

Multi-Omics in SC

sc DNA methylation using scRRBS

sc chromatin accessibility using scATACseq

sc Proteomics using CITEseq

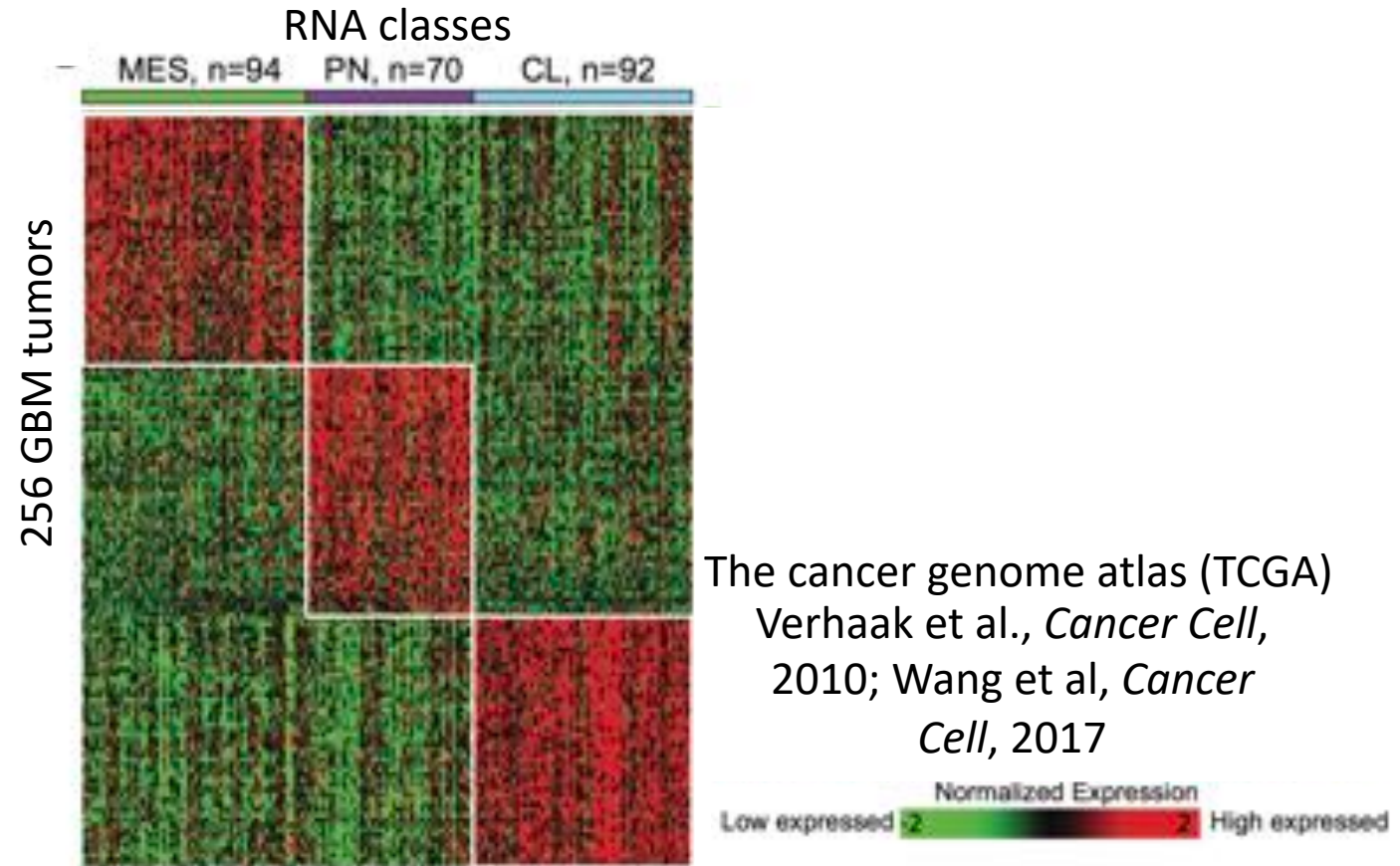
scDNAmethylation

Why do we need scDNA
methylation?

Glioma tumors exhibit inter-tumor heterogeneity

Glioma brain tumors:

- Main types:
 - Glioblastoma (GBM) ~90%
 - IDH-mutant glioma 8-9%
- Life expectancy hardly changed over the last 40 years of research
- Highly heterogenous

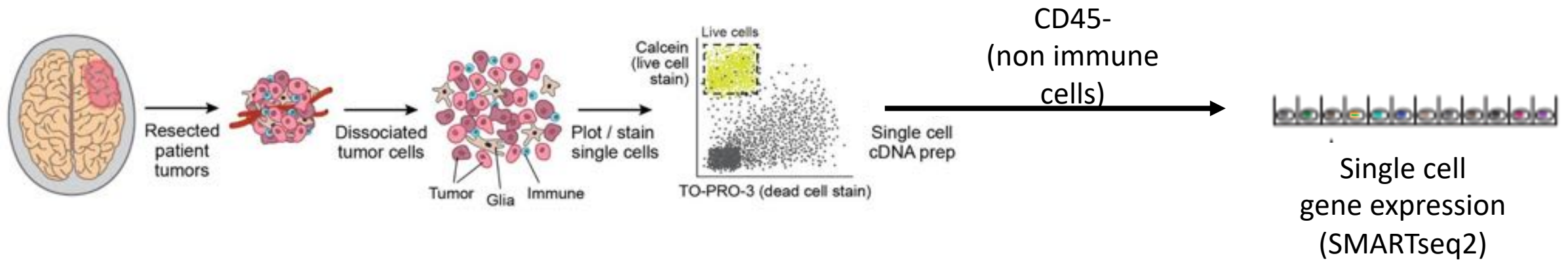


Is the heterogeneity driven by:

the tumors are composed of different cell types?

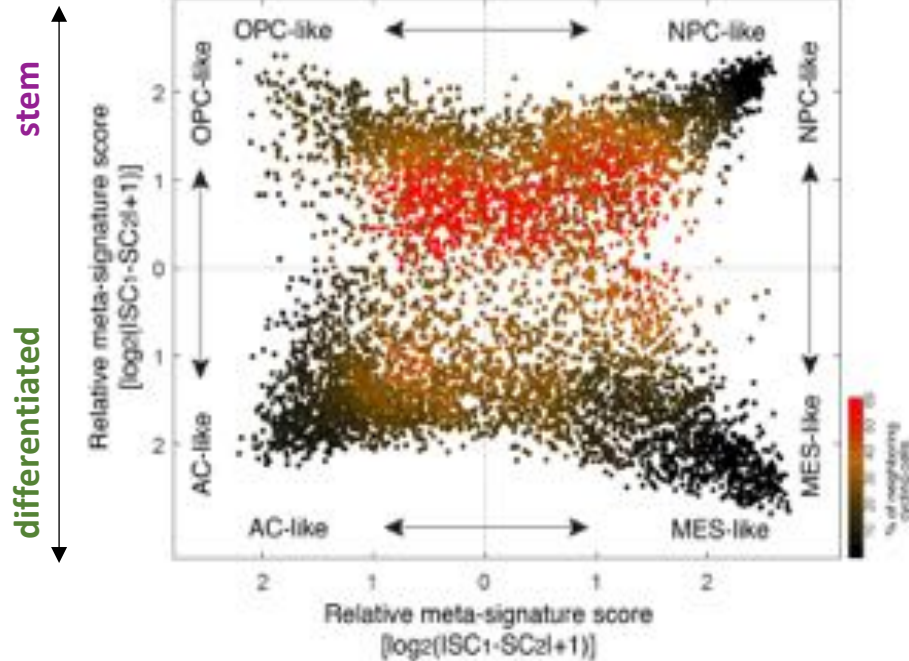
the tumors have the same cell types mixed in different proportions?

Revealing glioma compositions via single cell RNA sequencing

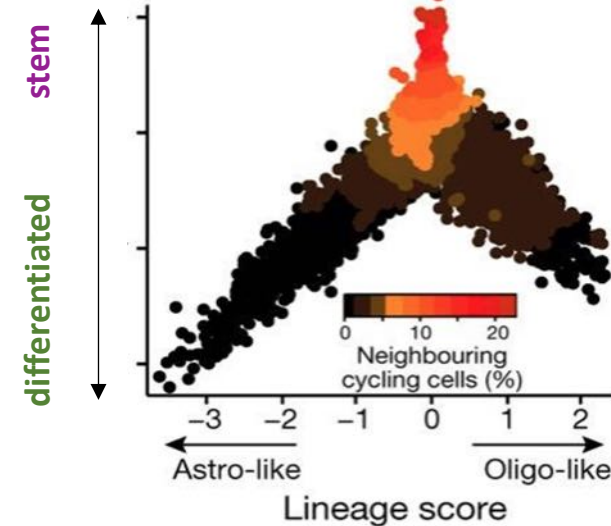


single cell RNAseq reveals intra-tumor heterogeneity

Glioblastoma



IDH-mutant glioma



Is the heterogeneity driven by:

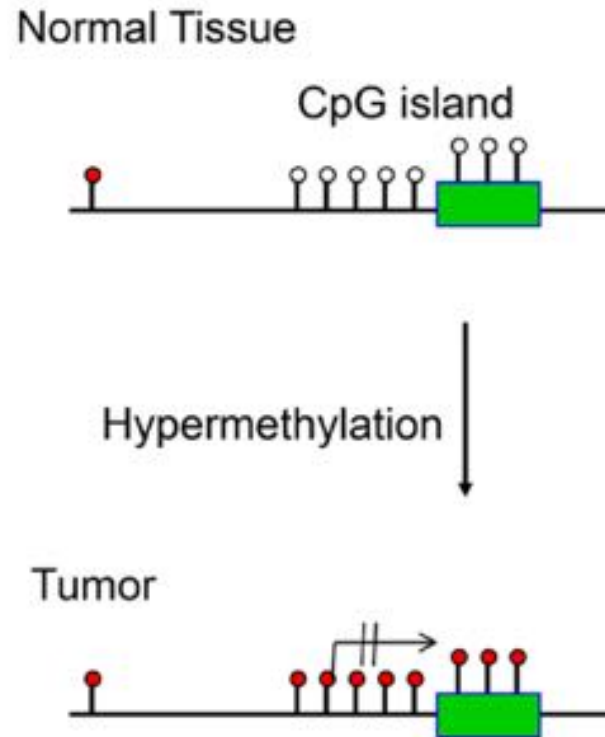
~~the tumors are composed of different cell types~~

the tumors have the same cell types mixed in different proportions!

Patel et al. 2014;
Tirosh et al. 2016;
Venteicher et al.
2017;
Neftel et al. 2019

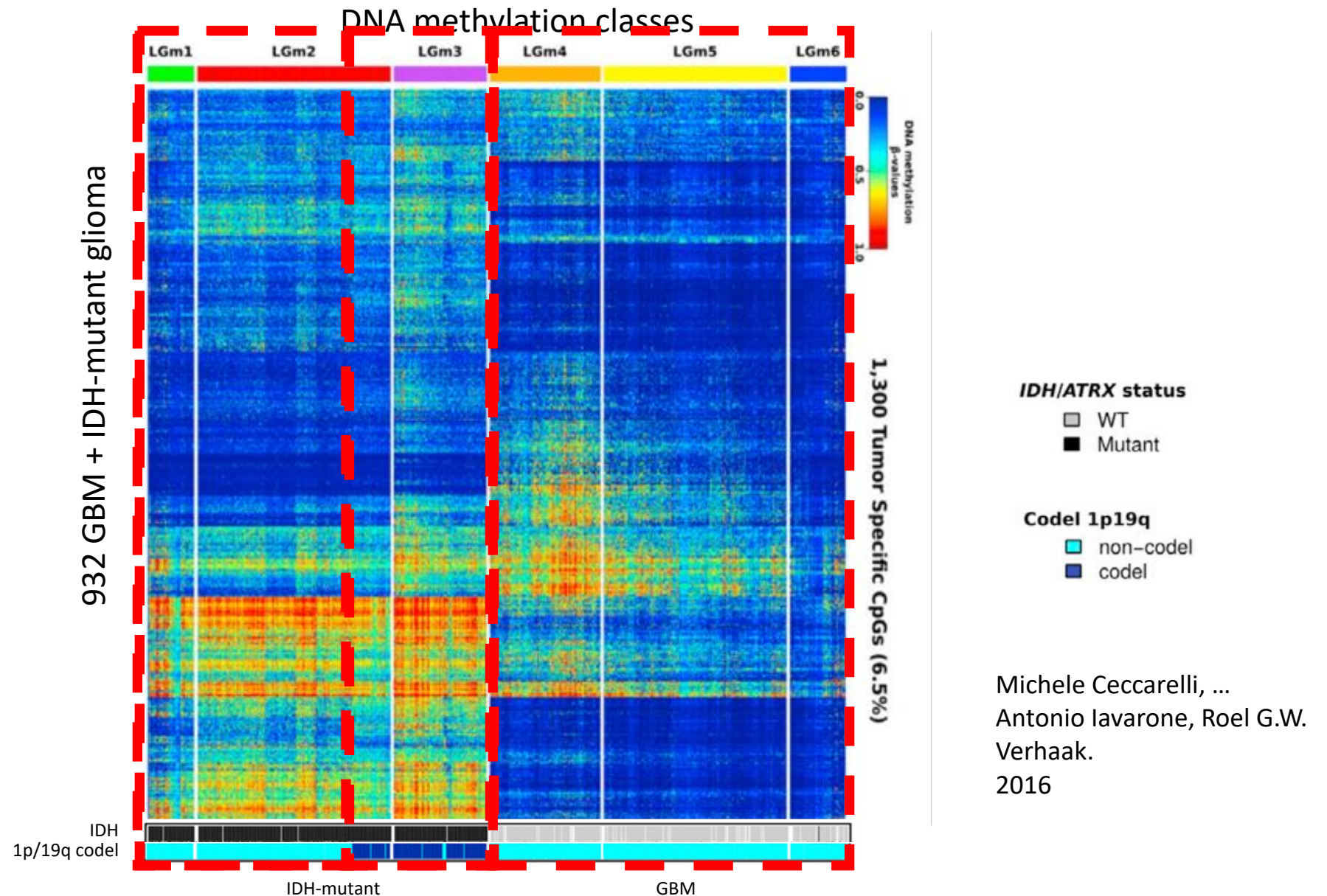


DNA methylation in healthy tissue and disease



From Pfeifer et. al.,
2018

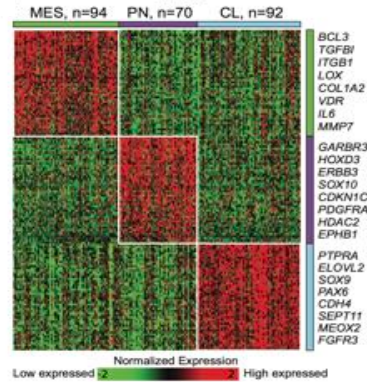
Inter-tumor DNA methylation heterogeneity



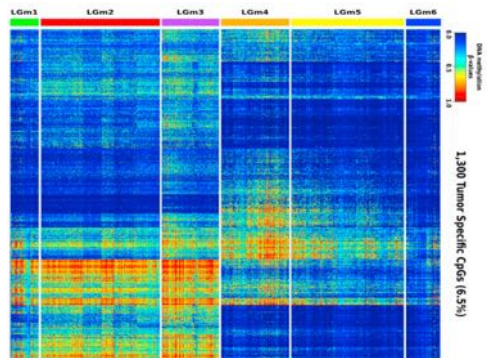
The goal: dissecting epigenetic identity of cellular states underlying glioma evolution

