

MUDANÇAS NOS PADRÕES DIÁRIOS DE ATIVIDADE AO LONGO DO ANO EM
TUCO-TUCOS DE VIDA LIVRE

CHANGES IN DAILY ACTIVITY PATTERNS THROUGHOUT THE YEAR IN FREE-LIVING
TUCO-TUCOS

Jefferson T. Silvério

Dissertação de Mestrado apresentada ao
Programa de pós graduação em Ciências, na
Área de Fisiologia Geral, IB, da Universidade
de São Paulo, como parte dos requisitos
necessários à obtenção do título de Mestre em
Ciências, na Área de Fisiologia Geral.

Orientadores: Gisele A. Oda

Verônica S. Valentinuzzi

Patricia Tachinardi Andrade
Silva

São Paulo

Fevereiro de 2022

MUDANÇAS NOS PADRÕES DIÁRIOS DE ATIVIDADE AO LONGO DO ANO EM
TUCO-TUCOS DE VIDA LIVRE

CHANGES IN DAILY ACTIVITY PATTERNS THROUGHOUT THE YEAR IN FREE-LIVING
TUCO-TUCOS

Jefferson T. Silvério

Banca Examinadora: Prof. Dr. Enrico L. Rezende

Dr. Francisco V. Dénes

Prof. Dr. Tiago G. de Andrade

SÃO PAULO, SP – BRASIL

FEVEREIRO DE 2022

Silvério, Jefferson T.

Mudanças nos padrões diários de atividade ao longo do ano em tuco-tucos de vida livre/Jefferson T. Silvério.
– São Paulo: Universidade de São Paulo/Instituto de Biociências, 2022.

XI, 92 p.

Orientadores: Gisele A. Oda

Verónica S. Valentinuzzi

Patricia Tachinardi Andrade Silva

Dissertação (mestrado) – Universidade de São Paulo/Instituto de Biociências/Programa de Ciências, na Área de Fisiologia Geral, 2022.

1. Ctenomys. 2. biologging. 3. hidden markov models. 4. chronobiology. 5. subterranean rodent. 6. neotropical rodent. 7. activity patterns. 8. diurnality. 9. cronobiologia. 10. roedor subterrâneo. 11. roedor neotropical. 12. padrões de atividade. 13. diurnalidade.

I. Oda, Gisele A., orient. II. Universidade de São Paulo, Ciências, na Área de Fisiologia Geral. III. Título.

“O sol afundou mais um pouco. As sombras se alongaram, atravessando a caldeira de sal. Linhas de cores vibrantes espalharam-se sobre o horizonte do poente. A cor foi se transformando num dedo de trevas a experimentar a areia. As sombras cor de carvão se alastraram, e o desmoronamento compacto da noite obliterou o deserto.”

— Frank Herbert, Duna.

Agradecimentos

Agradeço aos meus pais, Gerson e Dalva, e minha irmã, Beatriz, pelo apoio em um caminho de carreira que tem características tão particulares. Sem o apoio de vocês eu não teria conseguido trilhar esse caminho. Agradeço também a minha segunda família, Gilvan e Vânia, pelo acolhimento tão carinhoso nesse meu tempo em São Paulo.

Agradeço a minhas orientadoras Gisele e Verônica por terem me acolhido no laboratório. Pela dedicação nesses anos, mesmo à distância, e pelos ensinamentos em cronobiologia e em trabalho em grupo. Agradeço também a minha orientadora Patricia por ter me deixado participar desse projeto e pelas discussões, scripts, extrações e ideias sobre tucos e reprodução e pela companhia nas coletas de campo.

Agradeço aos colegas de laboratório Danilo, Milene e Giovane pela ajuda nas coletas de campo, discussões, desabafos e pela companhia e refeições compartilhadas no CRILAR. Obrigado também ao Danilo pelas conversas sobre tecnologia. Obrigado a Johana que me ajudou a resolver diversos problemas em espanhol e também me ajudou nas coletas de campo. Obrigado também aos amigos da minha “coorte” de pós na fisiologia: Beatriz e Mila.

Agradeço aos amigos do *Alô Ciência?*: Arnoni, Amanda, Beatriz, Caramelo, Camila, Leandro, Lucas e Marx pelas discussões sobre ciência e por toda companhia durante os últimos anos. Fazer parte desse grupo me deu uma visão muito mais abrangente de ciência e da sociedade. É um prazer dividir um planeta com vocês.

Agradeço também aos comentaristas de BBB: Arnoni, Bala, Caramelo, Limão, Marx e Zattar. Vocês fizeram esses últimos anos trancados em casa muito melhores.

Agradeço ao time de ultimate Frisbee que me ajudou a manter a sanidade nos

últimos anos e que também me deram uma visão muito diferente sobre o esporte e dureza mental. Um agradecimento especial aos treinadores Paolo, Gus e Nat. Agradeço também ao Alex por todo tempo dedicado na MoVer-se que me ajudaram a conseguir sentar, correr e jogar sem me preocupar com tantas dores.

I also thank James Kenagy for taking interest in me and my project and for accepting being part of my committee. I enjoyed our email exchanges and the teaching moments in CRILAR. Thank you for all the personal, scientific and writing advice and references. It was a privilege to get to know you.

Obrigado também aos amigos feitos no LASC: Guada, Mayara, Julia e Nathalia pela companhia no congresso e pelas conversas depois dele.

Por último, agradeço principalmente a Aline que foi a pessoa que mais esteve presente nessa jornada e foi essencial para o término dessa tese. Você não só me ajudou em planejamentos e ideias, mas também manteve meus pés no chão durante todo esse tempo. É um prazer dividir minha vida com você.

O presente trabalho foi realizado com apoio da Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – Brasil (CAPES) – Código de Financiamento 001.

RESUMO

Os padrões temporais de atividade no tuco-tuco de Anillaco (*Ctenomys aff. knighti*), um roedor subterrâneo, foram estudados extensivamente em laboratório e arenas semi-naturais. No entanto, essas condições controladas não reproduzem completamente o que os animais fazem na natureza. O presente estudo amplia nosso entendimento dos padrões de atividade dos tuco-tucos e descreve sua atividade diária em condições de vida livre. Nós coletamos dados de 21 tuco-tucos usando acelerômetros e sensores de luz ao longo de 2019 e 2020, em todas as estações do ano. Além disso, usamos modelos ocultos de Markov (HMMs) para classificar os dados do acelerômetro em três estados comportamentais distintos que correspondem, aproximadamente, a repouso e atividades de média e alta intensidade. Os registros do sensor de luz confirmam uma mudança anual no padrão temporal diário de emergência da superfície, a qual já havia sido descrita para animais em arenas semi-naturais. Os registros de acelerômetro confirmam que os tuco-tucos são diurnos em campo, sem alteração no padrão temporal diário ao longo do ano. Em contrapartida, em julho, os tuco-tucos apresentaram uma redução significativa nos níveis de atividade diária. Em termos de mudanças de estado comportamental, os tuco-tucos apresentaram mudanças significativas no tempo alocado em diferentes estados ao longo do ano. Em julho, houve uma redução no tempo diário gasto em atividades de alta intensidade. No entanto, observamos que os tuco-tucos não repousam mais em julho; em vez disso, eles alocam mais tempo em atividades de média intensidade. Esses resultados sugerem que as mudanças anuais nos níveis de atividade diária se devem a mudanças no tempo gasto em diferentes estados comportamentais. Em geral, confirmamos que os tuco-tucos são diurnos e apresentam mudanças anuais em seus níveis gerais de atividade, o que provavelmente é reflexo de uma mudança comportamental anual.

ABSTRACT

Temporal patterns of activity of the Anillaco tuco-tuco (*Ctenomys* aff. *knighti*), a neotropical subterranean rodent, have been extensively studied in laboratory and semi-natural enclosures. However, the laboratory-controlled settings do not fully reproduce what animals do in nature. The present study extends our understanding of tuco-tucos activity patterns and describes their daily activity in free-living conditions. We collected data from 21 tuco-tucos using animal-borne accelerometers and light loggers during 2019 and 2020, covering all seasons. Additionally, we used hidden Markov models (HMMs) to classify the accelerometer data into three distinct behavioral states that roughly correspond to rest, medium, and high-intensity activity. Lightlogger data confirms the annual change in the daily temporal pattern of surface emergence already described for animals in semi-natural enclosures. Our accelerometer data confirm that tuco-tucos are diurnal in the field, with no change in the daily temporal pattern throughout the year. Nevertheless, in July, tuco-tucos displayed a significant reduction in daily activity levels. In terms of behavioral state changes, tuco-tucos displayed significant changes in the time allocated in different states throughout the year. In July, there was a reduction in the daily time spent in high-intensity activity. However, we observed that tuco-tucos do not rest more in July; instead, they allocate more time in medium-intensity activities. These results suggest that the annual changes in daily activity levels are due to changes in the time spent in different behavioral states. All in all, we confirm that tuco-tucos are diurnal and exhibit annual changes in their general activity levels, which is probably a reflection of an annual behavioral change.

Sumário

Introdução Geral	1
O oscilador circadiano e os ritmos endógenos	1
Tuco-tucos como sistema cronobiológico	3
1 Chronobiology in the wild: toolkit to study daily rhythms in free-living animals	6
1.1 Introduction	7
1.2 Chronobiological studies in free-living animals	8
1.2.1 Populational Rhythms	8
1.2.2 Individual behavioral rhythms	10
1.2.3 Physiological rhythms	13
1.3 Conclusion	14
1.4 Acknowledgements	15
2 Changes in daily activity patterns throughout the year in free-living tuco-tucos (<i>Ctenomys</i> sp.)	16
2.1 Introduction	16
2.2 Methods	20
2.2.1 Study Species	20
2.2.2 Study Site	21
2.2.3 Animal Capture and Handling	21
2.2.4 Biologgers	23
2.2.5 Data Processing	25

2.2.6	Surface Emergence & Time On Surface	26
2.2.7	General Activity	27
2.2.8	Behavioral State Classification	27
2.2.9	Daily Time-Activity Budgets & Behavioral States Patterns	30
2.2.10	Aboveground Activity	30
2.2.11	Diurnality Index	31
2.2.12	Circadian Rhythmicity and Period Estimation	31
2.2.13	Statistical tests	32
2.3	Results	33
2.3.1	Time On Surface	33
2.3.2	General Activity	35
2.3.3	Behavioral State Classification	37
2.3.4	Daily Behavioral States Patterns	39
2.3.5	Diurnality	41
2.3.6	Daily Time-Activity Budgets	42
2.3.7	Aboveground Activity	44
2.3.8	Circadian Rhythmicity	45
2.4	Discussion	47
Considerações Finais		52
Apêndice		54
A Animals		55
B Anillaco's Environment		56
B.1	Anillaco's Plant Community	56
B.2	Anillaco's Weather	58
B.3	Anillaco's Yearly Daylength Changes	59
C Individual Actograms		60
C.1	Surface Emergence Actograms	61

C.2	VeDBA Actograms	62
C.3	High Activity Actograms	63
C.4	Medium Activity Actograms	64
C.5	Low Activity Actograms	65
D	Accelerometer Data Exploration	66
D.1	Assessment of Static Acceleration Smoothing Window	66
D.2	VeDBA Summary and Distributions	68
E	Hidden Markov Model	71
E.1	AIC	71
E.2	Estimated Parameters	71
E.3	Pseudo-residuals	72
E.4	Stationary Probabilities	73
E.5	Transition Probabilities	74
F	Individual Circadian Analysis	75
F.1	Individual Autocorrelation Plots	76
F.2	Individual Period Estimation	77
Referências		78

Introdução Geral

Quando olhamos ao nosso redor, é difícil não notar a existência de uma diversidade de ciclos ambientais: os ciclos diários, como o do claro/escuro associado ao dia e noite; os ciclos anuais, como a variação anual de temperatura, e outros ciclos, como o lunar e o ciclo das marés. Não é de se admirar, então, que ao longo da evolução tenham surgido mecanismos que possibilitaram aos organismos se adaptarem temporalmente a essas mudanças cíclicas e previsíveis do ambiente. Assim, a maioria dos organismos desenvolveu sistemas temporais endógenos, como o sistema circadiano, associado ao ciclo diário de 24 horas. Esse sistema é responsável pela geração e sincronização de diversos ritmos biológicos, alguns bastante perceptíveis, como o padrão diário de atividade. Outros ritmos biológicos, invisíveis aos olhos, controlam funções importantes na vida dos animais, é o caso da variação diária da concentração de diversos hormônios no plasma (Moore-Ede, Sulzman, and Fuller 1982).

O oscilador circadiano e os ritmos endógenos

A endogenicidade dos ritmos biológicos foi demonstrada pela primeira vez em 1729 pelo astrônomo Jean Jacques de Mairan. De Mairan investigava o movimento foliar de uma planta que fechava suas folhas durante a noite e realizou um experimento no qual colocou essa planta em uma caixa fechada, sem entrada de luz. Ele observou que, mesmo sem uma pista ambiental indicando quando era dia ou noite, o ritmo de abertura e fechamento das folhas se manteve. Desde então os ritmos biológicos vêm sendo estudados com mais atenção, levando ao estabelecimento da cronobiologia como disciplina a partir de 1960, com o propósito de investigar a origem e manutenção dos

ritmos biológicos endógenos e como eles são sincronizados pelos ciclos ambientais (Moore-Ede, Sulzman, and Fuller 1982).

Em mamíferos, os ritmos circadianos são gerados por um oscilador endógeno central, localizado nos núcleos supraquiasmáticos do hipotálamo (Moore-Ede, Sulzman, and Fuller 1982). Assim como no experimento de De Mairan, os ritmos biológicos endógenos em mamíferos se mantêm mesmo em condições laboratoriais constantes, com período próximo, mas não idêntico, ao do ciclo ambiental associado. Essa é uma propriedade importante dos ritmos biológicos, e mostra que esses ritmos são endógenos e possuem um período intrínseco. O período do ritmo em condições constantes é denominado período de livre curso (τ), e corresponde ao período endógeno do oscilador circadiano na ausência de pistas temporais ambientais. O valor de τ se aproxima, mas não é igual ao valor do período do ciclo ambiental com o qual ele é associado. No caso de ritmos diários o valor do tau fica em torno de 24 horas, variando de espécie para espécie, e por isso esses ritmos são conhecidos como ritmos circadianos (Circa = cerca de, dien = dia) (Moore-Ede, Sulzman, and Fuller 1982).

Outra propriedade importante dos ritmos circadianos, além da endogenicidade, é a sua sincronização por ciclos ambientais diários. Para grande parte dos animais, o principal agente sincronizador do oscilador circadiano é o ciclo claro-escuro. A sincronização acontece pela transmissão da informação temporal cíclica aos osciladores por meio de fotorreceptores, que compõem a via aferente do sistema circadiano (Figure 1). Em mamíferos, particularmente, os fotorreceptores estão presentes exclusivamente na retina e a informação luminosa percebida pelas células retinianas é transmitida para o oscilador circadiano por meio do trato retino hipotalâmico (Moore-Ede, Sulzman, and Fuller 1982). Assim, apesar do τ de um ritmo circadiano ser ligeiramente diferente de 24 horas, há um ajuste do período desse ritmo com o período do ciclo ambiental resultante do processamento da informação luminosa cíclica diária. Dessa maneira, o animal é capaz de manter um ritmo sincronizado, com período de 24 horas, mantendo também estável a relação de fase – que é a distância temporal entre duas fases de referência – desse ritmo com o ciclo ambiental.

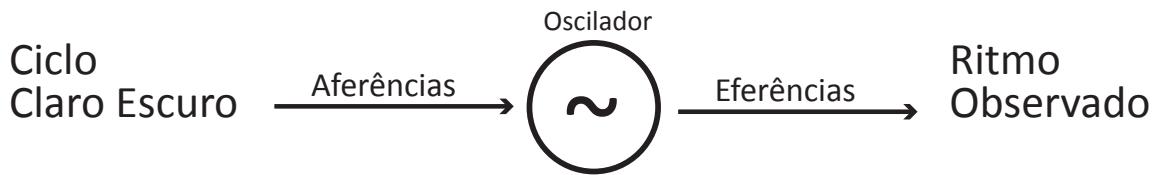


Figura 1: Figura esquemática do mecanismo de sincronização do oscilador circadiano. O ciclo claro-escuro é percebido por células fotorreceptoras presentes na retina. A informação luminosa é transmitida para o oscilador através das vias aferentes, que em mamíferos é o trato retino hipotalâmico. O oscilador circadiano se sincroniza ao ciclo luminoso por meio da informação nervosa proveniente das vias aferentes. O oscilador circadiano mantém uma marcação interna do tempo e transmite essa informação temporal para diversos órgãos e tecidos, o que resulta no ritmo que é observado em diversas variáveis fisiológicas e comportamentais.

Tuco-tucos como sistema cronobiológico

A família Ctenomyidae está presente em grande parte da América do Sul se estendendo desde o sul do Peru até Tierra del Fuego, Argentina, e do leste até o sudoeste do Brasil, ocupando uma diversidade de habitats, desde áreas florestais até regiões áridas (Lacey, Patton, and Cameron 2000). A família apresenta apenas um gênero vivente, *Ctenomys*, que é o gênero de roedor subterrâneo com maior número de espécies descritas. Espécies desse gênero são tipicamente solitárias, apresentam diversas adaptações morfológicas ao hábito de vida subterrâneo e são chamadas popularmente de tuco-tucos, pela semelhança desse nome com o som produzido pela vocalização desses animais (de Freitas, Gonçalves, and Maestri 2021). A natureza subterrânea, a grande distribuição geográfica e a relativa facilidade de manutenção em laboratório, tornam esses roedores sistemas muito atraentes para o estudo cronobiológico. A população de animais estudados pelo Laboratório de Cronobiologia Binacional é descrita em trabalhos anteriores como pertencentes à espécie *Ctenomys aff. knight* mas a descrição e nomenclatura específica está em andamento. Essa população ocorre na região em torno de Anillaco, La Rioja, Argentina, e possui o nome popular de tuco-tuco de Anillaco.

Em laboratório, o ritmo de atividade do tuco-tuco de Anillaco é sincronizado pelo ciclo diário de claro/escuro e apresenta padrão noturno (Flôres et al. 2021; Valentimuzzi et al. 2009). Estudos realizados em arenas seminaturais revelaram que os ritmos de

atividade dessa espécie também se sincronizam com o ciclo claro-escuro ambiental, mesmo com os animais vivendo em galerias subterrâneas (Jannetti et al. 2019; Tomotani et al. 2012). Durante esses estudos em arenas seminaturais, foi observado que a exposição natural à luz desses animais acontece principalmente quando os tuco-tucos saem de suas tocas para forragear ou escavar seus túneis. Embora essa exposição à luz seja irregular, ela já é suficiente para sincronizar o oscilador desses animais (Flôres et al. 2013; Flôres et al. 2021). Um outro resultado importante observado nos trabalhos com tuco-tucos de Anillaco é a diferença entre as fases de atividade em laboratório e nas arenas seminaturais. No laboratório, os picos de atividade estão concentrados na fase de escuro e indicam um comportamento noturno (Tomotani et al. 2012). Nas arenas, a situação é diferente, os picos de atividade estão mais concentrados durante o dia e indicam um comportamento diurno (Jannetti et al. 2019).

Os primeiros estudos de tuco-tuco em arenas seminaturais tinham como objetivo investigar a exposição à luz e os padrões de saídas dos tuco-tucos de suas galerias subterrâneas (Tomotani et al. 2012). No entanto, a resposta a algumas perguntas demandava também o estudo da atividade dos tuco-tucos no subterrâneo. Esse tipo de investigação se tornou possível graças à miniaturização de sensores de movimento, que são pequenos o suficiente para serem usados em colares colocados em pequenos roedores. Esses tipos de sensores são conhecidos como *biologgers* e seu uso como *bio-logging*. Um tipo de *bilogger* comumente utilizado para o registro de movimento é o acelerômetro, sensor que registra movimentos por meio de aceleração do corpo e que também estão presentes em celulares, sendo os responsáveis pela detecção da orientação do aparelho. Em animais, os acelerômetros são fixados em colares e registram o movimento do animal continuamente em alta frequência. Dessa forma, esses sensores são muito interessantes para os estudos de cronobiologia e de ritmos de atividade. Os primeiros registros de acelerômetro em tuco-tucos foram feitos em arenas seminaturais por Jannetti et al. (2019). Os resultados desse estudo apontam que em condições seminaturais os tuco-tucos são majoritariamente diurnos e passam por volta de 60% de seu dia ativos.

