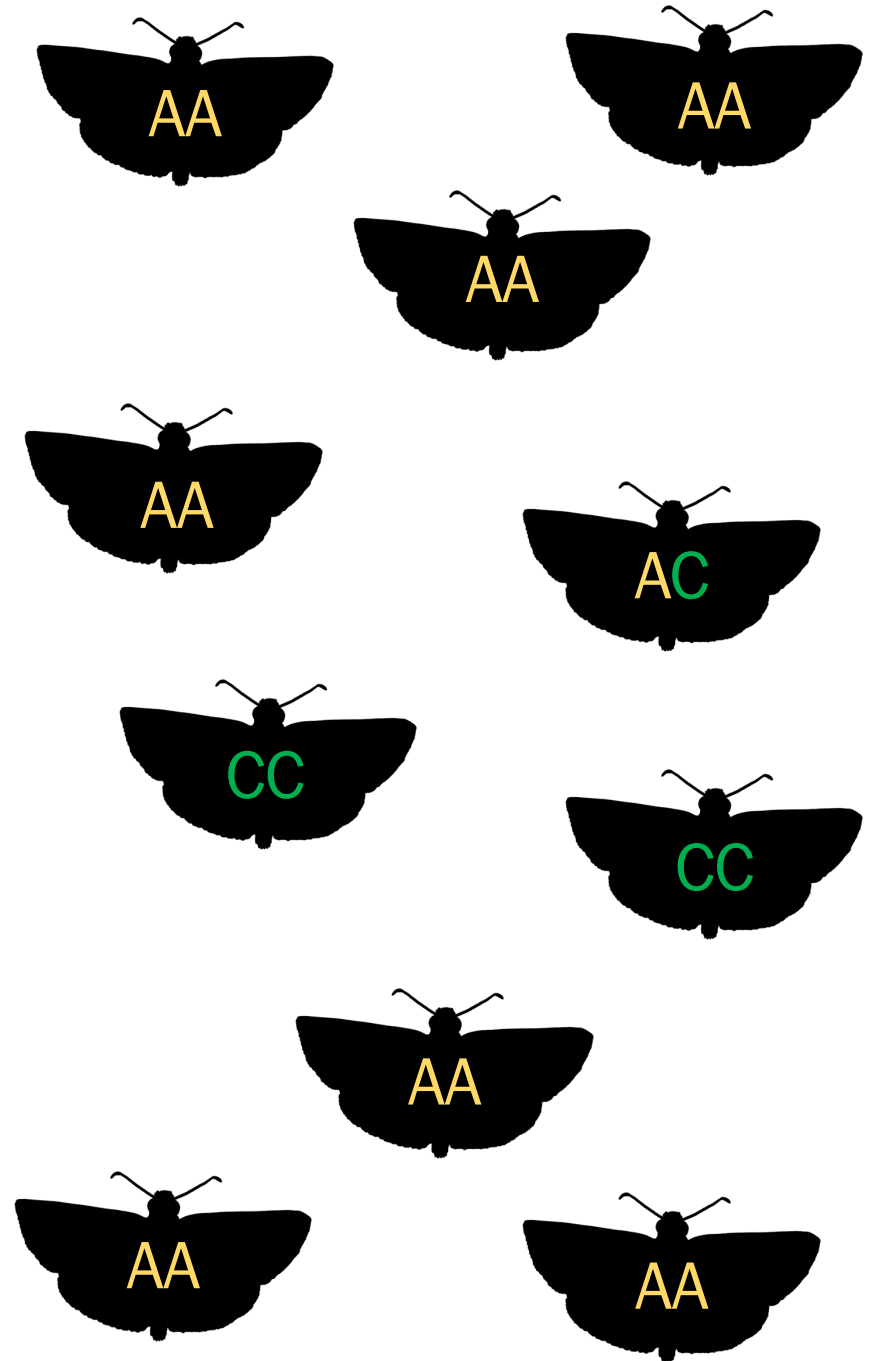
Find the R scripts at:

