

1 **Title: A derivation error exists in the current implementation of the Johnson et al. (1942) modified**
2 **Arrhenius function that affects leaf carbon balance models**

3 **Running Title: Derivation error in modified Arrhenius model**

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20

21 **Abstract**

22 Understanding biological temperature responses is crucial to predicting global carbon fluxes. The
23 current approach to modeling photosynthetic temperature responses in large scale modeling efforts
24 uses a modified Arrhenius equation. We rederived the modified Arrhenius equation from the source
25 publication and uncovered a missing term that was dropped between 1942 and 2002. We compare
26 fitted temperature response parameters between the new and old derivation of the modified
27 Arrhenius equation. We find that most parameters are minimally affected, though small errors still
28 exist. We then scaled the impact of these small errors to whole plant carbon balance and found that
29 the impact of the rederivation of the Arrhenius on modelled daily carbon gain causes a meaningful
30 deviation of ~1.8%. This suggests that the error in the derivation of the modified Arrhenius equation
31 has impacted predictions of carbon fluxes at larger scales. We argue that it is time to move beyond
32 the modified Arrhenius paradigm since the current implementation is categorically incorrect and to
33 use more thermodynamically-grounded temperature response equations going forward.

34 *Keywords: Arrhenius, temperature, photosynthesis, gas exchange, modeling, carbon balance*

Introduction

Globally, photosynthesis and autotrophic respiration are the largest biological carbon fluxes, with photosynthesis removing ~120 Gt C year⁻¹ and autotrophic respiration releasing ~60 Gt C year⁻¹ (Amthor, 2000; Ciais et al., 2013). Given the temperature sensitivity of these large carbon fluxes, understanding how photosynthesis and respiration respond on acute, acclimatory, and adaptive timescales is crucial for predicting vegetative and carbon cycle responses to future global climates (Rogers et al., 2017; Stinziano et al. 2018). Biological temperature responses including photosynthesis and respiration are typically assumed to be exponential or peaked (Way and Yamori, 2014; Smith & Dukes, 2017; Kumarathunge et al., 2019). Exponential responses are usually modelled based on an Arrhenius-type curve (Arrhenius, 1915):

$$f(T) = Ae^{\frac{-E_a}{RT}} \quad \text{Equation 1}$$

$$f(T) = k_{25}e^{\left[\frac{E_a(T-298.15)}{RT_{298.15}}\right]} \quad \text{Equation 2}$$

where $f(T)$ is the rate of the process at temperature, A is a pre-exponential factor, T in K, k_{25} is the rate of the process at 298.15 K, E_a is the activation energy in J mol⁻¹, R is the universal gas constant of 8.314 J mol⁻¹ K⁻¹, and 298.15 is the reference temperature in K. As for peaked responses, while a few options are available (Kruse et al., 2008; Hobbs et al., 2013; Heskell et al., 2016), the most commonly implemented version is the modified Arrhenius model of Johnson et al. (1942) as presented in Medlyn et al. (2002):

$$f(T) = k_{25}e^{\left[\frac{E_a(T-298.15)}{RT_{298.15}}\right]} \left[\frac{1+e^{\frac{298.15\Delta S-H_d}{298.15R}}}{1+e^{\frac{T\Delta S-H_d}{TR}}} \right] \quad \text{Equation 3}$$

where H_d is the deactivation energy in J mol⁻¹, and ΔS is the entropy of the process in J mol⁻¹.

Equation 3 is used for modeling the temperature responses of photosynthetic capacity (based on Farquhar et al., 1980): maximum carboxylation capacity of rubisco, V_{cmax} , maximum electron transport capacity, J_{max} , and related kinetics as well as Sharkey (1985) and Harley & Sharkey (1991) in their

calculation of triose phosphate utilization capacity (TPU). These parameters are then used in ecophysiological studies to understand thermal acclimation of photosynthesis (see Kattge & Knorr, 2007; Smith & Dukes, 2017; and Kumarathunge et al., 2019 for examples). Furthermore, this equation is also used in terrestrial biosphere models to predict the future state of the Earth system (e.g. Rogers et al., 2017).

Due to its ubiquity, we revisited the original Johnson et al. (1942) modified Arrhenius function to rederive Equation 3. In the process of this rederivation, we uncovered a term that was dropped sometime between Johnson et al. (1942) and Medlyn et al. (2002) which causes a systematic error in the application of Equation 3 in calculations covering individual species (e.g. Medlyn et al., 2002) to global scale processes (e.g. Rogers et al., 2017). We then refit a freely available dataset (Kumarathunge et al., 2019), with both versions of the modified Arrhenius model, and fed the temperature response fits through a carbon balance model to estimate the impact of the derivation error on modelled plant carbon balance. We predicted that the derivation error would cause substantial variation in fitted temperature response parameters, and that these differences would propagate through to modelled daily carbon balance.

Materials & methods

Rederation of the modified Arrhenius equation

Johnson et al. (1942, equation 24) describe the temperature response of the light intensity of a luciferase reaction as:

$$I = \frac{c''T e^{\frac{-\Delta H^\ddagger}{RT}}}{1 + e^{\frac{\Delta S}{R}} e^{\frac{-\Delta H}{RT}}} \quad \text{Equation 4}$$

where I is the intensity of the luciferase reaction, c'' is not explicitly defined in Johnson et al. (1942), but presumably represents a second derivative of the rate, T is the temperature in K, R is the universal gas constant of 8.314 J mol⁻¹ K⁻¹, ΔH[‡] is the activation energy in J mol⁻¹, ΔH is the deactivation energy in J mol⁻¹, and ΔS is the entropy in J mol⁻¹. We can relativize the equation to a reference temperature:

$$\frac{I}{I_{25}} = \frac{\frac{c''Te^{-\frac{\Delta H^\ddagger}{RT}}}{1+e^{\frac{\Delta S}{R}}e^{-\frac{\Delta H}{RT}}}}{\frac{c''_{298.15}e^{-\frac{\Delta H^\ddagger}{R298.15}}}{1+e^{\frac{\Delta S}{R}}e^{-\frac{\Delta H}{R298.15}}}} \quad \text{Equation 5}$$

94

$$\frac{I}{I_{25}} = \frac{c''Te^{-\frac{\Delta H^\ddagger}{RT}}}{c''_{298.15}e^{-\frac{\Delta H^\ddagger}{R298.15}}} \frac{1+e^{\frac{\Delta S}{R}}e^{-\frac{\Delta H}{R298.15}}}{1+e^{\frac{\Delta S}{R}}e^{-\frac{\Delta H}{RT}}} \quad \text{Equation 6}$$

96

$$\frac{I}{I_{25}} = \frac{T}{298.15} e^{\frac{-\Delta H^\ddagger}{RT} - \frac{-\Delta H^\ddagger}{R298.15}} \frac{1+e^{\frac{\Delta S}{R} + \frac{-\Delta H}{R298.15}}}{1+e^{\frac{\Delta S}{R} + \frac{-\Delta H}{RT}}} \quad \text{Equation 7}$$

98

$$I = I_{25} \frac{T}{298.15} e^{\left[\frac{-\Delta H^\ddagger_{298.15} - -\Delta H^\ddagger T}{RT298.15} \right]} \left[\frac{1+e^{\frac{298.15\Delta S + -\Delta H}{298.15R}}}{1+e^{\frac{T\Delta S + -\Delta H}{TR}}} \right] \quad \text{Equation 8}$$

100

$$I = I_{25} \frac{T}{298.15} e^{\left[\frac{\Delta H^\ddagger(T-298.15)}{RT298.15} \right]} \left[\frac{1+e^{\frac{298.15\Delta S - \Delta H}{298.15R}}}{1+e^{\frac{T\Delta S - \Delta H}{TR}}} \right] \quad \text{Equation 9}$$

102

103 Harmonizing the notation scheme to that typically used in plant ecophysiology:

104

$$f(T) = k_{25} \frac{T}{298.15} e^{\left[\frac{E_a(T-298.15)}{RT298.15} \right]} \left[\frac{1+e^{\frac{298.15\Delta S - H_d}{298.15R}}}{1+e^{\frac{T\Delta S - H_d}{TR}}} \right] \quad \text{Equation 10}$$

106

107 Note the difference between Equations 3 and 10: the term $T / 298.15$ is missing from Equation 3. This
108 introduces multiple systematic errors:

- 109 1) fitted parameters E_a , H_d , and ΔS ;
- 110 2) scaling the rate variable using the wrong equation;
- 111 3) acclimation of E_a , H_d , and/or ΔS as errors due to 1) at each acclimation temperature.