Apesar do conhecimento adquirido sobre a atividade dos tuco-tucos em laboratório e arenas seminaturais, essas condições não reproduzem totalmente a complexidade do ambiente natural em que os tuco-tucos vivem. Por esse motivo, não sabemos se os resultados em arenas seminaturais refletem os comportamentos naturais dos tuco-tucos na natureza, o que só pode ser explorado em estudos com animais em vida livre. Assim, esta dissertação se propõe a implementar uma metodologia de campo para o estudo dos ritmos de atividade de tuco-tucos na natureza. Também temos como objetivo investigar e identificar quais padrões de atividade os tuco-tuco de Anillaco apresentam em seu ambiente natural, dentro do contexto ecológico da espécie, e se existe alteração anual desses padrões. Além disso, é também proposto o uso de *Hidden Markov models* para a classificação dos dados de acelerômetro, o que amplia de maneira significativa as interpretações que podemos fazer com esse tipo de dado.

O primeiro capítulo, intitulado “Chronobiology in the wild: toolkit to study daily rhythms in free-living animals,” foi publicado em uma edição especial da revista *Sleep Science* como parte da premiação de melhores pôsteres apresentados durante o XV LASC (Latin American Symposium on Chronobiology, 2018). Esse artigo traça um panorama geral do estudo de ritmos biológicos, em especial de atividade, em animais de vida livre e as tecnologias que permitem que esses estudos sejam realizados atualmente. O segundo capítulo, no formato de artigo, ainda será submetido a uma revista da área. Neste capítulo, realizamos o primeiro estudo da atividade geral e da saída à superfície em tuco-tucos de vida livre. Esse trabalho foi feito com o objetivo de confirmar estudos e observações anteriores feitas em arenas seminaturais, além de investigar possíveis mudanças anuais na atividade dessa espécie dentro do seu contexto ecológico.

Capítulo 1

Chronobiology in the wild: toolkit to study daily rhythms in free-living animals

Jefferson T. Silvério, Patricia Tachinardi

Published in Sleep Science - Volume 13, Supplement 2. 2020.

Since chronobiology's foundation has been laid out in the 60s, tremendous progress has been made regarding our knowledge about the nature of the circadian clock, its molecular bases and its synchronization by photic and non-photic stimuli. The majority of these studies have been done in laboratory settings, which, although important, lack information about the adaptive value, ecological significance and plasticity of the biological rhythms in the wild. This concern has been raised several times, along with the development of the field. Methodological difficulties have been the biggest challenges of field studies. However, the recent development of new techniques is opening a wide range of opportunities to investigate biological rhythms in free-ranging animals. In this review, we promote an ecological approach to biological clocks and highlight some methods and newer technologies that can be used to study biological

clocks in the wild, along with some examples.

Keywords: biological rhythms; wild-clocks; field and laboratory; bilogger; free-ranging; activity patterns.

1.1 Introduction

Endogenous circadian clocks enable animals to anticipate changes in physiology and behavior to the daily changes in the environment synchronizing their internal state with the 24-hour environmental cycles. Our knowledge of the function and mechanism of these clocks has grown extensively in different levels of organizations such as molecular, cellular, tissue and organs. Most of the studies of the mechanisms and nature of the clock itself has been done in the laboratory, under controlled conditions (Enright 1970; Morgan 2004). Using the laboratory approach, several endogenous circadian rhythms were described in a diverse range of organisms, from mammals to birds, insects and the molecular clocks of bacteria. However, the physiology and behavior of a laboratory animal can be unrepresentative of what happens in the field (Calisi and Bentley 2009; Willmer, Stone, and Johnston 2005). Several authors have brought the argument that we know very little about the ecological significance of circadian rhythms (Enright 1970; Halle and Stenseth 2000; Helm et al. 2017). The central argument is that to understand the adaptive significance of circadian rhythms, the diversity of temporal strategies and how flexible circadian rhythms are, we must turn to ecological studies in free-living animals. To fill this gap, several recent studies on animal chronobiology have been carried out in free-living animals. These studies investigate the interplay between the endogenous circadian pacemaker and the biotic and abiotic stimuli acting on the plasticity of overt daily rhythms. These studies have suggested that biological rhythms in the wild are much more flexible than previously thought (Hut et al. 2012).

The advancement of chronobiological studies in the field has long been challenged by the difficulty of measuring biological rhythms in nature. In the laboratory, locomotor

activity and body temperature (T_b) rhythms have been widely used due to ease of measurement and replicability. Commonly, activity is measured by counting running wheel revolutions, by detecting movement with an infrared motion sensor placed above the animal's cage or by implanting a wireless activity sensor in the animal (Blanchong et al. 1999). Measurements of T_b, in turn, are more invasive and can require more expensive equipment. Usually, T_b sensors are implanted surgically in the peritoneal cavity of the animal. These sensors can either be loggers, in which the data is stored in the device or transmitters, which send data to receivers outside the animal (Weinert and Waterhouse 1998). When studying free-living animals, however, many of these commonly used methods to study locomotor activity and T_b rhythms are very difficult or simply impossible to be deployed, being it because of technical or financial drawbacks.

Recent technological advances, such as the miniaturization of sensors and logging devices, have opened a wide variety of opportunities to study biological rhythms in free-living animals. In this mini-review, we will discuss some of the methods being used to assess behavioral and physiological rhythms of free-living animals in both populational and individual levels. We will focus on methods used in the research of terrestrial mammals, although many of them can also be deployed to other taxa and to aquatic and flying animals (Dominoni et al. 2017). It is important to note that most methods require handling the animal for implanting or externally attaching a device. Thus, researchers should always take into account any possible effects this manipulation can have on the animals and on the data obtained (Chmura, Glass, and Williams 2018).

1.2 Chronobiological studies in free-living animals

1.2.1 Populational Rhythms

Most of the early investigations of clocks in the wild have been carried out focusing on populational rhythms, using observational and trapping methods (Daan and Slopsema

1978; Halle and Stenseth 2000; Kenagy 1976). Using live-traps to infer activity rhythms can be done setting a grid of traps in the study site and then counting the number of captures for a chosen time interval. For example, using hourly record of vole (*Microtus arvalis*) trappings in the wild it was possible to record the pattern of daily activity in these species and their ultradian components (M. Gerkema and Daan 1985). If a higher temporal resolution is required, it is possible to combine live-trapping and direct observations to record the exact time the animals were trapped. Using this approach it was shown that Kangaroo rats (genus *Dipodomys*) display seasonal change in their start and end time of activity, which might be related to the length changes of sunset (Kenagy 1976). Both observational and live-trapping methods are limited in the range of habitats and diversity of animals they can be applied to. Observational studies have to be done in diurnal species living in open habitats, otherwise, it becomes unfeasible to observe the study species. Small nocturnal animals and animals living in habitats with dense vegetation are not well suited for this technique. On the other hand, trapping studies face the problems of how easily individuals of the study species can be trapped (Kenagy 1976). Both of the above mentioned examples were done in species that are relatively easy to capture in live traps. Kangaroo rats, for example, do not exhibit an aversion to the traps and manipulation. Voles, in turn, leave recognizable tracks aboveground that can guide trap placement and increase trapping success.

More recently, the use of time-stamped camera-trap images has expanded the study of population activity rhythms to species that cannot be easily observed or captured (Frey et al. 2017; Rowcliffe et al. 2014). Moreover, this non-invasive technique can be used to simultaneously monitor several species that occur in the same area. For example, Mendes and colleagues (Mendes et al. 2020) did an extensive camera-trap study to evaluate the changes in daily activity rhythms of 17 rainforest-dwelling mammal species in response to human disturbance. They found out that there was a shift in the timing of activity in highly disturbed areas, with some species becoming more nocturnal and others more diurnal. Species that were susceptible to preying or hunting were more likely to show activity time shifts. Possible drawbacks of using camera-traps are

camera placement bias, detectability of the animals in the camera and the assumption that all individuals of the population will display peak activity at the same time (Frey et al. 2017; Rowcliffe et al. 2014).

1.2.2 Individual behavioral rhythms

In general, to study individual daily activity patterns it is necessary to continuously follow and record movement and behavior of the same individual for many days (Daan 1981), which can be very challenging in the field. Individual monitoring can be done using direct observations (Everts et al. 2004; Tomotani et al. 2012), but this method restricts the monitoring of the animal to the times when it is visible. For continuous measurements, it is usually required to attach a device to the animal. Among the earliest measurements of individual activity rhythms using animal-borne devices were those using radio telemetry (Ables 1969; Hut, van Oort, and Daan 1999). When using this technique, the animal is equipped with a radio telemetry collar that continuously transmits a pulse in a certain radiofrequency that can be picked up by a radio receiver tuned to the same frequency. This way it is possible to locate the animal in the field during defined time intervals. Daily activity patterns are inferred by the size of spatial displacements between these time points throughout day and night. Due to the availability of small and lightweight transmitters, this technique can be used on mammals of nearly any size (Naef-Daenzer 2005). Telemetry studies are often done manually, thus being very labor-intense and time-consuming. There are automated telemetry systems, which are a feasible alternative to measure activity patterns, as long as the technical and financial requirements to set up the system can be met and errors due to small body size and large home ranges are taken into account (Kays et al. 2011). Another method that can be used to assess activity patterns based on animal movement are satellite trackers, which previously could only be used in large-sized animals, but are currently being miniaturized (Wikelski and Cooke 2006). An important advantage of this technique is that data can be obtained remotely, with no need to locate the animal after the tag is deployed.

Individual-level rhythms can also be measured with newer biologging devices, which are attached to the animal's body and can record onboard physiological and behavioral parameters including activity, Tb, heart rate and skin humidity (Ropert-Coudert and Wilson 2005; Rutz and Hays 2009). Given the advancements in the biologging technologies, devices are now smaller and capable of recording multiple parameters in a frequency of hundreds of points per second, providing insight into the details of the animal's life (Ropert-Coudert and Wilson 2005). Due to the onboard memory, the main drawback of biologgers is the need to recapture the animals to obtain the data.

Examples of biologging devices used to monitor animal behavior in chronobiological studies include lightloggers and accelerometers. Lightloggers are devices that were originally designed to be used in geolocation in bird migration studies but are particularly interesting for chronobiology. Lightloggers are being used to conduct research on a number of small rodents, recording their temporal patterns of exposure to light. Studies that are benefitted by the use of lightloggers include research on diurnal animals that retreat to nests, subterranean or semi-fossorial animals and hibernating animals in which exposure to light means the animals are active above ground. Accelerometers, in turn, are tri-axial devices that continuously record fine-scale movement. The data generated by accelerometers can be used to quantify gross activity timing, identify behaviors based on movement patterns, quantify time-activity budgets and indirectly derive energy expenditure (Brown et al. 2013; Rutz and Hays 2009; Ropert-Coudert and Wilson 2005).

Lightloggers were used to investigate the daily and seasonal patterns of activity in wild red squirrels (*Tamiasciurus hudsonicus*) and arctic ground squirrels (*Urocitellus parryii*) (Williams et al. 2014). Using these devices, they could calculate the total time outside of the nest, the number of activity bouts per day, time of the first emergence in the morning and return to nest at night. They showed that the level of activity was flexible and correlated with changes in the thermoregulatory conditions of the environment. On a different rodent species, the tuco-tuco (genus *Ctenomys*), a combi-

nation of lightloggers and accelerometry was used to investigate the activity patterns of a subterranean rodent in outdoor enclosures (Jannetti et al. 2019). In this study, accelerometers were used to record gross motor activity while lightloggers recorded time on surface. A combination of both methods, therefore, could discriminate levels of aboveground activity and below-ground activity when the animal is inside its burrow system. The study showed that time of day and temperature were the main environmental conditions modulating time spent on the surface (Jannetti et al. 2019) and that there was a shift from diurnal to nocturnal levels of activity when the same animals were transferred from outdoor enclosure to controlled laboratory conditions (Jannetti et al. 2019; Tachinardi et al. 2015). Although this study was done in outdoor enclosures, this approach can also be used in free-ranging animals. Shifts in the timing of activity are also known to happen in other species of rodents besides *Ctenomys* sp. (Hut et al. 2012; Kronfeld-Schor et al. 2017) and are among the most dramatic discrepancies between rhythms in the field and lab. Biologgers are a helpful addition to approach this phenomenon at the individual level and boost this investigation. The examples provided here highlight possible scenarios where the use of biologging devices can be used to reveal the intimacy and complexity of rhythms in the wild, investigating chronobiological questions and the flexibility of circadian and seasonal rhythms.

Other alternative technologies can be used to assess rhythms in the wild, such as RFID (Radio Frequency Identification) and newly developed ones such as the BATS system. RFID are small tags that can be used to identify individuals in a population. Passive RFID tags do not require their battery to operate and are powered by the energy from the reader's radio waves. They are largely used to identify farm and laboratory animals, being very small and attached either externally or subcutaneously. RFID tags can be used to record activity rhythms by placing readers in locations that the animals visit regularly, such as nests or feeders. This setup allows the recording of the rhythmicity of visits to a specific location where the reader is placed (van der Vinne et al. 2019). Although this technique is usually deployed in outdoor enclosures it could be used in free-living animals that have fixed nests or feeding locations. The

BATS system is an interesting technology that is fully automated (Duda et al. 2018; S. P. Ripperger, Carter, Page, et al. 2019). It is composed of small lightweight proximity sensors and a set of receiver stations that are distributed in the study site. The proximity sensors can record interactions between two animals in a sampling rate of seconds and then transmit these data to the base stations. This system can also be used to derive movement trajectory even in structurally complex habitats (S. P. Ripperger, Carter, Page, et al. 2019). Ripperger and colleagues have used this system to study the behavioral ecology of bats, such as the mother-offspring interactions in noctule bats (*Nyctalus noctule*) (S. Ripperger et al. 2019) and the social structure of the common vampire bats (*Desmodus rotundus*) (S. P. Ripperger, Carter, Duda, et al. 2019). Although the BATS system was developed to investigate a different set of questions it seems to be a great new technology to assist in the study of reproduction, social synchronization and the social influences on biological rhythms.

1.2.3 Physiological rhythms

Biologgers can also be used to assess daily rhythms of physiological variables, enabling the investigation of physiological adjustments to the changes in environmental conditions faced by free-living animals. Tb is the physiological variable most studied in wild chronobiology. It is usually measured and recorded by temperature loggers implanted into the abdomen. This method was used to monitor Tb in free-living arctic ground squirrels (*Urocitellus parryii*) throughout the year, showing that in these animals Tb is arrhythmic during hibernation but displays an entrained 24-hour rhythm in the active season even during the weeks of constant sun (Williams et al. 2011). Monitoring of Tb was also used to record dramatic shifts from diurnally to nocturnality when golden spiny mice (*Acomys russatus*) are transferred from large outdoor enclosures to the laboratory (Levy, Dayan, and Kronfeld-Schor 2007). This study is a clear example of how the assessment of physiological rhythms can be important in the investigation of the plasticity of biological rhythms, since the same physiological variable can be measured in both conditions, as opposed to measuring different types of behavioral rhythms un-

der field and laboratory conditions (e.g. foraging in the field and wheel running in the laboratory).

In the laboratory, energy expenditure rhythms can be assessed by measuring hourly rates of oxygen consumption using respirometry chambers, for instance, which is not possible in free-living animals. Since heart rate can be a proxy to energy expenditure, it is a valuable physiological variable in chronobiological studies involving energetics. Heart rate sensors can be attached to the skin, implanted subcutaneously (Tøien et al. 2011) or even placed non-surgically in the reticulum of ruminants (Signer et al. 2010). These devices can either be loggers (Tøien et al. 2011) or transmitters (Signer et al. 2010).

Brain electrical activity is also of interest to chronobiological studies since it can be used to investigate sleep patterns. Recently, miniature loggers have been developed to measure electrophysiological brain activity (Massot et al. 2019; Vyssotski et al. 2006). Studies using these loggers have, for instance, provided insights into the ecology of sleep in sloths (Rattenborg et al. 2008) and even indicated that birds can sleep during flight (Rattenborg et al. 2016).

1.3 Conclusion

In this mini-review, we highlighted some, but not all, methods that can be applied in chronobiological studies in the wild. There is a range of new devices and techniques currently being developed that could be applied to free-living animals to assess biological rhythms. The combination of different devices can also be used to discriminate and investigate how some behaviors or physiology interact, even at the cellular and molecular levels (Jansen et al. 2016). The fast methodological advancement and the increasing interest of chronobiologists in field studies are key aspects to narrow the gap between what we know about the circadian clocks in the lab and what we know about their functional significance in nature.

1.4 Acknowledgements

We would like to thank Gisele Oda for the revision and suggestions that improved the manuscript; Danilo Flôres, Milene Jannetti and Giovane Improta for discussion of everything chronobiology; and the organizing committee of the XV Latin American Symposium on Chronobiology. JTS was funded in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Finance Code 001, CAPES scholarship (88882.377383/2019-01) and additional grant from Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) (2017/19680-2). PT was funded by FAPESP fellowship 2017/22973-1.

Capítulo 2

Changes in daily activity patterns throughout the year in free-living tuco-tucos (*Ctenomys* sp.)

2.1 Introduction

Many environmental factors, such as light, have cyclic 24-hour variations in the form of day-night cycles. These cyclic environmental changes can impose time-dependent restrictions and trade-offs on an animal's physiology and behavior. Animals have evolved an internal timekeeping system to cope with these daily environmental changes, which involves the circadian clock. The circadian clock generates a variety of physiological, metabolic, and behavioral daily rhythms — such as hormonal rhythms, body temperature rhythms, and activity rhythms (Moore-Ede, Sulzman, and Fuller 1982). Rhythms driven by the circadian clock, and thus generated endogenously, are referred to as circadian rhythms (i.e., circa 24-hour). One main characteristic of the circadian clock is synchronization by daily environment cycles, acting as an internal timekeeper and maintaining a stable temporal relationship between the internal physiology and the external environmental cycle. The presence of circadian clocks is thought to enable organisms to anticipate daily environmental challenges (Jabbur, Zhao, and Johnson

2021; Pittendrigh 1993), so changes in physiology and behavior occur in anticipation of predictable environmental changes. The reactive response to environmental factors, not driven by the circadian clock, is also important in shaping activity rhythms. Animals are constantly interacting with multiple environmental cycles or non-cyclic challenges and, not surprisingly, can also respond acutely to these external stimuli by activity inhibition or stimulation. In nocturnal mammals, for example, light has both effects; the light-dark cycle synchronizes the circadian clock while light itself acutely inhibits the animals' activity. Consequently, in nature, the observed daily rhythm, such as the activity rhythm, is an outcome of the interaction of these two mechanisms: the synchronized circadian clock, driving endogenous rhythms, and the direct response to environmental stimulus.

Circadian clock and biological rhythms studies have been majorly conducted in laboratory conditions, which is a powerful approach that enabled the characterization of the clock and its mechanism of synchronization. Studies on circadian rhythms relied heavily on manipulating the light-dark cycle and controlling for other variables to investigate synchronization. However, there are limitations to laboratory approaches, especially when we want to extrapolate what we learned in the laboratory to an ecological context (Enright 1970; Halle and Stenseth 2000; Helm et al. 2017). In nature, animals are exposed to multiple cyclic and non-cyclic environmental challenges, including biotic factors such as predation, competition, social interaction, and food availability; and abiotic factors such as temperature, humidity, and light. Consequently, the observed daily rhythm in nature has significantly more influence from direct responses to external stimuli than in the laboratory (Hut et al. 2012; Kronfeld-Schor and Dayan 2003). For example, interspecific competition between *Dypomys microps* and *Dypomys merriami* appears to drive activity of *D. merriami* to the last part of the night (Kenagy 1973). Another more extreme example is the golden spiny mouse, *Acomys russatus*. This species completely changes its activity pattern from nocturnal to diurnal in areas where the common spiny mouse, *Acomys cahirinus*, is absent (Kronfeld-Schor and Dayan 2003; Shkolnik 1971). A second aspect that hinders the understanding of

the significance of circadian rhythms in nature is that the behavior and physiology of laboratory animals might be unrepresentative of those of wild animals (Calisi and Bentley 2009). A growing number of rodents have completely different activity patterns between laboratory and nature. Some species exhibit nocturnal running wheel activity in the laboratory, but field observations reveal diurnal activity in nature (Hut et al. 2012). There are also cases where the same animal transferred from the field to the laboratory completely changes its activity pattern, as is also the case for *Acomys russatus*. The reasons for this dramatic change in activity patterns are still unknown. Still, it is hypothesized that changes to a more diurnal activity pattern might have energetic advantages, helping reduce thermoregulatory costs (Hut et al. 2012). There has also been a lot of effort in advancing the studies of chronobiology in the wild to understand better the ecology and adaptive significance of circadian rhythms (Dominoni et al. 2017; Helm et al. 2017; Kronfeld-Schor, Bloch, and Schwartz 2013). Overall, these examples illustrate that it is hard to translate some aspects of chronobiology from the lab to the wild and that more field studies are still needed to better disentangle the influences and relevance of the endogenous clock and the direct responses to external stimulus in natural conditions.