https://github.com/jpjahner/congen_workshop

01_beta_simulate.R

02_explore_analyses.R

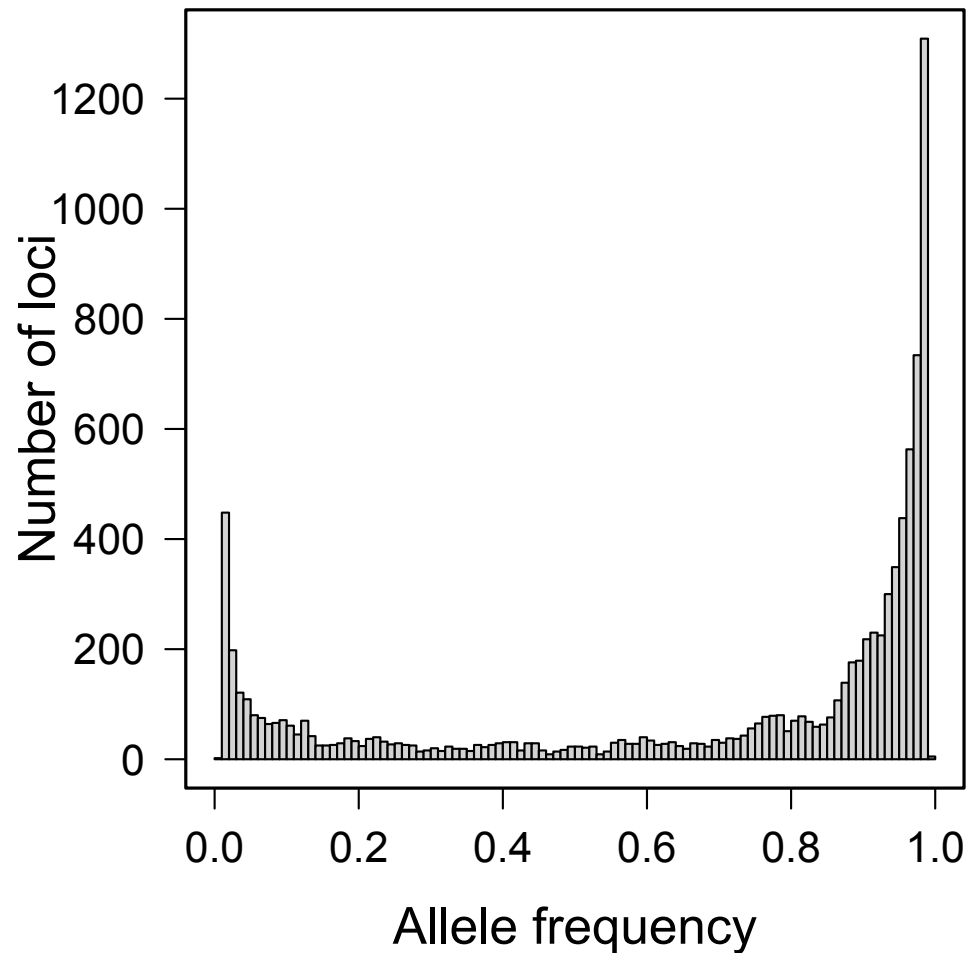
Allele frequency of A:
 $15 / 20 = 0.75$

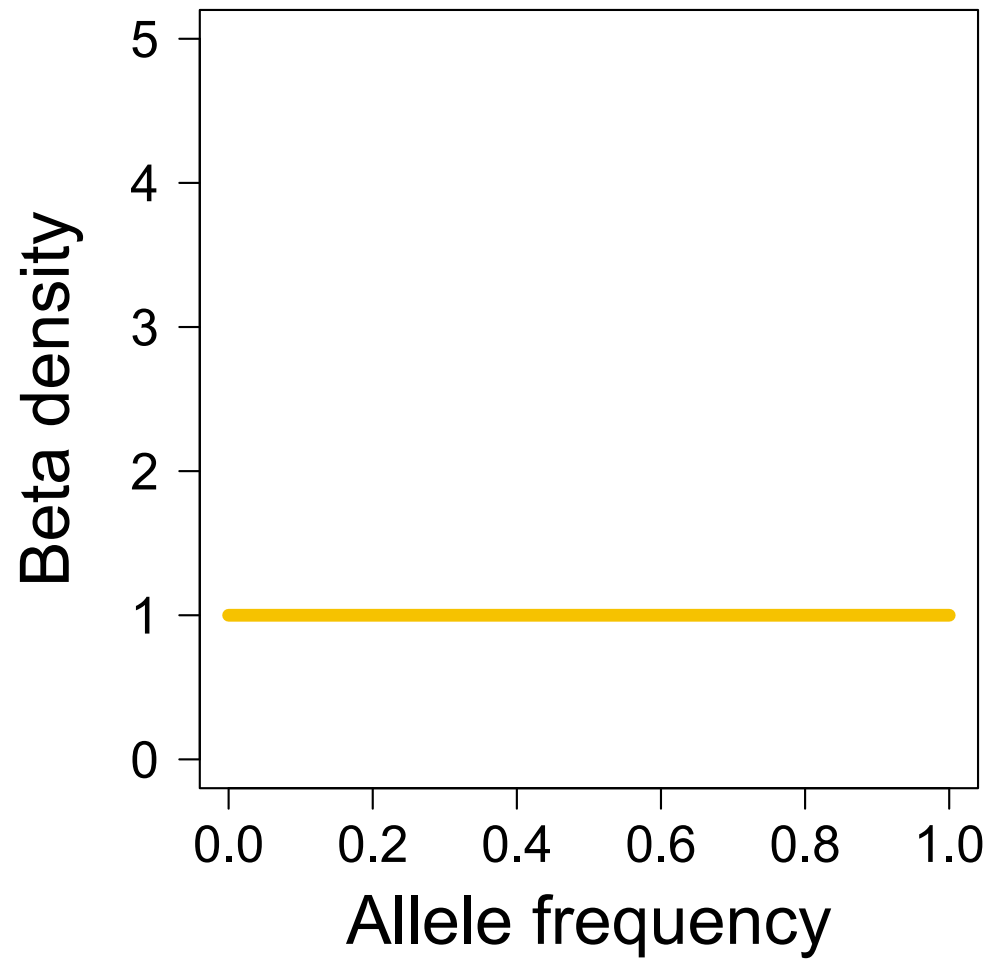


Influence of dams on sauger population structure and hybridization with introduced walleye

