

CHROMATIN ARCHITECTURE ABERRATIONS IN PROSTATE CANCER AND ACUTE  
LYMPHOBLASTIC LEUKEMIA

by

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# Chapter 1

## Discussion & Future Directions

Each of the previous chapters have presented a story interrogating multiple components of the chromatin architecture, how they interact with each other, and the plethora of computational and experimental methods required to unravel this architecture. ?? identifies and validates *cis*-regulatory elements (CREs) of the *FOXA1* gene, a critical transcription factor (TF) that regulates prostate cancer (PCa) development and regulates androgen receptor (*AR*) expression to control disease progression. ?? expands on these ideas to investigate how the three-dimensional genome organization impacts gene regulatory networks and how genetic aberrations can alter this organization to promote oncogenesis. ?? develops a mathematical and computational framework to reduce uncertainty about how individual aberrations in chromatin architecture impact gene expression. Finally, ?? identifies the strong relationship between genetic and epigenetic profiles in B-cell acute lymphoblastic leukemia (B-ALL) relapse and investigates how DNA methylation (DNAm) changes and revision to a more stem-like chromatin state may underlie disease relapse. Together, the work presented in this thesis demonstrates that different components of the chromatin architecture, the genome, molecular chromatin modifications, and three-dimensional organization, can all individually contribute to cancer development and progression. Moreover, this thesis demonstrates that aberrations in these components work together to drive disease. These multiple components of the chromatin architecture need to be studied in tandem to understand the origins of cancer and how to develop curative treatments for it.

## 1.1 Implications of non-coding single nucleotide variants targeting a single gene

In ??, I used gene essentiality screening data from multiple cell lines to prioritize the *FOXA1* TF as a critical factor across PCa cell lines. I also made use of the concept that single nucleotide variants (SNVs) converge on CREs of important genes in a given tumour type to predict how these mutations may impact candidate CREs for the *FOXA1* gene. *FOXA1* is also an important TF in breast cancers ??. Similar investigations into the impact of SNVs in breast tumours may identify the impact of aberrations to the CREs of *FOXA1*. Identifying important genes in this manner is not limited to *FOXA1* and breast and prostate tumours. Critical genes may be identified in other cancer types using clustered regularly interspaced short palindromic repeat (CRISPR) screens or massively-parallel reporter assays (MRPAs). Similarly SNVs are not the only chromatin aberrations that can affect TF binding or gene regulation. Other chromatin aberrations may accumulate in CREs of important genes in a similar fashion. Complex structural variants (SVs), changes in DNAm, or histone modifications may only need to accumulate in the set of CREs for a given gene, rather than be recurrent in a single element, to affect its expression. Interpreting chromatin aberrations in cancer in light of this plexus-based approach may aid in identifying driver events for cancer by aggregating previously unrelated events together. These approaches are not limited to prostate tumours and can serve as a starting point to identify important genes in other cancers, more generally.

## 1.2 Implications of three-dimensional organization and enhancer hijacking in prostate cancer

In ??, my co-authors optimized a low-input Hi-C method to interrogate genome organization in cryo-preserved prostate tissue slides. I then demonstrated that this could produce a high quality Hi-C library and helped produce that largest collection of genome organization data in prostate tumours to date. This technological step forward opens the door for profiling the three-dimensional genome in cancer patients without relying in cell lines or other models, and may be a critical step in moving personalized medicine forward. We add to existing evidence that SVs can, but rarely, alter 3D structure in disease [1–8]. Elucidating when and how SVs impact genome organization, then, is still an area that requires investigation. Developments in statistical methods, such as those discussed in ??, may help identify the effects of individual, non-recurrent SVs. Subclonality of SVs

may interfere with the ability to detect rearranged domains in bulk Hi-C measurements. Thus, developments in high throughput sequencing and microscopy measurements in single cells, such as ORCA [9] and STORM [10], as well as organoid or explant models that recapitulate the chromatin state of the original tumour, may help in identifying the effect of such events []. This work also adds to our ability to detect chromatin interactions between promoters and enhancers in patient samples, allowing for better characterization of gene regulatory networks for each and every gene. Given the benefits of plexus-based approaches to interpreting aberrations in the chromatin architecture, this work serves as a foundation on which to integrate gene regulatory networks with chromatin aberrations in cancers more generally. This foundation can be extended to studying the evolution of these networks, their genome organization, and their resiliency between species or over time as tumours respond to therapeutic interventions [].

### 1.3 Implications of DNA methylation changes in relapse

- prioritize role of stem cells in disease relapse
- use increasing DNAm as a potential biomarker of relapse
- may be able to use blood-based DNAm detection to create a non-invasive test for this development
- may be able to treat B-ALL patients with de-methylating agents if gains in DNAm are observed to prevent relapse

### 1.4 Summary and concluding remarks

- work does not focus on a single disease, should extend this type of analysis to all cancers, since they all appear to harbour aberrations affecting multiple components of the chromatin architecture
- multi-pronged approach of computational, statistical, and molecular, and microscopy methods optimized for low-input samples targeting the set of DNA elements and their relationships to each other in individual patients to develop personalized medicines and treat cancer at its origins in the chromatin