112 Here we focus on the impact of 1 and 2 on modeling whole-plant carbon balance.

113

114 Data analysis

115 Using data from Kumarathunge et al. (2019), available from Kumarathunge et al. (2018), we fit

116 Equations 3 and 10 to the data, both setting H_d to 200 kJ mol^{-1} and allowing H_d to be fit. We used the

117 R package {minpack.lm} (Elzhov et al., 2016) with starting parameters of $E_a = 40 \text{ kJ mol}^{-1}$, $\Delta S = 0.650 \text{ kJ}$
 118 mol^{-1} , $k_{25} = \text{mean}(\text{parameter})$, and H_d (when fitted) varying from 1 to 500 kJ mol^{-1} , followed by the
 119 {BIC} function to select the best model based on Bayesian Information Criteria. We obtained 92
 120 successful curve fits for the fixed H_d case, and 84 successful curve fits for the fitted H_d case. This
 121 allows us to explore the impact of the missing term on the output data under a typical fitting scenario
 122 (H_d is fixed) and under the full fitting scenario.

123

124 **Modeling**

125 We modelled the impact of the equations 3 and 10 on daily net plant carbon balance under
 126 conditions of $H_d = 200 \text{ kJ mol}^{-1}$ and fitted H_d . Data for leaf area, root and shoot masses, as well as leaf
 127 dark respiration at 25 °C were taken for white spruce (*Picea glauca*) from Stinziano & Way (2017),
 128 while stomatal conductance model parameters were calculated with the gas exchange data reported
 129 in Stinziano & Way (2017). Mean data were taken from the control treatment at weeks 0 and 12 to
 130 provide contrasting biomass allocation patterns such that week 0 is a low respiration scenario and
 131 week 12 is a high respiration scenario. These different respiration scenarios were used to reduce bias
 132 in any conclusions regarding the impact of Equations 3 and 10 on carbon balance, as the ratio of
 133 photosynthesis to respiration may alter the sensitivity of carbon balance to the Arrhenius equation
 134 used. Root respiration for white spruce was taken from Weger and Guy (1991) and we assumed that
 135 stem respiration was equal to root respiration (Table 1).

136

137 **Table 1. Parameters used in modeling daily carbon gain.**

Parameter	Group	Value	Reference
Respiration	R_{dark}	$2.78 \mu\text{mol m}^{-2} \text{s}^{-1}$	Stinziano & Way, 2017
	R_{day}	$0.7 * R_{\text{dark}}$	Ayub et al., 2011
	R_{root}	$0.0095 \mu\text{mol g}^{-1} \text{s}^{-1}$	Weger & Guy, 1991
	R_{stem}	$0.0095 \mu\text{mol g}^{-1} \text{s}^{-1}$	Assumed
	Q_{10}	2.015	Atkin & Tjoelker, 2003
Γ^*	25 °C	$42.75 \mu\text{mol mol}^{-1}$	Bernacchi et al., 2001
	E_a	$37.83 \text{ kJ mol}^{-1}$	Bernacchi et al., 2001
K_m	25 °C	$718.4 \mu\text{mol mol}^{-1}$	Bernacchi et al., 2001
	E_a	$65.51 \text{ kJ mol}^{-1}$	Bernacchi et al., 2001
α		0.8	Norman & Campbell, 1998
ϕ		0.08	Norman & Campbell, 1998
Leaf Area	Low R	0.015 m^2	Stinziano & Way, 2017
	High R	0.025 m^2	Stinziano & Way, 2017

Stem Mass	Low R	0.496 g	Stinziano & Way, 2017
	High R	2.523 g	Stinziano & Way, 2017
Root Mass	Low R	0.498 g	Stinziano & Way, 2017
	High R	5.072 g	Stinziano & Way, 2017

Q₁₀: thermal sensitivity coefficient; Γ^* : photorespiratory CO₂ compensation point; K_m: Michaelis-Menten constant for rubisco; α : absorbance of photosynthetically activation radiation; ϕ : maximum quantum efficiency of photosynthetic electron transport; E_a: activation energy; R_{dark}: leaf respiration in the dark; R_{day}: leaf respiration in the light; R_{root}: root respiration; R_{stem}: stem respiration; Low R: low respiration scenario; High R: high respiration scenario.

For the full model structure and equations, please see the accompanying R package {arrhenius.comparison} ("arrhenius.comparison_1.0.0.tar.gz"; Stinziano & Murphy, 2020) (see Table 2 for equations). Briefly, we linked the Medlyn et al. (2011) stomatal conductance model with the Farquhar et al. (1980) C₃ photosynthesis model, assuming infinite mesophyll conductance to CO₂ as these assumptions were used in fitting the data from Kumarathunge et al. (2018). Photosynthetic capacity, both maximum rubisco carboxylation capacity, V_{cmax}, and maximum electron transport rate, J_{max} were scaled to temperature using either Equation 3 or 10, while respiration was scaled according to (Atkin & Tjoelker, 2003). Photosynthesis and respiration were summed across each modelled day to calculate daily plant carbon assimilation.

Table 2. Equations used in modeling daily carbon uptake.

Equation	Reference
$f(T) = k_{25} Q_{10}^{\frac{T-298.15}{10}}$	Atkin & Tjoelker, 2003
$f(T) = k_{25} e^{\left[\frac{E_a(T-298.15)}{RT298.15} \right]}$	Arrhenius, 1915
$f(T) = k_{25} e^{\left[\frac{E_a(T-298.15)}{RT298.15} \right]} \left[\frac{1 + e^{\frac{298.15\Delta S - H_d}{298.15R}}}{1 + e^{\frac{T\Delta S - H_d}{TR}}} \right]$	Johnson et al., 1942; Medlyn et al., 2002
$f(T) = k_{25} \frac{T}{298.15} e^{\left[\frac{E_a(T-298.15)}{RT298.15} \right]} \left[\frac{1 + e^{\frac{298.15\Delta S - H_d}{298.15R}}}{1 + e^{\frac{T\Delta S - H_d}{TR}}} \right]$	Johnson et al., 1942; this study
$W_c = V_{cmax} \frac{C_i - \Gamma^*}{C_i + K_m}$	Farquhar et al., 1980

$$W_j = \min \left(J_{max}, \alpha \phi Q_{in} \frac{C_i - \Gamma^*}{C_i + 2\Gamma^*} \right)$$

Farquhar et al., 1980; Way et al., 2011

$$A_{gross} = \min(W_c, W_j)$$

$$A_{net} = A_{gross} - R_{day}$$

$$g_s = g_0 + 1.6 \left(1 + \frac{g_1}{\sqrt{VPD}} \right) \left(\frac{A_{net}}{C_a} \right)$$