RNA

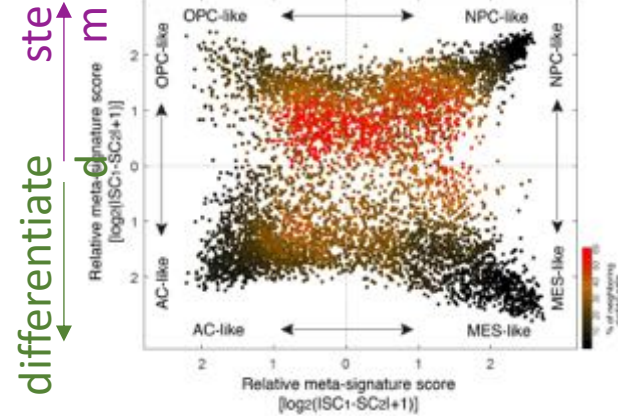
inter-tumor heterogeneity



DNA methylation



intra-tumor heterogeneity



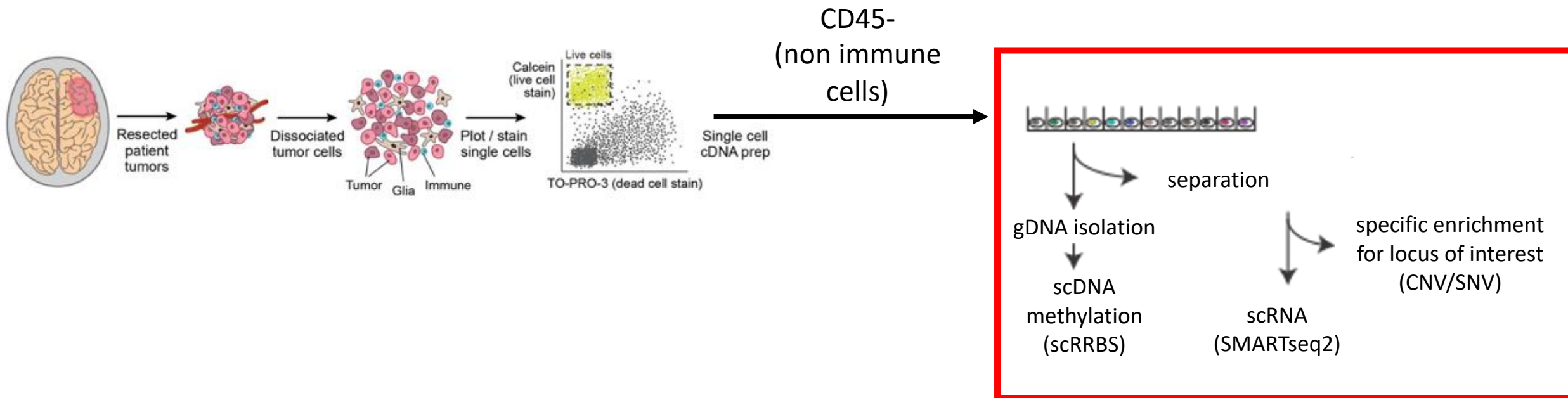
Is there a link between the RNA function and DNA methylation?



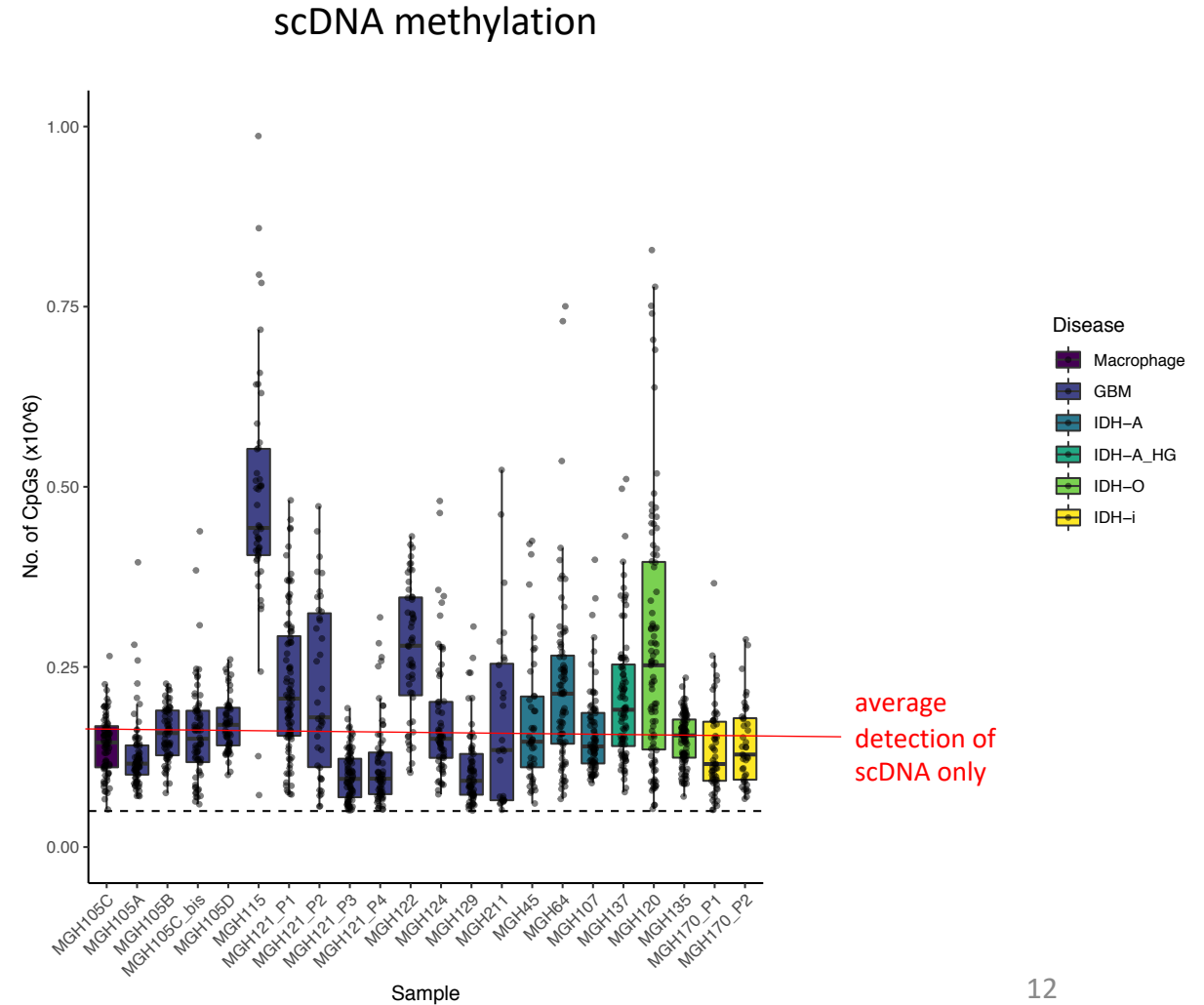
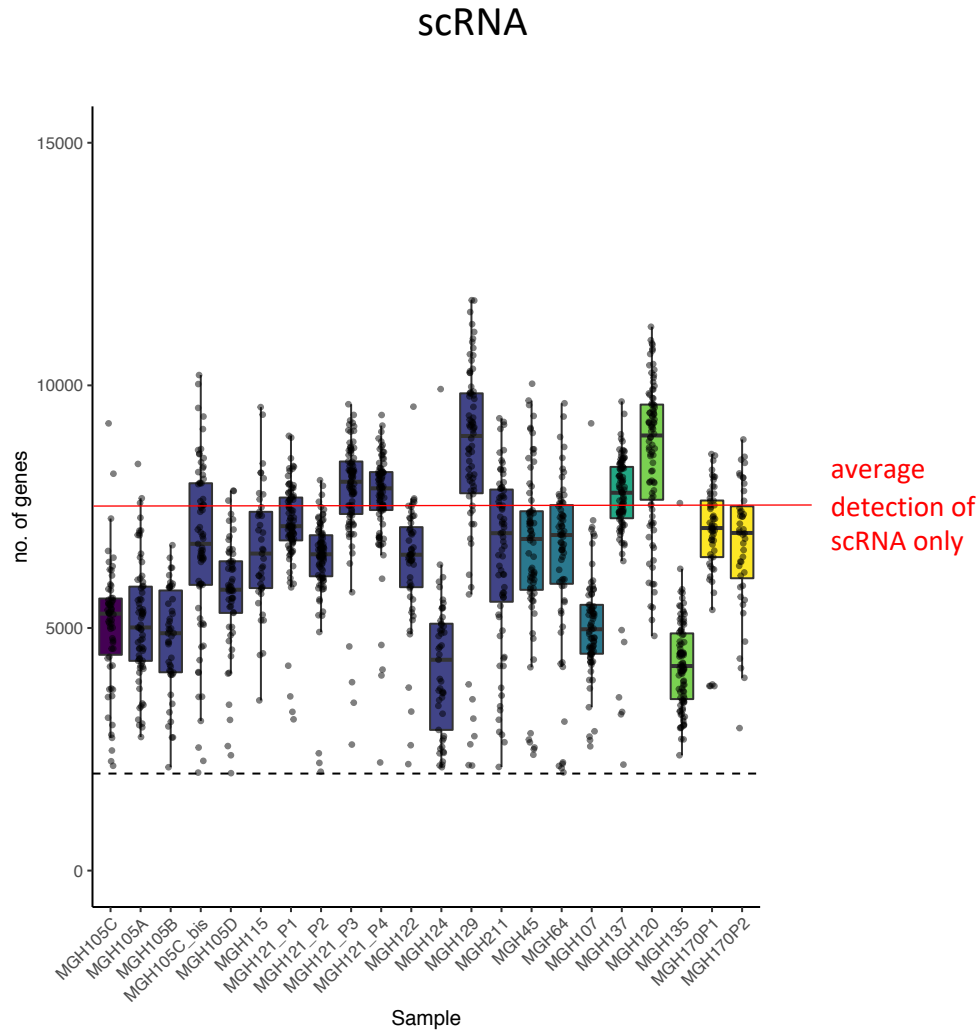
What is driving the DNA methylation heterogeneity?

Using scDNA methylation to dissect
epigenetic events driving cellular states

Simultaneously interrogating DNA methylation and RNA in the **same single cells**



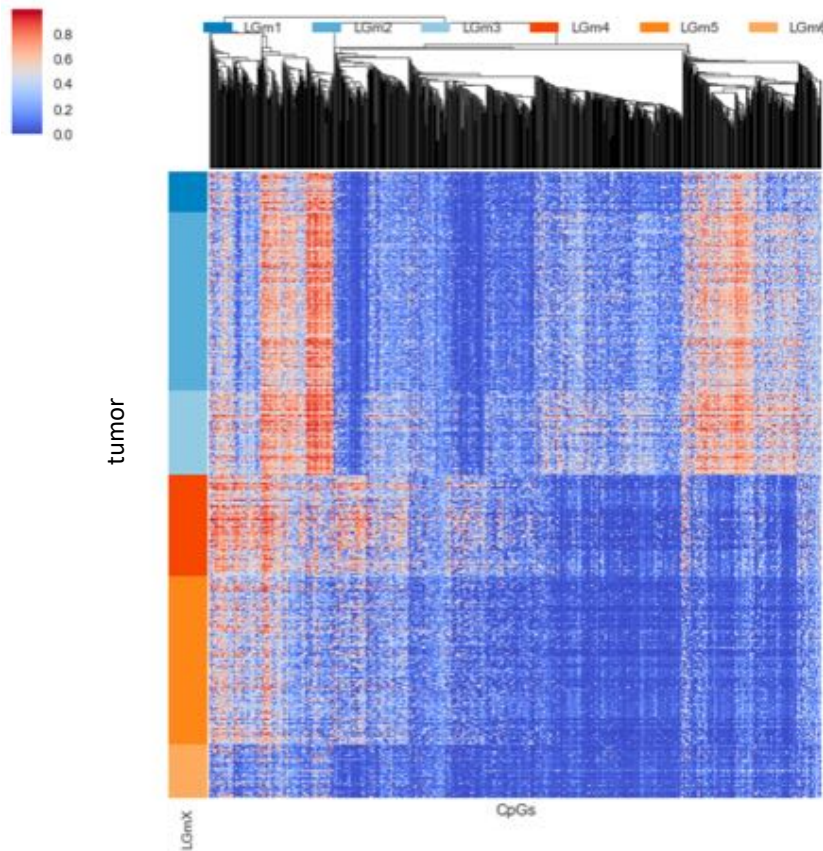
Number of features using scDNAseq compared to number of features using scRNAseq