Daily activity rhythms studies require a continuous recording of individual activity for a prolonged time; thus, it renders studies in the wild a practical and technological challenge (Silvério and Tachinardi 2020). Recording individual fine-scale activity data on some species of animals is hard, especially because some species are difficult to capture, observe or record for extended periods. More recently, however, advancements and miniaturization of animal-borne sensors allowed researchers to investigate fine-scale activity and physiology in free-living animals (Rutz and Hays 2009; Wilmers et al. 2015). These sensors, also known as *biologgers*, have enabled the remote fine-scale measurements of animal activity with relatively low disturbance. Two kinds of biologgers widely used in ecological and, more recently, in chronobiological studies are lightloggers and accelerometers. Lightloggers, as the names imply, record the light exposure of animals; they are often used in migration studies when the timing of light

exposure is of importance. Accelerometers, in turn, are devices that record animal movement in three dimensions, which can be used to calculate and quantify general activity, energy expenditure, and estimate movement paths (Brown et al. 2013). Albeit fine-scale time-series data from biologists is ecologically richer, it also requires more complex methods and techniques to be analyzed (Brown et al. 2013). One analytical tool used to deal with such data type is the Hidden Markov Model (HMM), which can be used, for example, to model the dynamics between rest and active states (Brett T. McClintock et al. 2020). All in all, the possibility of recording individual fine-scale data and the current statistical methods available opens a lot of opportunities to study activity rhythms in free-living animals.

The Anillaco tuco-tuco (*Ctenomys* aff. *knighti*) is an example of rodent species that has been studied in the laboratory and semi-natural enclosures and exhibit distinct activity patterns between these conditions. The genus *Ctenomys* is endemic to South America, distributed in diverse habitats, from deserts to high altitudes (de Freitas, Gonçalves, and Maestri 2021). Species from this genus are subterranean and spend most of their time inside their tunnels, going out to forage and remove soil from their tunnel systems (de Freitas, Gonçalves, and Maestri 2021). The Anillaco tuco-tuco occurs around the village of Anillaco, in the province of La Rioja, a semi-arid region in the northwest of Argentina. In the laboratory, standard chronobiological experiments showed that their running wheel activity rhythm is driven by the circadian clock and synchronized by the light-dark cycle (Tachinardi et al. 2014; Valentinuzzi et al. 2009). In these controlled settings, tuco-tucos are nocturnal, running on the wheel almost exclusively during the night (Valentinuzzi et al. 2009). However, in semi-natural enclosures, visual observations and automated recordings of light exposure and activity show that the Anillaco tuco-tuco emerges to the surface during the day (Jannetti et al. 2019; Flôres et al. 2016; Tomotani et al. 2012). Hence, it is diurnal in semi-natural enclosures, as opposed to what is recorded in the laboratory. In semi-natural enclosures, it has also been shown that surface emergence is correlated with environmental temperature, changing seasonally. In winter, tuco-tucos generally go out of their tunnels during the

middle of the day, in the warmest hours (Jannetti et al. 2019). In contrast, the hottest hours of midday are avoided in the summer, and surface emergence occurs mainly at the beginning and end of the day (Flôres et al. 2021). Although laboratory and semi-natural enclosures have been crucial to understanding the chronobiology and activity patterns of tuco-tucos, they do not replicate the whole complexity of natural habitats. Using available *biologgers* we can now record activity and surface emergence simultaneously in free-living tuco-tucos. Using such technologies in nature, we might shed light on the range of biotic and abiotic factors shaping tuco-tucos activity patterns in their complex natural habitat and validate the recorded activity patterns in semi-natural enclosures.

In the present work, our first aim is to develop a field methodology to study free-living tuco-tucos using *biologgers*. Our second aim is to describe the daily activity and surface emergence patterns in free-living Anillaco tuco-tucos using *biologgers*, additionally describing the annual changes in such patterns. To do this, we deployed animal-borne accelerometers and light loggers on 21 Anillaco tuco-tucos in different months of the year during 2019 and 2020. We used the collected data from these bio-loggers to describe tuco-tuco's daily activity and confirm whether they are diurnal in the field. We were also interested in establishing if tuco-tuco's daily activity patterns remain the same throughout the year. Lastly, we applied HMM to classify tuco-tucos activity and analyze any possible behavior change throughout the year.

2.2 Methods

2.2.1 Study Species

The studied *Ctenomys* population lacks a formal phylogenetic and taxonomic classification but there are some lines of evidence suggesting that the study area is occupied by a single unidentified species (Amaya et al. 2016). In other studies, the studied *Ctenomys'* species has been referred informally as the Anillaco tuco-tuco (Amaya et al. 2016) and as *Ctenomys* aff. *knighti* (Tomotani et al. 2012) or *Ctenomys* cf. *knighti* (Valentinuzzi et al. 2009).

2.2.2 Study Site

Fieldwork was conducted at a site located approximately 5km away from the village of Anillaco, in the province of La Rioja, Northwest Argentina. The study site (-66.95°, -028.80, 1325m, approximately 75m²; Figure 2.1) is located within the Monte Desert biome and surrounded by the Velasco mountain range. This area has little human disturbance and no artificial light source. The Monte Desert is characterized as open shrubland dominated by Zygophyllaceae (*Larrea cuneifolia* Cav., *Tricomaria usillo*), Fabaceae (*Prosopis torquata*, *Senna aphylla*) and Cactaceae (*Trichocereus* spp, *Tephrocactus* spp) (Abraham et al. 2009; Fracchia et al. 2011; Aranda-Rickert and Fracchia 2011). At the study site, a non-extensive survey of the plant community divided into three transects showed a dominance of the families Zygophyllaceae (*Larrea cuneifolia*, *Tricomaria usillo*), Poaceae (*Microchloa indica*, *Aristida mendocina*) and Fabaceae (*Zuccagnia punctata*) (Appendix Figure B.1). The climate is arid with marked daily and seasonal cycles in temperature and rainfall (Appendix Figure B.2). The monthly mean temperature ranges from 12°C in the winter months to 23°C in the summer months, with a clear difference in the daily temperature range (Abraham et al. 2009). The mean annual rainfall ranges from 145 to 380mm, concentrated almost exclusively in the summer months (Fracchia et al. 2011).

2.2.3 Animal Capture and Handling

A total of 30 tuco-tucos were part of the present study and received *biologging* collars. Trapping was conducted in four different campaigns to the study site. We made three campaigns in 2019 during March-April (Autumn), July (Winter), and October (Spring). A fourth campaign was made in February 2020 (Summer). A fifth campaign was planned to occur in May 2020 but was canceled due to the COVID-19 outbreak. Tuco-tucos were captured using a custom-made PVC pipe trap (35cm length, 10cm diameter) with a spring-loaded aluminum door at one end and a PVC-lid at the other. Before setting the traps, we scouted the study site for active tuco-tuco burrows. Active burrows could be identified by the presence of freshly excavated soil mounds at the burrow's

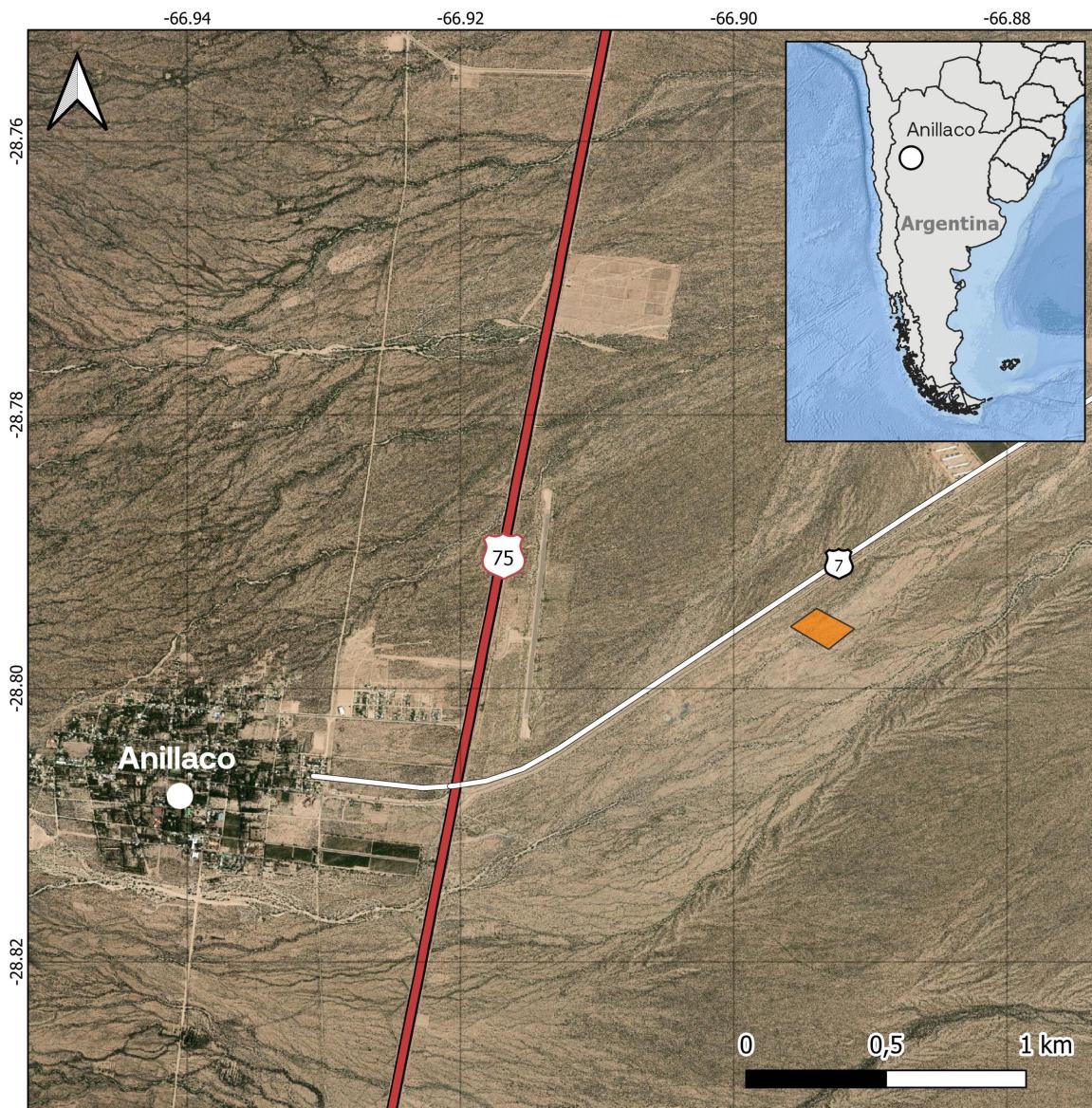


Figure 2.1: Study site location (orange polygon) at the Monte Desert, approximately 5km away from the village of Anillaco, northwest of Argentina. Study site has an area of approximately 75 m². Geographical information retrieved from ESRI satellites (Datum SIRGAS 2002 UTM 19s).

entrance. Once found, burrows were excavated to open the access to the underground tunnels, and a trap was placed horizontally at the burrow's entrance following the tunnel's orientation. We placed traps at all active burrows found at the study site, limited to a max of 20 traps available. Traps were set in the field during the morning and checked every 2 hours. Traps were checked for the last time at dusk and then taken out if no animal had been caught.

After capture, adult tuco-tucos (>120g) were lightly anesthetized to be carefully examined and received a biologging collar. We used a clear plastic anesthesia chamber (318.5cm³) with a clip-on lid and a cotton ball affixed inside the chamber, out of the animal's reach. The cotton ball received approximately 0.5 mL of isoflurane before transferring the animal from the trap to the chamber. While in the chamber, we observed animals for breathing, blinking, and loss of righting reflex. Once the tuco-tucos could not right themselves, we removed them from the chamber. Anesthetized animals were weighted (CSseries, OHAUS, ± 1 g precision), sexed, assessed for reproductive status, marked with a subcutaneous identification PITTag (Passive Integrative Transponder. Allflex, Brasil), and fitted with a collar bearing biologgers (See Biologgers).

We released animals in the same burrow they were initially captured. Tuco-tucos were left in the field for 5-18 days before being recaptured for collar recovery. All animal captures, procedures and animal handling were authorized by the local authorities at *Dirección General de Ambiente y Desarrollo Sustentable – Secretaría de Ambiente del Ministerio de Producción y Desarrollo Local – La Rioja, Argentina* (#00501-17). All procedures were also approved by the Ethics Committee at the *Instituto de Biociências* (#308-2018) and *Faculdade de Medicina Veterinária* (#2045300519) of the *Universidade de São Paulo*.

2.2.4 Biologgers

Accelerometers (Axy-4, TechnoSmart, Italy) and lightloggers (W65, Migrate Technology, UK) were used to record the general motor activity and light exposure, respectively. These biologgers were attached to a collar made of a cable tie inserted through silicon tubing [Jannetti et al. (2019); Williams et al. (2014); Figure 2.2]. A telemetry



Figure 2.2: Collar setup and example of field deployment. Upper photo shows the complete collar setup with accelerometer, lightlogger and the telemetry transmitter. Bottom photo shows a tuco-tuco wearing a collar. In the bottom photo it is possible to see the acceleromer and the lightlogger attached to the collar.

transmitter (SOM-2011. Wildlife Materials, USA) was also attached to the collar to assist animal with animal location during recapture and minimize sensor loss. The complete collar setup (accelerometer, lightlogger, and telemetry) weighted approximately 6g. Collars without the lightlogger weighted 5.3g. All accelerometers recorded tri-axial acceleration at a 10Hz sampling frequency and 4G sensitivity. Lightloggers were set to sample illuminance every minute but only registered the maximum sampled value every 5 minutes. The lightlogger recording range was 1-19000 lux.

2.2.5 Data Processing

Data were recorded on board of the sensors and later downloaded and converted to raw text files using software provided by the manufacturers. Acceleration data was used to measure general motor activity. Tri-axial acceleration data was first reduced to one dimension using the Vectorial Dynamic Body Acceleration (VeDBA, (Qasem et al. 2012)). VeDBA, measured in gravitational g, is commonly used as a proxy for the animal's activity level and energy expenditure (Williams et al. 2016). VeDBA was calculated by: (i) Estimating the effect of the gravitational force over the accelerometer, also known as static acceleration. We can estimate the static acceleration by applying a moving average over the raw acceleration data. There is no consensus over the number of points used to calculate the moving average, which can depend on the study species and device's recording frequency. In this study, we used a 4-second moving average after following the methodology proposed by Shepard et al. (2008) (Appendix D.1). (ii) Calculating the dynamic acceleration corresponding to the animal's movement in each axis. We calculated the dynamic acceleration by subtracting the static acceleration from the raw data. (iii) Lastly, we calculate the VeDBA by the vectorial sum of the dynamic acceleration over the device's axes (1.1). Once VeDBA was calculated, we downsampled the 1Hz acceleration data by taking the median over a 1-minute non-overlapping sliding window.

$$VeDBA = \sqrt{X^2 + Y^2 + Z^2} \quad (2.1)$$

We used light exposure to analyze patterns of surface emergence, the time tuco-tucos spend on the surface and further classify VeDBA data points as above or below ground. We classified each 5-minutes data point as above or below ground using a 2 lux threshold (Jannetti et al. 2019).

We merged accelerometer and lightlogger data according to the date and times of recordings using purposely written R scripts (R Core Team 2020). Because we did not synchronize recording time between both devices to the minute, lightlogger recording times were rounded to the nearest 5 minutes to merge both data streams.

We classified animal's data points as occurring during the day or night based on the day length of the recording dates. We calculated day length with the *maptools* package in R (Bivand and Lewin-Koh 2020), which uses the National Oceanic and Atmospheric Administration (NOAA) equations for estimating twilight times. We used civil twilight times, defined as the times in which the sun's center is 6° below the horizon, as thresholds to calculate day length and classify datapoints as occurring during the day or night time. Annual day length variation at the study site can be seen in the Appendix (Appendix Figure B.3). Daytime and nighttime intervals were defined from the estimated twilights for each day, using the same method.

To exclude any effects that capture and recapture can have on the animal's activity, we removed all datasets' first and last days. We also excluded the data corresponding to the days we were attempting to recapture the animal in cases where the recapture attempts took longer than one day. Animals that had data excluded due to recapturing efforts were FEV05 (5 days), JUL16 (5 days), and JUL23 (2 days).

2.2.6 Surface Emergence & Time On Surface

We visualized the daily temporal pattern of surface emergence using the 5-minute aboveground data (see Data Processing). We visually analyzed the temporal pattern of surface emergence by calculating the mean hourly number of surface emergences, which was calculated by binning the surface emergence data into 1-hour intervals for each animal and then calculating the mean number of emergences for each time interval, across all registered days for each animal.

To analyze tuco-tuco's daily time on surface, we calculated the mean time on surface per animal and the percentage of time on surface relative to daytime. Time on surface was calculated presuming that tuco-tucos spent each 5-minute data point classified as aboveground entirely outside the animal's burrow. Accordingly, we deduced the daily time on surface by summing the number of daily aboveground data points multiplied by 5 minutes and then calculating the mean daily time on surface per animal. In the same manner, we also calculated the percentage of the daytime spent on the surface

per day and then calculated the mean percentage per animal across all registered days.

2.2.7 General Activity

General activity was analyzed using the calculated VeDBA (see Data Processing). We visually analyzed the general activity temporal patterns by calculating the mean hourly VeDBA. The mean hourly VeDBA was calculated by summing the hourly VeDBA for each animal and day of recording. Next, we calculated the mean of the VeDBA sum for each 1-hour bin.

Daily VeDBA levels were analyzed relative to the daily mean VeDBA and the percentage of VeDBA during the daytime. Daily mean VeDBA was calculated by: (1) grouping the data per animal and summing the total VeDBA of each day and; (2) for each animal taking the mean of the calculated total VeDBA per day. The mean percentage of VeDBA during the daytime was calculated in the same manner, but we divided the daily sum of VeDBA during the daytime by the total sum of VeDBA and then calculated the mean per animal.

2.2.8 Behavioral State Classification

Hidden Markov Models

We used Hidden Markov Models (HMMs) to analyze further and classify the 1-minute VeDBA data. HMMs are a type of time series model; therefore, they take into account the temporal dependency of the observations (Leos-Barajas et al. 2017). HMMs are also well suited to model accelerometer data given their intrinsic temporal dependence (Leos-Barajas et al. 2017; A. Patterson et al. 2019).

HMMs are composed of two time-series: the observable *state-dependent process* (X_t), VeDBA in our case, and an underlying, or hidden, *state process* (S_t). The *state process* drives the observations and what we are interested in estimating, which roughly corresponds to behavioral states.

The *state process* follows the Markov Property and takes temporal dependency into

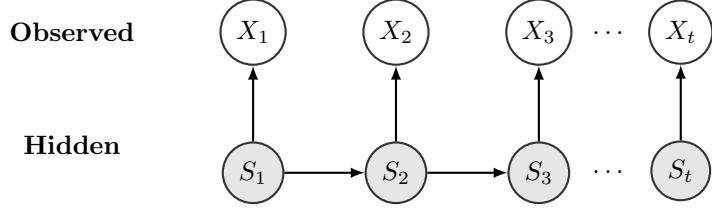


Figure 2.3: Basic dependence structure for a Hidden Markov Model. Hidden states sequence, or the state-process, is indicated by S . Observations, or the state-dependent process, are denoted by X .

account (Zucchini, Iain MacDonald, and Roland Langrock 2016). The Markov property denotes that a state S_t depends only on the previous state S_{t-1} (Zucchini, Iain MacDonald, and Roland Langrock 2016). In the case of accelerometer and animal movement studies, the states are representations of the animals' behavior and can take on finite numbers (N) of possible values. The number of states can be chosen *a priori* or based on model selection (Pohle et al. 2017). The changes in probabilities between states are also part of the HMM formulation, summarized by a Transition Probability Matrix that gives the probability of transitioning from the current state to a possible future state.

In the basic HMM formulation, the observable *state-dependent process* comes from a mixture of N distributions, one for each state. These distributions come from a common distribution family (e.g., Normal, Weibull, or Gamma), and each one has its own set of parameter values. The active distribution is determined by the system's state at a given time t . Therefore, the observations are a realization from one of these distributions. The distribution parameters, state transition probabilities, and other model parameters can be estimated by numerical maximization of the Likelihood (Zucchini, Iain MacDonald, and Roland Langrock 2016). With the model parameters in hand, the most probable state sequence can be found by the Viterbi algorithm (Brett T. McClintock et al. 2020; Zucchini, Iain MacDonald, and Roland Langrock 2016).

Model Formulation and State Classification

In our models, we have chosen VeDBA as our activity metric. We determined *a priori* a possible number of three different states ($N = 3$). This decision was made based on our research question, in the VeDBA distributions (Appendix D) and in the states' biological interpretability. We labeled the states as roughly corresponding to low, medium, and

high-intensity activity.