Medlyn et al., 2011

$$C_i = C_a - 1.6 \frac{A_{net}}{g_s}$$

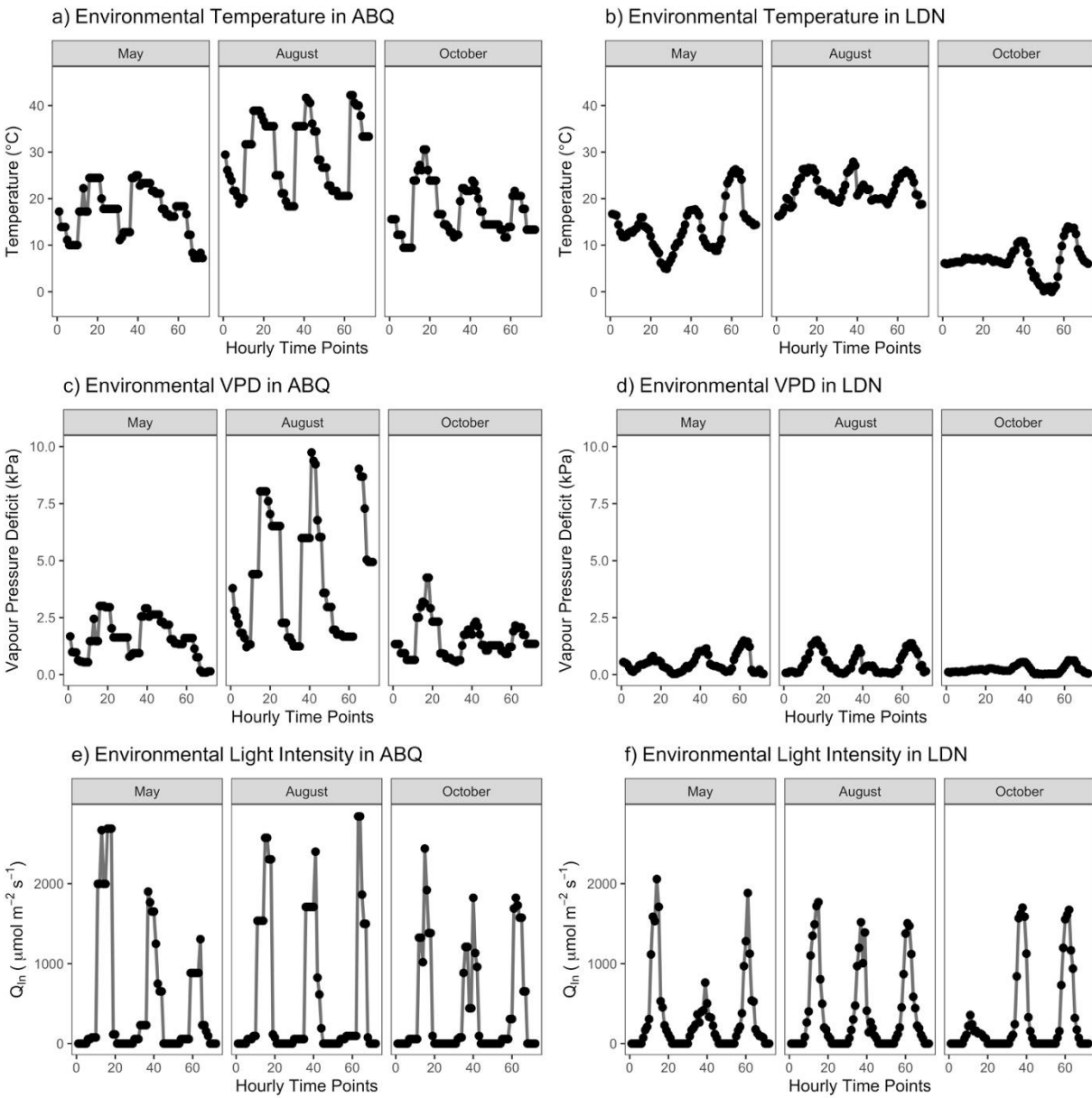
$$A_{plant} = A_{net} \times LA - R_{dark} \times LA - R_{stem} \times SM - R_{root} \times RM$$

$$C_{balance} = \sum A_{plant} \times 3600 \times \frac{12.01}{1,000,000}$$

f(T): rate of a process at a given temperature; T: temperature in K; k₂₅: rate of a process at 25 °C; Q₁₀: thermal sensitivity coefficient; E_a: activation energy in kJ mol⁻¹; ΔS: entropy parameter in kJ mol⁻¹; H_d: deactivation energy in kJ mol⁻¹; R: universal gas constant in 0.008314 kJ mol⁻¹ K⁻¹; W_c: rate of CO₂-limited carboxylation in μmol m⁻² s⁻¹; W_j: rate of RuBP regeneration-limited carboxylation in μmol m⁻² s⁻¹; V_{cmax}: maximum rate of rubisco carboxylation capacity in μmol m⁻² s⁻¹; J_{max}: maximum rate of electron transport in μmol m⁻² s⁻¹; C_i: intercellular CO₂ concentration in μmol mol⁻¹; Γ*: photorespiratory CO₂ compensation point in μmol mol⁻¹; K_m: Michaelis-Menten constant for rubisco in μmol mol⁻¹; α: absorbance of photosynthetically active radiation; φ: maximum quantum efficiency of electron transport; Q_{in}: incident photosynthetically active radiation; A_{gross}: gross CO₂ assimilation in μmol m⁻² s⁻¹; A_{net}: net CO₂ assimilation in μmol m⁻² s⁻¹; R_{day}: leaf day respiration in μmol m⁻² s⁻¹; g_s: stomatal conductance to water in mol m⁻² s⁻¹; g₀: intercept for the Medlyn et al. 2011 model; g₁: slope for the Medlyn et al. 2011 model; VPD: vapor pressure deficit in kPa; C_a: CO₂ concentration at the leaf surface in μmol mol⁻¹; A_{plant}: whole plant net CO₂ assimilation; LA: leaf area in m²; R_{dark}: leaf dark respiration in μmol m⁻² s⁻¹; R_{stem}: stem respiration in μmol m⁻² s⁻¹; R_{root}: root respiration in μmol m⁻² s⁻¹; SM: stem mass in g; RM: root mass in g; C_{balance}: whole plant daily carbon balance in g plant⁻¹ day⁻¹.

Modeling was performed on 18 total days of environmental data, with three days of data from three months (17th – 19th of May, August, and October, 2019) obtained from external irradiance sensors at the Biotron Experimental Climate Change Research Centre at the University of Western Ontario and

175 the remaining environmental data from Environment Canada historical climate data for South London
 176 (43.01°N, 81.27°W, altitude: 251 m) and the rooftop greenhouse at the University of New Mexico
 177 (35.08°N, 106.62°W, altitude: 1587 m) to capture different levels of environmental variability (Fig. 1).



178
 179 **Figure 1. Environmental data used to drive the model in Table 1 covering 3 days (17th, 18th, and 19th)**
 180 **of 3 months. (a,c,e) Albuquerque, NM, USA; (b,d,e) London, ON, Canada.**

181
 182 Overall, the modeling approach allows us to assess the relative differences between Equations 3 and
 183 10 under a low- and high- respiration scenario, fixing versus fitting H_d , and across different ranges of
 184 seasonal variability.

185

186 *Statistical analysis*

187 Data were analyzed using the {lm} function in R v.3.6.1 (R Core Team, 2019), regressing the data
188 obtained from Equation 10 against the data obtained from Equation 3. All code and data will be made
189 freely available on GitHub upon publication in the {arrhenius.comparison} R package (Stinziano &
190 Murphy, 2020).