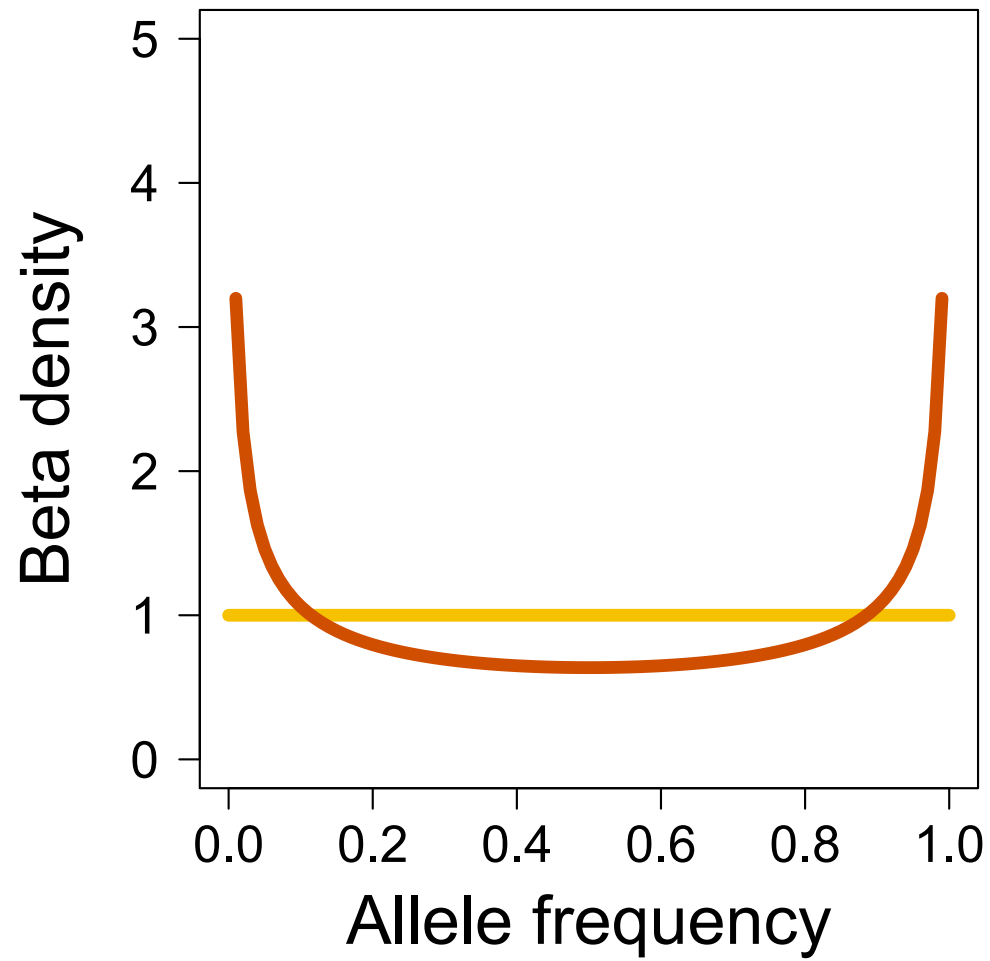
William C. Rosenthal^{1,2}  | Elizabeth G. Mandeville^{1,3} | Ashleigh M. Pilkerton^{2,4,5}  |
Paul C. Gerrity⁶ | Joseph A. Skorupski⁶ | Annika W. Walters^{2,4,5,7}  |
Catherine E. Wagner^{1,2}

Site Frequency Spectrum (SFS)