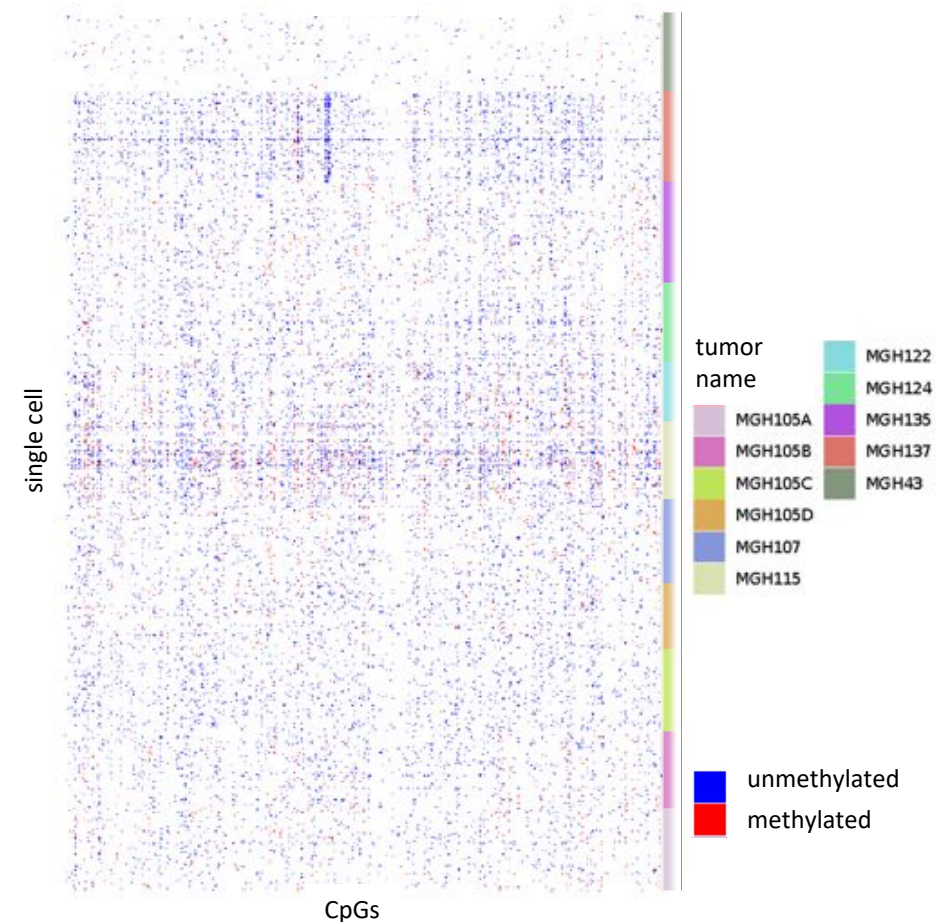
scDNA methylation is **really** sparse

Bulk DNA methylation

Verhaak DNA methylation data

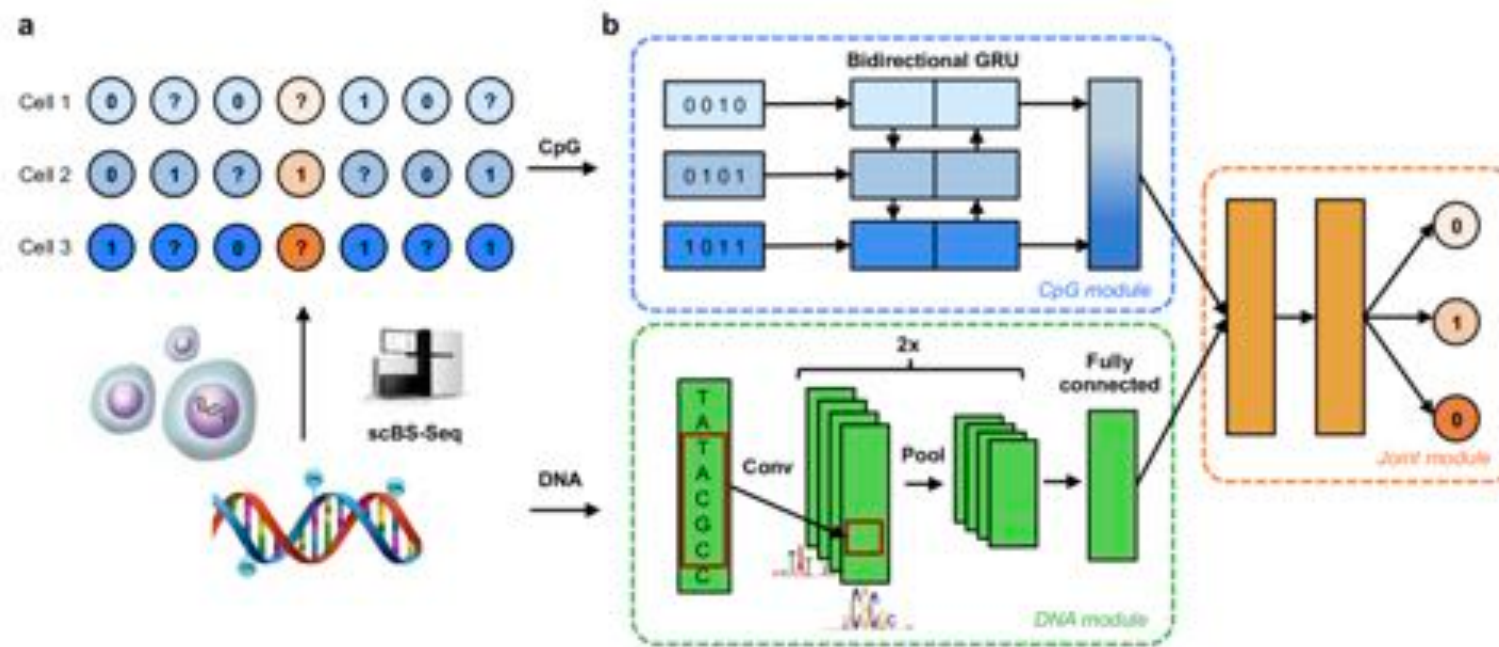


scDNA methylation

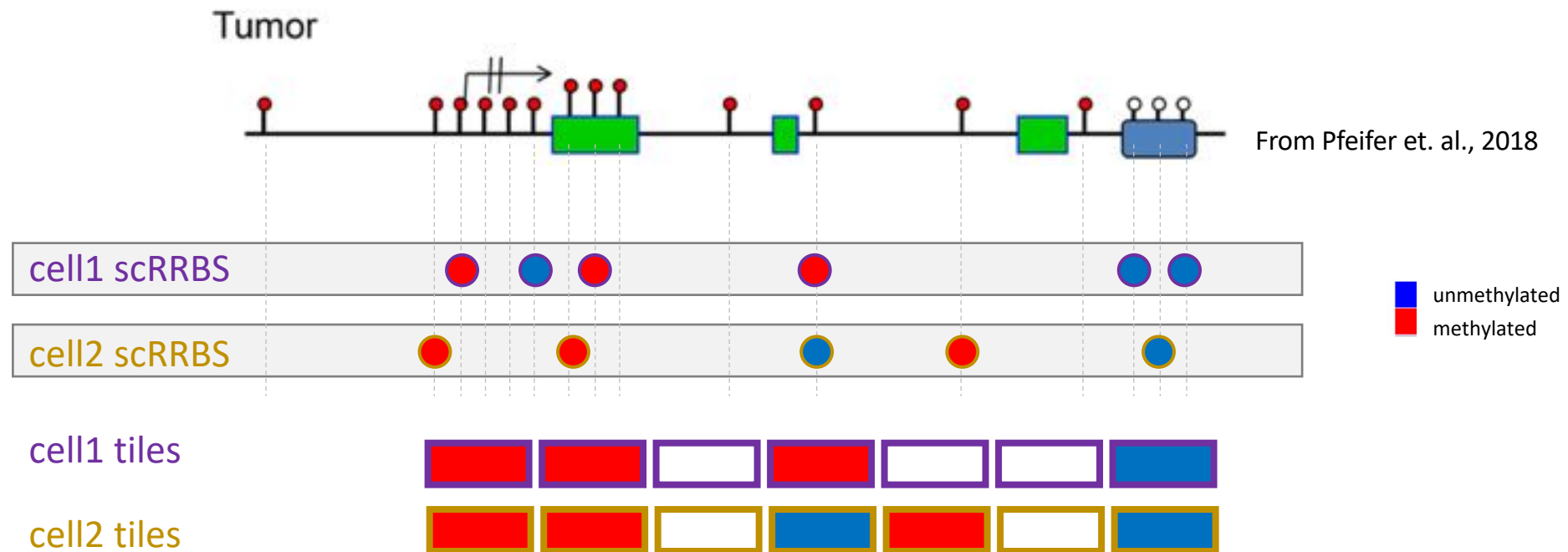


Impute sc DNA data: option1. machine learning techniques

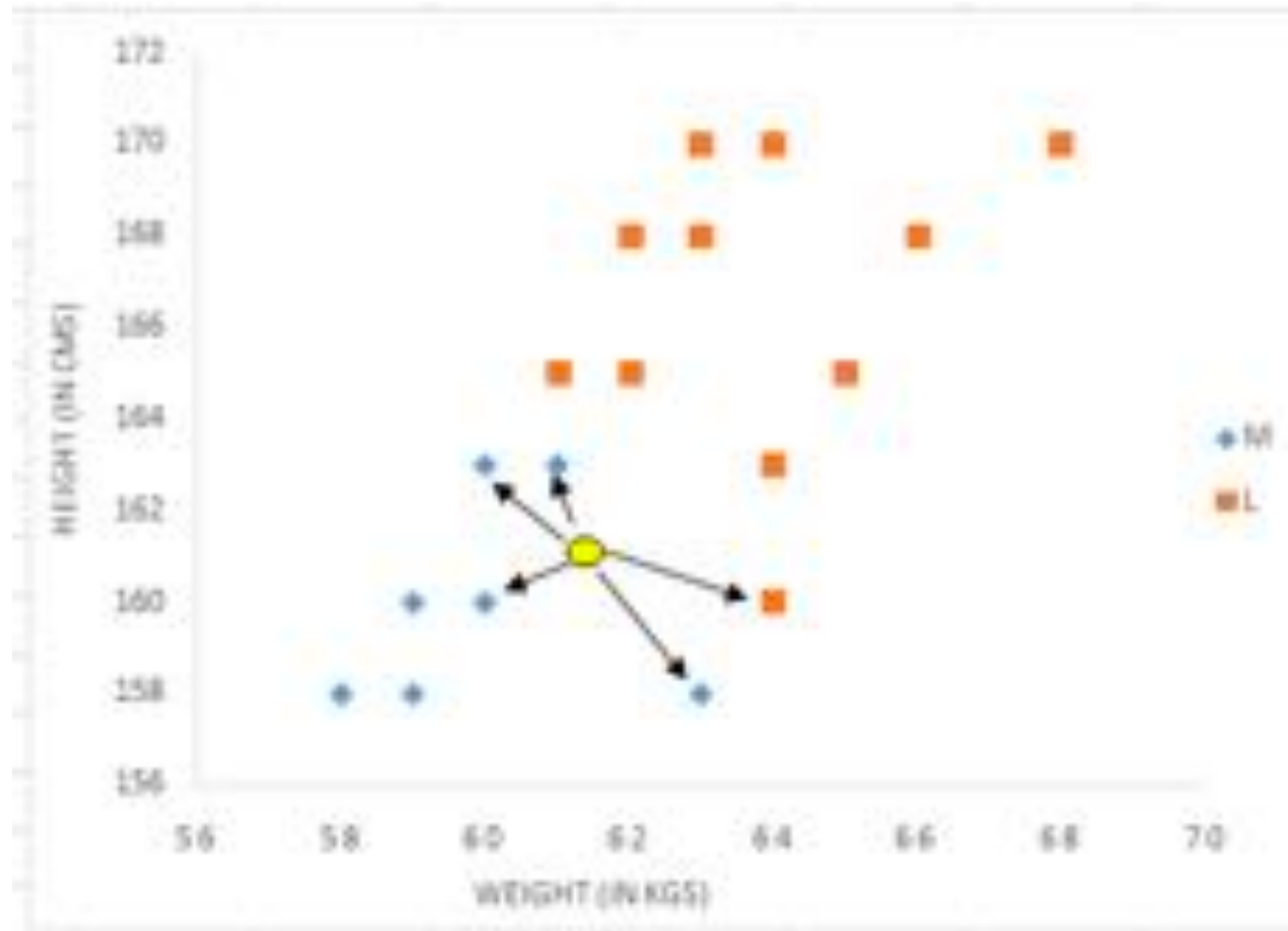
DeepCpG: Deep neural networks for predicting single-cell DNA methylation