191

192 **Results**

193 *Fixed H_d of 200 kJ mol⁻¹*

194 Incorporating the missing term into Equation 3 to produce Equation 10 caused small differences in
195 the fitting of E_a for V_{cmax} , with a slope of 0.997 ± 0.001 and intercept of $-2.162 \pm 1.19 \cdot 10^{-1}$ kJ mol⁻¹
196 ($F_{1,90} = 4.56 \cdot 10^5$, $R^2 = 0.9998$, $P < 2.2 \cdot 10^{-16}$) (Fig. 2b). For ΔS , the slope was $0.991 \pm 3 \cdot 10^{-3}$ and an
197 intercept of $0.005 \pm 2 \cdot 10^{-3}$ kJ mol⁻¹ (intercept approximately 1% of fitted ΔS) ($F_{1,90} = 1.23 \cdot 10^5$, $R^2 =$
198 0.9993 , $P < 2.2 \cdot 10^{-16}$) (Fig. 2f). Meanwhile V_{cmax25} was identical between both approaches with a
199 slope of $1.000 \pm 1.1 \cdot 10^{-5}$ (intercept not significant; $F_{1,91} = 8.2 \cdot 10^9$, $R^2 = 1.000$, $P < 2.2 \cdot 10^{-16}$) (Fig. 3b).
200 For J_{max} , there were small differences in the fitting of E_a for J_{max} , with a slope of $0.998 \pm 2 \cdot 10^{-3}$ and
201 intercept of $-2.230 \pm 1.08 \cdot 10^{-1}$ kJ mol⁻¹ ($F_{1,91} = 3.3 \cdot 10^5$, $R^2 = 0.9997$, $P < 2.2 \cdot 10^{-16}$) (Fig. 2d). For ΔS ,
202 the slope was $0.962 \pm 3 \cdot 10^{-3}$ and an intercept of $0.025 \pm 2 \cdot 10^{-3}$ kJ mol⁻¹ (intercept approximately 5%
203 of fitted ΔS) ($F_{1,90} = 1.1 \cdot 10^5$, $R^2 = 0.9992$, $P < 2.2 \cdot 10^{-16}$) (Fig. 2h). Meanwhile J_{max25} was identical
204 between both approaches with a slope of $1.000 \pm 1.2 \cdot 10^{-5}$ (intercept not significant; $F_{1,92} = 6.7 \cdot 10^9$,
205 $R^2 = 1.000$, $P < 2.2 \cdot 10^{-16}$) (Fig. 3d).

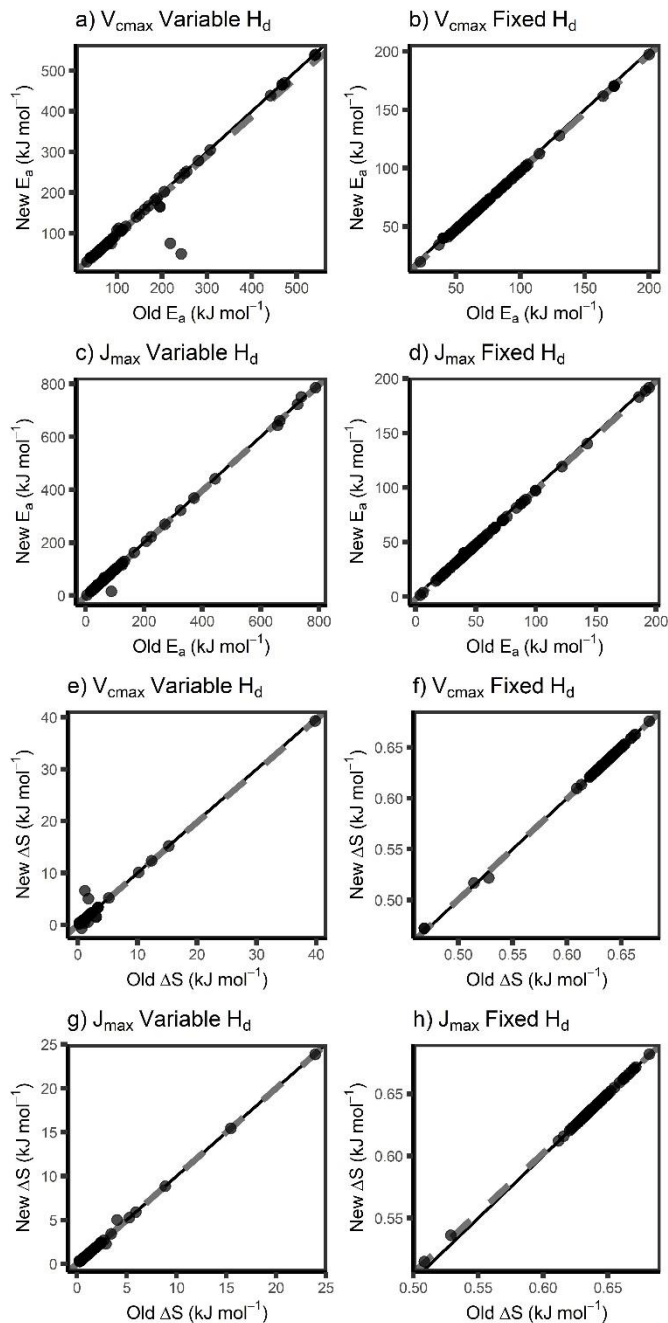


Figure 2 – The modified Arrhenius equation with the missing term (Equation 10) gives similar temperature response parameters as Equation 3 for E_a (a-d) and ΔS (e-h) for both V_{cmax} (a, b, e, f) and J_{max} (c, d, g, h) under scenarios where H_d is allowed to vary (a, c, e, g) and is fixed to 200 kJ mol⁻¹ (b, d, f, h). “New” indicates Equation 10, “Old” indicates Equation 3. E_a : activation energy, ΔS : entropy parameter, H_d : deactivation energy, V_{cmax} : maximum capacity of rubisco carboxylation, J_{max} : maximum rate of electron transport. Black line indicates 1:1 line and grey dashed line indicates respective modeled slopes and intercepts.

