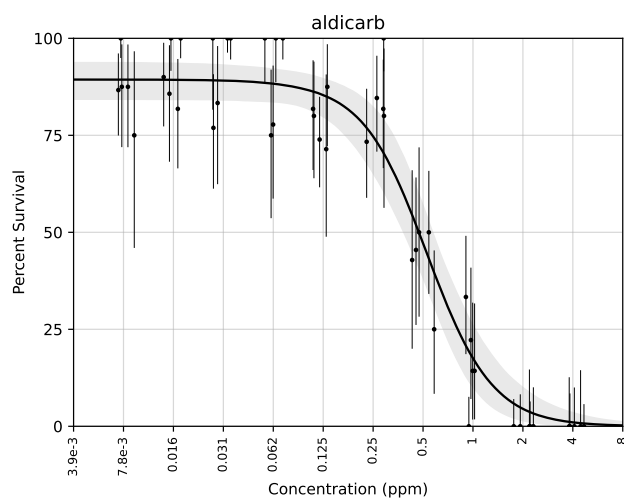
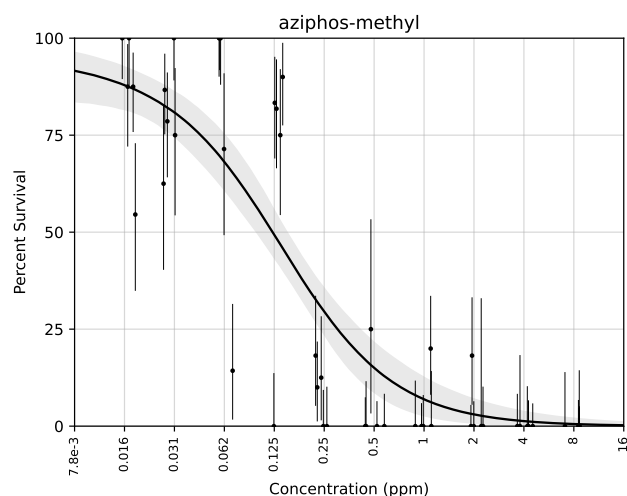


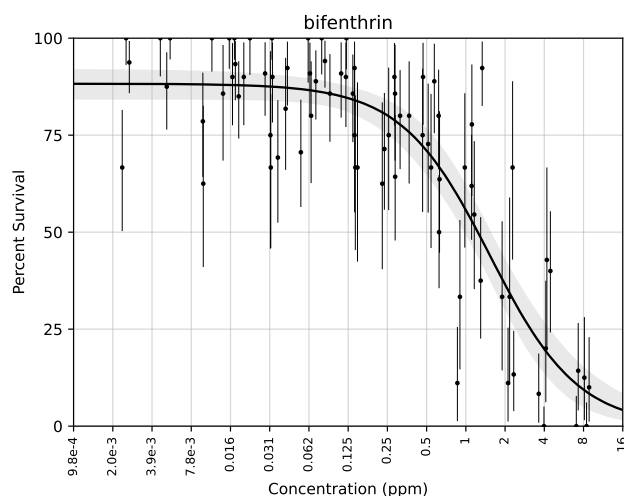
Acetamiprid LC_{50} : 1.39 ppm [0.992, 1.91]
4 biol. reps; 5 tech. reps; R^2 : 0.82



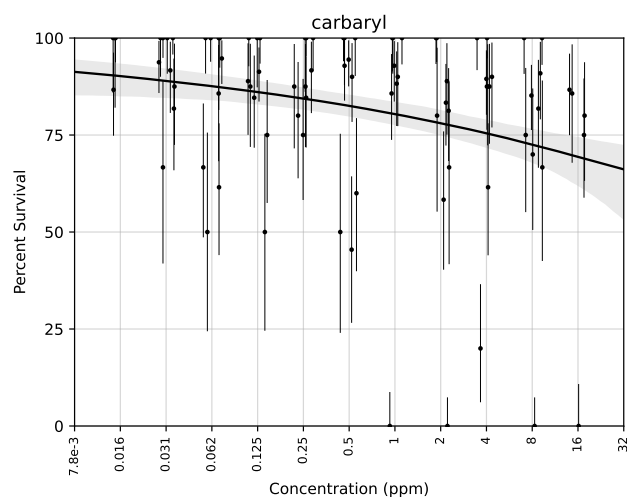
Aldicarb LC_{50} : 0.526 ppm [0.424, 0.645]
4 biol. reps; 5 tech. reps; R^2 : 0.941