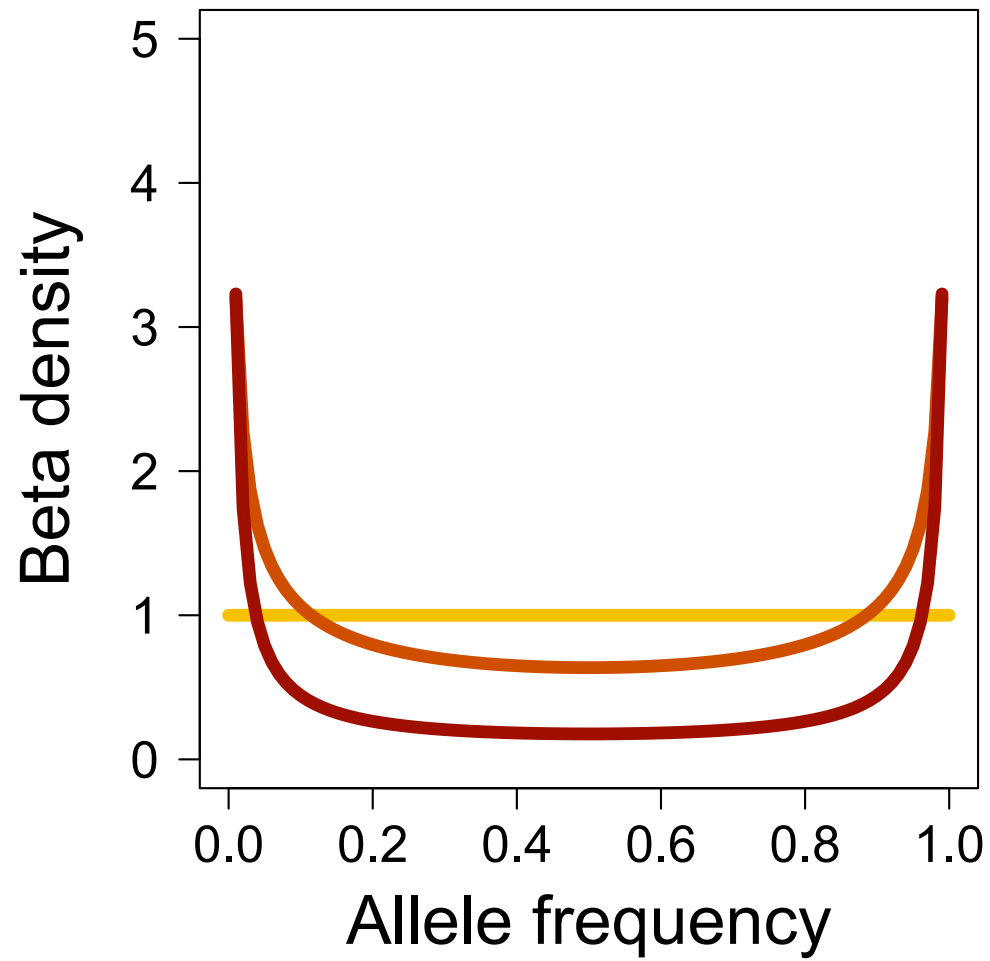




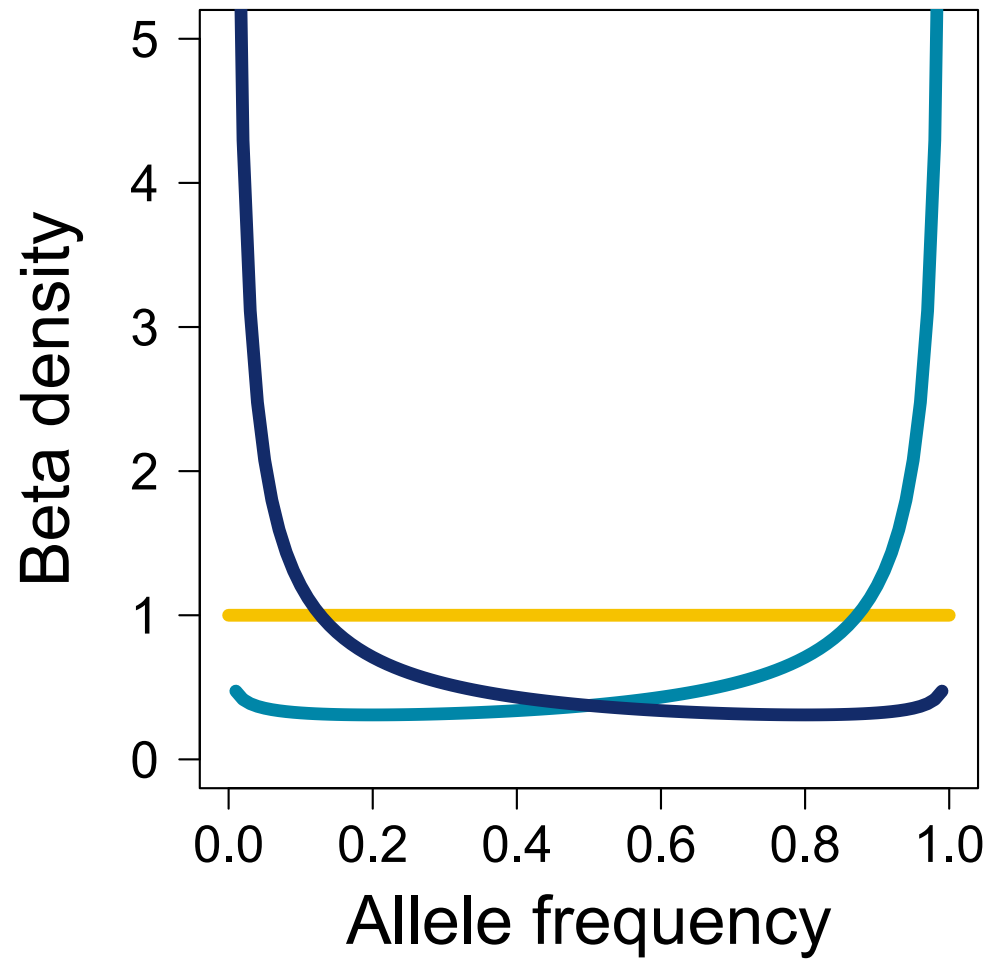
$$\alpha = 1 \quad \beta = 1$$



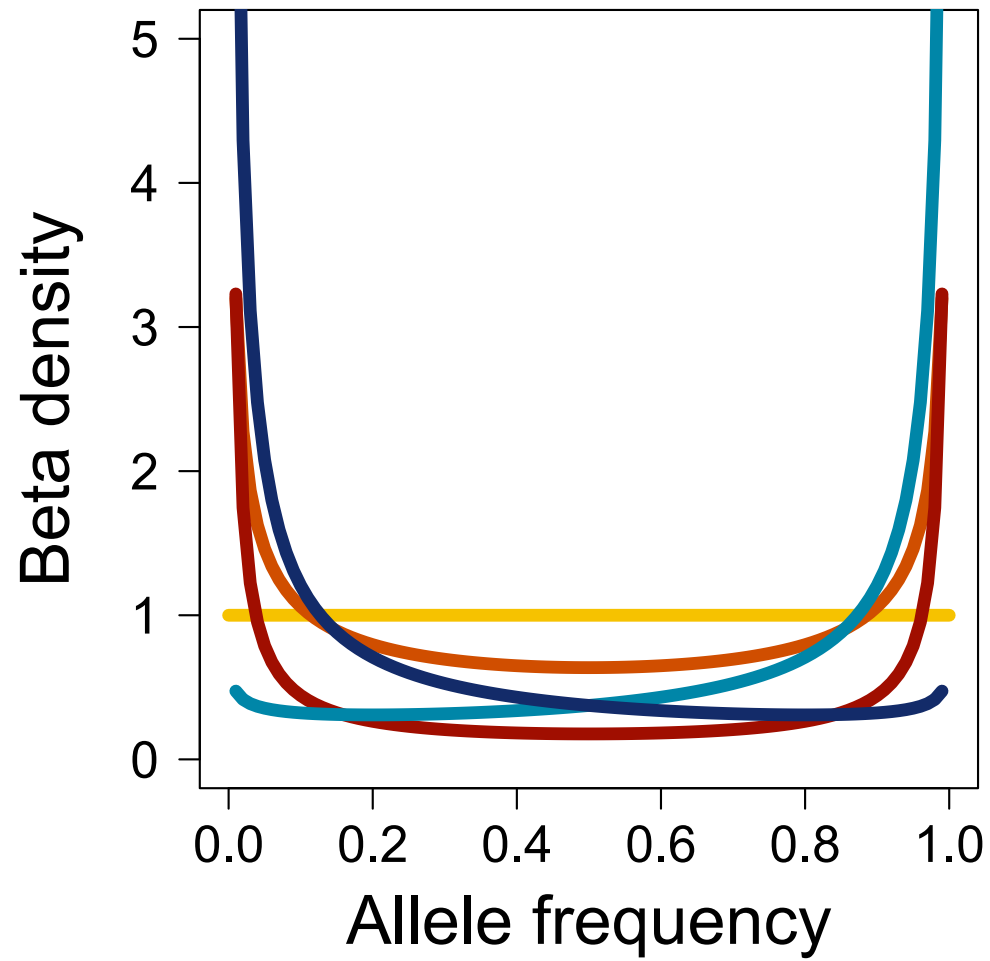
$\alpha = 1$ $\beta = 1$
 $\alpha = 0.5$ $\beta = 0.5$



