### **TINKER** simulations

Molecular Dynamics
Simulations
An EMBnet introductory course

10-11-2008

#### Jose R. Valverde EMBnet/CNB, CSIC

http://www.es.embnet.org

jrvalverde@cnb.csic.es

#### **Lecture Outline**

- TINKER introduction
- Protein simulations in vacuo
- Protein simulations in implicit solvent model
- Solvent simulations with periodic boundary conditions
- Protein simulations in explicit, all-atoms solvent model

# TINKER MD

TINKER introduction

#### **TINKER**

- Developed at Jay Ponder's lab
  - http://dasher.wustl.edu/tinker/
- Professional quality
- Main goal is development of algorithms
- Less extensive than other programs (e. g. CHARMM)
- Wider support for force fields
  - AMBERxx, AMOEBAxx, CHARMMxx, Dang, Dudek, Hoch, MMxx, OPLSxx, SMOOTHxx...
- Focus on ease of use
- Command line interface
- Free, with source code
- Integrated with GAMESS-US QM software (SIMOMM)

## Running TINKER

- Coordinates (XYZ)
  - Convert from/to PDB (pdbxyz, xyzpdb)
- Parameter file
  - .key file
    - tinker.key (always valid)
    - filename.key (filename is same as in filename.xyz)
  - keyword value
    - parameters /opt/structure/tinker/params/amber.prm
- Other files
  - Parameter files (potential energy functions/force fields)
  - Restart, output, trajectories
- Interactive command line
  - Easy: just answer
  - Answers may be given in the command line

## **Worth noting**

- Filename extension is not entered (tinker assumes it)
- Tinker asks for options not given in the command line
- Tinker outputs progress report to console
  - Worth saving for inspection/analysis
    - command model | tee model-command.log
    - command model > model-command.log &
- filename.key takes precedence over tinker.key
  - Use tinker.key for common options
  - User filename.key for model-specific options
- Existing output files are not overwritten
  - A sequential suffix is added (e. g. model.xyz\_2)

## TINKER MD

Protein model in vacuo

### **Initial setup**

- Create a subdirectory and move to it
  - Copy PDB coordinate file to your directory
    - e. g. cp /usr/local/tinker/test/calmodulin.pdb .
  - Look into tinker installation directory, folder 'params'
    - e. g. ls /usr/local/tinker/params/
  - Select a force field and put it into tinker.key
    - e. g. parameters /usr/local/tinker/params/charmm27.prm
  - Convert PDB coordinates to XYZ coordinates
    - e. g. pdbxyz calmodulin
    - Correct any discrepancies in atom names
      - PDB files tend to use atom names corresponding to the force field used during refinement
      - Not all force fields use the same atom names
  - Have an initial look with analyze

## **Initial structure optimization**

- Tinker offers various methods
  - minimize
  - optimize
  - newton
  - PSS
  - sniffer
- Minimization progresses until a target RMSD is reached
  - Use target RMSD of 1.0
  - Use chosen program interactively or in batch mode
    - minimize calmodulin 1.0
    - newton calmodulin A A 1.0
- Speed up using cutoffs in tinker.key
  - cutoff 15.0

## **Heating**

- Tune computation in tinker.key
  - cutoff 15.0
  - lights
- Reduce output clutter in tinker.key
  - archive
- Run ANNEAL to heat progressively the system to desired temperature
  - Heat from 1 to 298 K
  - Do not perform previous equilibration (0 steps)
  - Apply heat linearly
  - Use a 1.0 fs timestep and save data every 0.1 ps
  - Keep atomic weights
  - Run in batch mode
    - anneal calmodulin 1 298 0 2000 L 1.0 0.1 0.0

### **Equilibration**

- Keep the system at target temperature until equilibrium is reached
- Run DYNAMIC to simulate 3ps at 298K and monitor energies
  - Continue using same parameters
  - Run in batch mode
    - dynamic calmodulin 3000 1.0 0.1 298 > equil.out
  - After completion check system evolution
    - grep 'Temperature' equil.out > temperature.data
    - grep 'Total Energy' equil.out >totalenergy.data
    - **@** ...
    - Import into a graphical program and plot fluctuations
  - Visualize the trajectory
    - e. g. with gopenmol

#### **Production**

- Gather thermodinamical statistics and trajectory
- Start from equilibrated structure
- Run DYNAMIC to simulate 5 ps at 298 K
  - Timestep: 1fs
  - Number of steps: 5000
  - Take snapshots every 100 steps (100 fs = 0.1 ps)
  - Run in batch mode
    - dynamic calmodulin 5000 1.0 0.1 298 > prod.log &
- Monitor progress
  - Visualize trajectory
  - Watch out for strong energy fluctuations

### **Analysis**

- Visualize trajectory
- Extract relevant data
  - grep 'Total Energy' 2ech-prod.log > te.out
  - cat te.out | sed -e 's/ Total Energy//g' > te.1
  - cat te.1 | sed -e 's/K.\*//g' > te.2
  - cat te.2 | sed -e 's/ //g' > te.dat
- Plot data
  - Import into plotting program
- Perform distance geometry calculations
  - DISTGEOM
- Compute time dependent correlations
  - CORRELATE