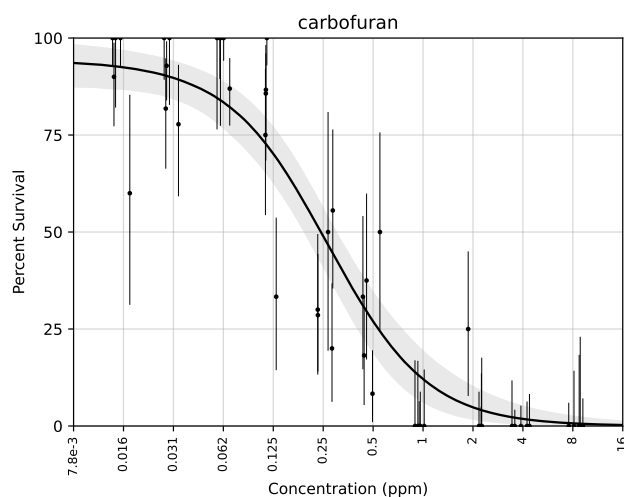
Aziphos-methyl LC_{50} : 0.134 ppm [0.101, 0.176]
4 biol. reps; 5 tech. reps; R^2 : 0.763



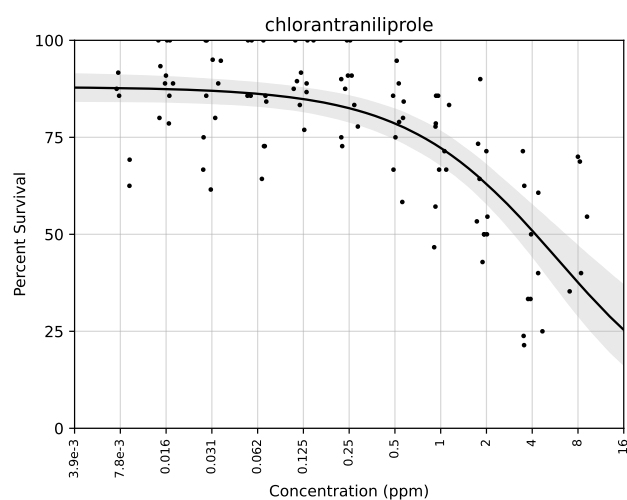
Bifenthrin LC_{50} : 1.52 ppm [1.2, 1.94]
7 biol. reps; 8 tech. reps; R^2 : 0.764



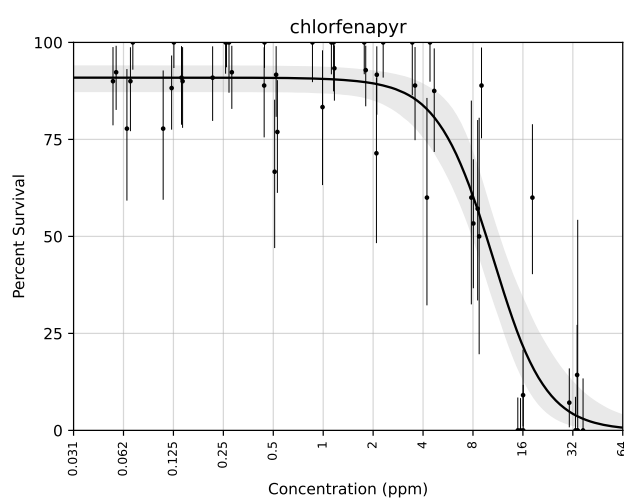
Carbaryl LC_{50} : 778 ppm [42, 2.29e4]
8 biol. reps; 9 tech. reps; R^2 : 0.0779