$\alpha = 1 \quad \beta = 1$
 $\alpha = 0.5 \quad \beta = 0.5$
 $\alpha = 0.1 \quad \beta = 0.1$



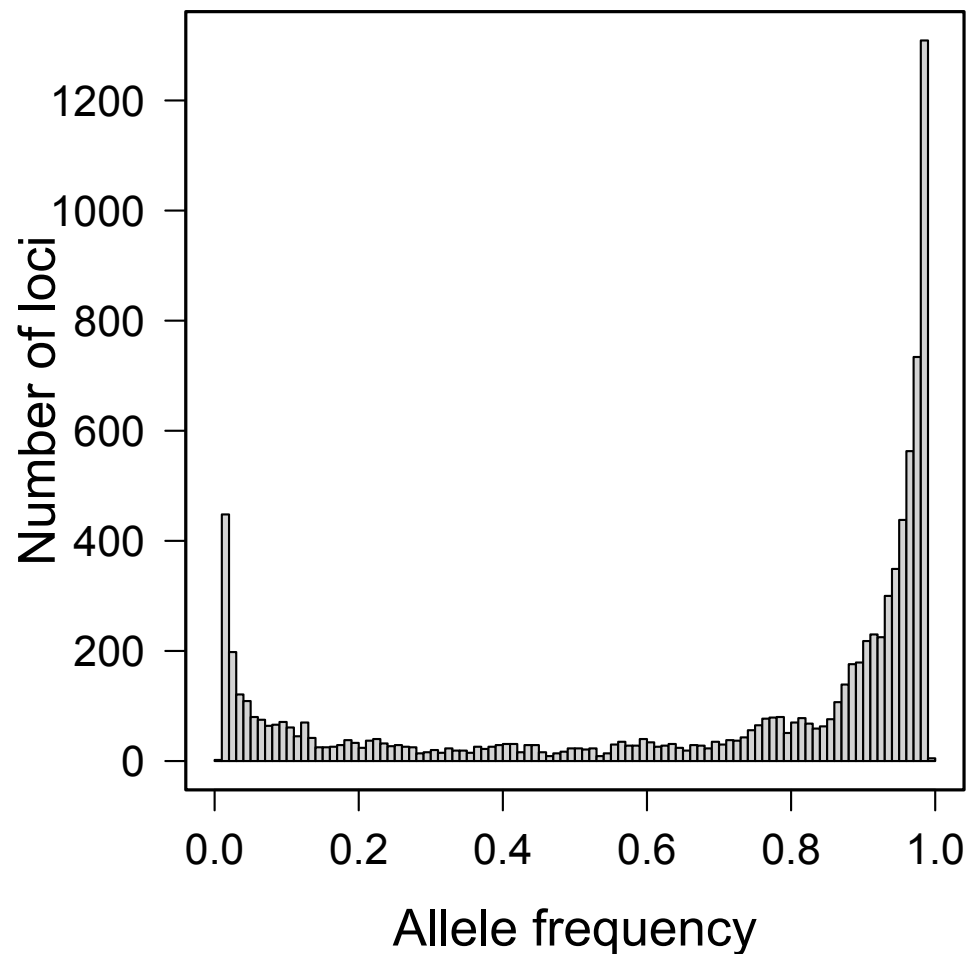
$\alpha = 0.2$ $\beta = 0.8$
 $\alpha = 0.8$ $\beta = 0.2$
 $\alpha = 1$ $\beta = 1$