Impute sc DNA data: option2. create tiles with average methylation in the tile

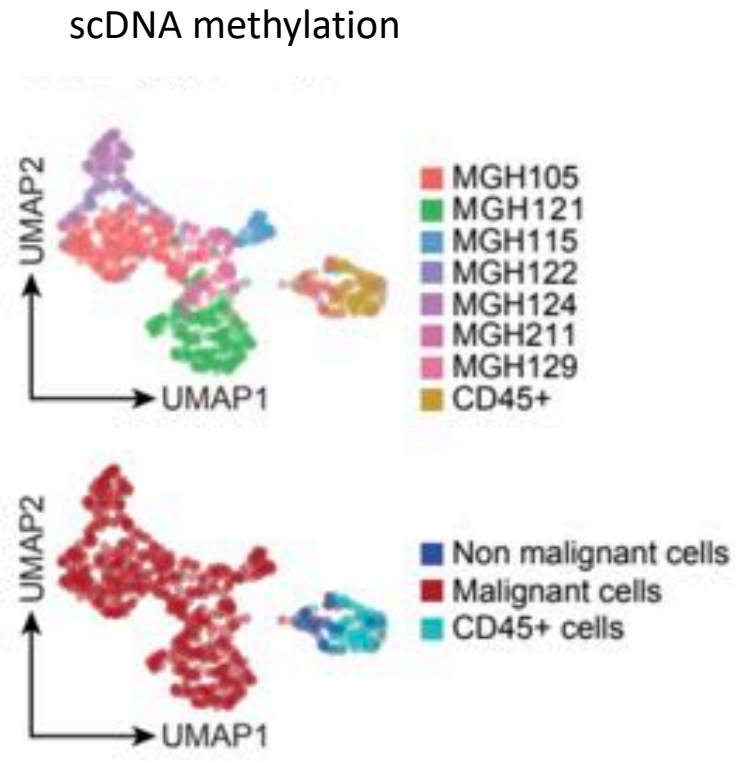


Impute sc DNA data: option3. KNN

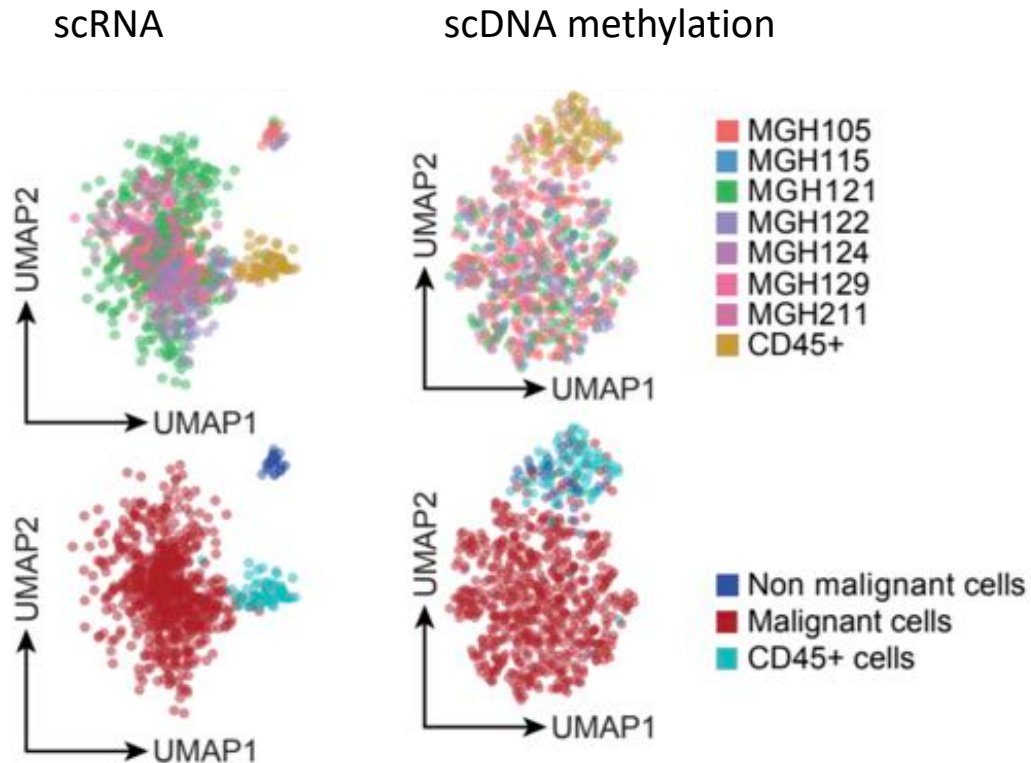


scDNA methylation clustering separates non-malignant cells from malignant cells

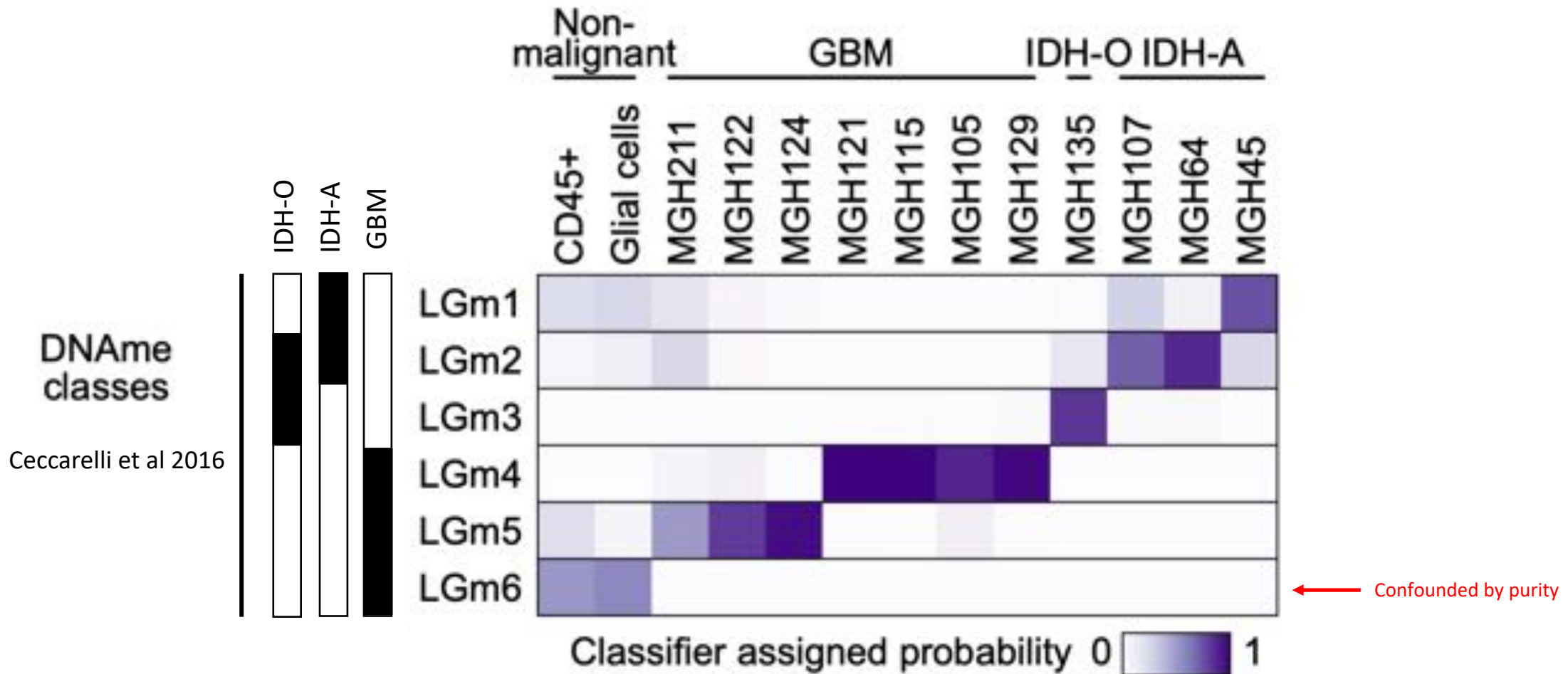
without batch correction



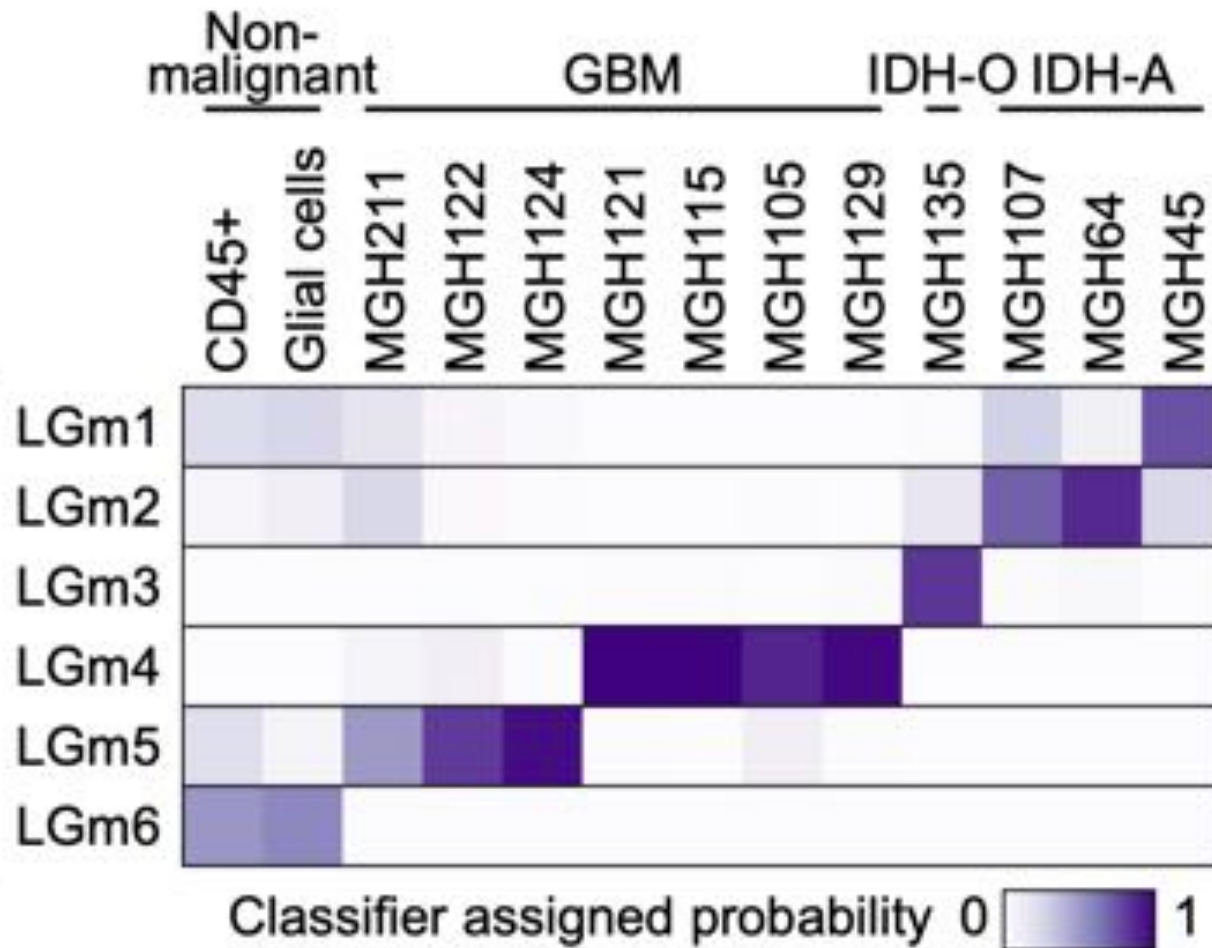
with batch correction



Mapping the tumors to previously defined classes recapitulates and refines previous classification

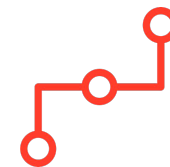


Mapping the tumors to previously defined classes recapitulates and refines previous classification

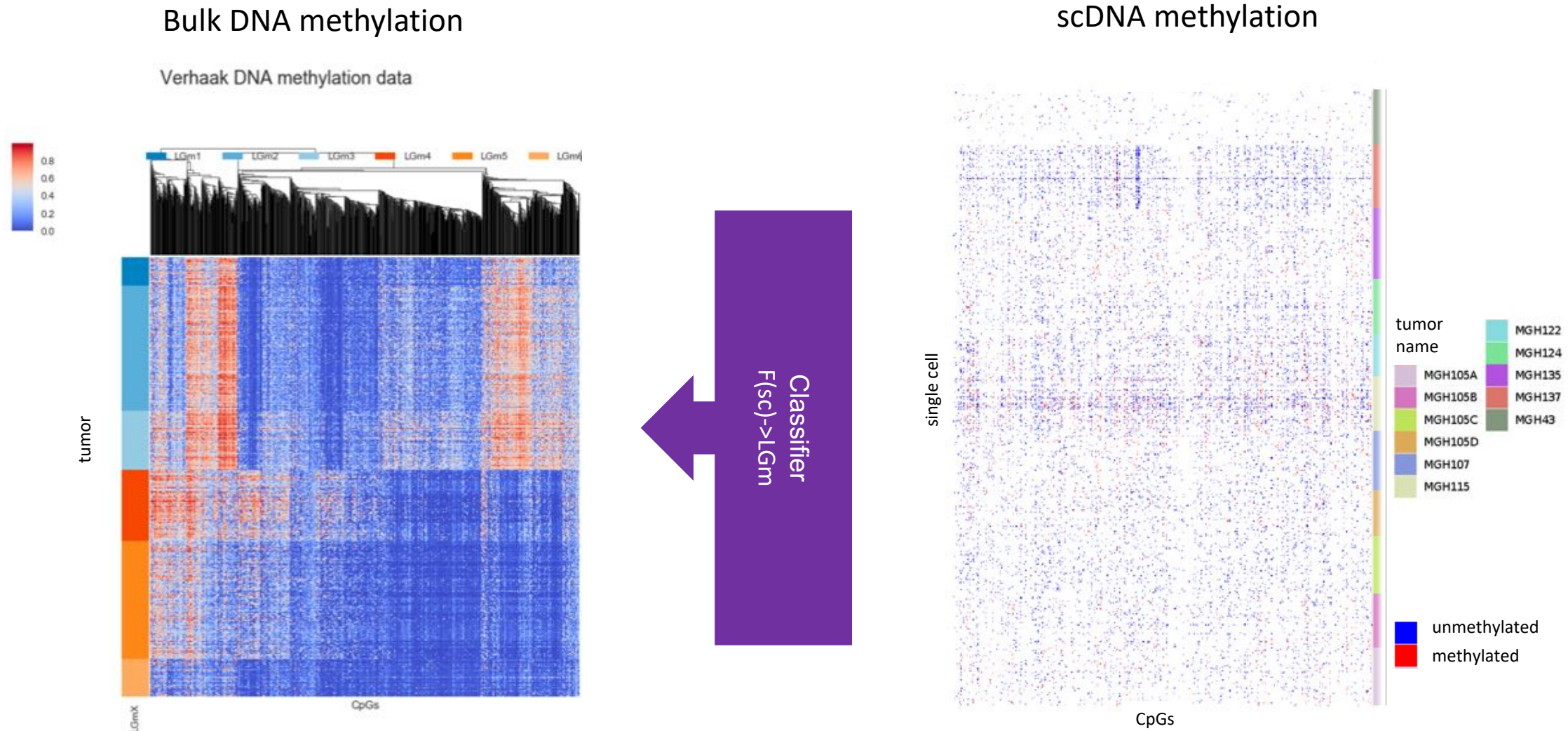


Translating single cell RRBS data to bulk 450K data is possible via machine learning techniques:

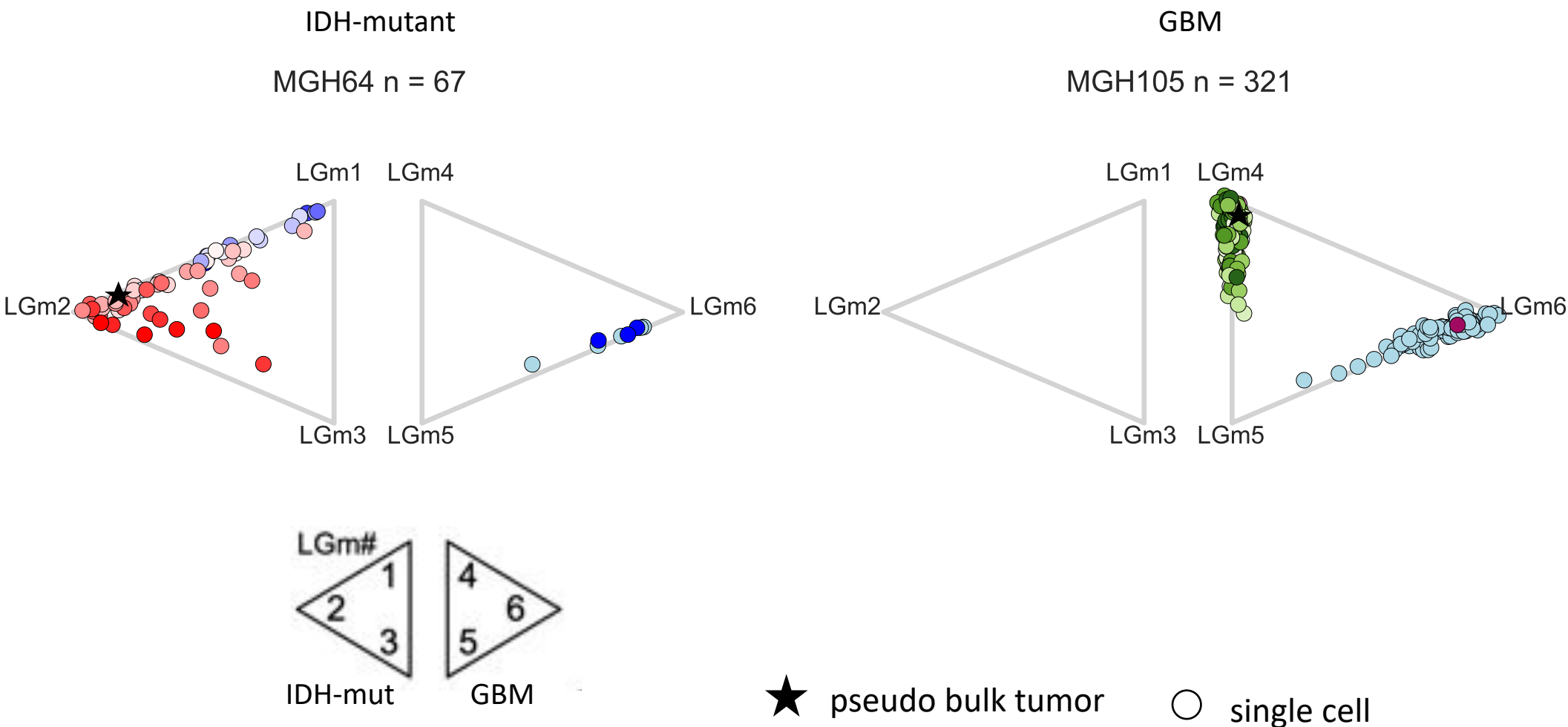
- We map scRRBS probes to the 450K probes
- Train a logistic regression classifier to translate from scRRBS to 450K



Mapping the tumors to previously defined classes recapitulates and refines previous classification



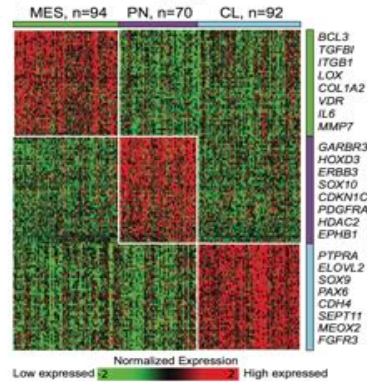
Gliomas are heterogenous on the DNA methylation level



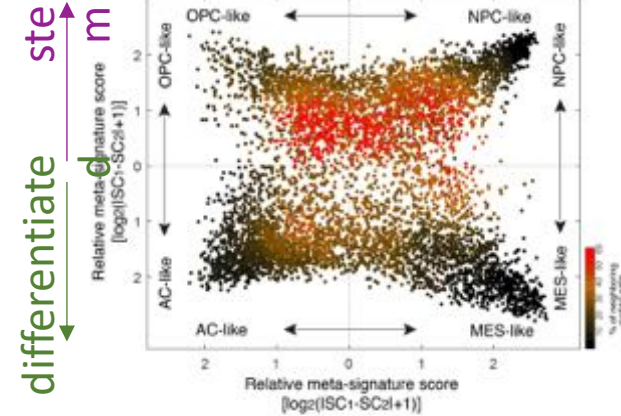
The goal: dissecting epigenetic identity of cellular states underlying glioma evolution