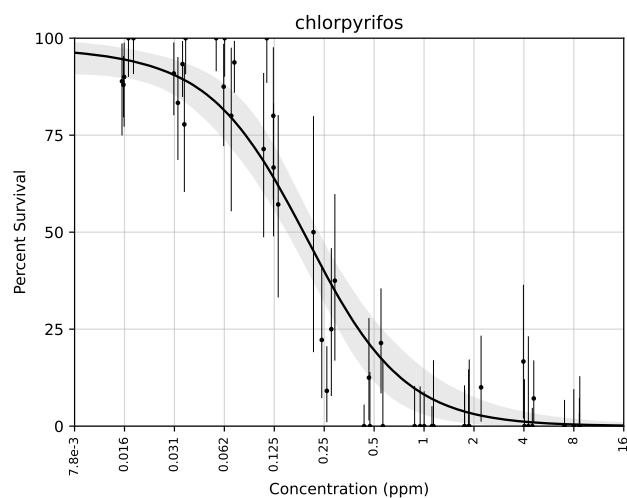
Carbofuran LC_{50} : 0.263 ppm [0.203, 0.335]
4 biol. reps; 5 tech. reps; R^2 : 0.888



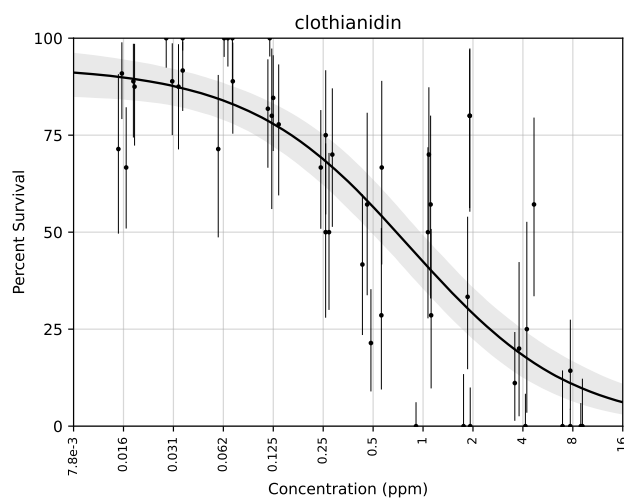
Chlorantraniliprole LC_{50} : 5.75 ppm [3.86, 9.2]
9 biol. reps; 10 tech. reps; R^2 : 0.541



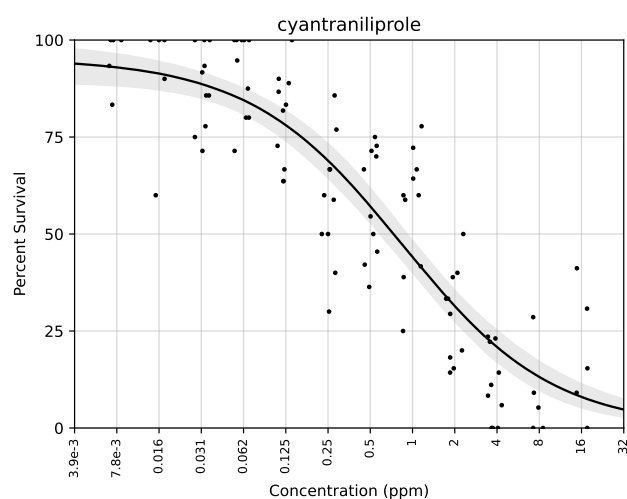
Chlorfenapyr LC_{50} : 10.4 ppm [8.63, 12.6]
4 biol. reps; 5 tech. reps; R^2 : 0.869



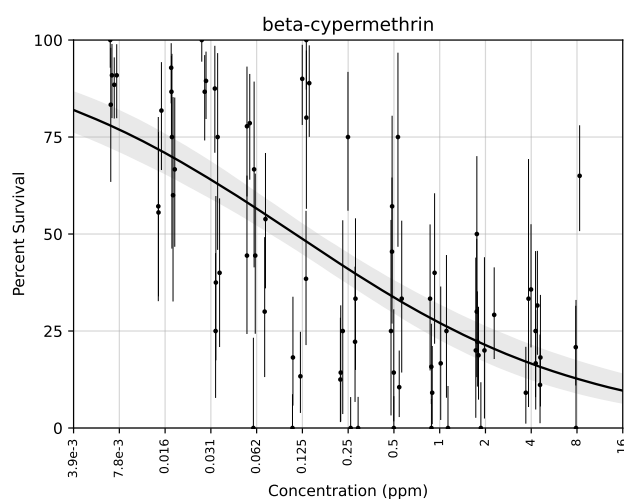
Chlorpyrifos LC_{50} : 0.196 ppm [0.154, 0.243]
4 biol. reps; 5 tech. reps; R^2 : 0.923