$\alpha = 0.2$ $\beta = 0.8$
 $\alpha = 0.8$ $\beta = 0.2$
 $\alpha = 1$ $\beta = 1$
 $\alpha = 0.5$ $\beta = 0.5$
 $\alpha = 0.1$ $\beta = 0.1$

Influence of dams on sauger population structure and hybridization with introduced walleye

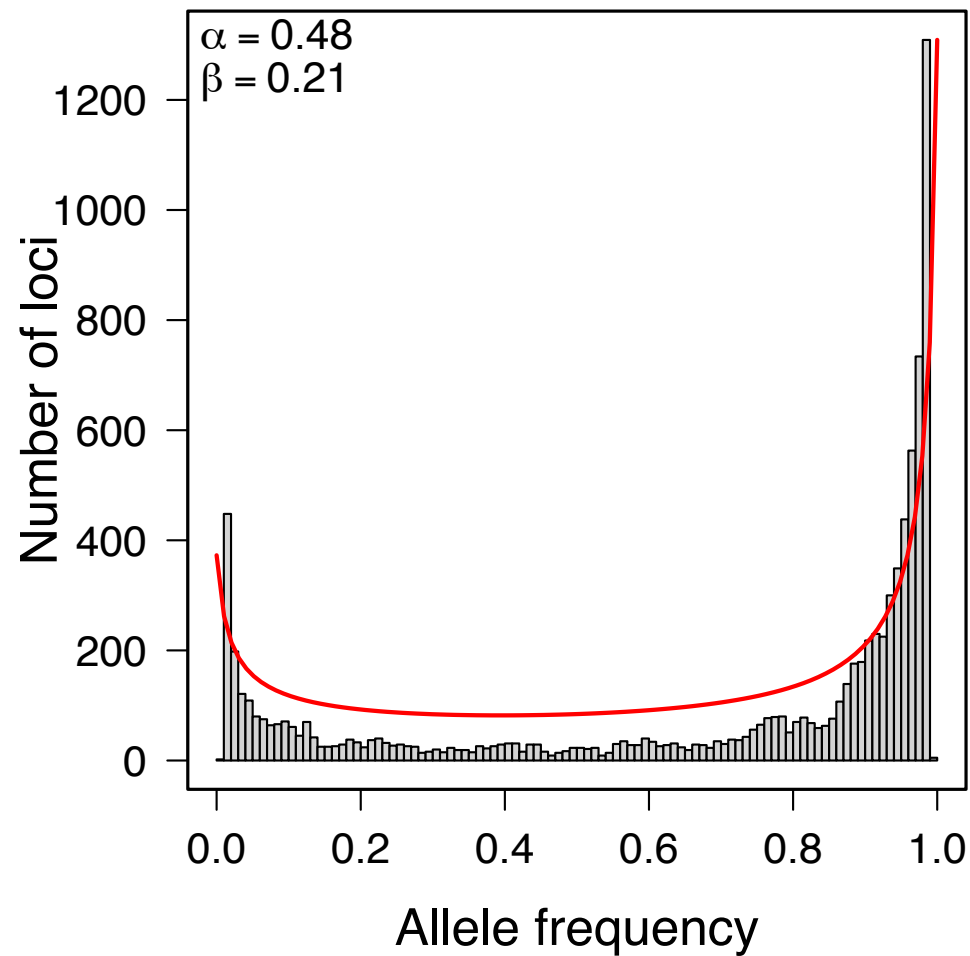
William C. Rosenthal^{1,2}  | Elizabeth G. Mandeville^{1,3} | Ashleigh M. Pilkerton^{2,4,5}  |
Paul C. Gerrity⁶ | Joseph A. Skorupski⁶ | Annika W. Walters^{2,4,5,7}  |
Catherine E. Wagner^{1,2}