HMMs can be fitted individually (e.g., van de Kerk et al. 2015) or to a pool of animals (Langrock et al. 2012). The models can also include covariate effects that modify either the *state-dependent* distribution parameters or the transition probabilities (T. A. Patterson et al. 2009; Langrock et al. 2012). Using a “complete pooling” approach, we fitted a 3-state HMM to the 1-minute VeDBA data. Using complete pooling in our case means that the *state-dependent* distribution parameters are common to all animals. Therefore, we assume that individuals are independent, but behaviors are the same for all individuals and across the year. However, given that the season/month of the year seems to be an important feature influencing the VeDBA distribution (Appendix D), we included month as a covariate in the *state process*. Hence, we let the probability of changing from one state to another vary with the season/month of the year. We also fitted an empty model with no covariate effects and used Akaike’s Information Criteria (AIC) to select the model with the best fit to the data.

Models were fitted using the *momentuHMM* package in R (Brett T. McClintock and Michelot 2021). We used the gamma distribution, parametrized with mean and standard deviation, to model VeDBA. The gamma distribution is a flexible distribution that accommodates positive right-skewed data. Appropriate starting values for the likelihood maximization of the model’s parameters were found following procedures suggested by Michelot and Langrock (2019). Season was included as a categorical variable; its influence over the transition probabilities was summarized using stationary probabilities plots (Leos-Barajas et al. 2017). The most probable state sequence was decoded using the Viterbi algorithm (Zucchini, Iain MacDonald, and Roland Langrock 2016). We checked model assumptions and goodness of fit by visual inspection of the pseudo-residuals (Zucchini, Iain MacDonald, and Roland Langrock 2016). The decoded sequence was used to conduct other *post-hoc* analysis of time spent in each state, aboveground activity, diurnality and rhythmicity.

2.2.9 Daily Time-Activity Budgets & Behavioral States Patterns

We used the labeled behavioral state data to visually analyze how the behaviors are organized in time. We visualize the temporal patterns in behavioral states using the same methods we used in the surface emergence patterns. We visualized the temporal pattern in each behavioral state by using a Von Mises circular kernel density estimate (Rowcliffe et al. 2014). The kernel density estimate was done at the population level, using the pooled data from all animals of each month of the year.

To analyze tuco-tuco's behavioral states, we calculated their mean daily time and the mean daily proportion spent in each behavioral state (i.e., time-activity budget). Similar to what was done in the general activity, we summarized and reduced the VeDBA data to one point estimate per animal. The mean daily time in a state was calculated by first grouping the data per animal and calculating the daily time spent in each state by summing the labeled data points of each state. Since the data has 1-minute intervals, we take that the number of labeled points per day corresponds to the time spent in each behavioral state. Then, we calculated the mean daily time in each state per animal by taking the mean of the previously calculated daily time in each state. We also calculated the time-activity budget per animal in the same manner, except we divided the time spent in each state by 1440 minutes.

2.2.10 Aboveground Activity

We calculated the prevalence of each behavioral state when animals were aboveground, outside of their tunnels. The aboveground activity was defined as any activity in which animals were exposed to light with intensity greater than 2 lux (see Data Processing). Using the combined state-labeled and light logger data, we calculated the total time spent in each state on the surface and the percentage relative to the total time spent aboveground. Similar to the procedures used in the general activity analysis, we also calculated only one point estimate per animal. The mean time spent in each state was calculated by first summing the time spent in each state per day and then taking the mean per animal. We calculated the percentage of time spent in each state in the same

manner by calculating the percentage of time spent in each state relative to the total time aboveground and then taking the mean of these percentages per animal.

2.2.11 Diurnality Index

We defined diurnality index (DI) as the percentage of daytime the animals spent in one of the states relative to the total time spent in the same state during both daytime and night-time, corrected by the daylength of each month (Halle and Stenseth 2000; Jannetti et al. 2019). The DI ranges from 0 to 1, with 0 meaning that all activity happens during the night and 1 meaning the opposite, that all activity happens during the day.

Diurnality calculation is shown in equation (2.2), where ts_{day} and ts_{night} are the time spent in the state during the day and night respectively. L_{day} and L_{night} are the day length and night length, determined by the civil twilight, calculated using the *maptools* package in R (Bivand and Lewin-Koh 2020).

$$Diurnality = \frac{ts_{day}/L_{day}}{ts_{day}/L_{day} + ts_{night}/L_{night}} \quad (2.2)$$

2.2.12 Circadian Rhythmicity and Period Estimation

We used autocorrelation analysis (Levine et al. 2002; Dowse 2009) and Lomb-Scargle periodograms to assess the robustness and periodicity of activity rhythms. The autocorrelation was calculated by comparing the data to itself lagged by a unit of time. The autocorrelation coefficient ranges from 0 to 1, and it is higher as both time series are more similar. When visually analyzing the autocorrelation plot, recurring peaks indicate periodic data. The height of the peak shows how robust the rhythms are (Dowse 2009). The rhythm's robustness, also referred to as the Rhythmicity Index (RI), is defined as the autocorrelation coefficient at the third peak of the autocorrelation plot (i.e., the height of the third peak).

Before estimating the RI, we applied a 3-hour low-pass Butterworth filter to remove

periodicity lower than 3 hours in the data. Autocorrelation plots were first visually analyzed and labeled as rhythmic if they showed recurring peaks in the 24-hour range. We classified animals with no recurring peaks as arrhythmic in the circadian range. Next, we estimated each behavioral state's period for animals classified as rhythmic using the Lomb-Scargle periodogram (Leise 2017). For comparison with the labeled data, we also calculated the RI for the unlabeled VeDBA data

All analyses were done in R (R Core Team 2020). Butterworth filtering was done using the *dprR* package (Bunn 2008). We did autocorrelation function, and plots in base R. The peaks in the autocorrelation plots were found using the *pracma* package (Hans W. Borchers 2019). Lomb-Scargle periodograms were calculated using the *lomb* package (Ruf 1999).

2.2.13 Statistical tests

We used ANOVA and post-hoc Tukey's HSD to test for differences between month-groups in time on surface, general activity levels, time spent on behavioral states, time spent on aboveground activity, and diurnality. Rhythmicity was compared only between behavioral states; we did not perform a month-group comparison given that, as we classified some animals as arrhythmic, the sample number for each month-group was too low to perform a meaningful statistical analysis. We checked for normality before conducting the ANOVA tests by visual plots and with Shapiro-Wilk test.

We also visually compared daily patterns of general activity levels, time on surface, and behavioral states based on the temporal distributions along the 24-hours. We visualized the daily temporal distribution for general activity and surface emergence patterns by calculating the hourly mean. On the other hand, we visualized behavioral states' daily patterns using Von Mises circular kernel density estimates. All analyses were done in R using the base packages (R Core Team 2020).

2.3 Results

We captured and deployed collars to 20 females and 10 males; 13 collars had both sensors. We were able to recapture 24 tuco-tucos and recover 21 collars (Table 2.1), a total of 70% of success rate in collar recovery. One collar was found malfunctioning because one animal got predated. The other two lost collars fell or were taken out of the tuco-tuco's neck between the time of capture and recapture. A complete list of the number of recorded days per animal can be seen in the Appendix Table A.1.

Table 2.1: Number of captured animals and sensors deployed in the field. There was a higher number of females captured independent of the season. Not all recaptured tucos still had their collars. Some collar were taken out by the animals between the time of captured and recaptured. One tuco was predated and the collar was found 1km away from the initial capture burrow malfunctioning.

	Captured		Recaptured		Recovered Collars	Accelerometers	Lightloggers
	Males	Females	Males	Females			
February (Summer)	3	7	2	5	5	5	5
July (Winter)	4	5	4	5	8	8	6
March (Autumn)	0	2	0	2	2	2	0
October (Spring)	3	6	1	5	6	6	2

2.3.1 Time On Surface

Surface emergence shows a changing daily temporal pattern along the year. In July, the peak of surface emergence is concentrated in the middle of the day. In other months the peak of surface emergence shifts to the first hours of daylight (Figure 2.4A). Individual surface emergence actograms can be seen in appendix C.1.

The total time on surface shows no difference along the year. The overall mean and standard deviation of daily time on surface was 71.98 ± 29.21 , with no significant differences among month groups (ANOVA, $F = 2.148$, $p = 0.167$). In contrast, the daily percentage of the time on surface relative to day length was significantly different between groups (ANOVA; $F = 4.38$, $p = 0.0429$). Post hoc Tukey's HSD shows a significant difference between July-February ($p = 0.035$). For these groups the mean daily percentage of time on surface is $13\% \pm 4\%$ for July, and $6\% \pm 4\%$ for February.

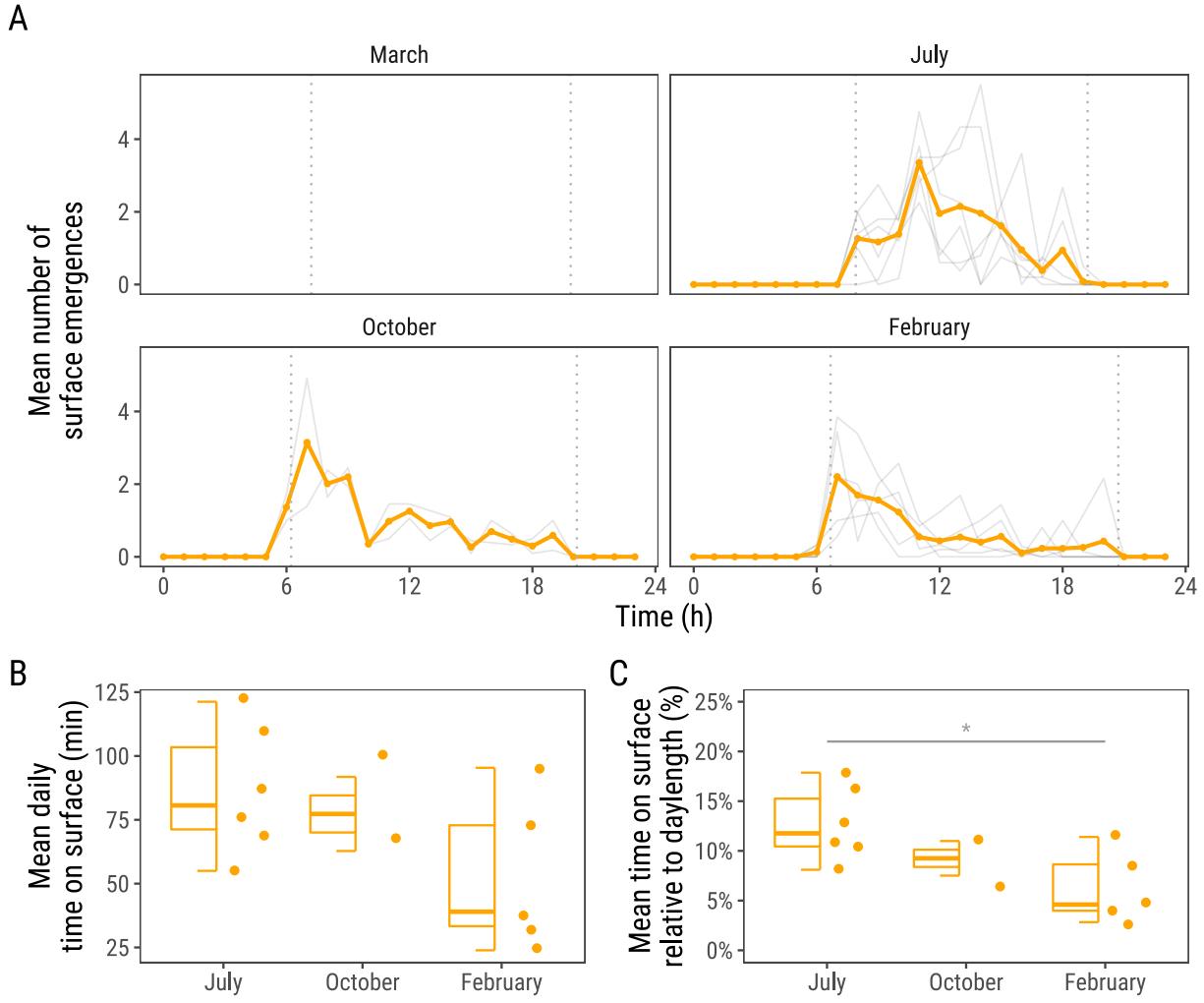


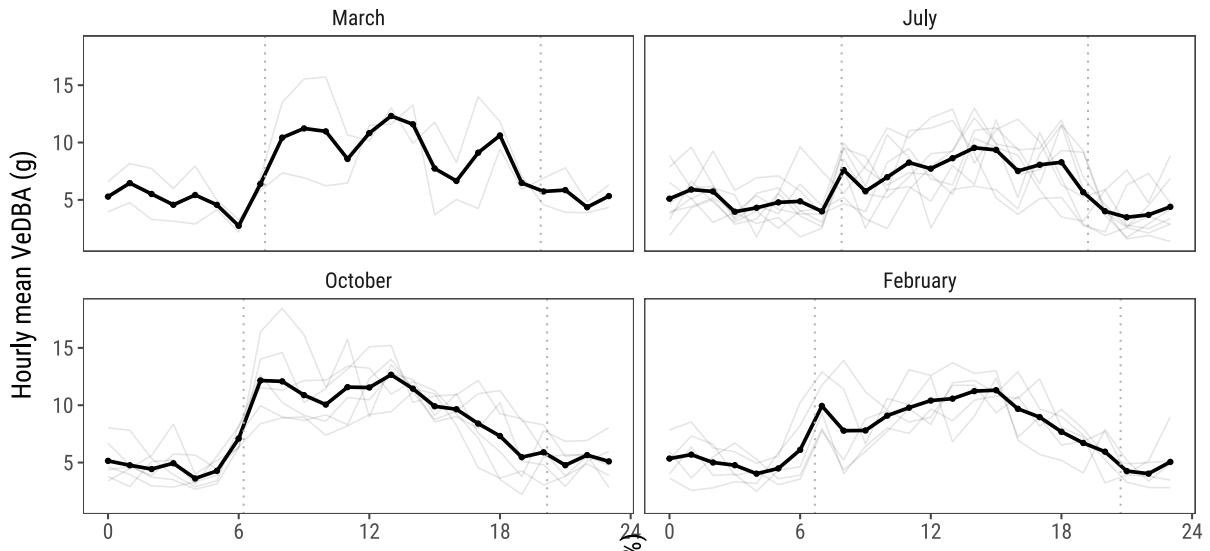
Figure 2.4: Tuco-tuco's daily patterns of surface emergence and average time on surface. (A) Daily temporal pattern of surface emergence. X-axis is time of the day in Anillaco, La Rioja (UTC-3). Light grey lines show individual temporal pattern of surface emergence. Orange line shows the mean of all animals. Dotted vertical lines show times of civil twilights. (B) Mean daily time on surface per animal. Each point is an animal's mean daily time on surface on registered days (ANOVA; $p = 0.167$). (C) Mean daily percentage of time on surface in relation to the daylength. Each point is an animal's mean daily percentage of time on surface across registered days (ANOVA; $p = 0.043$). Asterisks indicates pairwise significant statistical difference (Tukey's HSD; $p < 0.05$).

2.3.2 General Activity

Tuco-tuco's general activity follows a typical diurnal pattern. Hourly mean VeDBA is lower during the night time, rises just before dawn, remains generally higher during the daytime and falls just after dusk. The average daily VeDBA is significantly different among month-groups (ANOVA; $F = 7.182$, $p = 0.002$; Figure 2.5). There is a significant month-group difference between July-October and July-February (Tukey's HSD, $p < 0.05$). In both pairwise comparisons July's average daily VeDBA is lower, with a difference in means of 0.029 g in comparison to October and 0.019 g in comparison to February. Individual surface emergence actograms can be seen in appendix C.2.

The percentage of daytime VeDBA is also significantly different between months (ANOVA; $F = 8.288$, $p = 0.001$; Figure 2.5). There is a significant difference between July-October and July-February (Tukey's HSD, $p < 0.05$), with a lower July VeDBA level and mean difference of 13%.

A



B

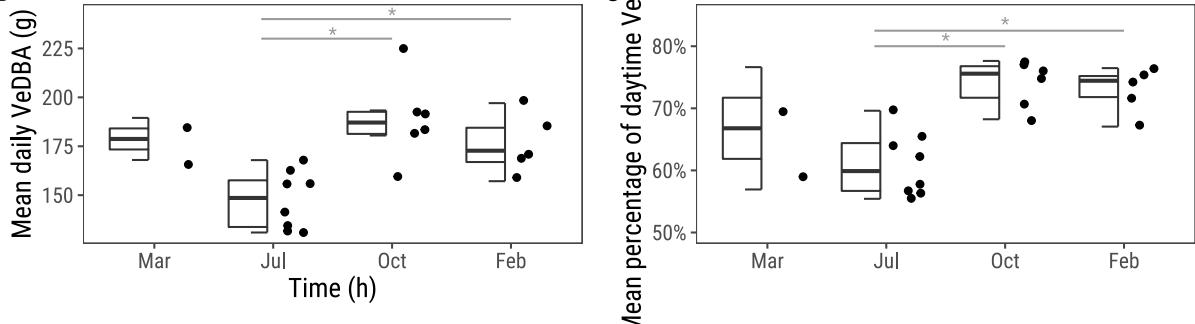


Figure 2.5: Tuco-tuco's daily temporal pattern of VeDBA and Mean daily VeDBA. (A) Daily temporal pattern of VeDBA binned by hour. Light gray horizontal lines show individual patterns. Black lines show hourly mean VeDBA of all animals. (B) Mean daily VeDBA. Each point is an animal's Mean daily VeDBA. There is a significant difference between month-groups (ANOVA, $p = 0.002$). In July tuco-tucos exhibited lower average daily VeDBA than October and February (Tukey's HSD; Jul-Oct = 0.002, Jul-Feb = 0.049). (C) Percentage of daytime VeDBA relative to total daily VeDBA. Points show mean daytime percentage of VeDBA in relation to the total VeDBA during the day. There is a significant group difference (ANOVA; $p = 0.013$). In July animals show lower VeDBA during the day than October and February (Tukey's HSD; Jul-Oct = 0.001, Jul-Feb = 0.006). Vertical dotted lines show time of civil twilights.

2.3.3 Behavioral State Classification

We fitted two different HMMs to VeDBA data, one empty model with no covariates and a second one with ‘*season*’ as a covariate in the transition probability matrix. The second model was selected based on informational criterion ($\Delta\text{AIC} > 2$; Appendix E).

The estimated state-dependent distributions are shown in Figure 2.6. We interpreted and labeled these states as low, medium, and high-intensity activity. The marginal distribution (Figure 2.6; dashed line) has good correspondence to the empirical VeDBA distribution. A visual analysis of the Pseudo-residuals (Appendix E) shows that the residuals deviate from the expected normal distribution, especially in the lower-end values. Still, there is significant residual autocorrelation. Nevertheless, the overall fitting seems to be reasonable. The estimated state-dependent parameters are shown in Appendix E.

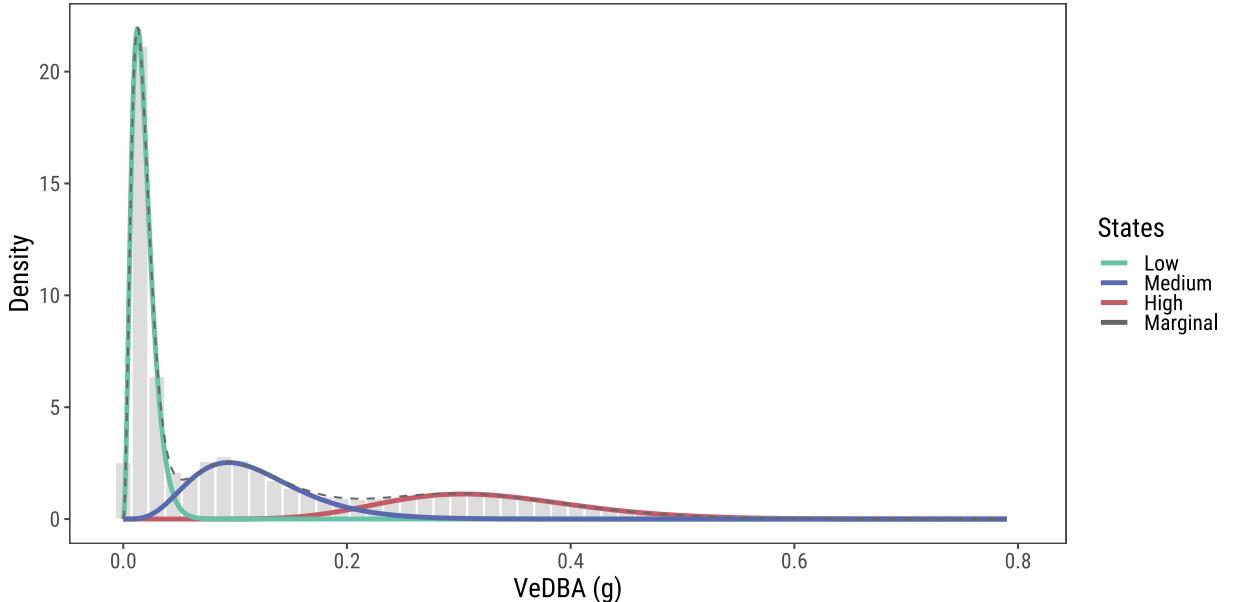


Figure 2.6: State-dependent distributions of the selected Hidden Markov model fitted to the VeDBA acceleration metric. Histogram, in grey, shows the empirical VeDBA from the pooled data of 21 Anillaco’s tuco-tuco. State-dependent gamma distributions are shown above the histograms. These distributions are weighted according to the proportion of observations assigned to each state.