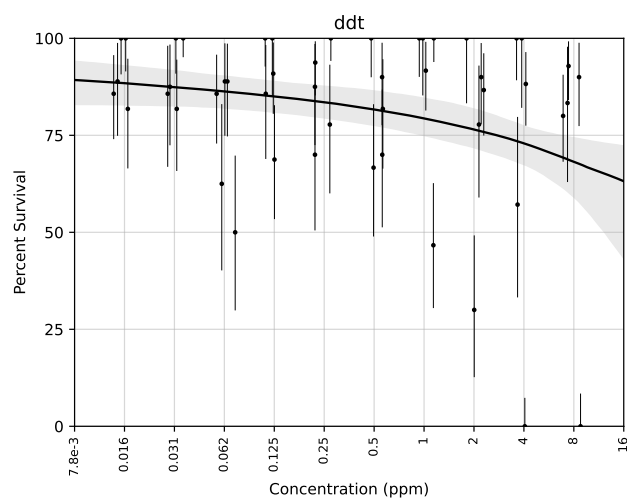
Clothianidin LC_{50} : 0.824 ppm [0.543, 1.23]
4 biol. reps; 5 tech. reps; R^2 : 0.693



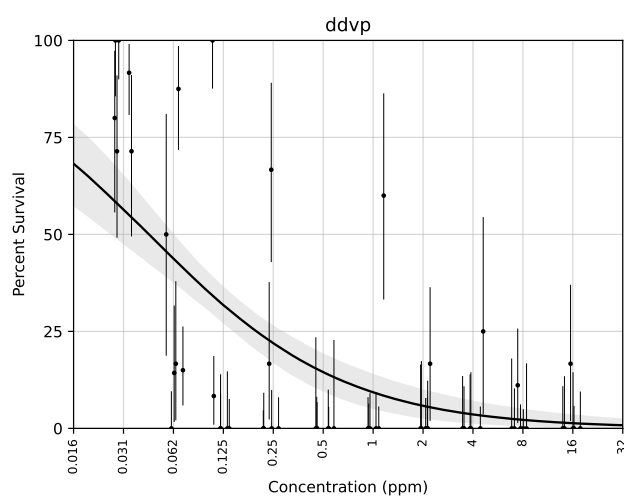
Cyantraniliprole LC_{50} : 0.828 ppm [0.613, 1.13]
9 biol. reps; 10 tech. reps; R^2 : 0.801



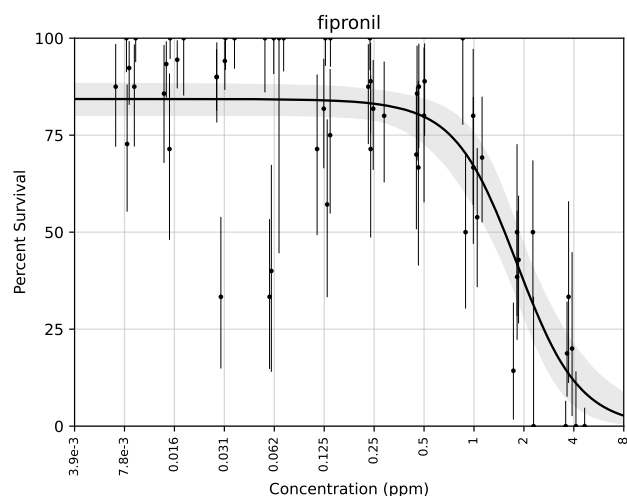
β -Cypermethrin LC_{50} : 0.112 ppm [0.0777, 0.157]
7 biol. reps; 8 tech. reps; R^2 : 0.437



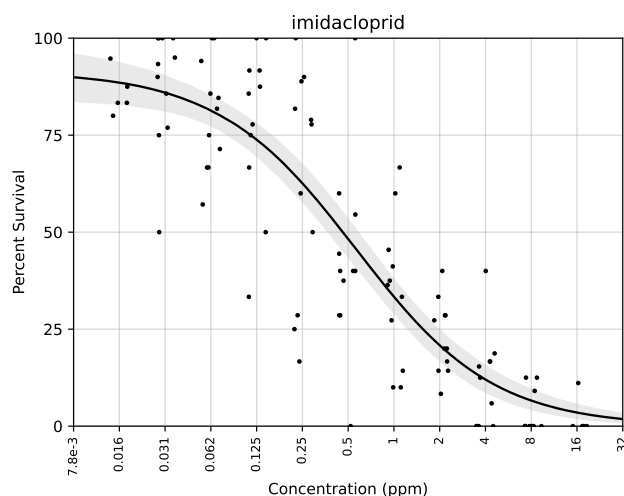
DDT LC_{50} : 117 ppm [10.2, 9.12e3]
 4 biol. reps; 5 tech. reps; R^2 : 0.0807