# TINKER MD

Protein in implicit solvent model

### Define implicit solvent model

- Select an implicit solvation model
  - ASP / SASA / ONION / STILL / HCT / ACE / GBSA
  - Add it to tinker.key
    - solvate still
  - State solvent dielectric constant
    - dielectric 80.0
  - State polarization effects
    - polarization direct
- Perform the whole simulation process using the new parameters (tip: use a separate directory)
  - Start from initial PDB
  - Minimize
  - Equilibrate
  - Production run
  - Analyze

## **TINKER MD**

Water in Periodic Boundary Conditions

#### Initialization

- Use a fresh directory
- Start with the coordinates for one molecule of water
  - water.xyz
- Choose a model for water (e. g. TIP3P)
  - TINKER contains a TIP3P model
  - You can build your own
  - Check atom name discrepancies
    - $e. g. O \rightarrow OT (101), H \rightarrow HT (88)$
- Generate a water box using XYZEDIT
  - xyzedit water
  - Place 27 molecules in a cubic box of 9.3125 Å side

### Heating

- Define periodic boundary conditions in tinker.key (or in water.key):
  - Box size
    - a-axis 9.3125
  - Cutoff distance (less than ½ the box size)
    - cutoff 4.0
- Fine tune computation (TIP3P is a rigid model)
  - integrate RIGIDBODY
  - group-molecule
  - neutral-groups
- Heat the system during 6 ps at NVT.
  - dynamic waterbox 6000 1.0 0.1 2 298
  - anneal waterbox 1 298 0 6000 L 1.0 0.1 0.0

### **Equilibration**

- After we have heated the system, we let it equilibrate for another 6ps at 298K (NVT)
  - dynamic waterbox 6000 1.0 0.1 2 298
- Analyze system evolution for equilibration
  - grep 'Temperature' water-equil.out > temperature.data
  - grep 'Total Energy' water-equil.out >totalenergy.data
  - **...**
  - Import into a graphical program and plot fluctuations
  - Visualize the trajectory
    - e. g. with gopenmol

#### **Production**

- Gather thermodinamical statistics and trajectory
- Start from equilibrated structure
- Run DYNAMIC to simulate 5 ps in the NVE ensemble
  - Timestep: 1fs
  - Number of steps: 5000
  - Take snapshots every 100 steps (100 fs = 0.1 ps)
  - Run in batch mode
    - dynamic waterbox 5000 1.0 0.1 1 > production.log &
- Monitor progress
  - Visualize trajectory
  - Watch out for strong energy fluctuations
- Analyze results.

## TINKER MD

Protein in explicit solvent model

#### Introduction

- Explicit solvent models are very expensive
  - Approximations and shortcuts have dramatic effects
  - Boundary Conditions are required to keep the solvent confined
    - Periodic Boundary Conditions (spherical)
    - Stochastic Boundary Conditions (Brownian motion)
  - Cutoffs and cell sizes reduce computation area
    - Reaction region
    - Bufer (Langevin motion) region
    - Reservoid (fixed) region

#### **Process**

- Preparation
  - Generate biomolecule coordinates input file
  - Compute energy
  - Minimize Energy
- Generate solvation sphere
  - Generate solvent cube
  - Soak biomolecule in solvent
  - Minimize water around biomolecule
  - Equilibrate water around biomolecule
- Heat-Equilibration-Production of entire system
- Analyze results.

## **Preparation**

- Same as we do for in vacuo simulations:
  - Grab PDB file
  - Convert to XYZ
  - Analyze structure
  - Minimize structure energy
- You can start from saved minimized coordinates from a previous in vacuo run.

#### **Adding water**

- Create a box of water
  - Get hold of a water molecule
  - Select force field (e. g. CHARMM27)
  - Fix atom name and definition discrepancies
    - e.g. O : OHT, type 101, H : HT, type 88
  - Run XYZEDIT
    - Number of molecules: 1600
    - Box size: 36.342
  - Translate box to center of mass using XYZEDIT
- Solvate biomolecule
  - Open with XYZEDIT
  - Translate to center of mass
  - Soak in water
  - No need to remove water outside the sphere
    - TINKER will ignore it once defined.

## Equilibration of water around fixed protein

- Open original biomolecule file
  - Note down molecule size (713 atoms)
- Define computation conditions in tinker.key
  - Use spherical boundary conditions
    - The system is centered around origin 0.0.0
    - The system size is ~32 A
      - sphere 0.0 0.0 0.0 18.0
      - wall
  - Exclude biomolecule by defining active region
    - Range from first (714) to last (5372) solvent atoms
      - active -714 5372
  - Stir the bottle to remove strong Van der Waals interactions
    - rattle
- Minimize energy and equilibrate as usual.

#### **MD** simulation

- Once the solvent has adapted to the biomolecule's environment we can run the simulation:
  - Remove restriction on the active region
  - Select a faster integration method
    - integrate verlet
- Heat to 298K
  - dynamic 2ech 1000 1.0 0.1 298
  - anneal 2ech 1 298 0 L 1000 1.0 0.1 0.0
- Equilibrate at 298K
  - dynamic 2ech 1000 1.0 0.1 298
- Production run at 298K
  - dynamic 2ech 5000 1.0 0.1 298
- Analyze