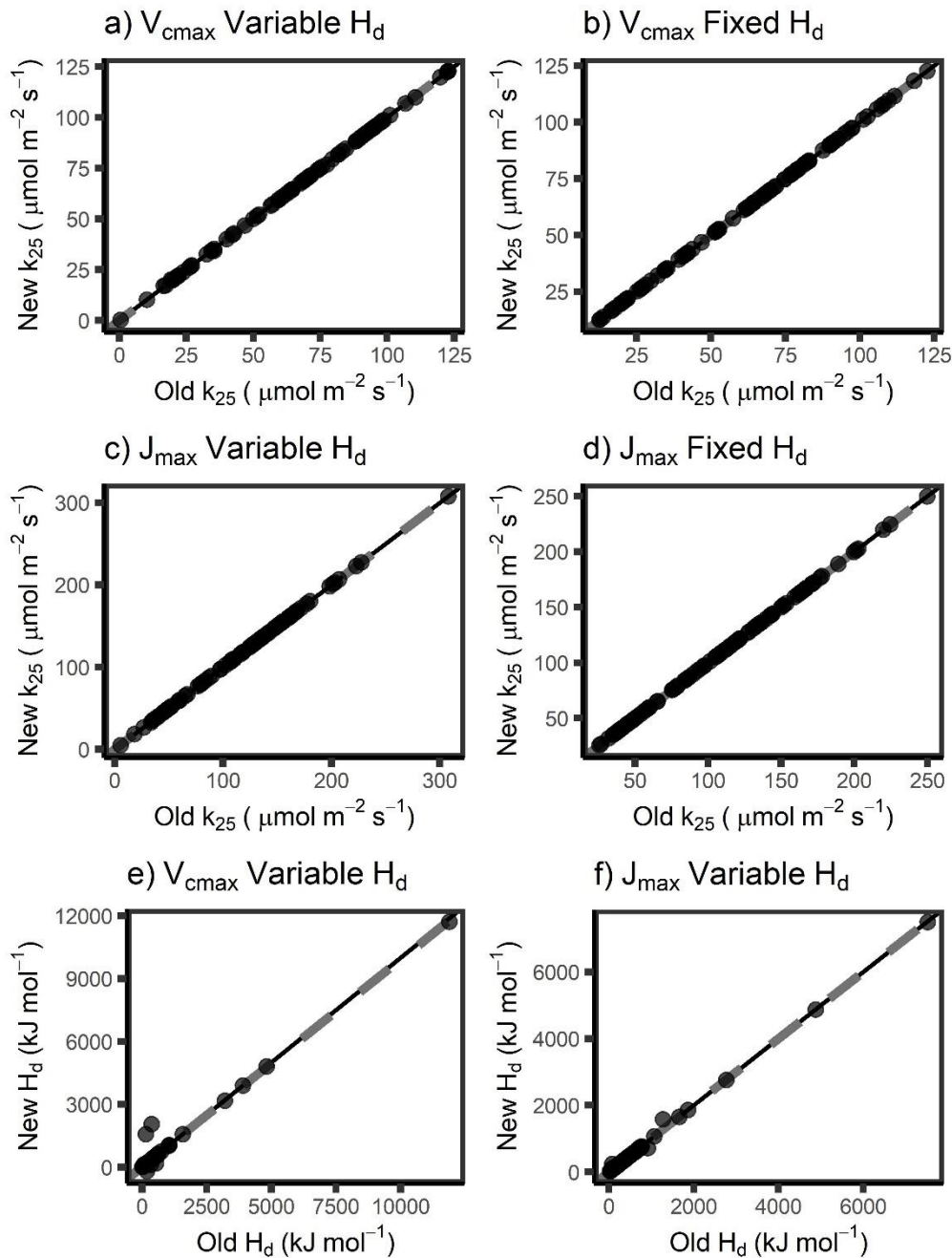


Figure 3 – The modified Arrhenius equation with the missing term (Equation 10) gives similar temperature response parameters as Equation 3 for k_{25} (a-d) and H_d (e, f) for both V_{cmax} (a, b, e) and J_{max} (c, d, f) under scenarios where H_d is allowed to vary (a, c, e, f) and is fixed to 200 kJ mol^{-1} (b, d). “New” indicates Equation 10, “Old” indicates Equation 3. k_{25} : rate of the process at 25 °C, H_d : deactivation energy, V_{cmax} : maximum capacity of rubisco carboxylation, J_{max} : maximum rate of electron transport. Black line indicates 1:1 line and grey dashed line indicates respective modeled slopes and intercepts.

222 *Variable H_d*

223 Incorporating the missing term into Equation 3 to produce Equation 10 caused small differences in
224 the fitting of E_a for V_{cmax} , with a slope of $0.966 \pm 2.6 \cdot 10^{-2}$ (intercept not significant; $F_{1,74} = 3005$, $R^2 =$
225 0.976 , $P < 2.2 \cdot 10^{-16}$) (Fig. 2a). For ΔS , the slope was $0.989 \pm 1.7 \cdot 10^{-2}$ (intercept not significant; $F_{1,74} =$
226 3339 , $R^2 = 0.9783$, $P < 2.2 \cdot 10^{-16}$) (Fig. 2e), while H_d had a slope of $0.992 \pm 1.9 \cdot 10^{-2}$ (intercept not
227 significant; $F_{1,74} = 2812$, $R^2 = 0.9744$; $P < 2.2 \cdot 10^{-16}$) (Fig. 3e). Meanwhile V_{cmax25} was nearly identical
228 between both approaches with a slope of $1.000 \pm 4 \cdot 10^{-3}$ (intercept not significant; $F_{1,74} = 7.98 \cdot 10^6$,
229 $R^2 = 1.000$, $P < 2.2 \cdot 10^{-16}$) (Fig. 3a). For J_{max} , there were small differences in the fitting of E_a for J_{max} ,
230 with a slope of $1.000 \pm 5 \cdot 10^{-3}$ and an intercept of $-3.119 \pm 1.089 \text{ kJ mol}^{-1}$ ($F_{1,80} = 3.6 \cdot 10^4$, $R^2 = 0.9978$,
231 $P < 2.2 \cdot 10^{-16}$) (Fig. 2c). ΔS was identical, the slope was $1.000 \pm 4 \cdot 10^{-3}$ (intercept not significant; $F_{1,81} =$
232 $6.4 \cdot 10^4$, $R^2 = 0.9987$, $P < 2.2 \cdot 10^{-16}$) (Fig. 2g), while H_d was identical with a slope of $1.000 \pm 4 \cdot 10^{-3}$
233 (intercept not significant; $F_{1,81} = 5.2 \cdot 10^4$, $R^2 = 0.9985$; $P < 2.2 \cdot 10^{-16}$) (Fig. 3f). J_{max25} was identical
234 between both approaches with a slope of $1.000 \pm 1 \cdot 10^{-3}$ (intercept not significant; $F_{1,81} = 5.8 \cdot 10^7$, R^2
235 $= 1.000$, $P < 2.2 \cdot 10^{-16}$) (Fig. 3c).

236

237 *Impacts on modelled net carbon balance*

238 In general, the differences in thermal response parameters were amplified when integrated at the
239 whole-plant level. For daily photosynthesis (A) measured at a fixed H_d of 200 kJ mol^{-1} , the slope for
240 the low respiration model was $0.977 \pm 7.666 \cdot 10^{-3}$ and an intercept of $0.002 \pm 9.366 \cdot 10^{-5}$
241 (approximately 2% of modeled A) ($F_{1,898} = 1.625 \cdot 10^6$, $R^2 = 0.9994$; $P < 2.2 \cdot 10^{-16}$) (Fig. 4b). Similarly,
242 the high respiration model for A had a slope of $0.977 \pm 7.666 \cdot 10^{-3}$ and an intercept of $0.004 \pm 1.591 \cdot$
243 10^{-3} (approximately 2% of modeled A) ($F_{1,898} = 1.625 \cdot 10^6$, $R^2 = 0.9994$; $P < 2.2 \cdot 10^{-16}$) (Fig. 4d). For
244 daily A measured at a variable H_d , the slope for the low respiration model was $0.983 \pm 9.909 \cdot 10^{-3}$ and
245 the intercept was $0.002 \pm 1.128 \cdot 10^{-3}$ (approximately 2% of modeled A) ($F_{1,556} = 9.834 \cdot 10^5$, $R^2 =$
246 0.9994 ; $P < 2.2 \cdot 10^{-16}$) (Fig. 4a). For the high respiration model of daily A , the slope was $0.983 \pm 9.909 \cdot$
247 10^{-3} and the intercept was $0.003 \pm 9.909 \cdot 10^{-4}$ (approximately 2% of modeled A) ($F_{1,556} = 9.834 \cdot 10^5$,
248 $R^2 = 0.9994$; $P < 2.2 \cdot 10^{-16}$) (Fig. 4c).