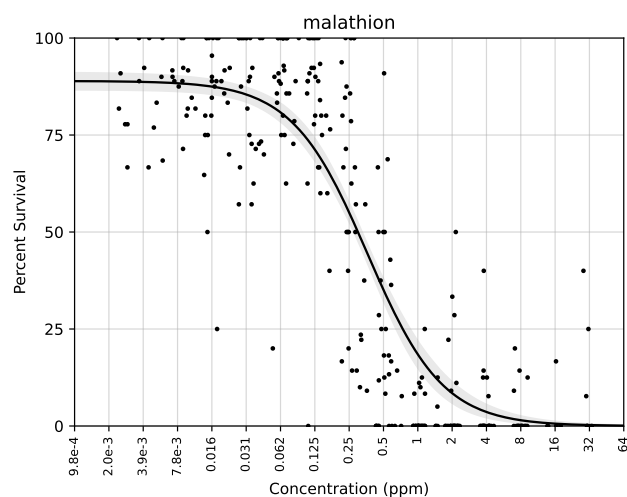
DDVP LC_{50} : 0.0445 ppm [0.0282, 0.065]
 5 biol. reps; 6 tech. reps; R^2 : 0.469



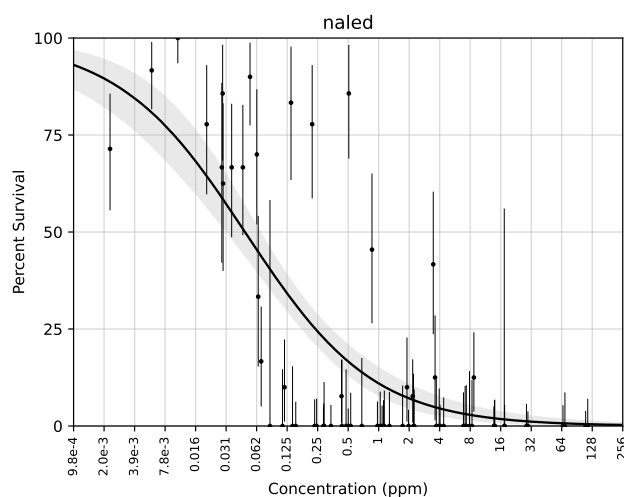
Fipronil LC_{50} : 1.8 ppm [1.48, 2.23]
 5 biol. reps; 6 tech. reps; R^2 : 0.676



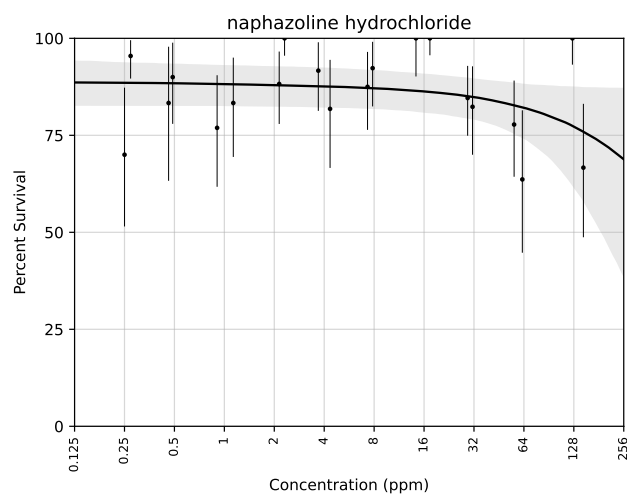
Imidacloprid LC_{50} : 0.562 ppm [0.403, 0.786]
 9 biol. reps; 11 tech. reps; R^2 : 0.776