RNA

inter-tumor heterogeneity

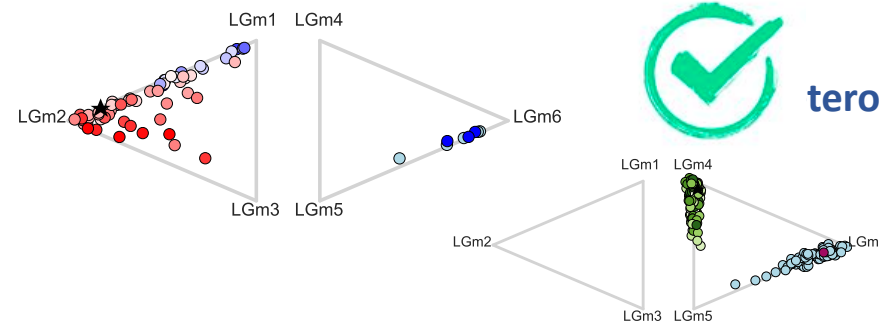
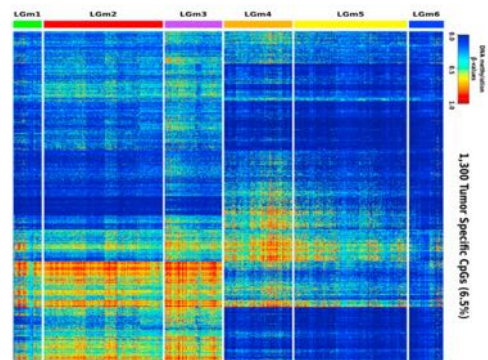


intra-tumor heterogeneity



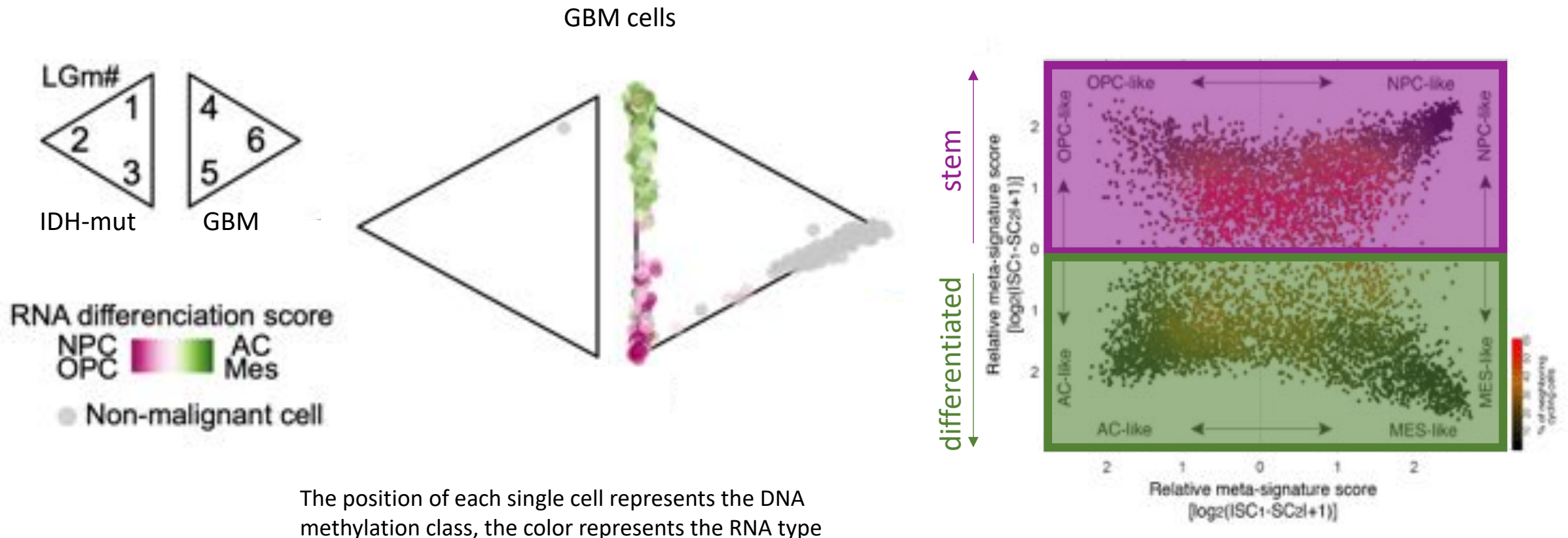
Is there a link between the RNA function and DNA methylation?

DNA methylation



The DNA methylation heterogeneity stems from different cells composition

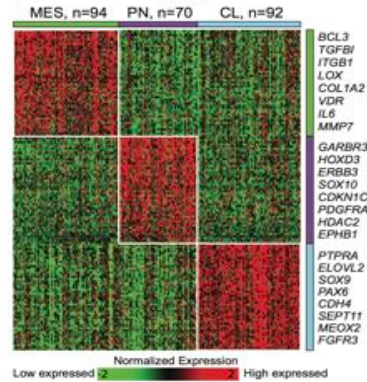
Finding association between DNAm state to RNA state



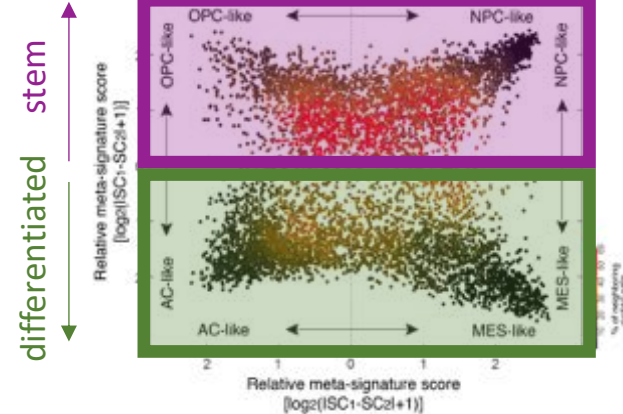
The goal: dissecting epigenetic identity of cellular states underlying glioma evolution

RNA

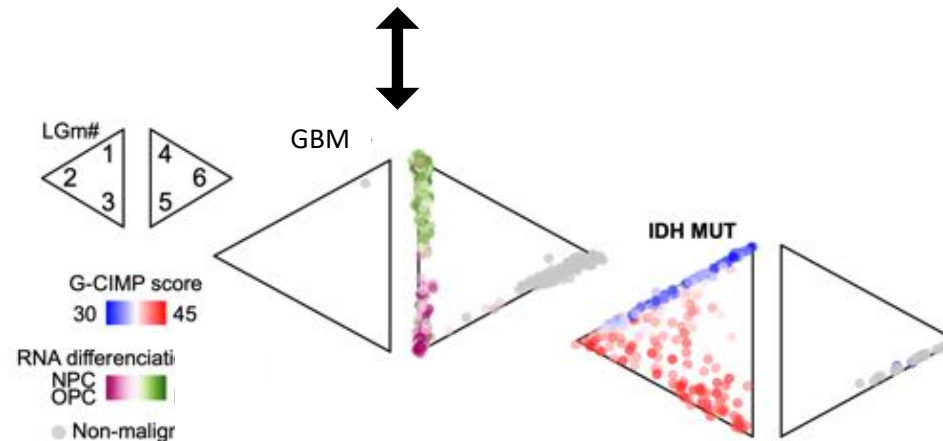
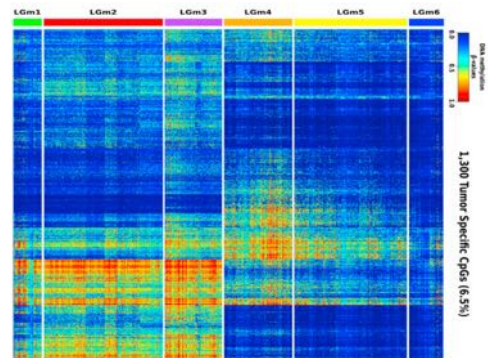
inter-tumor heterogeneity



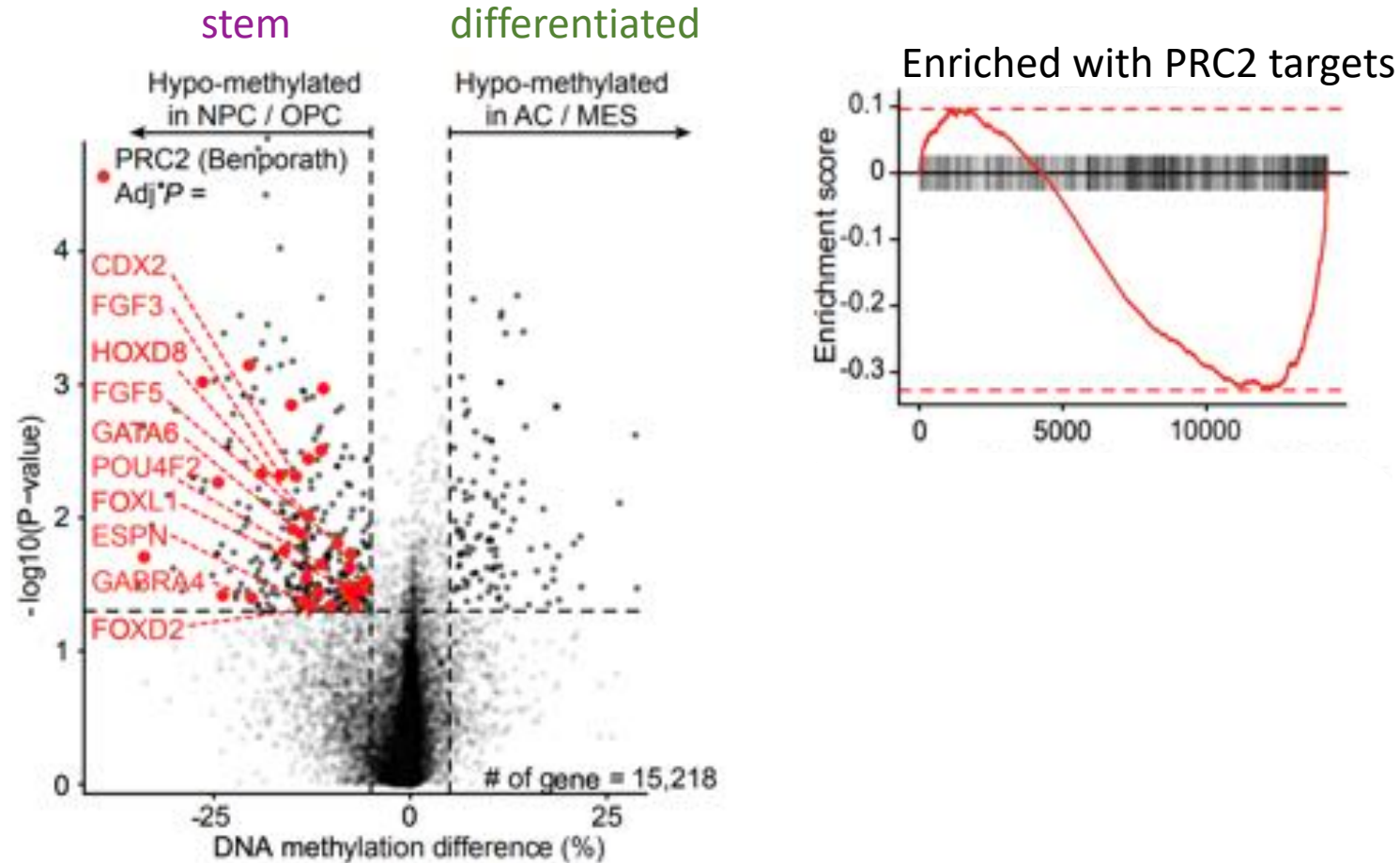
intra-tumor heterogeneity



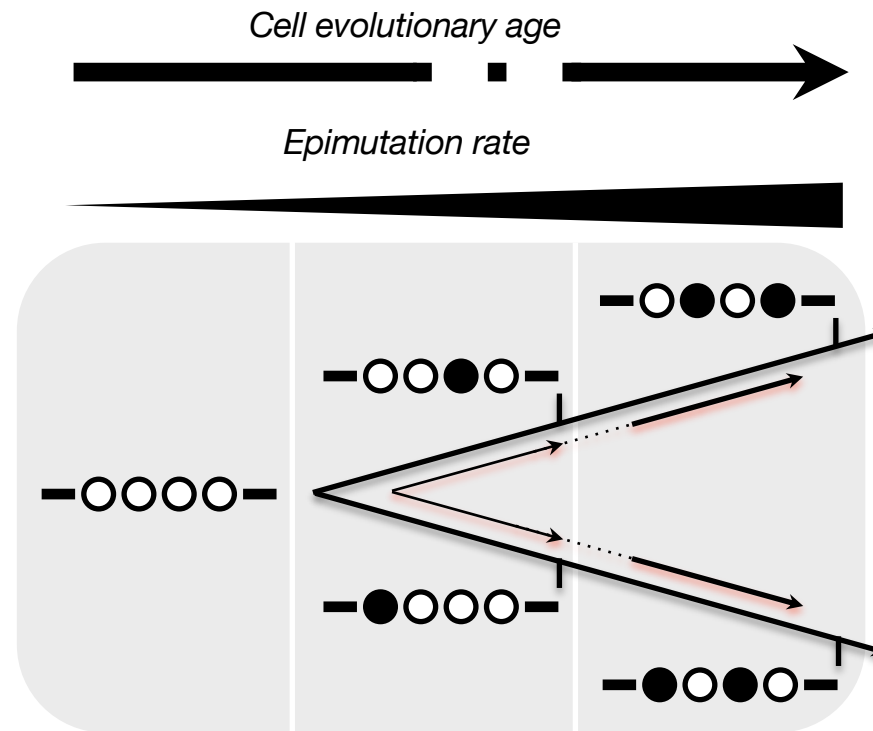
DNA methylation



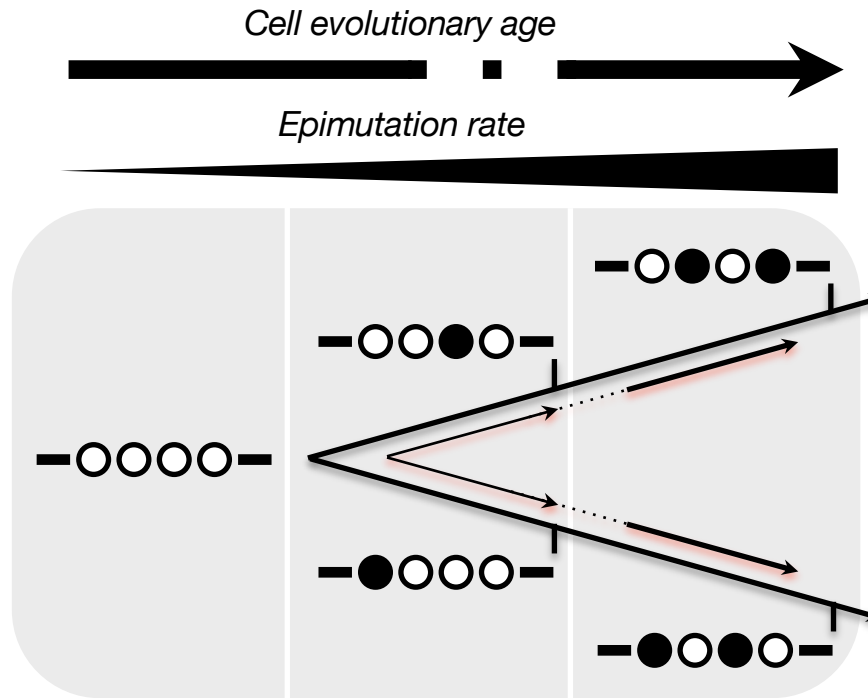
Differentially methylated events in single cells identifies PRC2 targets as hypo methylated in stem states