With the state-labeled data, we could dissociate and visualize the daily patterns of each different state. An example of how VeDBA levels and the behavioral states are related is shown in Figure 2.7.

A visual inspection of all individual actograms (Appendix C) indicates that the high activity state rhythm is more robust than the Medium Activity rhythm. Nonetheless, neither of these behavioral states occurs exclusively during the daytime. Despite being more concentrated during the daylight hours, the high-intensity state also occurs sporadically during the night. Medium-intensity state, in turn, is spread throughout the day and night.

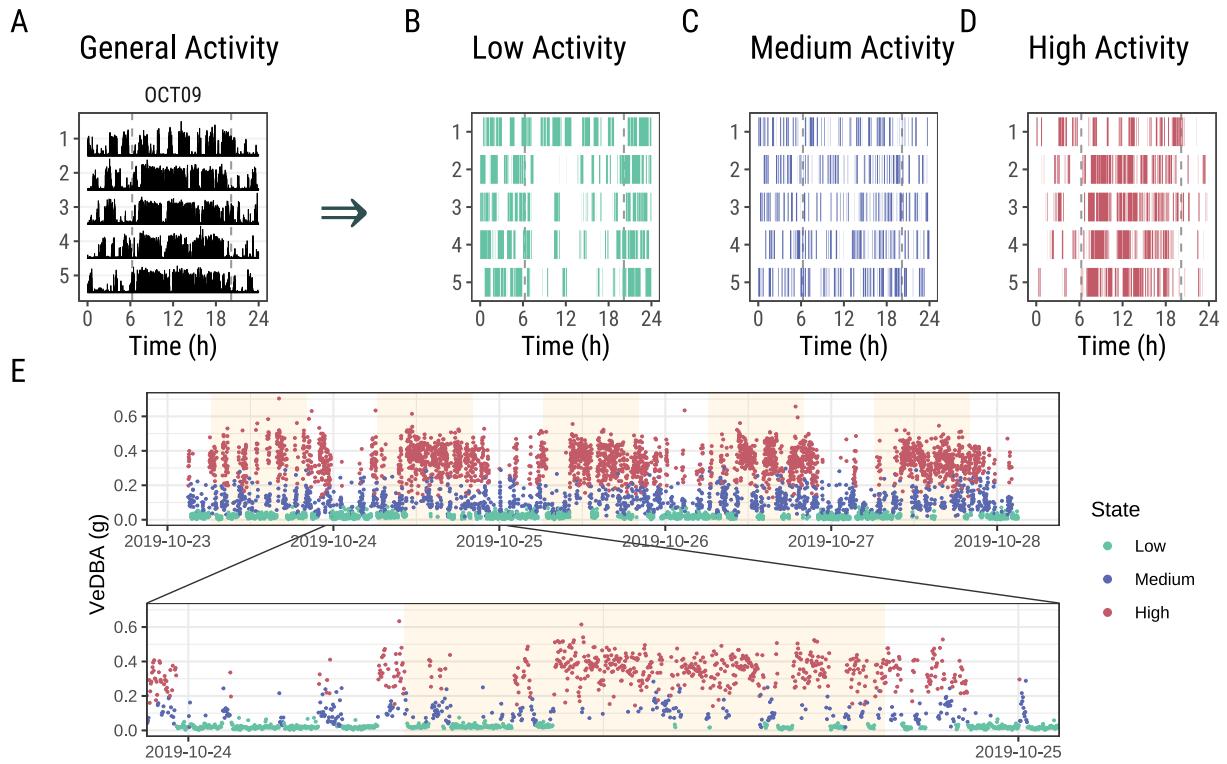


Figure 2.7: Actograms and time series plots of general activity, measured by VeDBA, and state-labelled data of a representative animal (#OCT09). The 5-day actograms show daily patterns of VeDBA (A) and of Low, Medium and High State occurrences (B, C and D) classified using HMM. X-axis represent time of day and each line on the Y-axis represents a day. Dashed lines show time of dawn and dusk. Medium Activity State shows no clear pattern of a daily rhythm, with episodes spread throughout the day. High Activity, on the other hand, has a higher concentration during daylight hours. The time series (D) shows 1-minute VeDBA measures colored by behavioral state. Light orange background shows daytime.

2.3.4 Daily Behavioral States Patterns

Daily activity patterns for each behavioral state are shown in Figure 2.8. These plots show that the timing of high-intensity activity follows a diurnal pattern. Medium-intensity activity, in contrast, is spread out along the 24h and does not follow a daily (24-hour) rhythm. It is important to note that the timing of peak occurrence of high-intensity activity does not appear to change dramatically along the year; in all the sampled Months, the peak of High Activity is around 14:00.

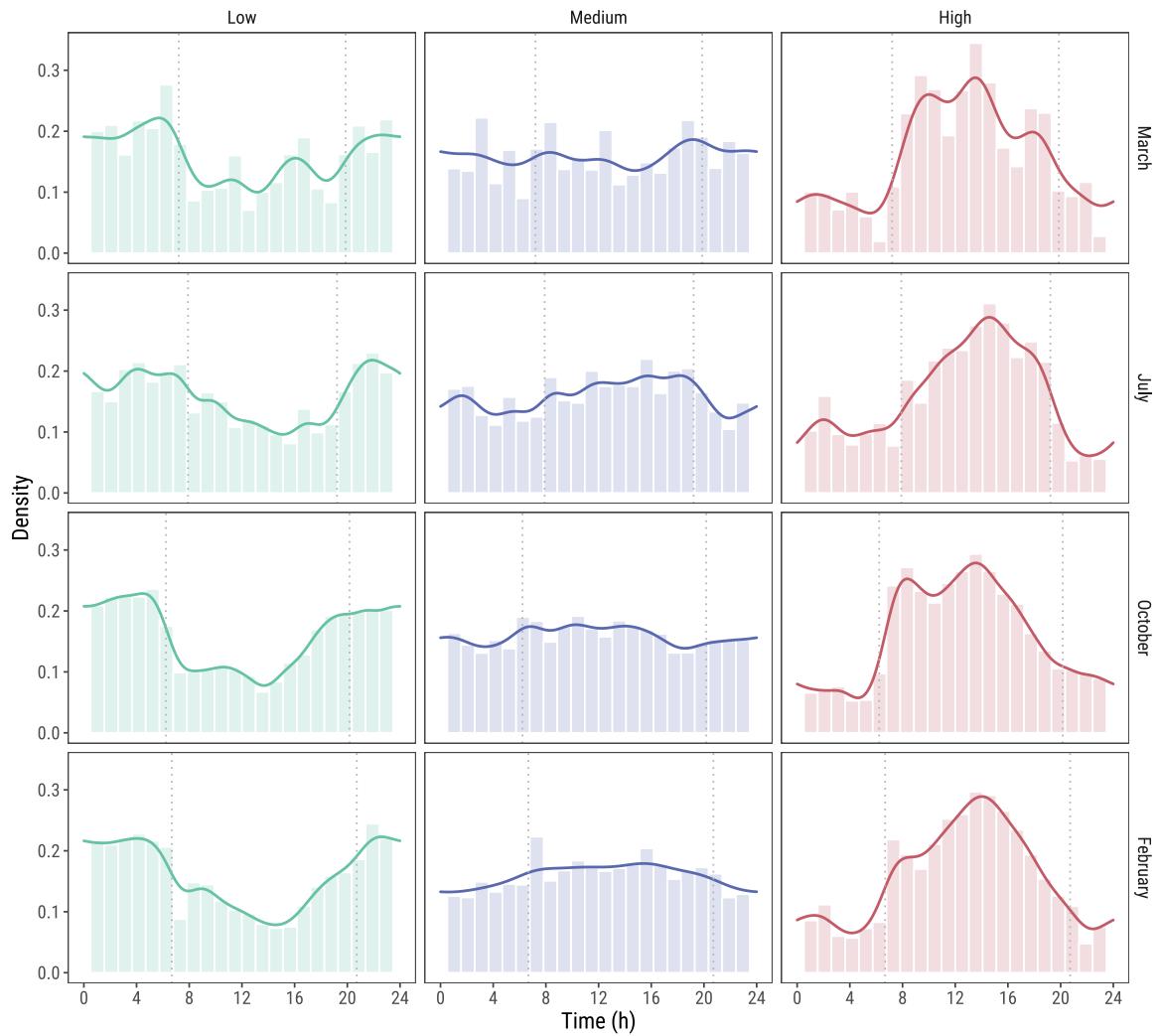


Figure 2.8: Density estimates of daily activity patterns of tuco-tucos' behavioral states. Solid lines indicate the Von-Mises circular kernel density estimates. Light-colored bars show observed distribution of each behavioral state occurrence. The x-axis is hour of the day in Anillaco, La Rioja (UTC-3). Dotted vertical lines show time of civil twilights. Visually, the low activity state is not exclusive of one phase of the day, occurring both during day and night, although, more concentrated during the night. Medium activity visually shows no clear daily rhythms, with medium activity occurring during day and night. High activity state shows a clear diurnal pattern independent of the time of the year, peaking at around 14:00.

2.3.5 Diurnality

High-intensity state is predominately diurnal ($DI > 0.5$). The average diurnality for the high-intensity state is higher than 0.7 for all month-groups. However, the medium-intensity state has a diurnality index that ranges from 0.5 in March to 0.56 in July and February. The low activity state is predominantly nocturnal, with Diurnality lower than 0.38 in all months. There is no statistical difference between months (ANOVA; $p > 0.05$ for all states; Figure 2.9).

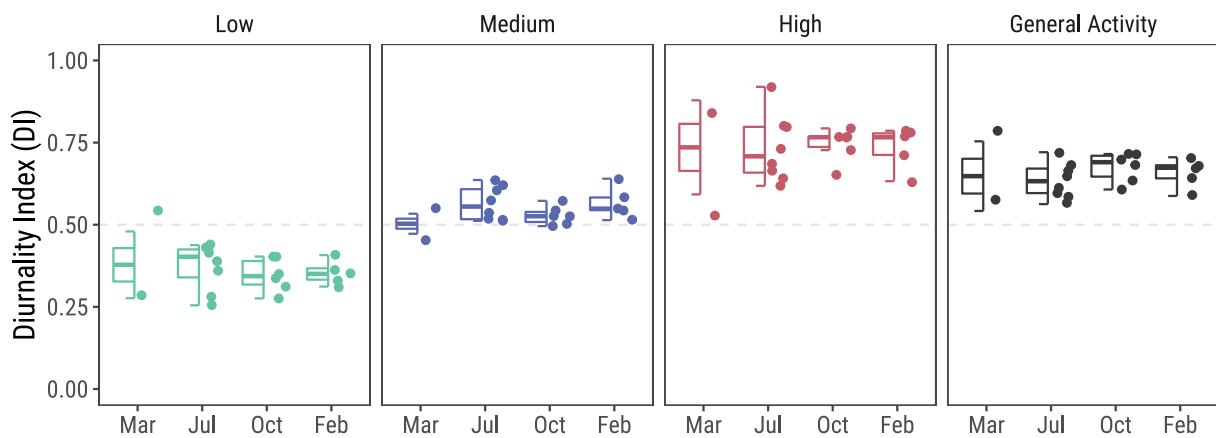


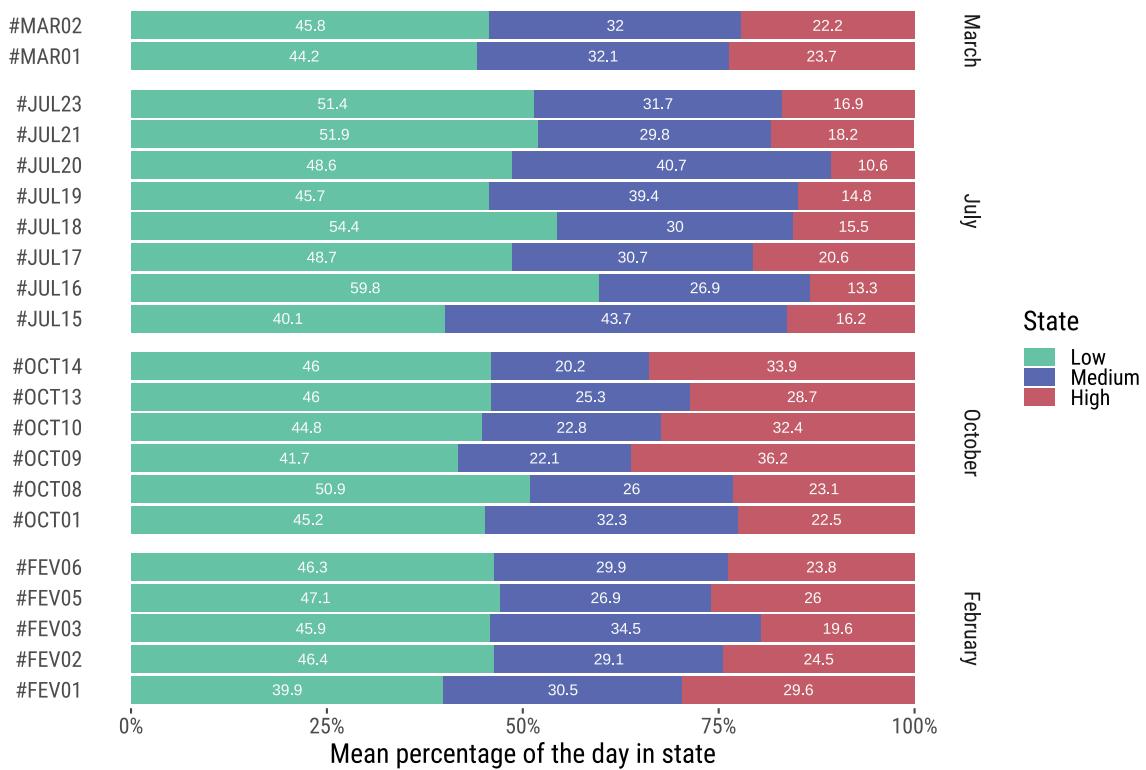
Figure 2.9: Distribution of calculated diurnality index (DI). Each point represents the DI of an animal. Values above the horizontal dashed line indicates that there is more activity during the day than during the night, while values below this line indicates that activity is more nocturnal. High Activity State is predominantly diurnal, with diurnality greater than 0.7 across all month-groups. General activity, measured by the unlabelled VeDBA, is also predominantly diurnal. Medium state is spread almost evenly between day and night. Low activity is concentrated during the night.

2.3.6 Daily Time-Activity Budgets

On average, tuco-tucos spent 680 ± 67 minutes per day in the low activity state, with no statistical difference between month-groups (ANOVA; $F = 1.93$, $p = 0.163$).

In contrast, there is a significant difference in the percentage of time spent in Medium and High Activity States (Medium: ANOVA, $F = 4.457$, $p = 0.0175$; High: ANOVA, $F = 13.62$, $p = < 0.001$). In the medium state there is a significant difference between July-October (Tukey's HSD, $p < 0.05$). In July, the mean time spent in the medium state was 491 ± 89.3 minutes, and in October, 357 ± 61.2 minutes. The high activity states show a significant difference between October-July and February-July (Tukey's HSD, $p < 0.01$). The daily mean time spent in high activity was: July, 227 ± 43.7 minutes; October, 424 ± 82 minutes; February, 355 ± 52.2 minutes.

A



B

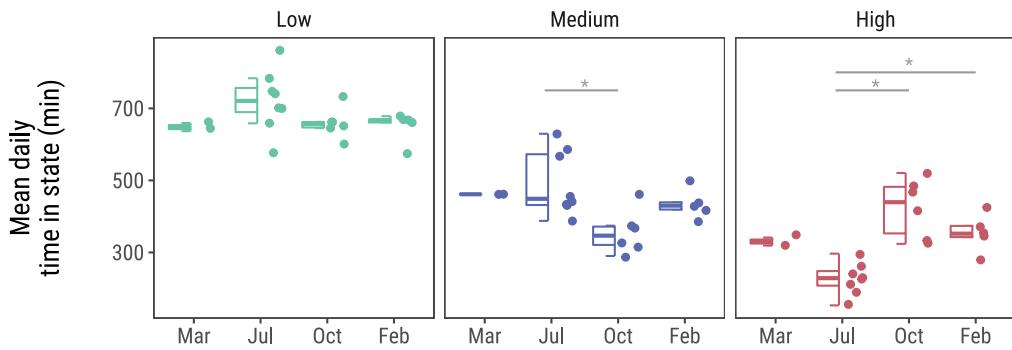
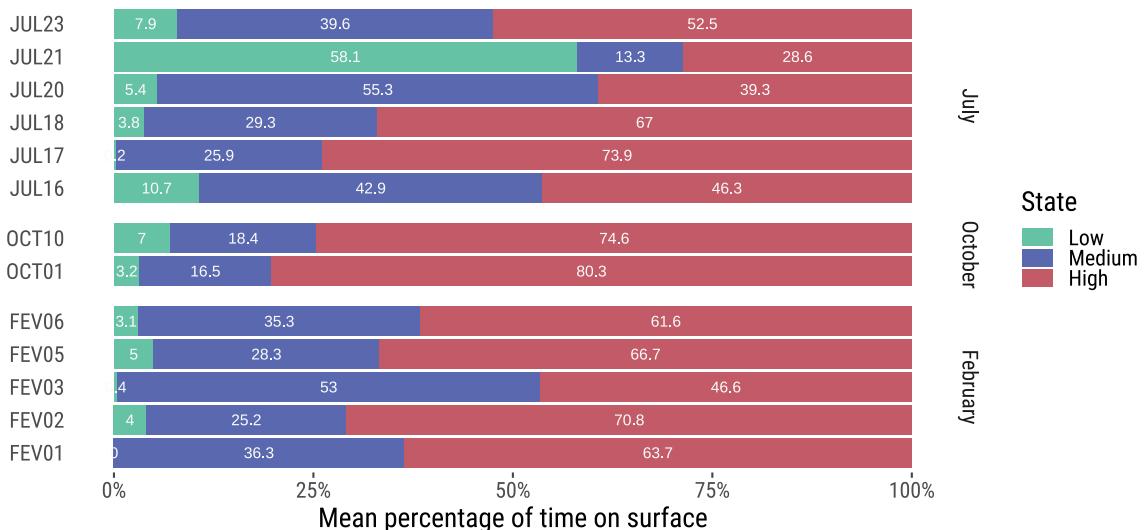


Figure 2.10: Time spent in each behavioral state and corresponding individual time-activity budgets. (A) Time-activity budget of the allocation of behavioral states per animal. (B) Mean daily time in each behavioral state per animal. Asterisks show statistical significance in pairwise comparison between Months. The mean percentage of time spent in the Medium Activity State, however, is higher in July in comparison with October (Tukey's HSD, $p < 0.05$). The mean percentage of time spent in the High Activity State is lower in July in comparison to October and February (Tukey's HSD, $p < 0.05$).

2.3.7 Aboveground Activity

Outside of their tunnels, when exposed to light, tuco-tucos are mostly in an active state, either high or medium state (Figure 2.11). There is no significant difference between month-groups within any state (Kruskal-Wallis, $p > 0.05$). The overall mean daily time on surface spent on the high activity state is 41.75 minutes, whether for the medium state it is 23.75 minutes. Low activity, in contrast, is the behavioral state that happens the least on the surface with a mean daily time of 6.39 minutes, considering all month-groups together.

A



B

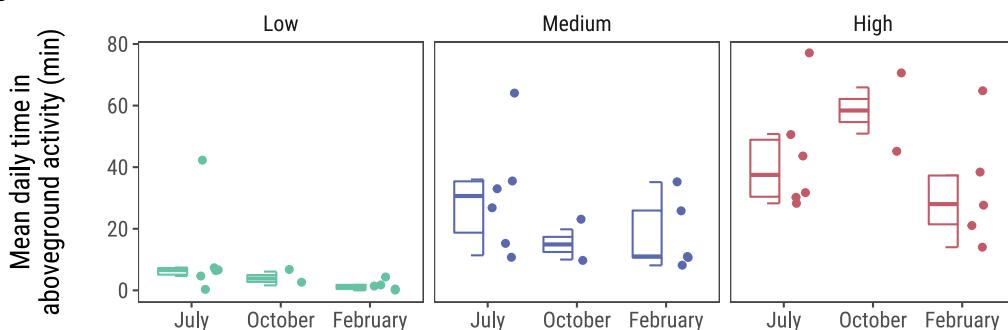


Figure 2.11: Mean time per animal and individual percentages of behavioral states exhibited on the surface. (A) Percentage of aboveground time spent in each behavioral state per animal in relation to the total time on surface. (B) Mean daily time of behavioral states exhibited aboveground per animal. There is no significant difference between month-groups within any state (Kruskal-Wallis, $p > 0.05$).