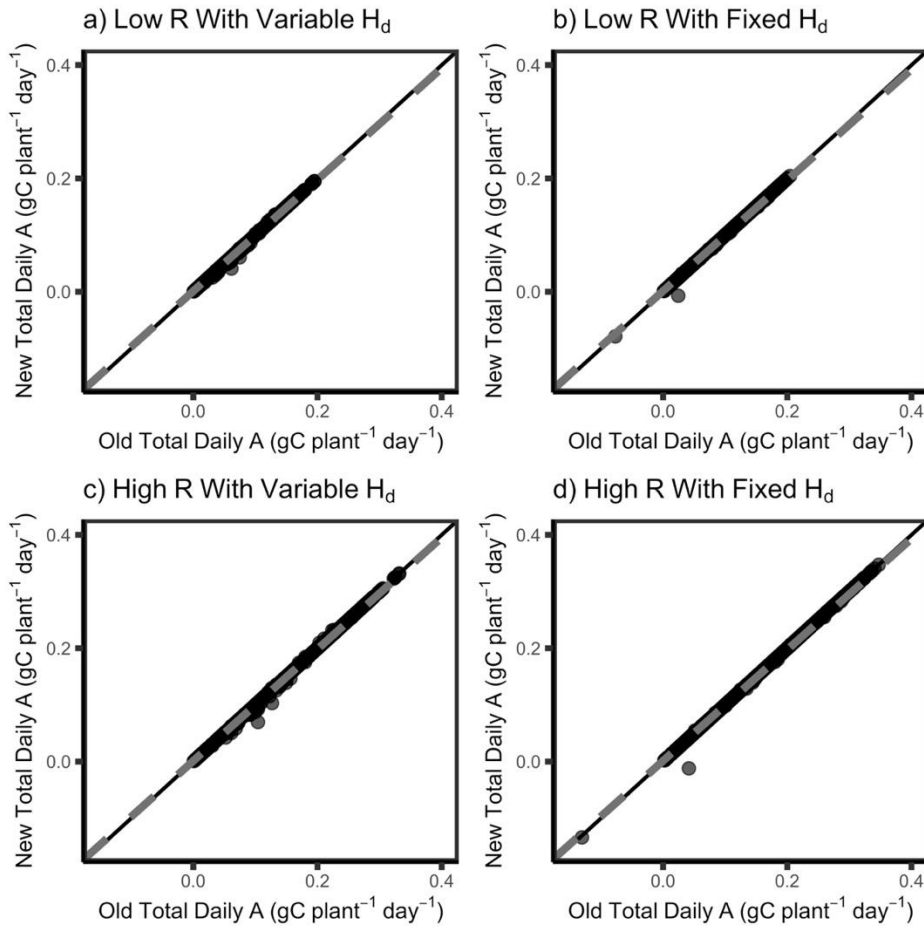
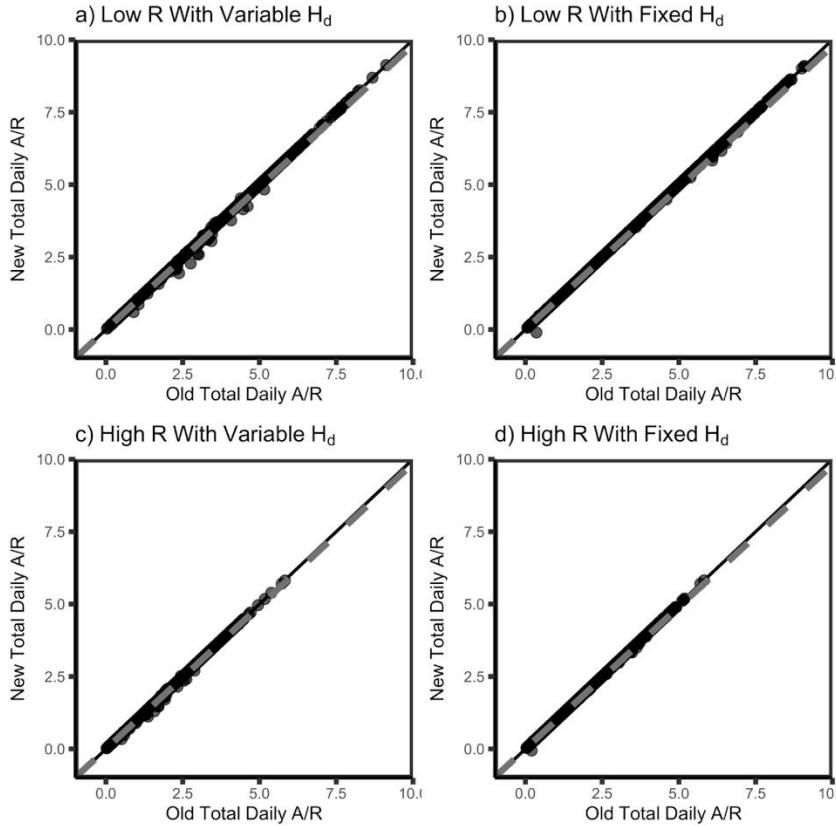


Figure 4 – The modified Arrhenius equation with the missing term (Equation 10) gives slight differences in modeled total daily photosynthesis compared to Equation 3 for low R (a,b) and high R (c,d) under scenarios where H_d is allowed to vary (a, c) and is fixed to 200 kJ mol^{-1} (b,d). “New” indicates Equation 10, “Old” indicates Equation 3. A: photosynthesis, Low R: low respiration, High R: high respiration. Black line indicates 1:1 line and grey dashed line indicates respective modeled slopes and intercepts.

The ratio of the total daily photosynthesis : respiration (A/R) was also considered when comparing models. Using a fixed H_d of 200 kJ mol^{-1} , the low respiration model of A/R had a slope of $0.993 \pm 5.634 \cdot 10^{-3}$ and the intercept was 0.025 ± 0.002 (approximately 1% of modeled A/R) ($F_{1,898} = 3.109 \cdot 10^6$, $R^2 = 0.9997$; $P < 2.2 \cdot 10^{-16}$) (Fig. 5b). The high respiration model of A/R was similar; the slope was 0.993 ± 5.628 and the intercept was 0.014 ± 0.001 (approximately 1% of modeled A/R) ($F_{1,898} = 3.115 \cdot 10^6$, $R^2 = 0.9997$; $P < 2.2 \cdot 10^{-16}$) (Fig. 5d). Using a variable H_d , the low respiration model of A/R had a slope of 0.994 ± 0.001 and the intercept was 0.030 ± 0.001 (approximately 1% of modeled A/R) ($F_{1,556} = 6.857 \cdot$

264 10^5 , $R^2 = 0.9992$; $P < 2.2 \cdot 10^{-16}$) (Fig. 5a). The high respiration model had a similar slope of $0.994 \pm$
 265 0.001 and the intercept was 0.017 ± 0.003 (approximately 1% of modeled A/R) ($F_{1,556} = 6.812 \cdot 10^5$, R^2
 266 $= 0.9992$; $P < 2.2 \cdot 10^{-16}$) (Fig. 5c).

267



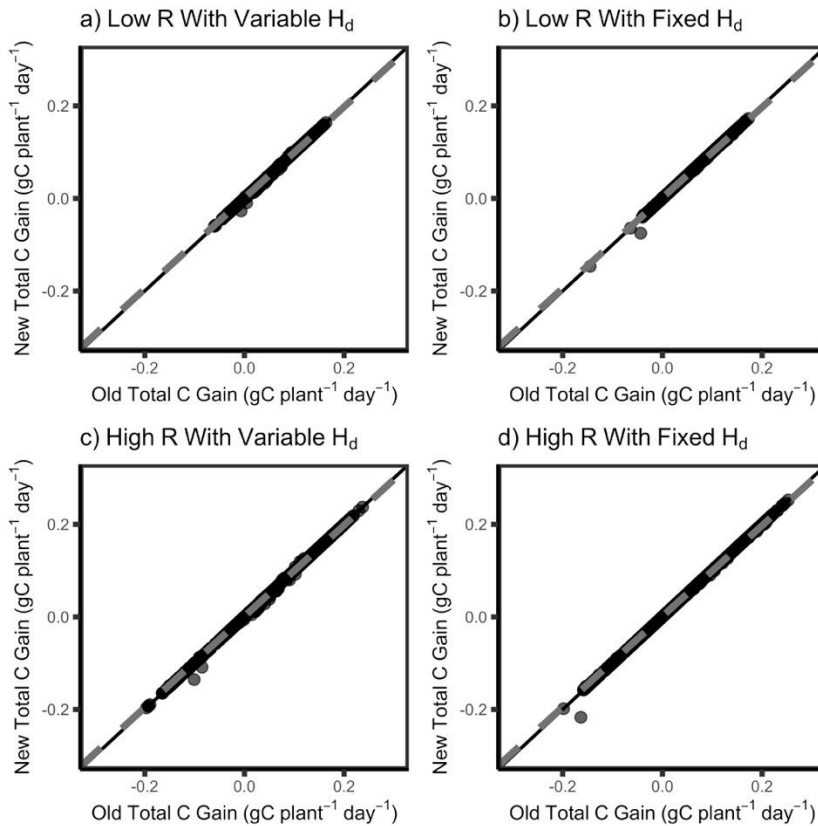
268

269 **Figure 5 – The modified Arrhenius equation with the missing term (Equation 10) gives slight**
 270 **differences in modeled total daily A/R compared to Equation 3 for low R (a,b) and high R (c,d) under**
 271 **scenarios where H_d is allowed to vary (a, c) and is fixed to 200 kJ mol^{-1} (b,d). “New” indicates**
 272 **Equation 10, “Old” indicates Equation 3. A: photosynthesis, R: respiration, Low R: low respiration,**
 273 **High R: high respiration. Black line indicates 1:1 line and grey dashed line indicates respective**
 274 **modeled slopes and intercepts.**