Epimutation rate - a molecular clock



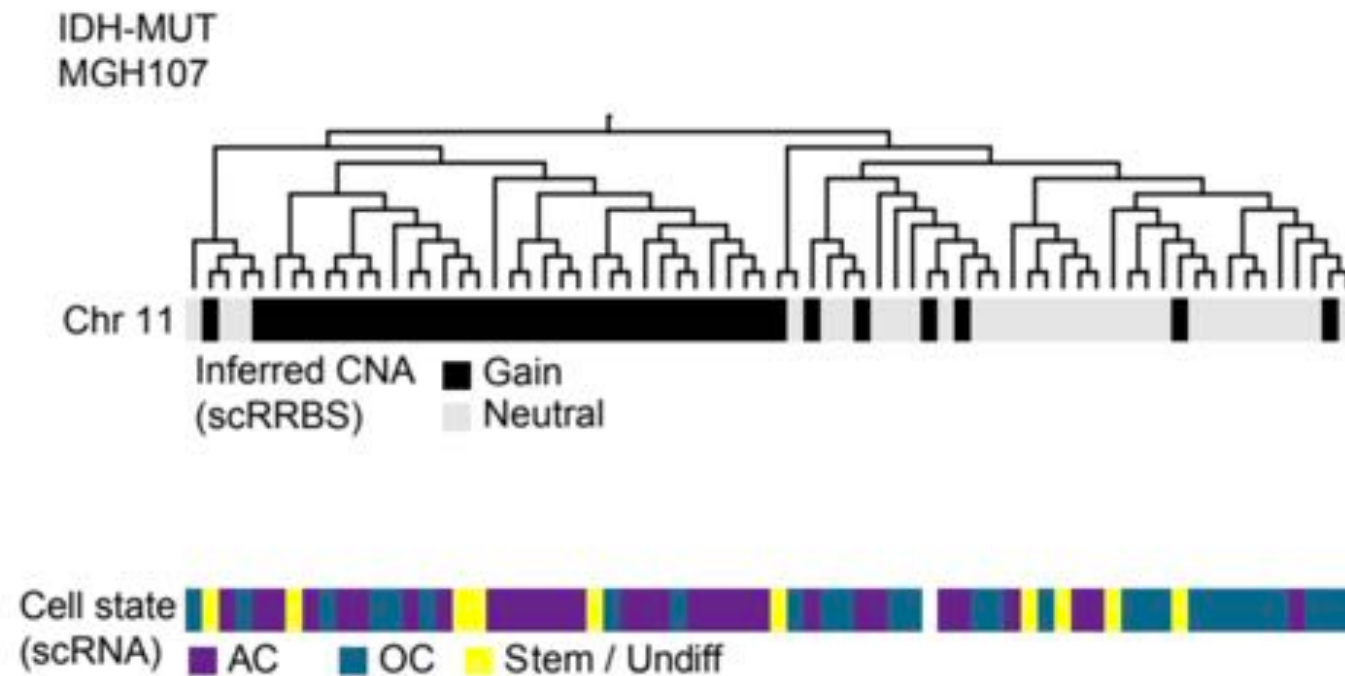
Motivation for epi-phylogenetic trees



Epimutation rate \gg mutation rate

Lineage reconstruction at low sequencing cost and despite single-cell data sparsity

epi-phylogenetic trees reveal tumorigenesis in glioma



scATACseq

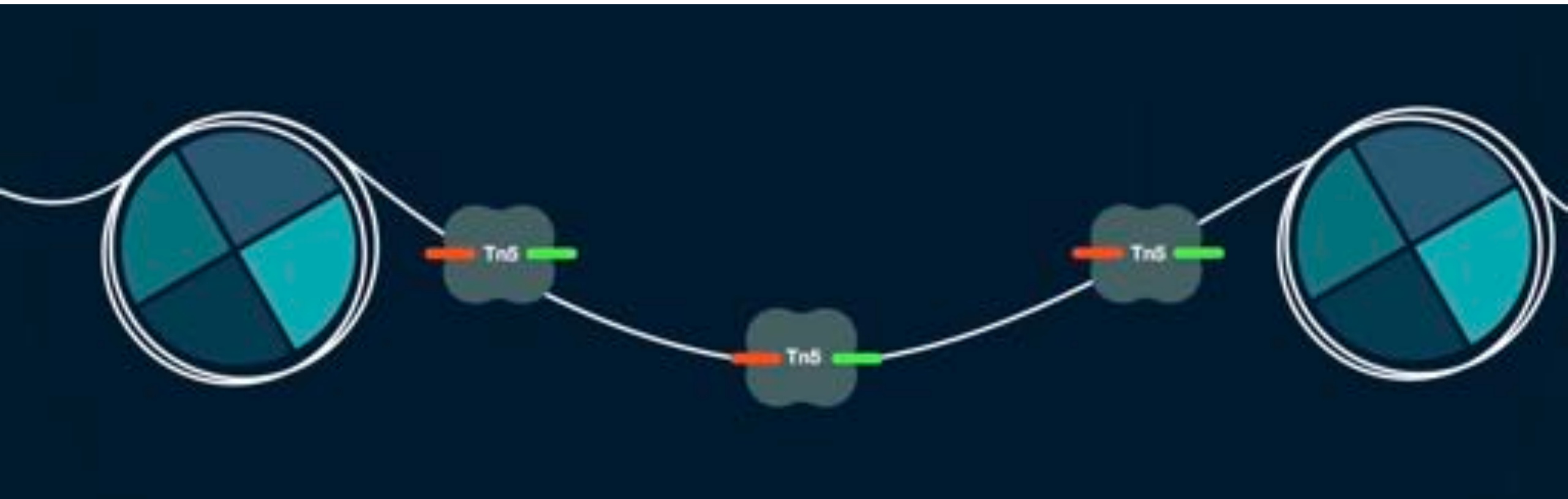
ATACseq measures accessible chromatin

In the cell nucleus, the chromosomes contain tightly packed chromatin material. Part of the chromatin is open and accessible to many regulatory factors who control the expression and suppression of a variety of genes.

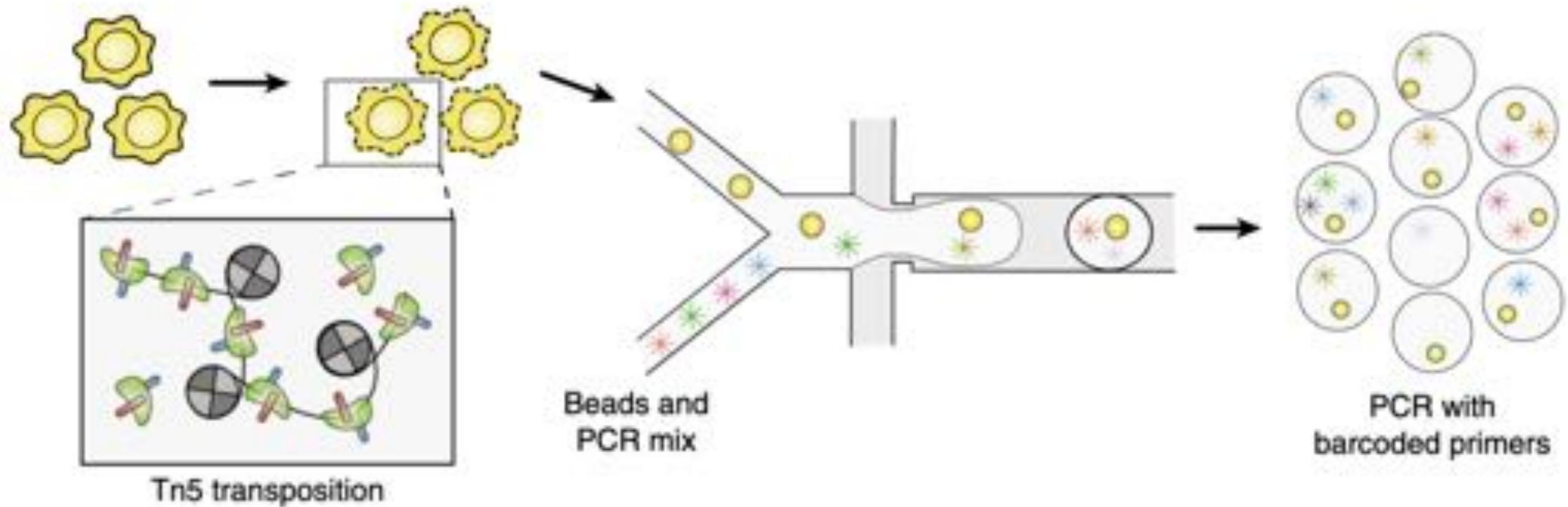


ATACseq measures accessible chromatin

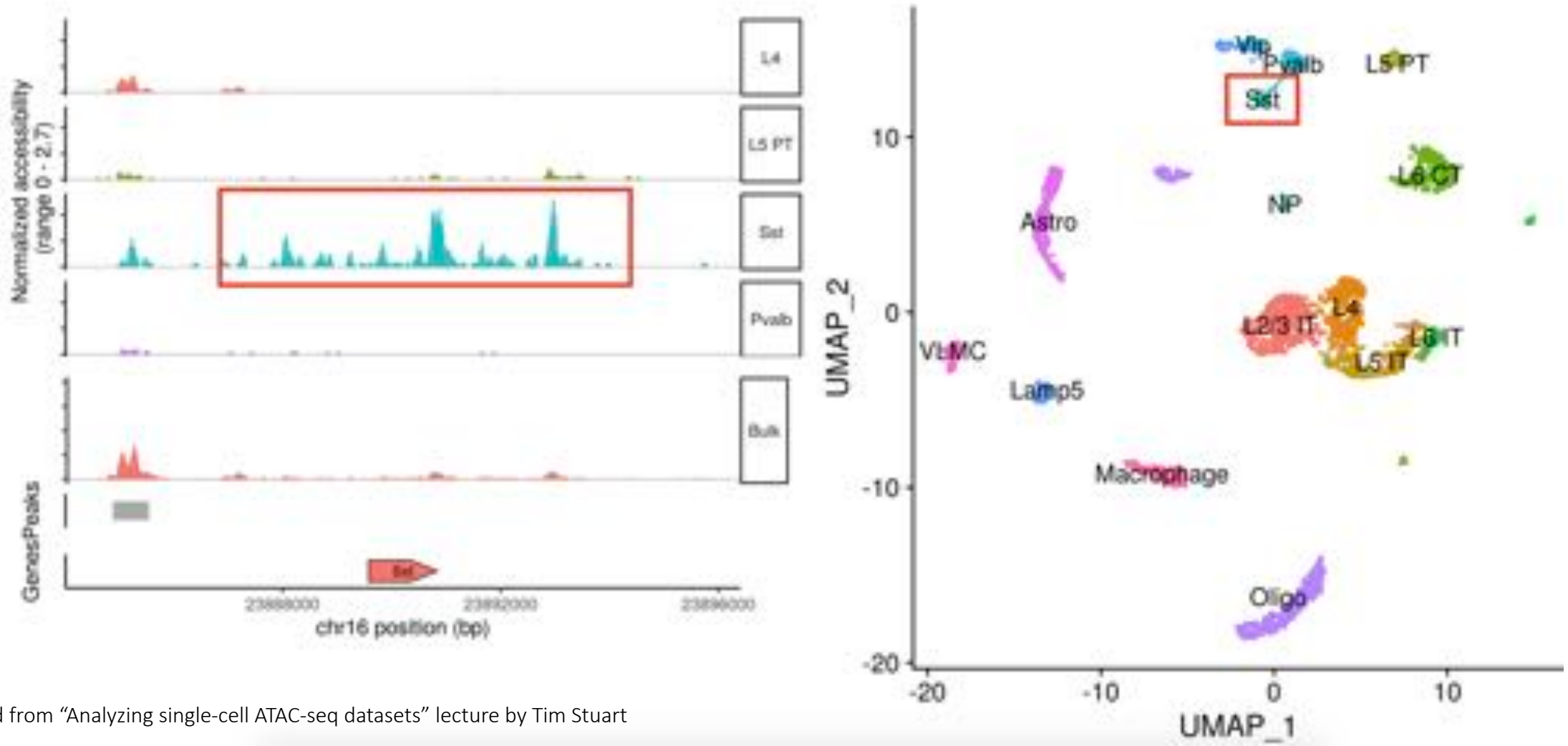
ATACseq (as well as scATACseq) measures how open this piece of DNA is. This openness is a proxy of how easily a transcription factor can bind these parts of the genome. ATACseq measures by using an enzyme called Tn5 transposase which binds open chromatin and inserts DNA sequencing adapters.



Each cell is captured in a droplet with cell barcode



single cell resolution is crucial



A full list of **all** unique fragments across all single cells, as opposed to only reads that map to peaks.

scATACseq data

1. Indexed fragment file

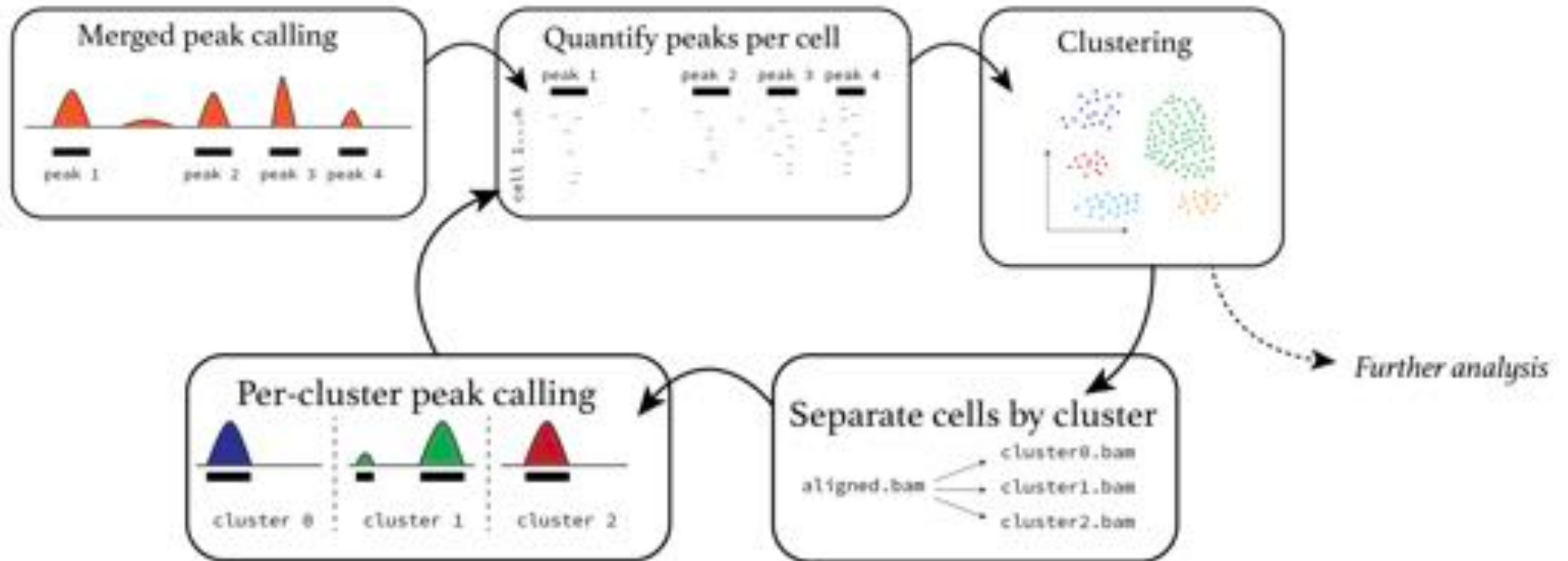
chrom	start	stop	barcode	reads
chr1	3000141	3000517	GGTTGCGAGCCGCAAA-1	3
chr1	3000159	3000373	CTCAGCTAGTGTCCT-1	1
chr1	3000431	3000621	GAAGTCTGTAACTC-1	1

2. Large sparse matrix

	AAACGAAAGAGTTTGA-1	AAACGAAAGCGAGCTA-1
chr1:565107-565550	.	.
chr1:569174-569639	.	.
chr1:713460-714823	.	2
chr1:752422-753038	.	.
chr1:762106-763359	.	4

Each value in the matrix represents the number of Tn5 cut sites for each single barcode (i.e. cell) that map within each peak