`dbeta(xvals, shape1=NA, shape2=NA)`

Influence of dams on sauger population structure and hybridization with introduced walleye

William C. Rosenthal^{1,2}  | Elizabeth G. Mandeville^{1,3} | Ashleigh M. Pilkerton^{2,4,5}  |
Paul C. Gerrity⁶ | Joseph A. Skorupski⁶ | Annika W. Walters^{2,4,5,7}  |
Catherine E. Wagner^{1,2}




```
## Function for simulating genetic data from the beta distribution
```

```
single_pop_sim <- function(nloci=NA, ninds=NA, alpha=NA, beta=NA){  
  genotypes <- matrix(NA, ninds, nloci)  
  for (i in 1:nloci){  
    afreq <- rbeta(1, alpha, beta)  
    for (j in 1:ninds){  
      genotypes[j,i] <- sum(rbinom(2, 1, afreq))  
    }  
  }  
  return(genotypes)  
}
```

```
## Function for simulating genetic data from the beta distribution
```

```
single_pop_sim <- function(nloci=NA, ninds=NA, alpha=NA, beta=NA){  
  genotypes <- matrix(NA, ninds, nloci)  
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    for (j in 1:ninds){  
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    }  
  }  
  return(genotypes)  
}
```




Draw an allele
freq from the
beta distribution


```
## Function for simulating genetic data from the beta distribution
```