275

276 Lastly, total daily carbon (C) gain was also compared between models with different thermal response
 277 parameters. Using a fixed H_d of 200 kJ mol^{-1} , the low respiration model of total C gain had a slope of
 278 $0.978 \pm 7.445 \cdot 10^{-4}$ and an intercept of $0.001 \pm 7.609 \cdot 10^{-5}$ (approximately 2% of modeled C gain)
 279 ($F_{1,898} = 1.726 \cdot 10^6$, $R^2 = 0.9995$; $P < 2.2 \cdot 10^{-16}$) (Fig. 6b). Likewise, the high respiration of total C gain
 280 had a slope of $0.979 \pm 7.261 \cdot 10^{-4}$ and an intercept of $0.001 \pm 1.116 \cdot 10^{-4}$ (approximately 2% of

281 modeled C gain) ($F_{1,898} = 1.821 \cdot 10^6$, $R^2 = 0.9995$; $P < 2.2 \cdot 10^{-16}$) (Fig. 6d). This can be compared to a
 282 variable H_d ; starting with the low respiration model, total C gain had a slope of $0.985 \pm 9.555 \cdot 10^{-4}$
 283 and an intercept of $0.001 \pm 1.223 \cdot 10^{-3}$ (approximately 1.5% of modeled C gain) ($F_{1,556} = 1.06 \cdot 10^6$, R^2
 284 $= 0.9995$; $P < 2.2 \cdot 10^{-16}$) (Fig. 6a). The high respiration of total C gain also had a slope of $0.985 \pm 9.110 \cdot$
 285 10^{-4} and an intercept of $0.001 \pm 1.290 \cdot 10^{-4}$ (approximately 1.5% of modeled C gain) ($F_{1,556} = 1.17 \cdot$
 286 10^6 , $R^2 = 0.9995$; $P < 2.2 \cdot 10^{-16}$) (Fig. 6c).
 287



288
 289 **Figure 6 – The modified Arrhenius equation with the missing term (Equation 10) gives slight**
 290 **differences in modeled total carbon gain compared to Equation 3 for low R (a,b) and high R (c,d)**
 291 **under scenarios where H_d is allowed to vary (a, c) and is fixed to 200 kJ mol^{-1} (b,d). “New” indicates**
 292 **Equation 10, “Old” indicates Equation 3. C: carbon, Low R: low respiration, High R: high respiration.**
 293 **Black line indicates 1:1 line and grey dashed line indicates respective modeled slopes and**
 294 **intercepts.**

295

296 Discussion

297 We sought to determine whether the missing term in Equation 3 had a meaningful impact on fitted
298 temperature response parameters due to its prevalence in photosynthetic temperature response data
299 and vegetation modeling (Kattge & Knorr, 2007; Duursma & Medlyn, 2012; Rogers et al., 2017; Smith
300 & Dukes, 2017; Stinziano et al., 2018; Stinziano et al., 2019; Kumarathunge et al., 2019). Our present
301 analysis suggests that there is no large impact on the fitted temperature response parameters for
302 V_{cmax} and J_{max} , with differences ranging between 0 and 3.8% depending on the parameter. In general,
303 fitting Equation 10 instead of Equation 3 results in slightly reduced values for E_a and ΔS , with
304 essentially no impact for k_{25} and H_d when H_d is fit. When H_d is fixed, there are no meaningful
305 differences in the fits except for ΔS for J_{max} , where Equation 10 values are ~4% lower than Equation 3
306 values, and intercepts in the E_a regressions of $\sim -2 \text{ kJ mol}^{-1}$, which represents <5% differences between
307 the two equations in most instances. These findings are promising in that one of the parameters to
308 which modelled carbon gain is particularly sensitive, H_d (Stinziano et al., 2018), is minimally affected
309 by the missing term. However, since temperature responses are non-linear, small changes in the
310 shape of an accelerating curve can have a strong impact on the integral of the response (Jensen,
311 1906). This caused the differences in temperature response parameters to cause $\sim 1.5 - 2.2\%$
312 difference in daily C balance. The daily C balance was most affected by the missing term in Equation 3
313 when comparing models where H_d was fixed, leading to a 2.2% change. Overall, comparisons of low-
314 respiration to high-respiration scenarios resulted in similar percent changes in daily C balance;
315 however, the variation was consistently larger under high-respiration across all variables measured.
316 Similar trends were found when comparing differences in daily photosynthesis and A/R.

317

318 Based on the above analysis, the impact of the missing term in the modified Arrhenius equation may
319 appear negligible based on the parameter outputs. However given that carbon balance is the time
320 integral of net CO_2 assimilation, this may lead to substantial impacts over a long time period. It is thus
321 possible that the apparently small differences we observed could accumulate to relatively large
322 carbon flux errors across large spatial and temporal scales with fluctuating temperatures. A growing
323 body of literature on using Optimality Theory for estimating $V_{\text{cmax}25}$ and $J_{\text{max}25}$ (Ali et al., 2016; Walker
324 et al., 2017; Smith et al., 2019) still relies on Equation 3 for temperature scaling. Given the impacts we
325 observed, Optimality Theory may be even more successful if the temperature scaling were done

326 either with Equation 10, or, as we recommend, macromolecular rate theory which is better grounded
327 in thermodynamics (e.g. Hobbs et al., 2013; Liang et al., 2018).

328

329 Since Equation 3 is incorrect, there are three arguments to support moving beyond the modified
330 Arrhenius paradigm: 1) there are new approaches to modeling biological temperature responses that
331 are better-grounded in thermodynamics (e.g. Macromolecular Rate Theory; Hobbs et al., 2013; Liang
332 et al., 2018); 2) uncertainties in model outputs may accumulate due to the missing term across large
333 spatial and temporal scales, and 3) the modified Arrhenius model as implemented is categorically
334 incorrect due to a missing term. Therefore, we conclude that future modeling efforts should move
335 beyond this incorrect paradigm.

336

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340

341 **Additional Information**

342 A version of this manuscript was posted on bioRxiv (manuscript ID: BIORXIV/2020/921973).

343

344 **References**

- 345 Ali AA, Xu C, Rogers A, Fisher RA, Wullschleger SD, Massoud EC, Vrugt JA, Muss JD, et al. 2016. A
346 global scale mechanistic model of photosynthetic capacity (LUNA V1.0). Geoscientific Model
347 Development 9:587-606.
- 348 Amthor JS. 2000. Direct effect of elevated CO₂ on nocturnal in situ leaf respiration in nine temperate
349 deciduous tree species is small. Tree Physiology 20: 139–144.
- 350 Arrhenius S. 1915. Quantitative laws in biological chemistry. Bell: London.
- 351 Atkin OK, Tjoelker MG. 2003. Thermal acclimation and the dynamic response of plant respiration to
352 temperature. Trends in Plant Science 8:343-351.
- 353 Ayub G, Smith RA, Tissue DT, Atkin OK. 2011. Impacts of drought on leaf respiration in darkness and
354 light in *Eucalyptus saligna* exposed to industrial-age atmospheric CO₂ and growth temperature.
355 New Phytologist 190: 1003-1018.