Defining features for scATACseq



scATACseq data

1. Indexed fragment file

chrom	start	stop	barcode	reads
chr1	3000141	3000517	GGTTGCGAGCCGCAAA-1	3
chr1	3000159	3000373	CTCAGCTAGTGTCACT-1	1
chr1	3000431	3000621	GAAGTCTGTAACTC-1	1

2. Large sparse matrix

	AAACGAAAGAGTTTGA-1	AAACGAAAGCGAGCTA-1
chr1:565107-565550	.	.
chr1:569174-569639	.	.
chr1:713460-714823	.	2
chr1:752422-753038	.	.
chr1:762106-763359	.	4

Use LSI instead of PCA

Originally developed for topic modeling / natural language processing (Deerwester et al. 1990)

First applied to scATAC-seq in 2015 (Cusanovich et al. *Science*)

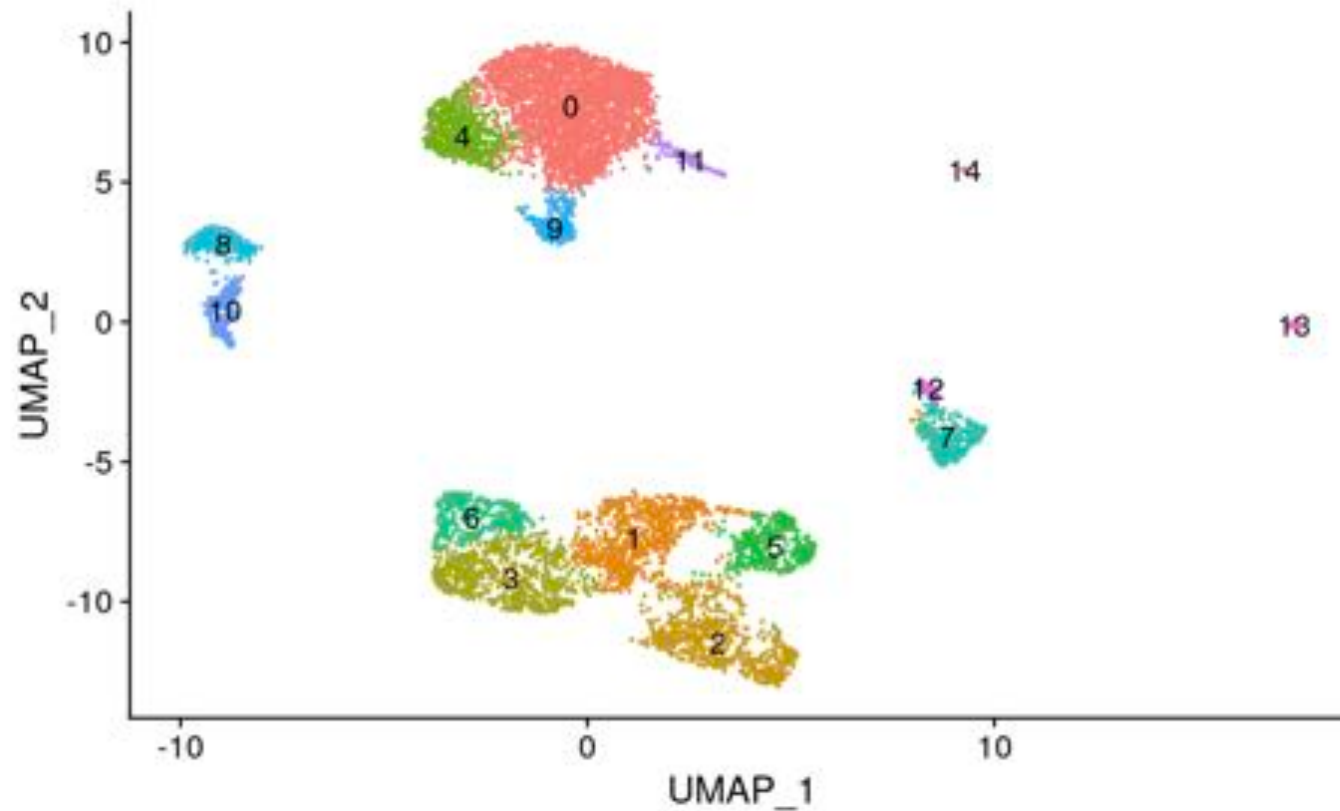
Cell = document, peak = word

LSI combines local (within-cell) and global (across-population) weighting functions then applies singular value decomposition

Multiple different local and global weighting functions that can be used

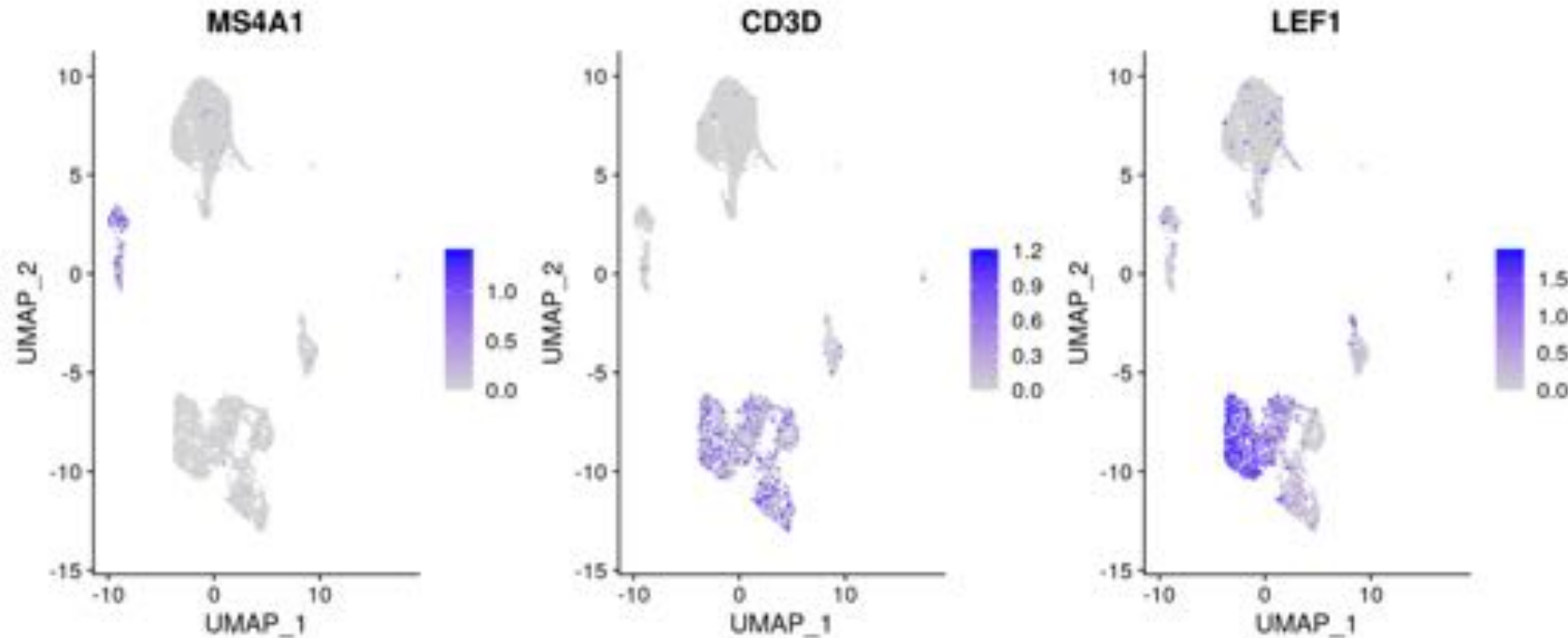
scATAcseq yields well separated clusters

```
pbmc <- RunUMAP(object = pbmc, reduction = 'lsi', dims = 2:30)
pbmc <- FindNeighbors(object = pbmc, reduction = 'lsi', dims = 2:30)
pbmc <- FindClusters(object = pbmc, verbose = FALSE, algorithm = 3)
DimPlot(object = pbmc, label = TRUE) + NoLegend()
```

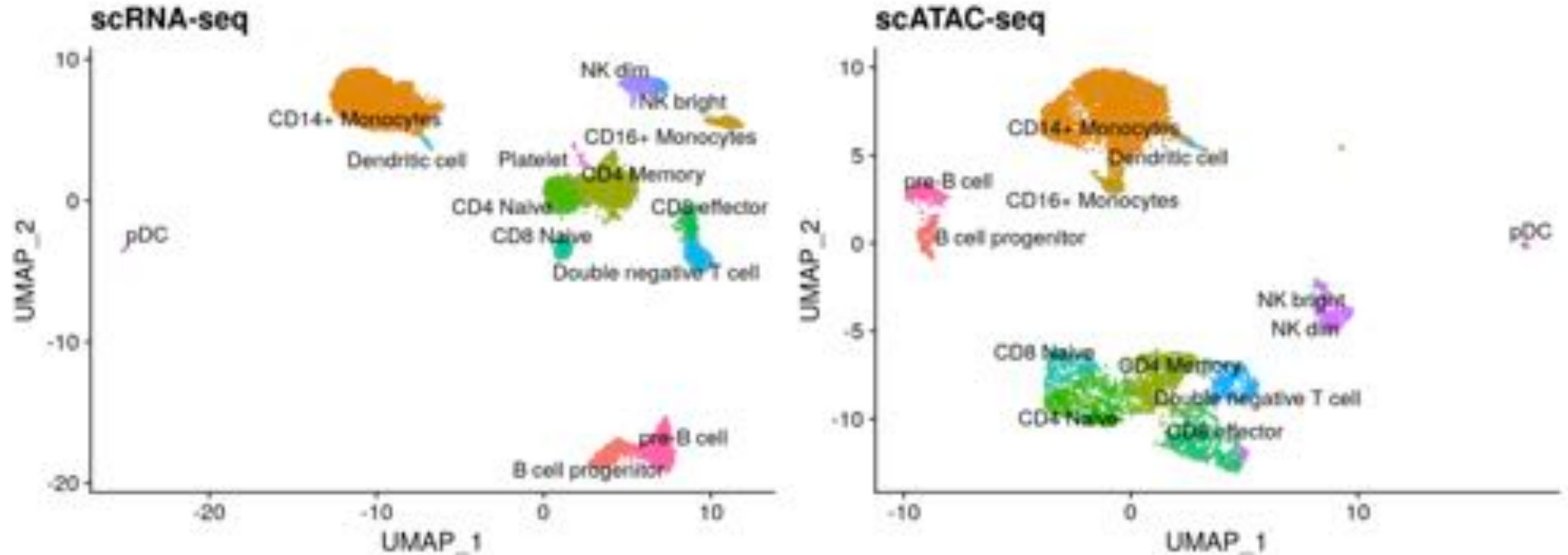


Use fragments file to infer gene activity and annotate clusters

Quantify the activity of each gene in the genome by assessing the chromatin accessibility associated with each gene: count the number of fragments for each cell that map to the promoter + gene body

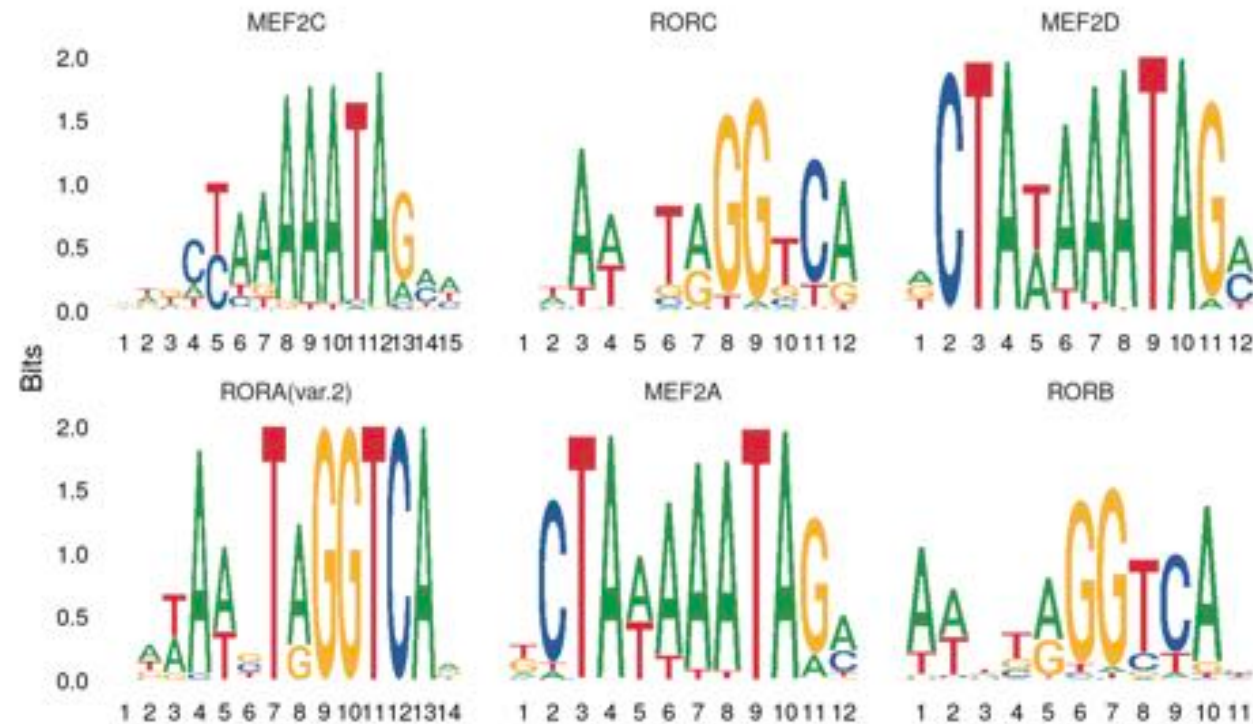


Integrating with scRNA-seq data using CCA



Finding overrepresented motifs

To identify potentially important cell-type-specific regulatory sequences, signac searches for DNA motifs that are overrepresented in a set of peaks that are differentially accessible between cell types.



Computing motif activities

ChromVAR identifies motifs associated with variability in chromatin accessibility between cells.

