DNA extraction

Needed equipment:



Mini-Centrifuge (ideally reaching 16.000 g)

A detailed protocol for Qiagen's Power Soil kit can be found in: https://www.qiagen.com/us/Resources/ResourceDetail?id=3d576814-4f1e-4e26-9c94-57d5dc2bb60a&lang=en

A brief summary follows:

1. Spin the PowerBead Pro Tube briefly to ensure that the beads have settled at the bottom.

Add up to 250 mg of sample and 800 µl of Solution CD1. Vortex briefly to mix.

- 2. Secure the PowerBead Pro Tube horizontally on a Vortex Adapter for 1.5–2 ml tubes Vortex at maximum speed for 10 min.
- 3. Centrifuge the PowerBead Pro Tube at 15,000 x g for 1 min.
- 4. Transfer the supernatant to a clean 2 ml Microcentrifuge Tube
- 5. Add 200 µl of Solution CD2 and vortex for 5 s.

- 6. Centrifuge at 15,000 x g for 1 min at room temperature. Avoiding the pellet, transfer up to 700 μ l of supernatant to a clean 2 ml Microcentrifuge Tube
- 7. Add 600 µl of Solution CD3 and vortex for 5 s
- 8. Load 650 μ l of the lysate onto an MB Spin Column and centrifuge at 15,000 x g for 1 min.
- 9. Discard the flow-through and repeat step 8 to ensure that all of the lysate has passed through the MB Spin Column.
- 10. Carefully place the MB Spin Column into a clean 2 ml Collection Tube
- 11. Add 500 μl of Solution EA to the MB Spin Column. Centrifuge at 15,000 x g for 1 min.
- 12. Discard the flow-through and place the MB Spin Column back into the same 2 ml Collection Tube.
- 13. Add 500 µl of Solution C5 to the MB Spin Column. Centrifuge at 15,000 x g for 1 min.
- 14. Discard the flow-through and place the MB Spin Column into a new 2 ml Collection Tube
- 15. Centrifuge at up to 16,000 x g for 2 min. Carefully place the MB Spin Column into a new 1.5 ml Elution Tube
- 16. Add $50-100 \mu l$ of Solution C6 to the center of the white filter membrane.
- 17. Centrifuge at 15,000 x g for 1 min. Discard the MB Spin Column.

If the portable centrifuge cannot provide the requested g, try increasing centrifugate time.