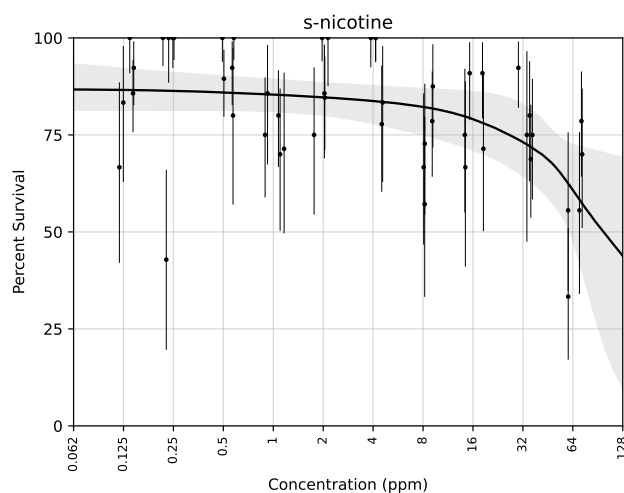
Malathion LC_{50} : 0.362 ppm [0.33, 0.396]
 16 biol. reps; 31 tech. reps; R^2 : 0.789



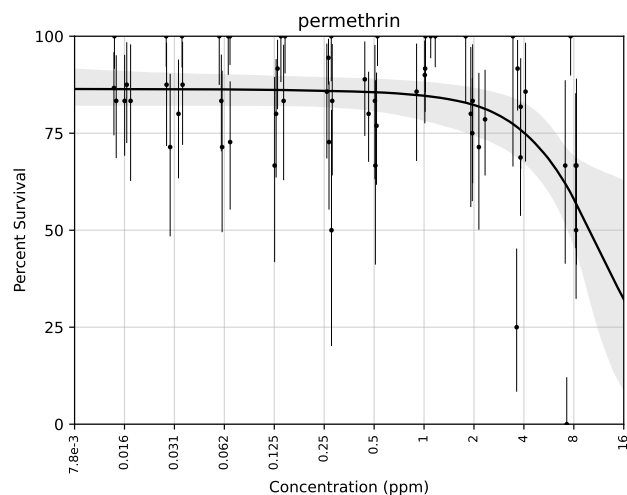
Naled LC_{50} : 0.0486 ppm [0.0321, 0.0718]
 7 biol. reps; 7 tech. reps; R^2 : 0.566



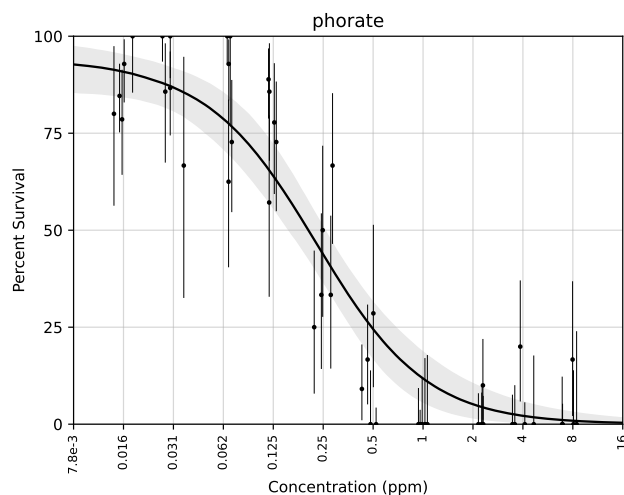
Naphazoline hydrochloride LC_{50} : 1.02e3 ppm [74.1, 3.07e17]
2 biol. reps; 2 tech. reps; R^2 : 0.0569



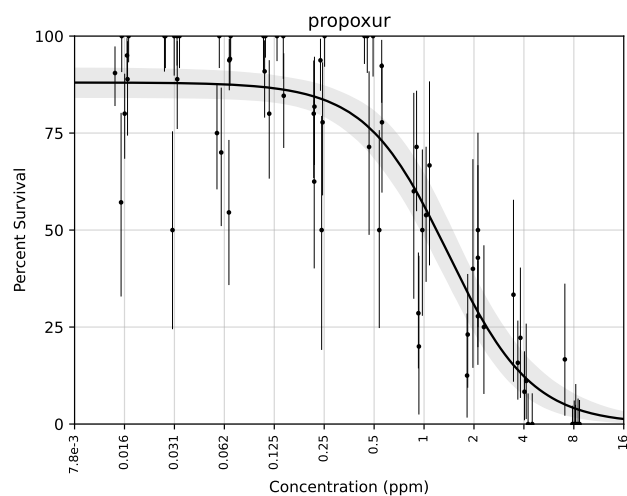
S-nicotine LC_{50} : 129 ppm [52, 4.05e3]
4 biol. reps; 5 tech. reps; R^2 : 0.272