2.3.8 Circadian Rhythmicity

Circadian rhythms are present in all states, as seen in the visual analysis of the autocorrelation plots (Appendix F). The percentage of animals classified as rhythmic is higher for the high activity state, where a total of 18 animals were classified as rhythmic (85.7%). In contrast, we classified 11 animals (52.4%) as rhythmic for the medium activity state (Table 2.2). We calculated the RI only for animals classified as rhythmic

Table 2.2: Number and percentage of animals classified as rhythmic.

State	Animals	Rhythmic	Percentage
Low	21	16	76.2
Medium	21	11	52.4
High	21	18	85.7
General Activity	21	16	76.2

(Figure 2.12A), which is why there is a different number of samples between each category. Nonetheless, the Medium Activity State is significantly different from the other states and VeDBA (Figure 2.12A; Kruskal-Wallis; $p < 0.05$). We did not test for differences between month-groups (See Methods).

We used Lomb-Scargle periodograms to estimate the periodicity of each state (Figure 2.12B). All estimated periods, independent of state, were in the 24-hour range. We also estimated RI and periodicity for unlabelled VeDBA. In comparison with state-labelled data VeDBA analysis are more similar to the high activity state. The number of animals classified as rhythmic based on the autocorrelation plot is similar between VeDBA and high activity state data. The distribution of RI shows no difference between High state and VeDBA (Figure 2.12 A).

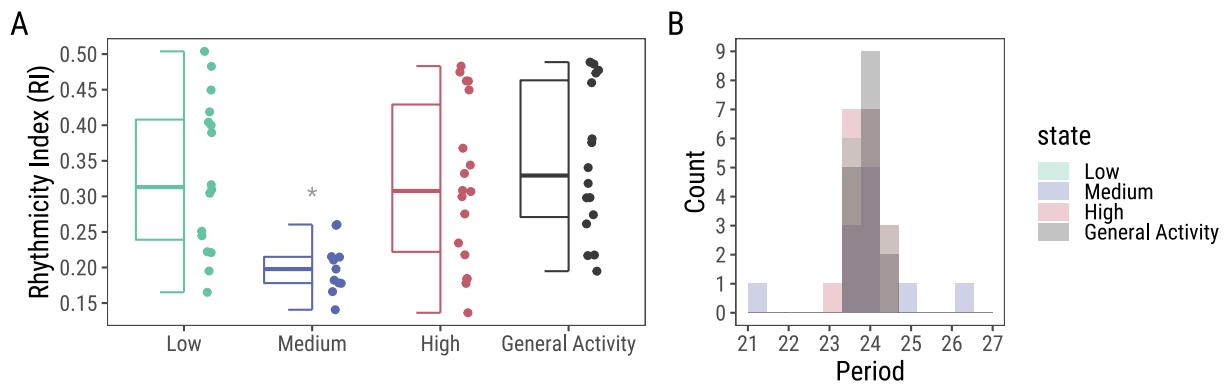


Figure 2.12: Rhythmicity metrics for estimated behavioral estates and unlabelled VeDBA. (A) Rhythmicity Index calculated per animal for state-labelled data and VeDBA. The distribution of the Medium State is statistically different from all other states and VeDBA (Kruskal-Wallis, $p > 0.05$). (B) Period estimation for behavioral states and VeDBA.

2.4 Discussion

In the present study, we aimed to establish a field methodology to use biologgers to assess daily activity rhythms in tuco-tucos and describe changes in these rhythms throughout the year. Regarding the field methodology, biologging studies in Anillaco's tuco-tucos are feasible and bear good recapture rates and recovery of the deployed sensors (Table 2.1). Our accelerometer data shows that tuco-tucos are diurnal, allocating the bulk of their movement during the daytime (Figure 2.5). In terms of the daily temporal pattern, the daily pattern of general activity was the same during all sampled months (Figure 2.5A), peaking at the middle of the day. Meanwhile, the temporal pattern of surface emergence had a clear difference along the year (Figure 2.4A). Using HMMs to classify the acceleration data, we found that high-intensity activities have a more robust diurnal pattern than the medium and low-intensity activities (Figure 2.8). Furthermore, time allocated in different behavioral states seems to change annually. In July, tuco-tucos decreased their time in high-intensity activities, replaced by medium-intensity activities (Figure 2.10). Taken together, the results show that free-living tuco-tucos are diurnal and display annual changes in total activity and time-activity budget.

The daily pattern of surface emergence showed changes throughout the year, even though the daily time on surface remained the same (Figure 2.4). The peak of surface emergences changed from the middle of daytime in July (Winter) to the first hours of the day in other months (Figure 2.4A). These pattern changes were also described in previous studies in semi-natural enclosures (Jannetti et al. 2019), where surface emergence patterns of tuco-tucos were different between summer and winter. It is proposed that part of these changes are endogenous due to seasonal synchronization to photoperiodic changes (Flôres et al. 2021). Additionally, environmental temperature also modulates these pattern changes due to thermoregulatory constraints (Jannetti et al. 2019; Flôres et al. 2021). Our results revealed that tuco-tucos, on average, spend the same amount of daily time on the surface in all months (Figure 2.4B). However, the percentage of time relative to day length showed changes between July and February (Figure 2.4C). This means that tuco-tuco extend or compress the range of their surface

time as the day length changes. Overall, we have confirmed that free-living Anillaco tuco-tucos exhibit changes in their daily surface emergence patterns and, additionally, that they do not change the total time on the surface throughout the year. Moreover, we showed that the surface emergence patterns in October are similar to what we see in February, which was not yet documented.

General activity patterns, differently from surface emergence, showed no changes in daily patterns throughout the year. In all months, general activity peaked around the middle of the day (Figure 2.5A). There was, however, a reduction in the total level of daily activity in July (Figure 2.5B), which might be related to the energetic challenges tuco-tucos face during winter (Tachinardi et al. 2017; Antinuchi et al. 2007; Jannetti et al. 2019). Two factors related to energetic constraints might play a role in this observed reduction in general activity: the environmental temperature and food. In July, the mean environmental temperature (Appendix Figure B.2A) is below the estimated thermo-neutral zone (TNZ: ~24°C-32°C) for the Anillaco tuco-tuco (Tachinardi et al. 2017). Additionally, July is also the month of the year with the lowest rainfall in Anillaco (Appendix Figure B.2B), which reduces primary productivity in the Monte Desert (Bisigato et al. 2009; Paruelo, Jobb'agy, and Sala 2001) and might affect tuco-tucos food quality and availability. Lower food quality, in particular, has been shown to reduce the resting metabolic rate in *Ctenomys talarum* (Antinuchi et al. 2007). Therefore, reducing activity might be energetically beneficial when facing challenging conditions, such as food reduction and lower temperatures. Interestingly, general activity patterns showed contrasting results to the ones in surface emergence. For the surface emergence, there was no difference in the amount of time spent on the surface but we did observe a change in its daily temporal pattern (Figure 2.4A). For general activity, on the other hand, we observed a reduction in total daily activity with no significant difference in daily temporal patterns (Figure 2.5). These results suggest that general activity and surface emergence might respond differently to environmental factors, or the interaction between direct response and the circadian clock might differ between these types of activity.

Using HMMs, we classified VeDBA into three distinct behavioral modes (low, medium, and high). It is essential to note that the states do not correspond directly to specific behaviors (e.g., feeding, foraging, or digging). However, they can be assumed to roughly correspond to behavioral states that can encompass a range of different behaviors (Leos-Barajas et al. 2017; Papastamatiou et al. 2018). Given that our approach is unsupervised, meaning that we did not match behavioral states with observations, we are limited in the biological interpretation of the classified states. The medium and high states are probably a good example of such limitation; despite knowing each state's intensity, we cannot pinpoint these states to any specific behavior. We believe, however, that the low-intensity state is majorly, but not exclusively, composed of resting behavior when the animal is generally in a low movement state. In general, the used HMMs had a good fit to our data. They allowed us to more thoughtfully analyze the observed reduction in daily general activity levels (Figure 2.5B). Due to the scope of this study, we opted for the simplest HMM model, using only season as a covariate. However, numerous extensions to HMMs might be interesting to answer other ecological questions Brett T. McClintock and Michelot (2021).

We analyzed the daily pattern and the time allocated for each state using the behavioral state's data. Our results show different daily activity patterns between states (Figure 2.8). Not surprisingly, the high-intensity state also showed a diurnal pattern, given the observed results in the general activity analysis. High-intensity activity had more than 70% of its occurrences during the daytime (Figure 2.8 and 2.9). Low-intensity, on the other hand, showed a nocturnal pattern, which is compatible with our interpretation of the low-intensity state as a resting state if we consider tuco-tucos to be diurnal (Figure 2.8 and 2.9). Unexpectedly, the medium-intensity state was distributed evenly between day and night (Figure 2.9), even though a few animals had a 24-hour rhythm in this state (Figure 2.12). Since we were only interested in the daily patterns in the present study, we did not test for periodicity in the ultradian range (periods much shorter than 24h). However, the observed pattern, lower rhythmicity index, and individual actograms (Appendix C) for the medium-intensity state might indicate an

ultradian rhythm, a behavior that is commonly observed in small rodents (Bloch et al. 2013; M. P. Gerkema et al. 1993; van Rosmalen and Hut 2021).

An advantage of using HMM to classify the VeDBA data is that it allowed us to extend our general activity analysis. The behavioral state data shows that despite animals displaying lower levels of general activity in July, they are not spending more time resting (Figure 2.5 and Figure 2.10). During July, tuco-tucos spent less time in a high-intensity state, substituted by more time in medium-intensity activities (Figure 2.10B). Although the meaning of such behavioral change is not clear, the reduction in high-intensity activity may also be related to a decrease in energy expenditure as discussed for general activity (Tachinardi et al. 2017; Antinuchi et al. 2007). Other than that, the increase in high-intensity activity in October and February (Figure 2.10B) might be related to tuco-tucos reproduction. Different species from the genus *Ctenomys* have a seasonal reproduction that generally occurs during Spring and Summer and sometimes extends to Autumn (Fanjul, Zenuto, and Busch 2006; Malizia and Busch 2009). The seasonal reproductive behavior of Anillaco tuco-tuco is under description (Tachinardi, in prep). Still, we believe that the Anillaco tuco-tuco follows a similar reproduction pattern to other species of the genus *Ctenomys*. Therefore, seasonal reproduction might be another factor contributing to the observed increase in high-intensity activity, especially in October, during Spring.

Lastly, tuco-tucos are mainly in a high-intensity state on the surface, which does not change throughout the year (Figure 2.11). The lack of annual change in the aboveground states is surprising (Figure 2.11B), given that the underlying total time spent in each state does change throughout the year (Figure 2.10). During visual observations, tuco-tucos spent their time aboveground foraging and, mostly, maintaining their tunnels (Tomotani et al. 2012), which includes opening plugged entrances and taking sand out of the underground tunnel system. We can speculate that this observed behavior might correspond to high-intensity behavioral states and that these might be activities that are needed year-round. However, we can not confirm such correspondence without grounding truth the models used in the present study. Tuco-tucos in nature, however,

seems to be spend less time in a low activity behavior in the surface than tuco-tucos in semi-natural arenas (Jannetti et al. 2019). However, this difference between animals in different conditions could be a rouding error in our calculation and still need to be quantified to confirm. Overall, we observe that the aboveground activity of free-living tuco-tucos is majorly composed of high-intensity activity.

In conclusion, our results show that free-living *Anillaco* tuco-tucos change their general activity levels and daily patterns of surface emergence throughout the year. The use of HMM enabled us to extend our analysis of general activity. It showed that the reduction in general activity in July is due to tuco-tucos reducing their time spent in high-intensity activities and increasing their time in medium-intensity activities. Overall, we validated, in nature, previous observations done in semi-natural enclosures showing that tuco-tucos are diurnal. We also showed that, throughout the year, *Anillaco* tuco-tucos exhibit changes in their daily activity levels, surface emergence patterns, and time-activity budgets.

Considerações Finais

Um dos objetivos principais desta dissertação era criar uma metodologia de campo para o estudo dos ritmos diários de atividade dos tuco-tuco de Anillaco na natureza, utilizando biologgers. Nesse sentido, consideramos que tivemos sucesso e que a metodologia de captura e recaptura apresentada no Capítulo 2 rendeu ótimos resultados, com um sucesso de recaptura e recuperação de dados maior do que 70%. Essa foi a primeira tentativa de estudo em campo de tuco-tucos no Laboratório de Cronobiologia Binacional e, além dos resultados discutidos no Capítulo 2, também possibilitou coleta de dados sobre a reprodução dessa espécie que serão publicados futuramente (Tachinardi et al., in prep.).

Os resultados discutidos no Capítulo 2, bem como, o uso de *biologging* (Capítulo 1) para investigar a atividade de tuco-tucos na natureza ainda pode responder muitas novas questões. Por exemplo, os resultados do Capítulo 2 mostram que as mudanças ao longo do ano entre saída à superfície e atividade geral são distintas. Então, novos estudos em campo e em arenas seminaturais podem elucidar de maneira mais clara a origem dessas diferenças e se esses atividade geral e saída à superfície respondem diferentemente aos desafios ambientais. Além disso, há uma redução na atividade geral dos tuco-tucos no Inverno, que volta a aumentar em outubro, na primavera. Como foi discutido no Capítulo 2, essas mudanças ao longo do ano podem estar relacionadas ao início da estação reprodutiva, assim como acontece em outras espécies. Portanto, futuros estudos de *biologging* focados no inverno e primavera e na diferença entre machos e fêmeas podem elucidar se essas mudanças anuais se repetem em outros anos e se elas estão relacionadas a comportamentos sexuais e reprodutivos.

O uso de *Hidden Markov models* (HMM) na análise dos dados de acelerometria possibilitou entender melhor as mudanças na atividade geral dos tuco-tucos. Anteriormente, os estudos com acelerômetros em tuco-tucos usaram um valor de limiar baseado na média diária da atividade para definir o que era atividade ou repouso (Jannetti et al. 2019). Esse método, apesar da simplicidade, limita os tipos de análise ou de comportamentos que podemos extrair dos dados, já que a definição de um limiar não é tão simples para mais do que dois comportamentos. Os HMM, em contrapartida, já são bastante utilizados para a análise de dados de acelerometria em animais (Brett T. McClintock et al. 2020; Leos-Barajas et al. 2017). No nosso caso, apesar de algumas limitações, o uso de HMMs permitiu, a partir da distribuição dos próprios dados, classificar três estados comportamentais utilizando a atividade geral (Capítulo 2). O uso futuro desse tipo de modelo nos permitirá explorar ainda mais a fundo como os comportamentos dos tuco-tucos mudam em diferentes estações ou quando submetidos a diferentes desafios ambientais. Outro dado inédito (Capítulo 2) foi o cruzamento dos dados de saída à superfície e dos estados comportamentais, que revelou diferenças comportamentais entre tuco-tucos de Anillaco em condição de vida-livre e semi-natural. Nesses resultados, vimos que os tuco-tucos passam relativamente pouco tempo em repouso quando estão na superfície quando comparados a tuco-tucos em arenas seminaturais (Jannetti et al. 2019), os quais apresentam maior tempo diário na superfície. Esses resultados apontam o uso dos HMMs como uma importante ferramenta em estudos cronobiológicos de campo, permitindo explorar as diferenças nos padrões diários de diferentes comportamentos. Além disso, a possibilidade de utilizarmos HMMs de forma supervisionada nas arenas e em laboratório pode permitir a identificação de comportamentos específicos, algo que não é possível com os dados atuais.

Apêndice

Apêndice A

Animals

Table A.1: List of all tuco-tucos captured, the corresponding attached biologgers and the number of days recorded

ID	Total Days recorded	Season	Sex	Capture Weight (g)	Accelerometer	Lightlogger	Recaptured	Collar Recovered
MAR01	6	March	f	120	Yes	No	Yes	Yes
MAR02	6	March	f	137	Yes	No	Yes	Yes
JUL15	6	July	f	110	Yes	No	Yes	Yes
JUL16	5	July	m	162	Yes	Yes	Yes	Yes
JUL17	8	July	f	132	Yes	Yes	Yes	Yes
JUL18	6	July	f	133	Yes	Yes	Yes	Yes
JUL19	4	July	m	220	Yes	No	Yes	Yes
JUL20	4	July	m	147	Yes	Yes	Yes	Yes
JUL21	4	July	f	152	Yes	Yes	Yes	Yes
JUL23	5	July	f	168	Yes	Yes	Yes	Yes
OCT01	18	October	f	202	Yes	Yes	Yes	Yes
OCT08	15	October	m	208	Yes	No	Yes	Yes
OCT09	7	October	f	144	Yes	No	Yes	Yes
OCT10	11	October	f	164	Yes	Yes	Yes	Yes
OCT13	11	October	f	189	Yes	No	Yes	Yes
OCT14	5	October	f	144	Yes	No	Yes	Yes
FEV01	5	February	f	175	Yes	Yes	Yes	Yes
FEV02	9	February	f	147	Yes	Yes	Yes	Yes
FEV03	7	February	m	123	Yes	Yes	Yes	Yes
FEV05	13	February	m	165	Yes	Yes	Yes	Yes
FEV06	9	February	f	153	Yes	Yes	Yes	Yes

Apêndice B

Anillaco's Environment

B.1 Anillaco's Plant Community

Following methods similar to (Aranda-Rickert, Diez, and Marazzi 2014) a non-extensive survey of the plant community was done in May 2019. Three perpendicular 50m transects were defined near the study site. A point-intercept method was used to record plant species present in the transects. Species right below the sampling points were registered in the data. Sampling points were defined every 1m along the 50m transects. Plant species were identified in the field by a Botanist, except for a few members of the Poaceae family.

The results for the plant survey is in line with what has been described in the literature for the region (Abraham et al. 2009; Aranda-Rickert, Diez, and Marazzi 2014; Fracchia et al. 2011). The results show a dominance of Zygophyllaceae, Poaceae and Fabaceae families. The relative frequency of plant families and species recorded in the area are shown in the graphs below (Fig. B.1).