# Glossary

**3C** chromatin conformation capture

**ALL** acute lymphoblastic leukemia

**AML** acute myeloid leukemia

**ANOVA** Analysis of Variance

**AR** androgen receptor

**ATAC-seq** assay for transposase-accessible chromatin sequencing

**B-ALL** B-cell acute lymphoblastic leukemia

**bp** basepair

**cDNA** complementary DNA

**ChIP-seq** chromatin immunoprecipitation sequencing

**CLL** chronic lymphocytic leukemia

**CML** chronic myeloid leukemia

**CMP** common myeloid progenitor

**CNV** copy number variant

**CPC-GENE** Canadian Prostate Cancer Genome Network

**CpG** CG dinucleotide

**crRNA** CRISPR RNA

**CRE** *cis*-regulatory element

**CRISPR** clustered regularly interspaced short palindromic repeat

**CTCF** CCCTC-binding factor

**CUT&RUN** cleavage under targets and release using nuclease

**DEPMAP** Cancer Dependency Map

**DHS** DNase I hypersensitive sites

**DLBCL** diffuse large B-cell lymphoma

**DMR** differentially methylated region

**DNA** deoxyribonucleic acid

**DNAme** DNA methylation

**DNase-seq** DNase I hypersensitive sequencing

**dRI** disease relapse-initiating

**Dx** diagnosis

**EarlyProB** early progenitor B cell

**FDR** false discovery rate

**FN** false negative

**FP** false positive

**FOX** forkhead box

**GLM** generalized linear model

**GMP** granulocyte-macrophage progenitor

**GO** gene ontology

**gRNA** guide RNA

**HSC** hematopoietic stem cell

**HSPC** hematopoietic stem and progenitor cell

**IDH** isocitrate dehydrogenase

**IID** independent and identically distributed

**ISUP** International Society of Urological Pathology

**JS** James-Stein

**KMT** histone lysine methyltransferase

**KO** knockout

**LDA** limiting dilution assay

**LMPP** lymphoid-primed multi-potent progenitor

**MeCapSeq** DNA methylation capture sequencing

**MEP** megakaryocyte-erythrocyte progenitor

**MNase-seq** micrococcal nuclease sequencing

**MSE** mean square error

**mCRPC** metastatic castration-resistant prostate cancer

**MDS** myelodysplastic syndrome

**MLP** monocyte-lymphoid progenitor

**MPP** multi-potent progenitor

**MRPA** massively-parallel reporter assay

**NSG** NOD scid gamma

**OLS** ordinary least squares

**mRNA** messenger RNA

**PCa** prostate cancer

**PDX** patient-derived xenograft

**PreProB** pre-progenitor B cell

**ProB** progenitor B cell

**PSA** prostate-specific antigen



**Rel** relapse

**RNA** ribonucleic acid

**RNAi** RNA interference

**RNA-seq** RNA sequencing

**shRNA** small hairpin RNA

**siRNA** small interfering RNA

**SNV** single nucleotide variant

**SRA** Sequence Read Archive

**SNF** similarity network fusion

**SV** structural variant

**T2E** *TMPRSS2-ERG*

**TAD** topologically associated domain

**TCGA** The Cancer Genome Atlas

**TET** ten-eleven translocation

**TSS** transcription start site

**TN** true negative

**TNM** tumour node metastasis

**TP** true positive

**TF** transcription factor

**tracrRNA** trans-activating CRISPR RNA

**UTR** untranslated region

**WES** whole exome sequencing

**WGBS** whole genome bisulfite sequencing

**WGS** whole genome sequencing

**WT** wild-type

# References

1. Ghavi-Helm, Y. *et al.* Highly Rearranged Chromosomes Reveal Uncoupling between Genome Topology and Gene Expression. En. *Nature Genetics*, 1. ISSN: 1546-1718 (July 2019).
2. Oudelaar, A. M. & Higgs, D. R. The Relationship between Genome Structure and Function. en. *Nature Reviews Genetics*. ISSN: 1471-0056, 1471-0064 (Nov. 2020).
3. Despang, A. *et al.* Functional Dissection of the Sox9–Kcnj2 Locus Identifies Nonessential and Instructive Roles of TAD Architecture. en. *Nature Genetics* **51**, 1263–1271. ISSN: 1061-4036, 1546-1718 (Aug. 2019).
4. Williamson, I. *et al.* Developmentally Regulated *Shh* Expression Is Robust to TAD Perturbations. en. *Development* **146**, dev179523. ISSN: 0950-1991, 1477-9129 (Oct. 2019).
5. Dixon, J. R. *et al.* Integrative Detection and Analysis of Structural Variation in Cancer Genomes. En. *Nature Genetics* **50**, 1388. ISSN: 1546-1718 (Oct. 2018).
6. Akdemir, K. C. *et al.* Disruption of Chromatin Folding Domains by Somatic Genomic Rearrangements in Human Cancer. en. *Nature Genetics*, 1–12. ISSN: 1546-1718 (Feb. 2020).
7. Li, Y. *et al.* Patterns of Somatic Structural Variation in Human Cancer Genomes. en. *Nature* **578**, 112–121. ISSN: 1476-4687 (Feb. 2020).
8. Iyyanki, T. *et al.* Subtype-Associated Epigenomic Landscape and 3D Genome Structure in Bladder Cancer. en. *Genome Biology* **22**, 105. ISSN: 1474-760X (Dec. 2021).
9. Mateo, L. J. *et al.* Visualizing DNA Folding and RNA in Embryos at Single-Cell Resolution. En. *Nature* **568**, 49. ISSN: 1476-4687 (Apr. 2019).
10. Bates, M., Jones, S. A. & Zhuang, X. Stochastic Optical Reconstruction Microscopy (STORM): A Method for Superresolution Fluorescence Imaging. en. *Cold Spring Harbor Protocols* **2013**, pdb.top075143. ISSN: 1940-3402, 1559-6095 (June 2013).