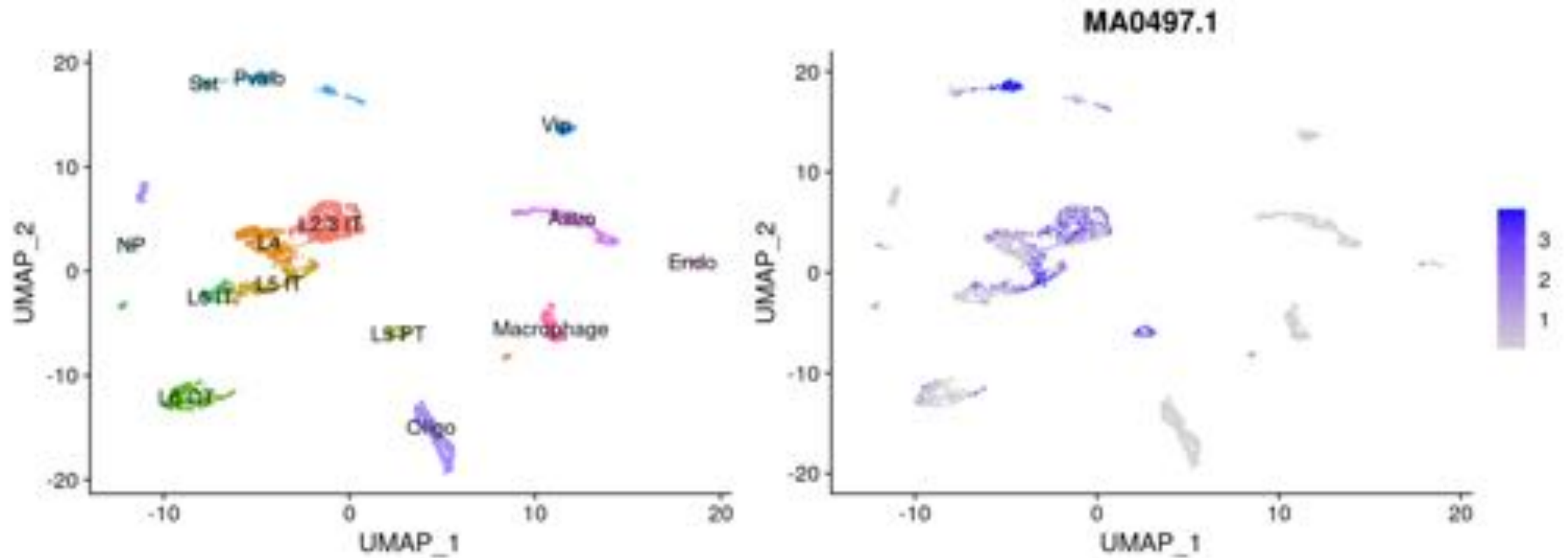


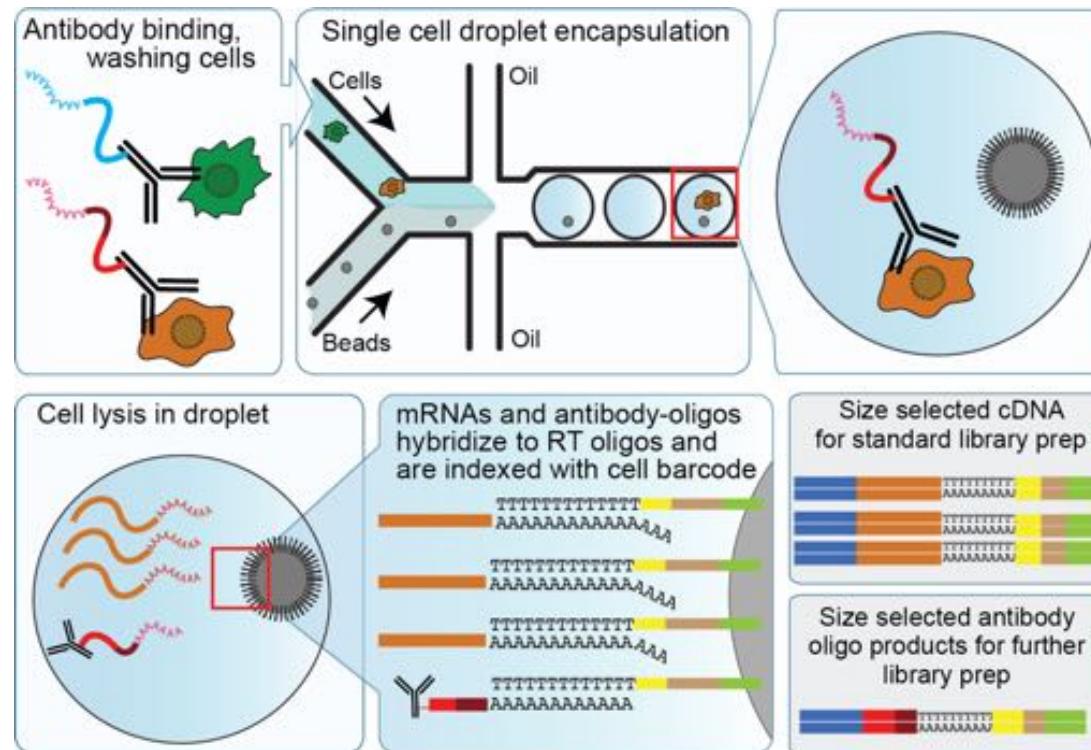
figure adopted from Motif analysis with Signac tutorial, https://satijalab.org/signac/articles/motif_vignette.html

Software solutions

<i>Signac</i>	Extension of Seurat of single-cell chromatin data	https://github.com/timoast/signac
<i>SnapATAC</i>	Analysis pipeline for single-cell ATAC-seq	https://github.com/r3fang/SnapATAC
<i>cisTopic</i>	Probabilistic modeling of cis-regulatory topics	https://github.com/aertslab/cisTopic
<i>chromVAR</i>	Chromatin variability across regions	https://github.com/GreenleafLab/ chromVAR https://github.com/caleblareau/gchromVAR
<i>CICERO</i>	Predicting the cis-regulatory landscape	https://github.com/cole-trapnell-lab/cicero- release

CITEseq

CITEseq protocol

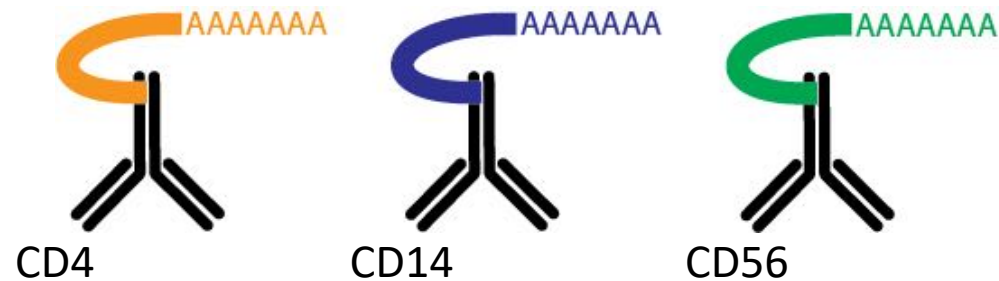


Count matrix1: scRNAseq

Count matrix2: CITEseq

Input

- List of cell surface proteins of interest:
- For example, I chose: CD4, CD14, CD56



Output

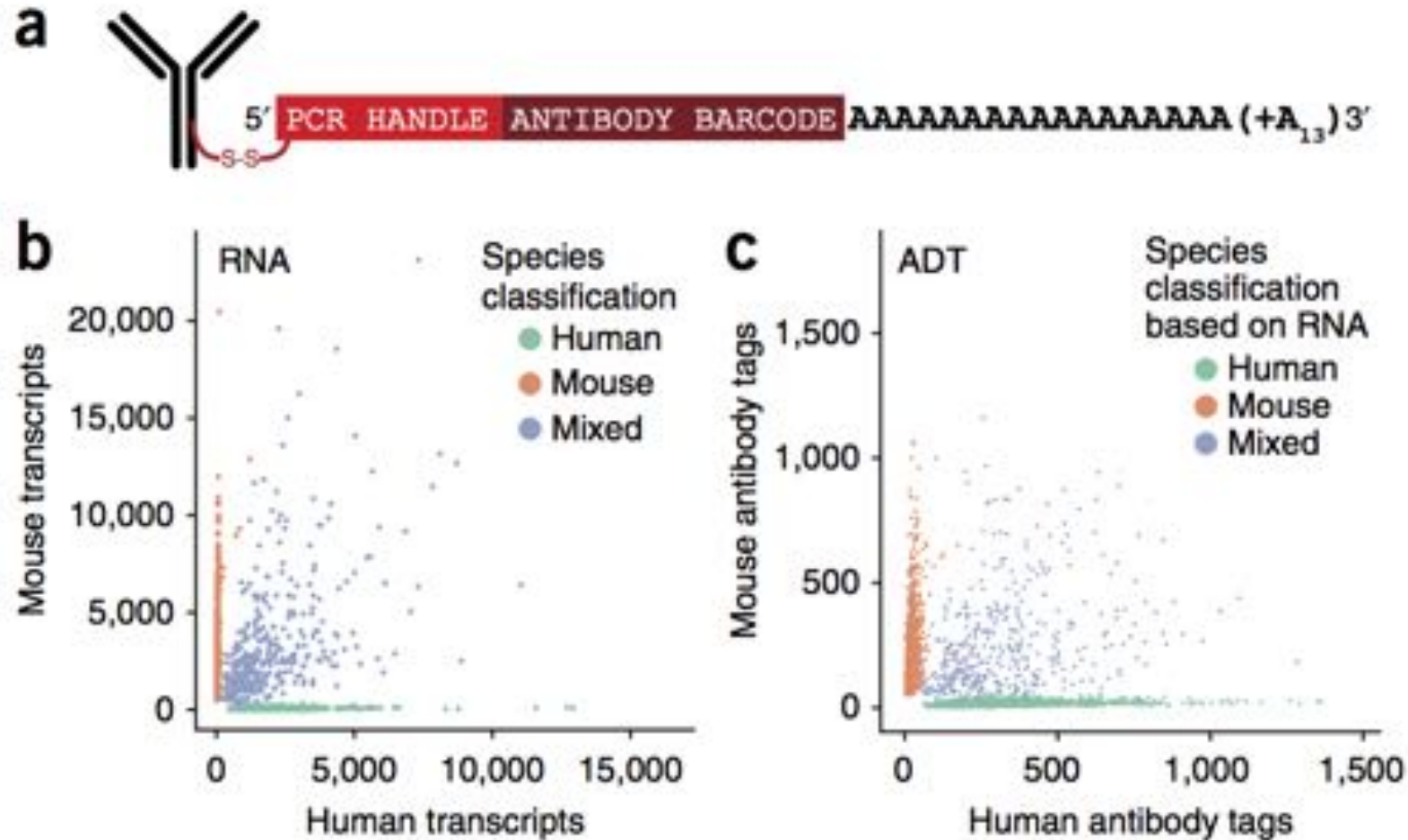
scRNAseq

	cell1	cell2	cell3	cell4
Gene1	0	0	0	4
Gene2	0	2	0	0
Gene3	0	0	0	0

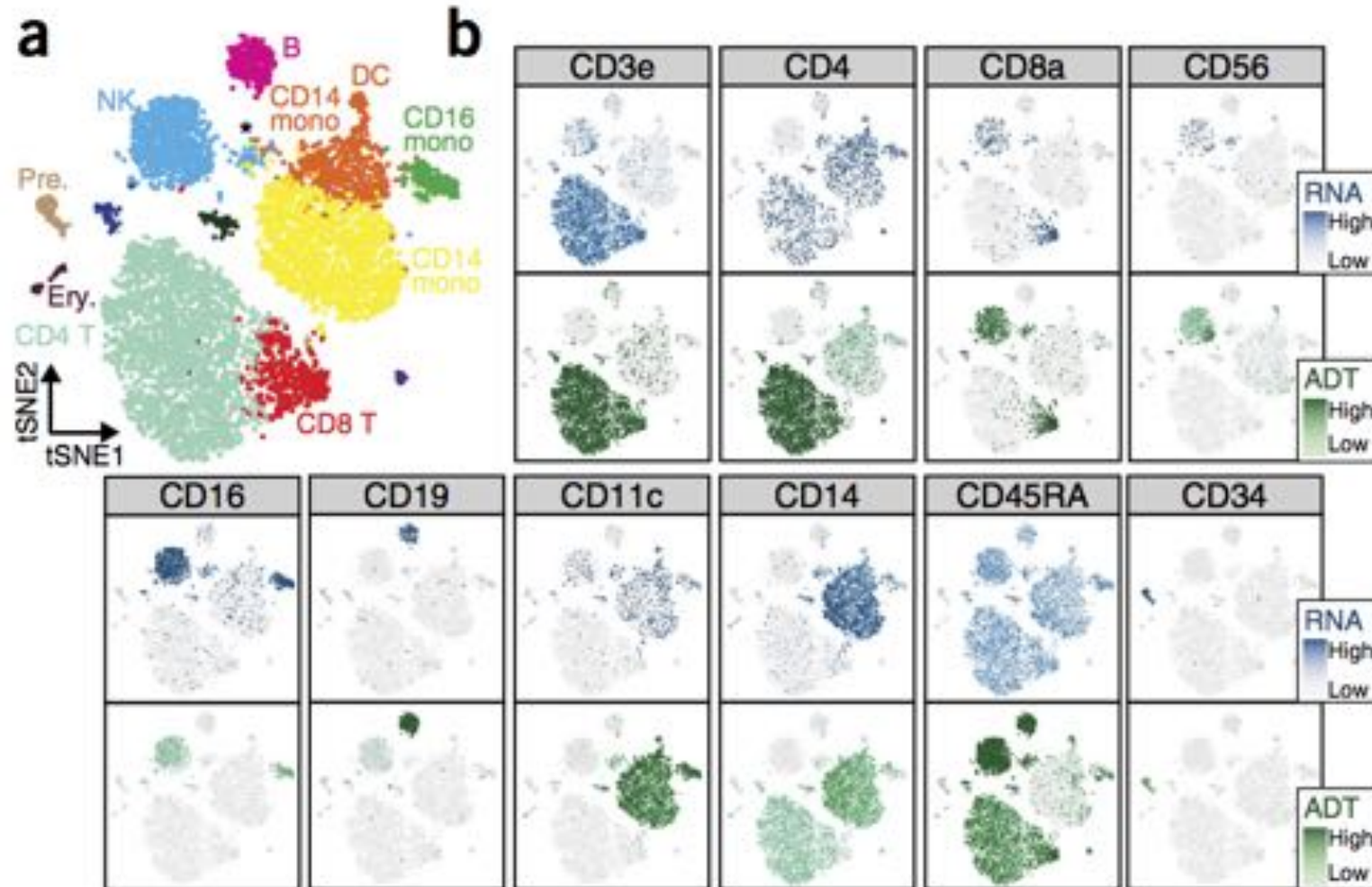
CITEseq

	cell1	cell2	cell3	cell4
Antibody for CD4	100	0	0	38
Antibody for CD14	0	0	45	0
Antibody for CD56	2	0	0	0

Simultaneous protein and RNA measurement in single cells

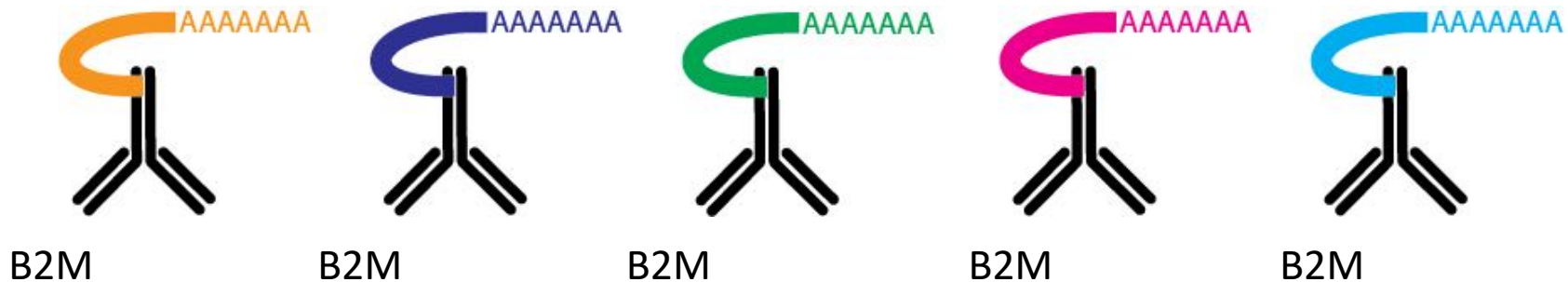


Simultaneous protein and RNA measurement in single cells



Cell hashing using CITEseq

- Cell Hashing is a method that enables sample multiplexing and super-loading on single cell RNA-sequencing platforms.
- Cell Hashing uses a series of oligo-tagged antibodies against ubiquitously expressed surface proteins to uniquely label cells from distinct samples, which can be subsequently pooled in one scRNA-seq run.



Overloading

By sequencing these tags alongside the cellular transcriptome, we can assign each cell to its sample of origin, and robustly identify doublets originating from multiple samples.