356 Bernacchi CJ, Singsaas EL, Pimentel C, Portis Jr AR, Long SP. 2001. Improved temperature response
 357 functions for models of rubisco-limited photosynthesis. *Plant Cell and Environment* 24:253-259.
 358 Ciais P, Sabine C, Bala G, Bopp L, Brovkin V, Canadell J, Chhabra A, DeFries R, Galloway J, Heimann M.
 359 2013. Carbon and Other Biogeochemical Cycles. In: Heinze C, Tans P, Vesala T, eds. *Climate*
 360 *Change 2013: The Physical Science Basis*. Cambridge, UK, and New York, NY, USA: Cambridge
 361 University Press.
 362 Duursma RA, Medlyn BE. 2012. MAESPA: a model to study interaction between water limitation,
 363 environmental drivers and vegetation function at tree and stand levels, with an example
 364 application to [CO₂] x drought interactions. *Geoscientific Model Development* 5:919-940.
 365 Elzhov TV, Mullen KM, Spiess A-N, Bolker B. 2016. minpack.lm: R Interface to the Levenberg-
 366 Marquardt nonlinear least-squares algorithm found in MINPACK, plus support for bounds. R
 367 package version 1.2-1. <https://CRAN.R-project.org/package=minpack.lm>
 368 Farquhar GD, et al. 1980. A biochemical model of photosynthetic CO₂ assimilation in leaves of C₃
 369 species. *Planta* 149: 78-90.
 370 Harley PC, Sharkey TD. 1991. An improved model of C₃ photosynthesis at high CO₂: reversed O₂
 371 sensitivity explained by lack of glycerate re-entry into the chloroplast. *Photosynthesis Research*
 372 27:169-178.
 373 Heskell MA, O'Sullivan OS, Reich PB, Tjoelker MG, Weerasinghe LK, Penillard A, Egerton JJG, et al.
 374 2016. Convergence in the temperature response of leaf respiration across biomes and plant
 375 functional types. *Proceedings of the National Academy of Sciences USA* 113: 3832-3837.
 376 Hobbs JK, Jiao W, Ester AD, Parker EJ, Schipper LA, Arcus VL. 2013. Change in heat capacity for enzyme
 377 catalysis determines temperature dependence of enzyme catalyzed rates. *ACS Chemical Biology*
 378 8:2388-2392.
 379 Jensen JLWV. 1906. Sur les fonctions convexes et les inégalités entre les valeurs moyennes. *Acta*
 380 *Mathematica* 30: 175-193.
 381 Johnson FH, Eyring H, Williams RW. 1942. The nature of enzyme inhibitions in bacterial luminescence:
 382 sulfanilamide, urethane, temperature and pressure. *Journal of Cellular and Comparative*
 383 *Physiology* 20:247-268.
 384 Kattge J, Knorr W. 2007. Temperature acclimation in a biochemical model of photosynthesis: a
 385 reanalysis of data from 36 species. *Plant, Cell & Environment* 30:1176-1190.

386 Kruse J, Hopmans P, Adams MA. 2008. Temperature responses are a window to the physiology of dark
 387 respiration: differences between CO₂ release and O₂ reduction shed light on energy
 388 conservation. *Plant, Cell Environment* 31: 901-914.

389 Kumarathunge DP, Medlyn BE, Drake JE, Tjoelker MG, Aspinwall MJ, Battaglia M, Cano FJ, Carter KR,
 390 Molly AC, Lucas AC, et al. 2019. Acclimation and adaptation components of the temperature
 391 dependence of plant photosynthesis at the global scale. *New Phytologist* 222:768-784.

392 Kumarathunge DP, Medlyn BE, Drake JE, Tjoelker MG, Aspinwall MJ, Battaglia M, Cano FJ, Carter KR,
 393 Molly AC, Lucas AC, et al. 2018. ACi-TGlob_V1.0: a global dataset of photosynthetic CO₂
 394 response curves of terrestrial plants. doi: 10.6084/m9.figshare.7283567.v1.

395 Liang LL, Arcus VL, Heskell MA, O'Sullivan OS, Weerasinghe LK, Creek D, Egerton JGG, et al. 2018.
 396 Macromolecular rate theory (MMRT) provides a thermodynamics rationale to underpin the
 397 convergent temperature response in plant leaf respiration. *Global Change Biology* 24:1538-
 398 1547.

399 Medlyn BE, Duursma RA, Eamus D, Ellsworth DS, Prentice IC, Barton CVM, Crous KY, et al. 2011.
 400 Reconciling the optimal and empirical approaches to modelling stomatal conductance. *Global*
 401 *Change Biology* 17:2134-2144.

402 Medlyn BE, Dreyer E, Ellsworth D, Forstreuter M, Harley PC, Kirschbaum MUF, Le Roux X, et al. 2002.
 403 Temperature responses of parameters of a biochemically based model of photosynthesis. II. A
 404 review of experimental data. *Plant, Cell & Environment* 25:1167-1179.

405 Norman JM, Campbell GS. 1998. An introduction to environmental biophysics. New York NY: Springer.

406 R Core Team. 2019. R: A language and environment for statistical computing. R Foundation for
 407 Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.

408 Rogers A, Medlyn BE, Dukes JS, Bonan G, von Caemmerer S, Dietze MC, Kattge J, et al. 2017. A
 409 roadmap for improving the representation of photosynthesis in Earth system models. *New*
 410 *Phytologist* 213:22-42.

411 Sharkey TD. 1985. Photosynthesis in intact leaves of C₃ plants – physics, physiology and rate
 412 limitations. *Botanical Review* 51:53-105.

413 Smith NG, Dukes JS. 2017. Short-term acclimation to warmer temperatures accelerates leaf carbon
 414 exchange processes across plant types. *Global Change Biology* 23:4840-4853.

415 Smith NG, Keenan TF, Prentice IC, Wang H, Wright IJ, Niinemets Ü, Crous KY, Domingues TF, et al.
416 2019. Global photosynthetic capacity is optimized to the environment. *Ecology Letters* 22:506-
417 517.

418 Stinziano JR, Murphy BK. 2020. *arrhenius.comparison*: comparing versions of the modified Arrhenius
419 equation. R package version 1.0.0.

420 Stinziano JR, Way DA. 2017. Autumn photosynthetic decline and growth cessation in seedlings of
421 white spruce are decoupled under warming and photoperiod manipulations. *Plant, Cell and*
422 *Environment* 40:1296-1316.

423 Stinziano JR, Way DA, Bauerle WL. 2018. Improving models of photosynthetic thermal acclimation:
424 which parameters are most important and how many should be modified? *Global Change*
425 *Biology* 24:1580-1598.

426 Stinziano JR, Bauerle WL, Way DA. 2019. Modelled net carbon gain responses to climate change in
427 boreal trees: impacts of photosynthetic parameter selection and acclimation. *Global Change*
428 *Biology* 25:1445-1465.

429 Walker AP, Quaife T, van Bodegom PM, De Kauwe MG, Keenan TF, Joiner J, Lomas MR, MacBean, et
430 al. 2017. The impact of alternative trait-scaling hypotheses for the maximum photosynthetic
431 carboxylation rate (V_{cmax}) on global gross primary production. *New Phytologist* 215:1370-1386.

432 Way DA, Yamori W. 2014. Thermal acclimation of photosynthesis: on the importance of adjusting our
433 definitions and accounting for thermal acclimation of respiration. *Photosynthesis Research*
434 119:89-100.

435 Weger HG, Guy RD. 1991. Cytochrome and alternative pathway respiration in white spruce (*Picea*
436 *glauca*) roots. Effects of growth and measurement temperature. *Physiologia Plantarum* 83:675-
437 681.

438

439

440 **Supplementary Information**

441 *Instructions for installing {arrhenius.comparison} in R for review purposes*

- 442 1. Download the arrhenius.comparison.tar.gz file
- 443 2. Set working directory in R to the directory that holds the arrhenius.comparison.tar.gz file using
- 444 `setwd()`
- 445 3. Run:
- 446 `install.packages("arrhenius.comparison_1.0.0.tar.gz", repos = NULL, type = "source")`
- 447 `library(arrhenius.comparison)`