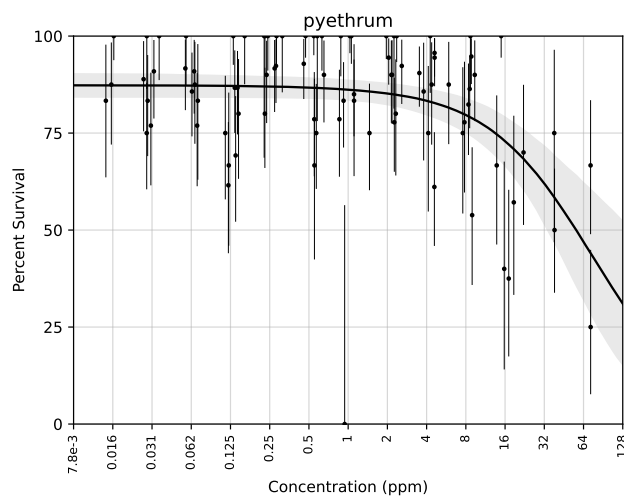
Permethrin LC_{50} : 11.9 ppm [7.42, 36.3]
5 biol. reps; 6 tech. reps; R^2 : 0.225



Phorate LC_{50} : 0.225 ppm [0.165, 0.295]
4 biol. reps; 5 tech. reps; R^2 : 0.897



Propoxur LC_{50} : 1.39 ppm [1.12, 1.74]
6 biol. reps; 7 tech. reps; R^2 : 0.824



Pyethrum LC_{50} : 74.1 ppm [38.5, 189]
8 biol. reps; 9 tech. reps; R^2 : 0.226

Data analysis was performed using the statistics module for the Merlin Data Analysis program. Live/dead counts from the bioassay were used to generate new survival probabilities using a Beta prior. The user-specified prior is Heldane's prior, the improper prior $\text{Beta}(0, 0)$, (set by `BETA_PRIOR`) and 3840 bootstrap iterations were used (set by `BOOTSTRAP_ITERS`). When either the live count or dead count was equal to 0, the prior the distribution $\text{Beta}(0.25, 0.25)$ (set by `BETA_PRIOR_0`) was used to avoid the sunrise problem. Correlation between wells in a replicate was modelled by generating multivariate normal random variables with correlation $\rho = 0.1$ (set by `RHO`), which were then converted to quantiles, and then back-converted to probabilities in the appropriate beta distribution.

Each iteration of bootstrapped dose-response data was fit to the curve

$$\theta_i = \frac{b_2}{(1 + \exp(b_0 + b_1 x_i))}$$

by maximizing the log-likelihood function, *i.e.*, solving

$$\underset{\mathbf{b}=(b_0, b_1, b_2)}{\text{argmax}} \left(f(\mathbf{b}) + \sum_{i=1}^n (1 - \theta_i) \ln(1 + \xi_i - b_2) + \sum_{i=1}^n \theta_i \ln b_2 - \sum_{i=1}^n \ln(1 + \xi_i) \right)$$

where

$$f(\mathbf{b}) = \frac{-(b_0^2 + b_1^2)}{2\sigma^2} + (\alpha - 1) \ln b_2 + (\beta - 1) \ln(1 - b_2)$$

and

$$\xi_i = e^{b_0 + b_1 x_i}.$$

Priors on parameters were $(b_0, b_1) \sim \mathcal{N}(\mathbf{0}, \sigma I_2)$ and $b_2 \sim \text{Beta}(\alpha, \beta)$, where $\sigma = 1000.0$, $\alpha = 1.5$, and $\beta = 1.001$, as defined by `LL_SIGMA`, `LL_BETA1`, and `LL_BETA2` in the `analysis_config.txt` file, respectively. Optimization of the log-likelihood function was performed using the optimized vector Broyden-Fletcher-Goldfarb-Shanno (BFGS) algorithm using a C interface to the GNU Scientific Library.

Credible intervals for the data points are shown at the 80% level when fewer than 10 replicates are used. The best-fit line is calculated as the median value of all fitted curves at a given concentration. The error region for the curve represents a 95% confidence region, as determined by quantiles of predicted survivals at each concentration.