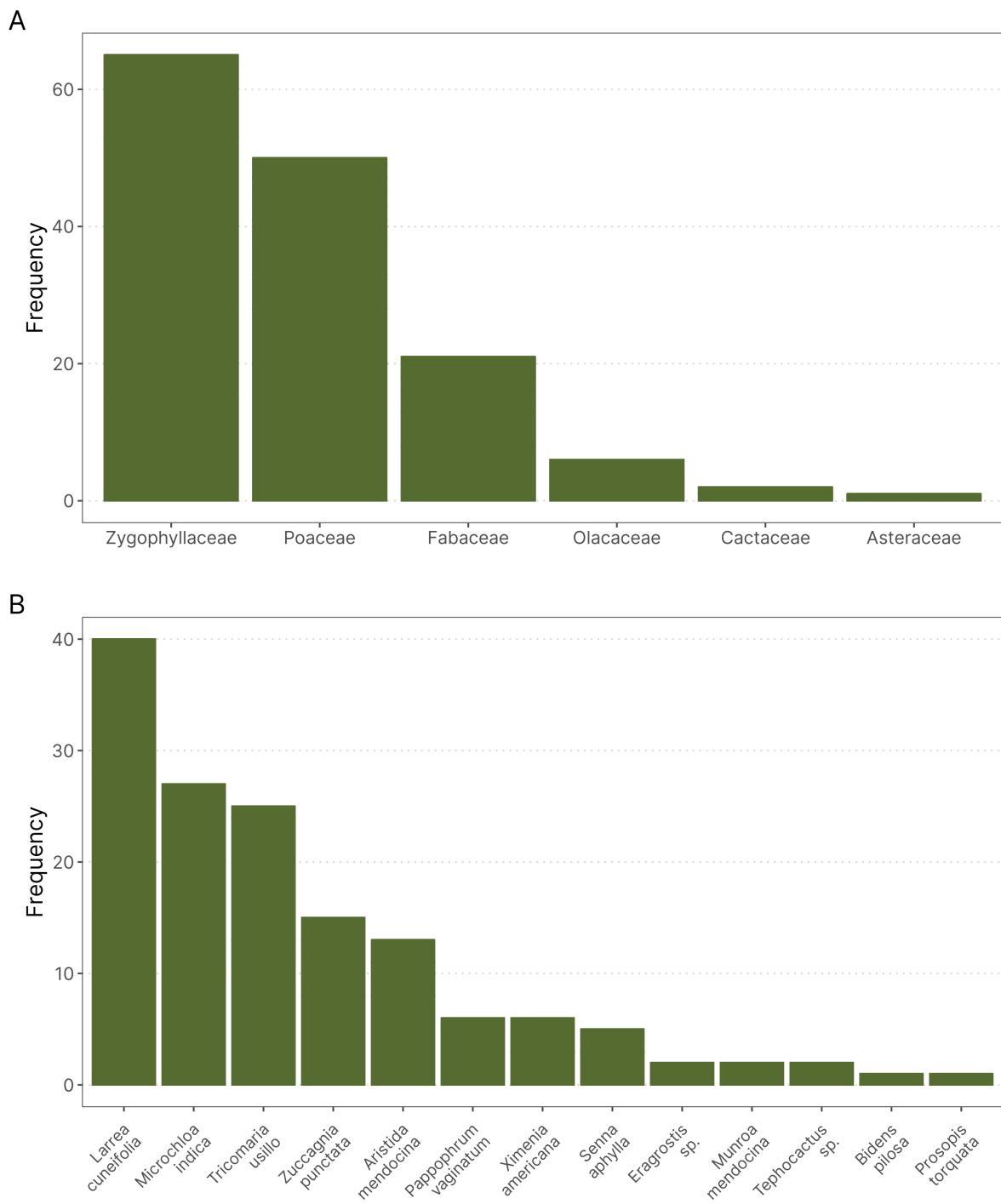


Figure B.1: Relative frequency of plants family (A) and species (B) in three transects near the study site, measured in 2019. The plant community is dominated by members of the Zygophyllaceae, Poaceae and Fabaceae families and is in accordance with what has been described in the literature. (n = 145)

B.2 Anillaco's Weather

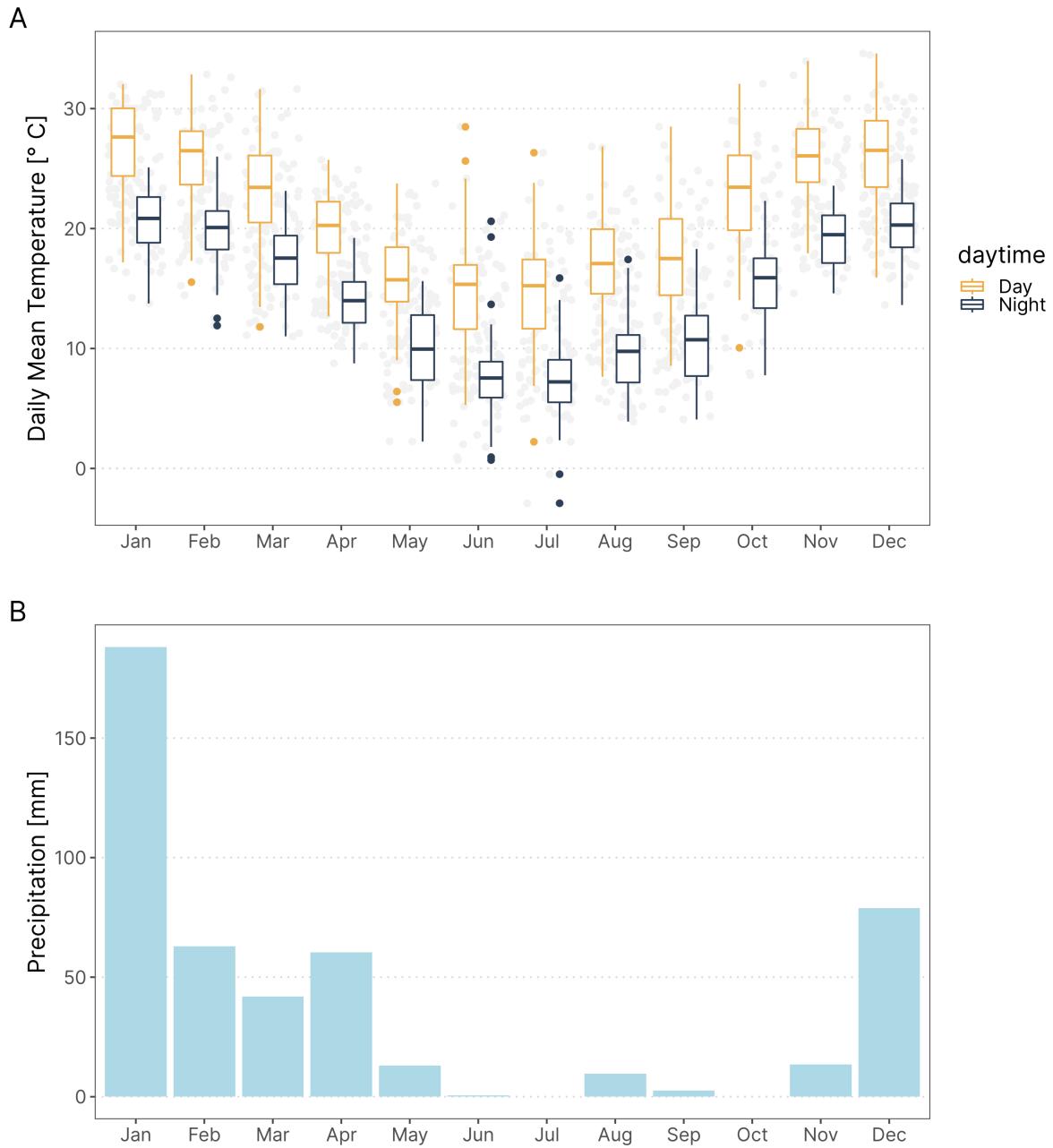


Figure B.2: Temperature and Rainfall yearly trends in Anillaco, Argentina. Data was collected in the years 2017 and 2019 from a weather Station (Vantage Pro 2, Davis Instruments. USA.) maintained in CRILAR, approximately 5km away from the study site.

B.3 Anillaco's Yearly Daylength Changes

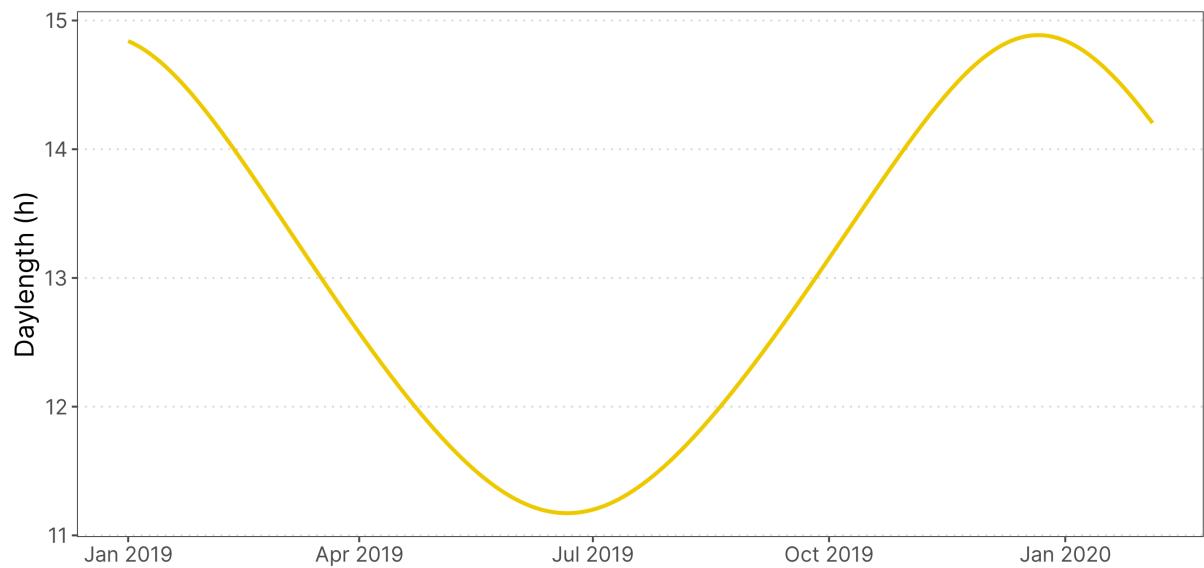


Figure B.3: Changes in day length changes across the year in Anillaco, La Rioja. Maximum duration of daytime, during summer, is 14 hours and 53 minutes. Minimum duration of daytime, during winter, is 11 hours and 10 minutes.

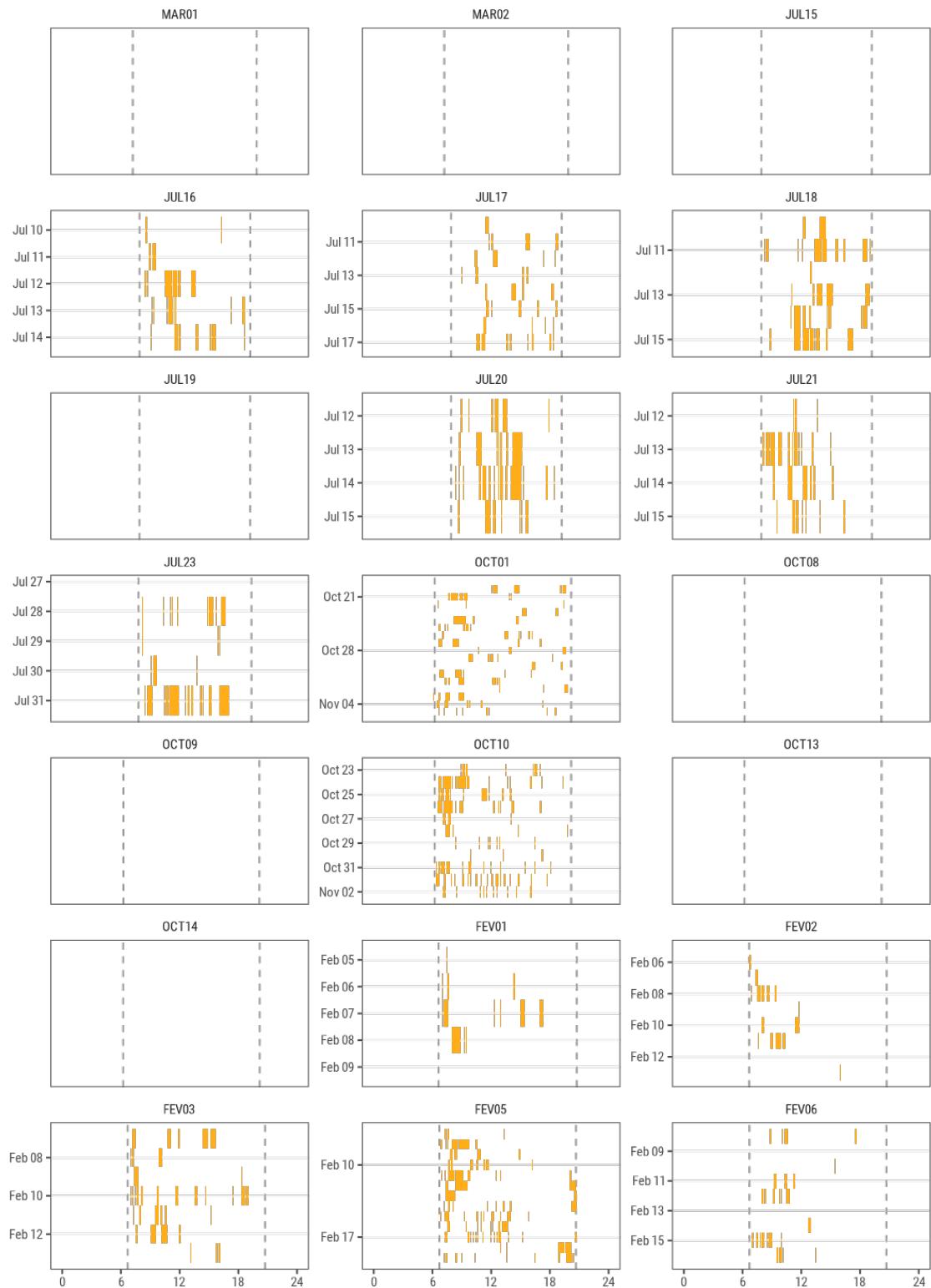
Apêndice C

Individual Actograms

Individual VeDBA and behavioral states actograms show only the first five days of data.

Surface emergence actograms show all days of recordings.

C.1 Surface Emergence Actograms



C.2 VeDBA Actograms

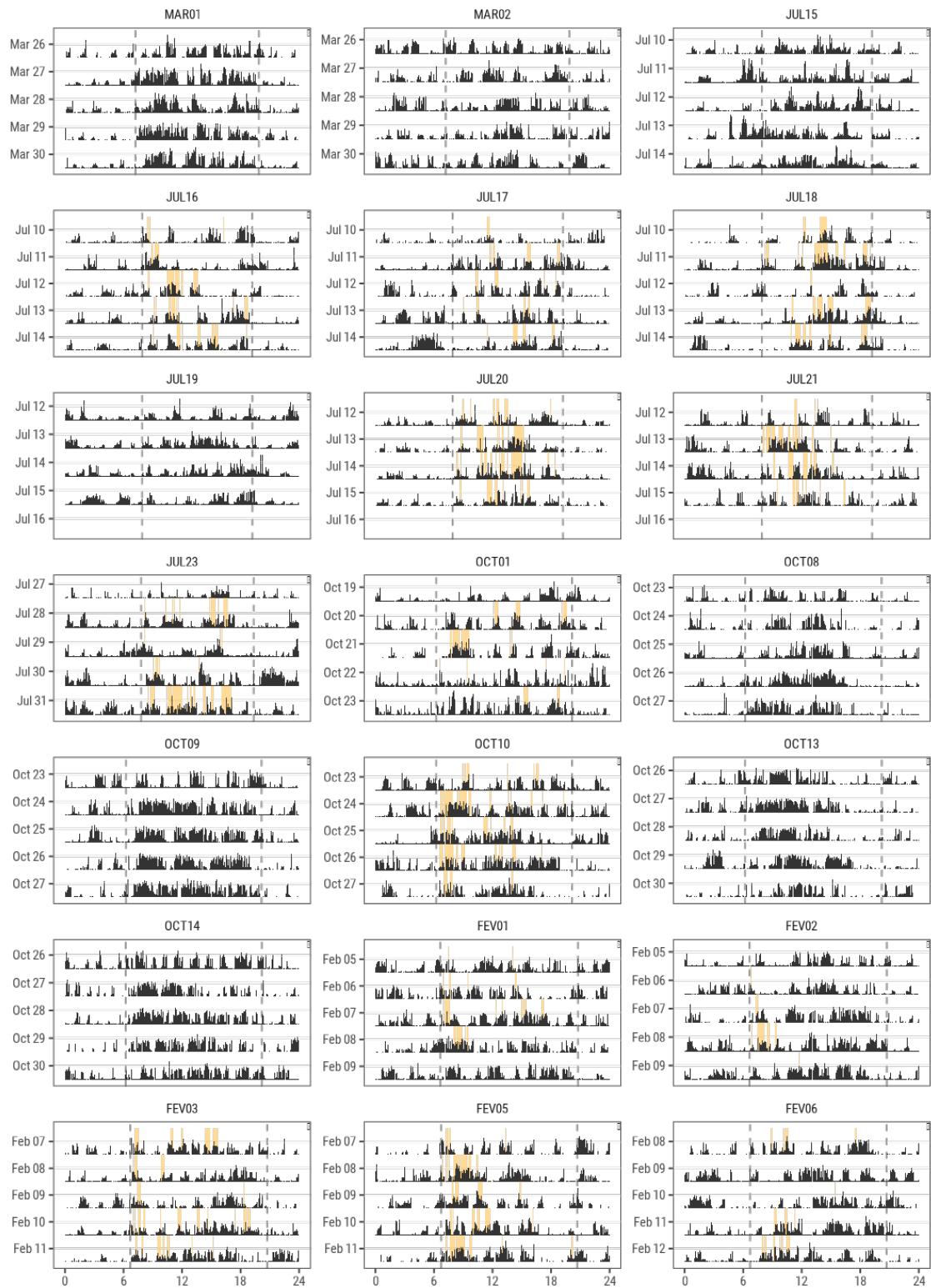
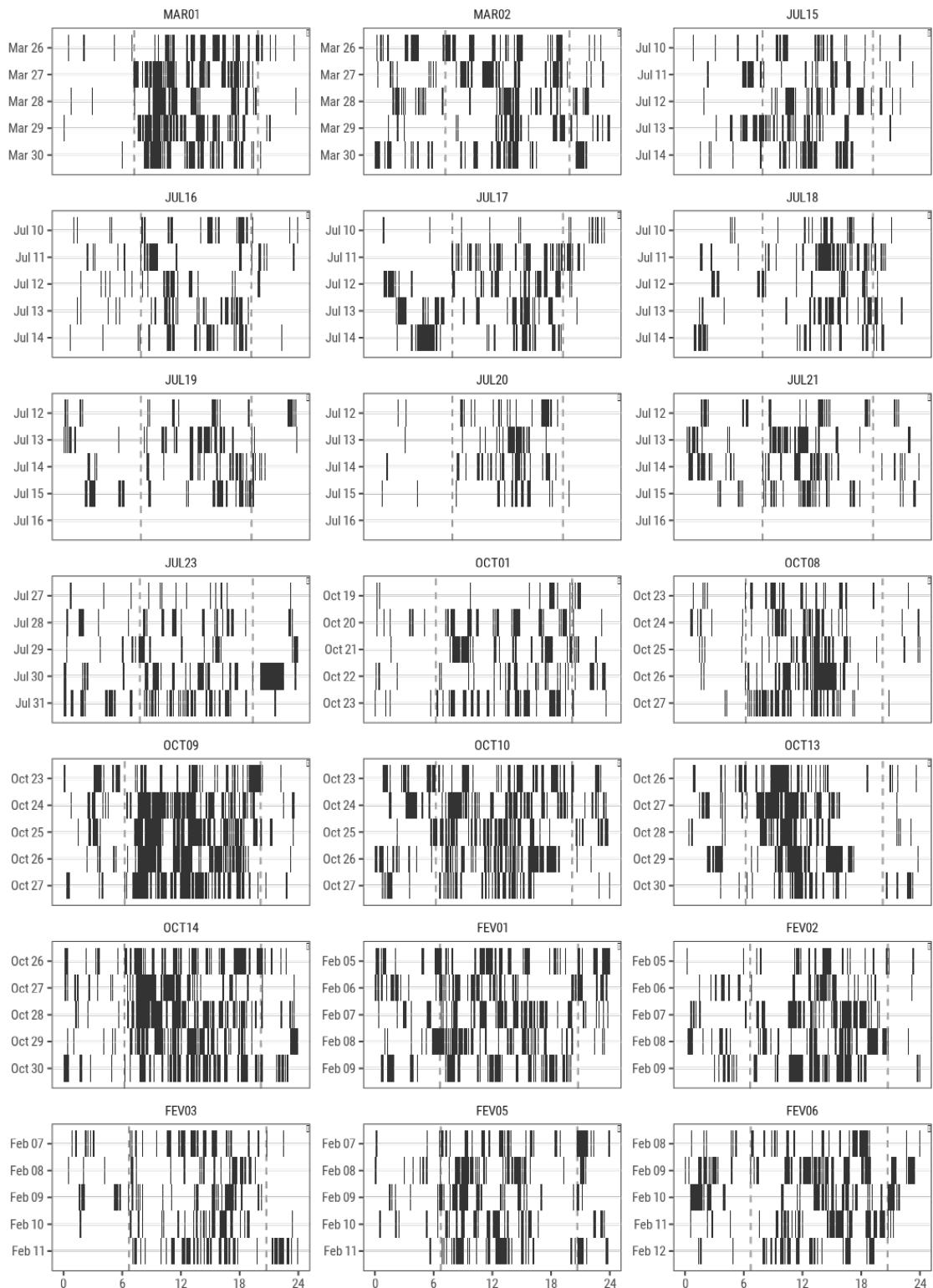
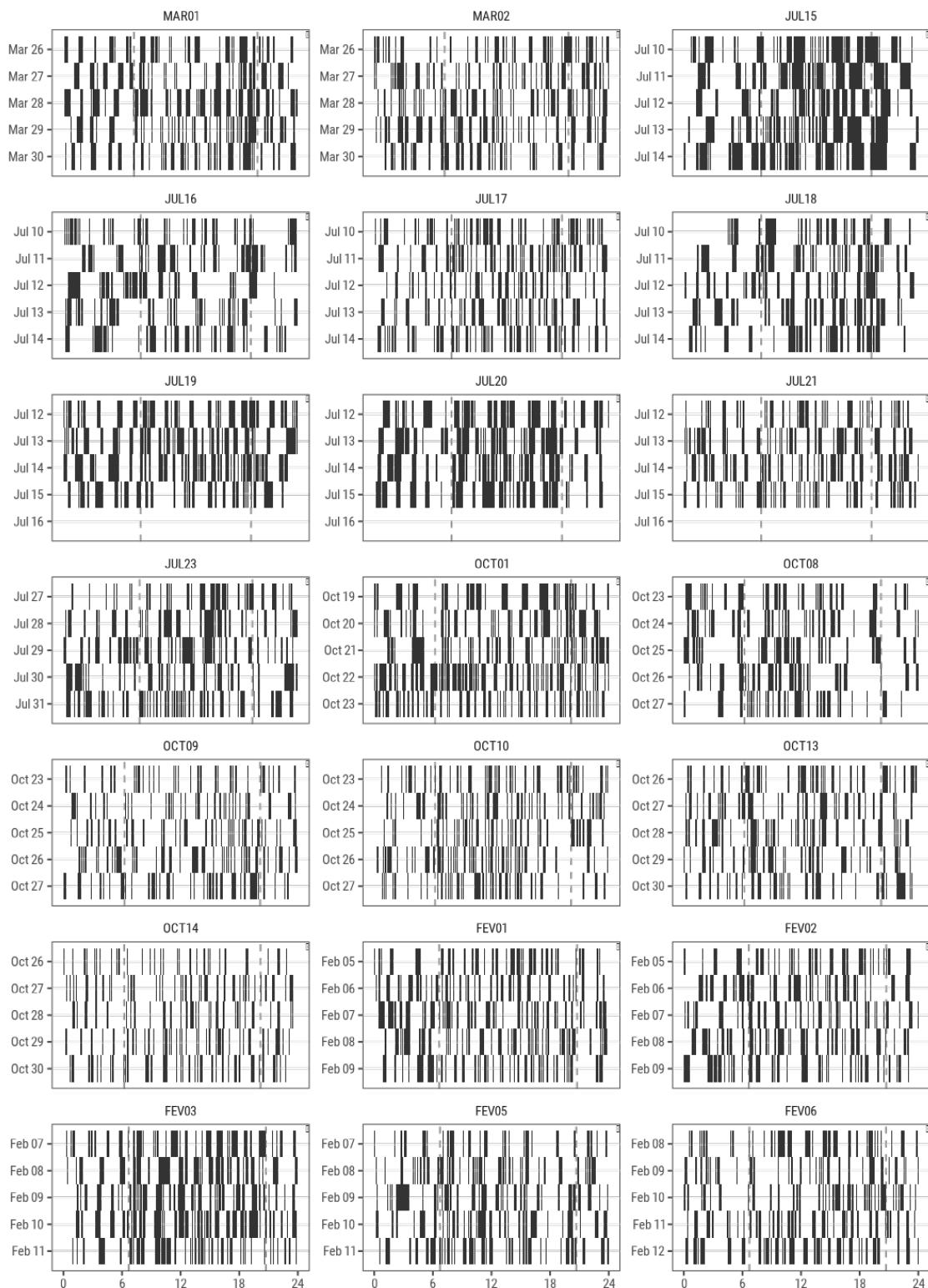


Figure C.1: VeDBA Actograms.