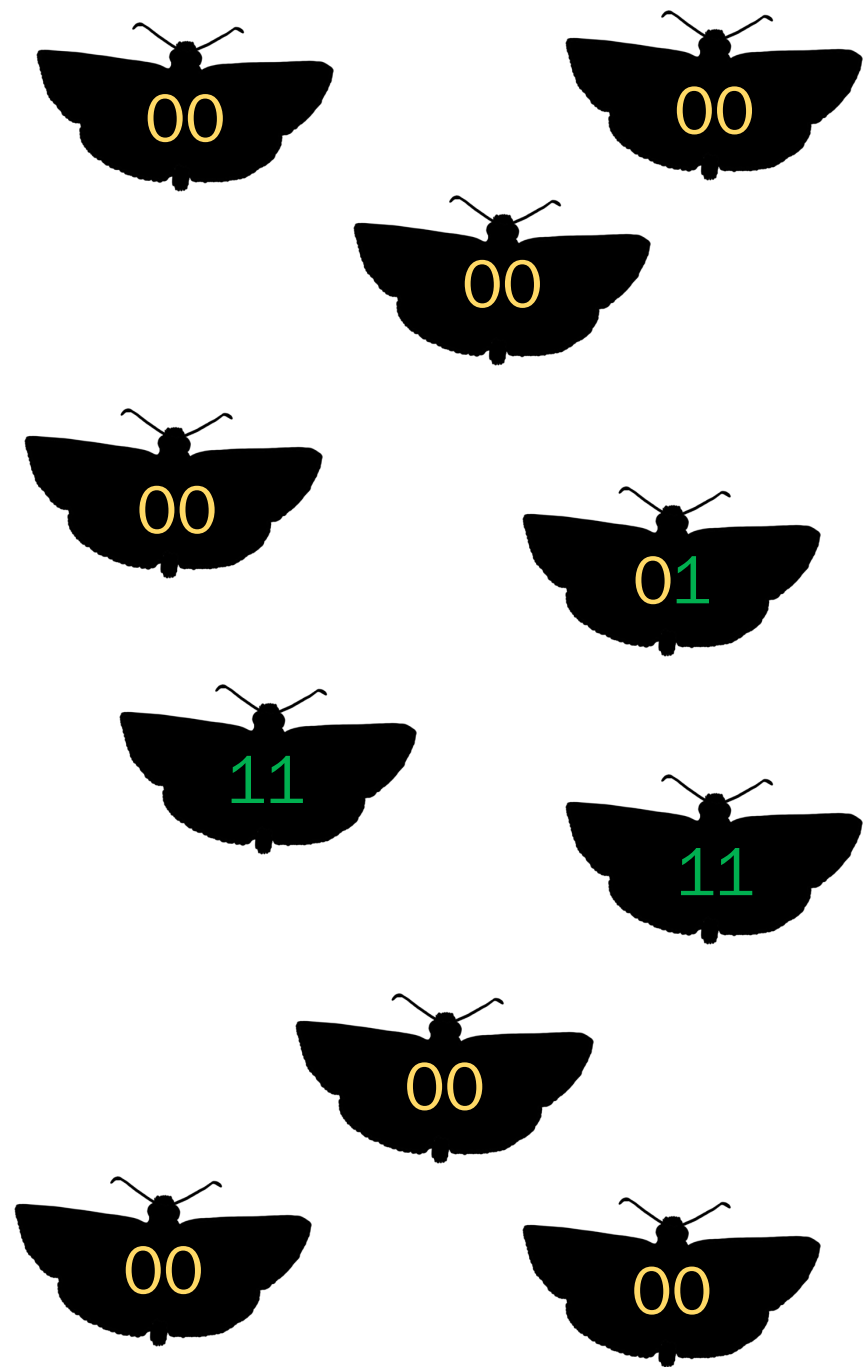
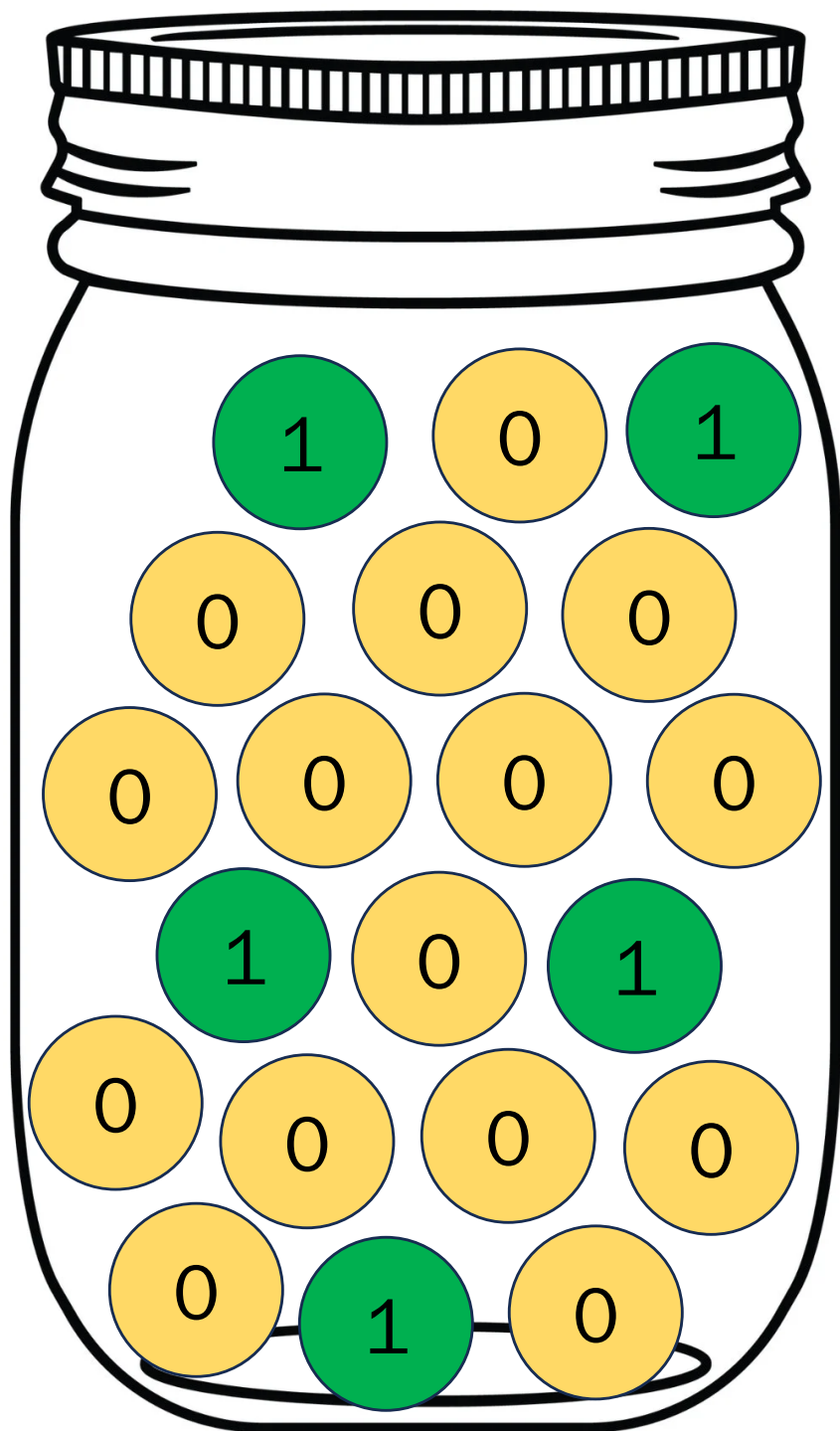
```
single_pop_sim <- function(nloci=NA, ninds=NA, alpha=NA, beta=NA){  
  genotypes <- matrix(NA, ninds, nloci)  
  for (i in 1:nloci){  
    afreq <- rbeta(1, alpha, beta)  
    for (j in 1:ninds){  
      genotypes[j,i] <- sum(rbinom(2, 1, afreq))  
    }  
  }  
  return(genotypes)  
}
```

Draw an allele
freq from the
beta distribution



Draw twice from the
binomial distribution to
determine genotype
(0, 1, or 2)





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Goals for the rest of our time:

1. Simulate genetic data using the beta distribution
2. Explore the data with three different genetic analyses

Goals for the rest of our time:

1. Simulate genetic data using the beta distribution
2. Explore the data with three different genetic analyses

How are genetic analyses
affected by the shape of the
site frequency spectrum?

How many genetic markers
do I need to detect a signal?