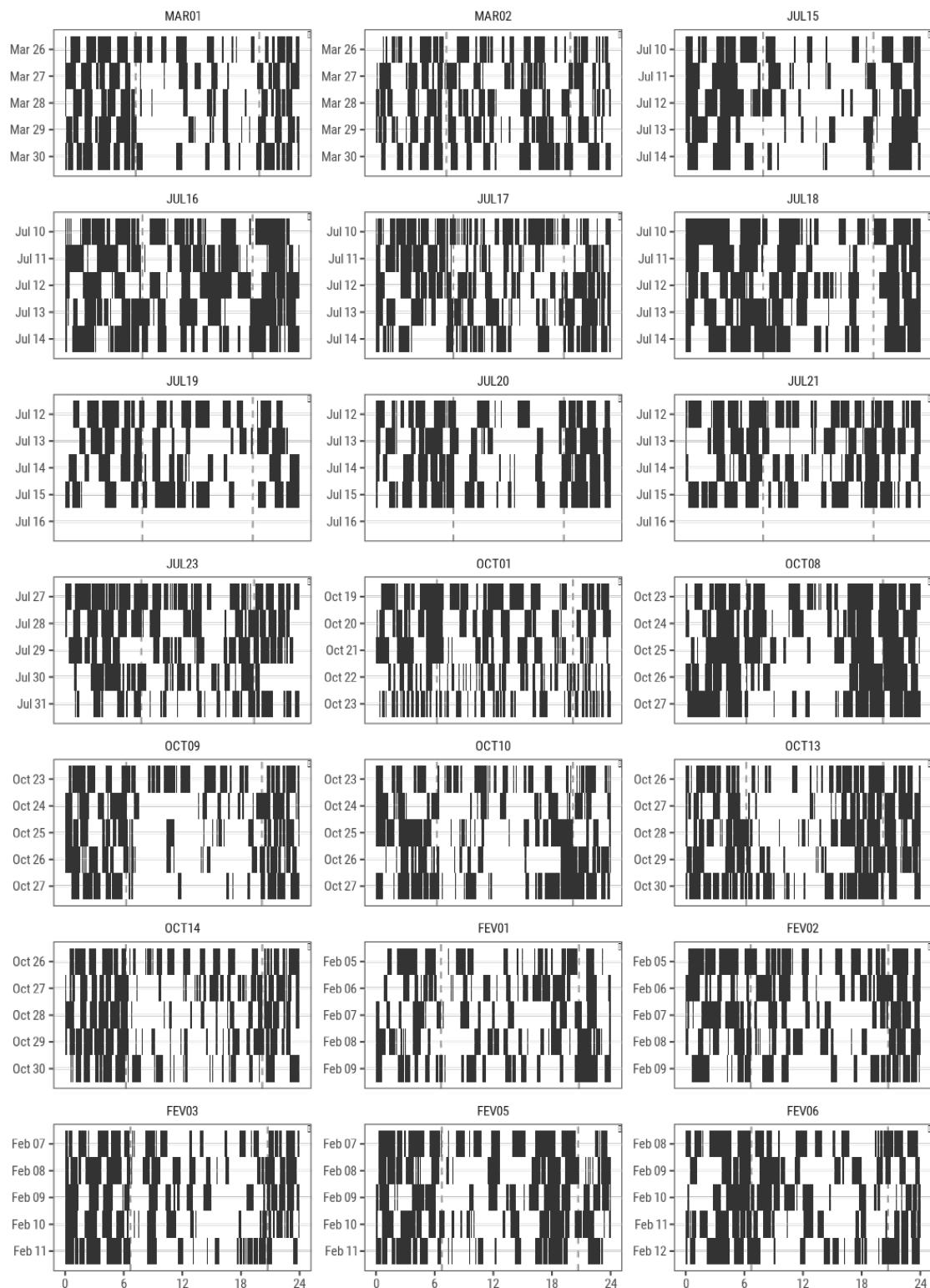
C.3 High Activity Actograms



C.4 Medium Activity Actograms



C.5 Low Activity Actograms



Apêndice D

Accelerometer Data Exploration

D.1 Assessment of Static Acceleration Smoothing Window

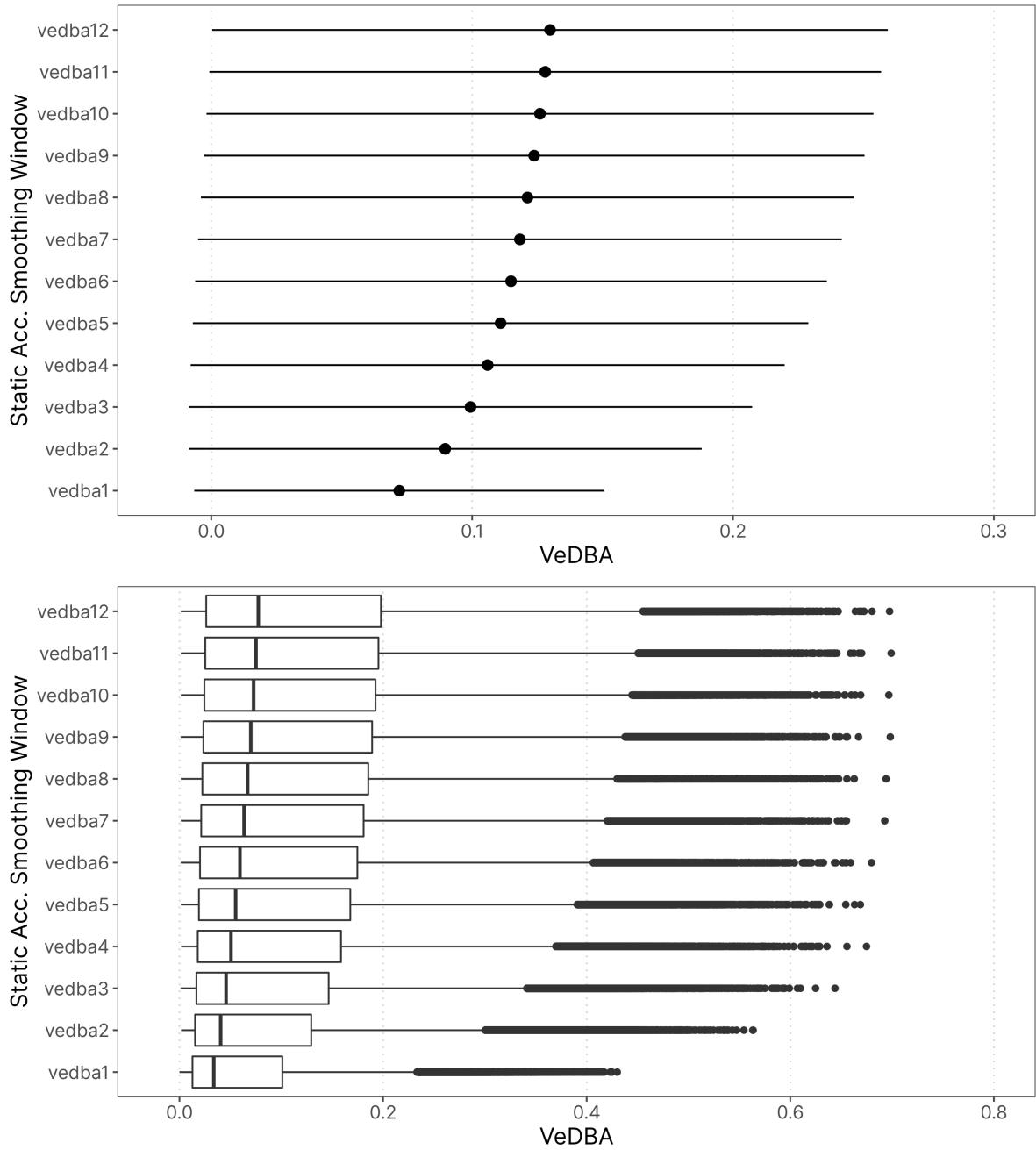


Figure D.1: Smoothing window assessment. We chose the 4-second moving average to proceed with analysis.

D.2 VeDBA Summary and Distributions

Table D.1: Summary table of individual VeDBA data.

ID	season	sex	Mean	Median	Max	Min	Range
MAR01	March	f	0.132	0.081	0.792	0.000	0.792
MAR02	March	f	0.117	0.069	0.728	0.000	0.728
JUL15	July	f	0.117	0.083	0.755	0.000	0.755
JUL16	July	m	0.091	0.032	0.740	0.001	0.739
JUL17	July	f	0.113	0.049	0.750	0.000	0.750
JUL18	July	f	0.093	0.031	0.623	0.000	0.623
JUL19	July	m	0.098	0.071	0.641	0.001	0.641
JUL20	July	m	0.091	0.054	0.624	0.000	0.624
JUL21	July	f	0.108	0.039	0.609	0.001	0.608
JUL23	July	f	0.108	0.042	0.711	0.001	0.710
OCT01	October	f	0.125	0.067	0.718	0.007	0.711
OCT08	October	m	0.110	0.041	0.624	0.000	0.624
OCT09	October	f	0.157	0.084	0.704	0.001	0.703
OCT10	October	f	0.134	0.071	0.680	0.000	0.680
OCT13	October	f	0.128	0.068	0.591	0.000	0.591
OCT14	October	f	0.132	0.066	0.661	0.007	0.655
FEV01	February	f	0.137	0.087	0.629	0.000	0.629
FEV02	February	f	0.116	0.069	0.635	0.000	0.635
FEV03	February	m	0.109	0.072	0.654	0.000	0.654
FEV05	February	m	0.128	0.076	0.646	0.007	0.639
FEV06	February	f	0.120	0.067	0.618	0.000	0.618

Table D.2: Summary table of VeDBA data grouped by season.

season	sex	median	mean	max	n.animals	n.points
March	f	0.076	0.124	0.792	2	17278
July	f	0.049	0.108	0.755	5	41755
July	m	0.040	0.093	0.740	3	18717
October	f	0.070	0.133	0.718	5	74875
October	m	0.041	0.110	0.624	1	21599
February	f	0.074	0.122	0.635	3	33117
February	m	0.074	0.121	0.654	2	28798

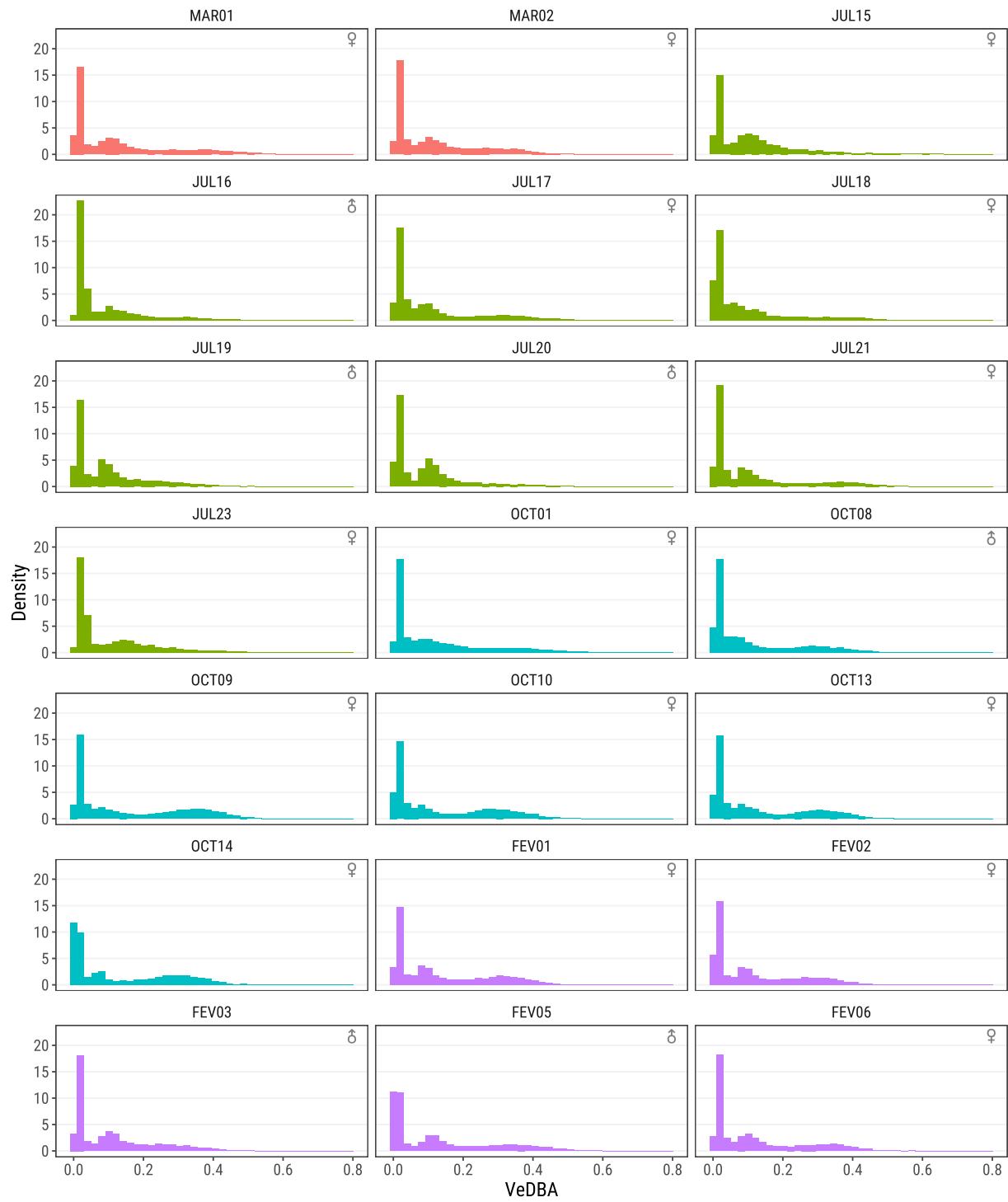


Figure D.2: Individual density histogram of VeDBA data. Colors represents different seasons.

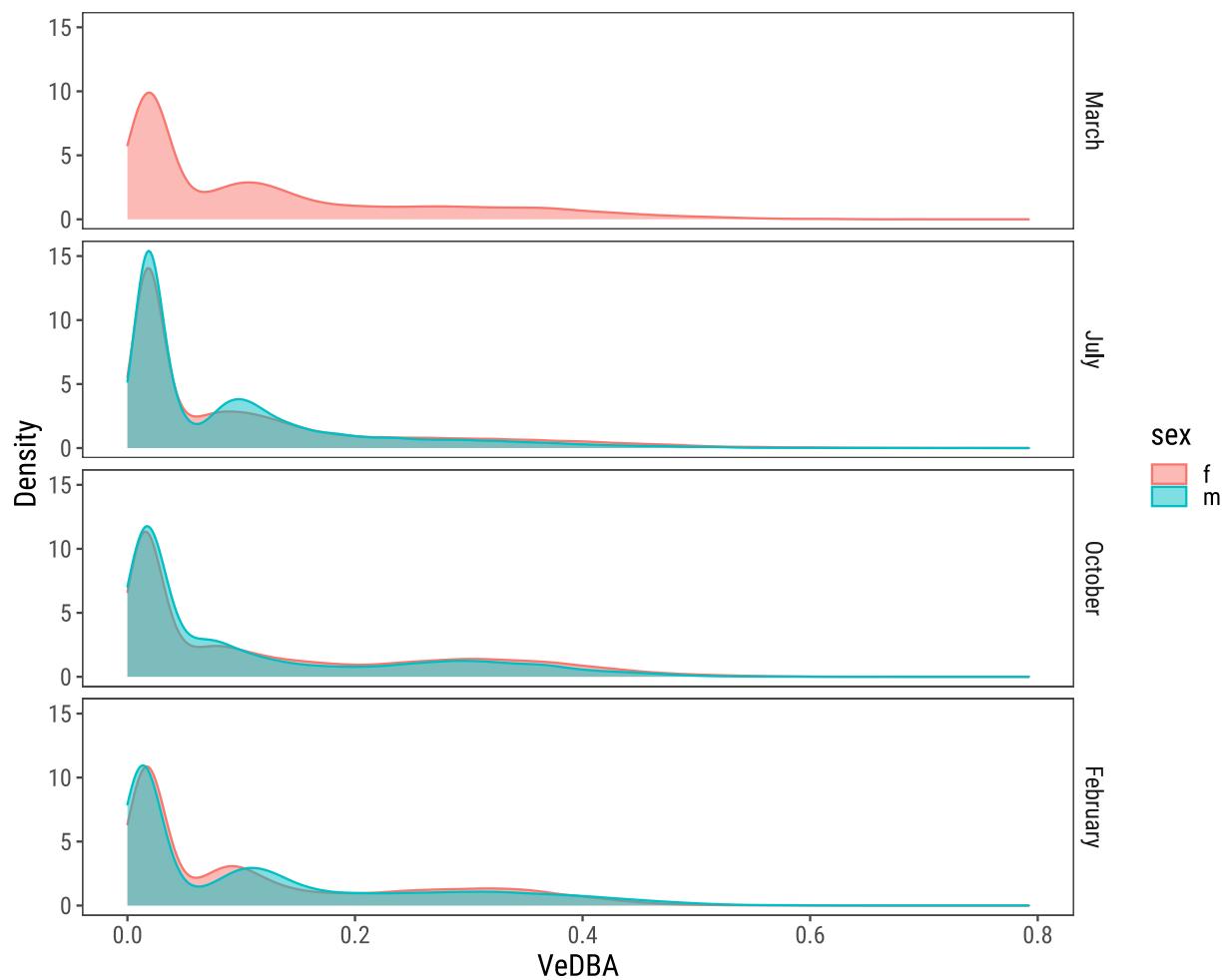


Figure D.3: Density plots of pooled VeDBA data.

Apêndice E

Hidden Markov Model

E.1 AIC

Table E.1: HMM models AIC table.

Model	Formula	AIC
m2	~season	-935612.2
m1	~1	-934800.8

E.2 Estimated Parameters

Table E.2: Gamma State-dependent distribution parameters, mean and standard deviation, estimated by a three-state Hidden Markov Model.

Parameter	State	Estimate	CI
mean	Rest	0.018	[0.018, 0.018]
mean	Medium	0.116	[0.116, 0.117]
mean	High	0.327	[0.326, 0.328]
sd	Rest	0.010	[0.01, 0.01]
sd	Medium	0.051	[0.05, 0.051]
sd	High	0.087	[0.086, 0.087]

E.3 Pseudo-residuals

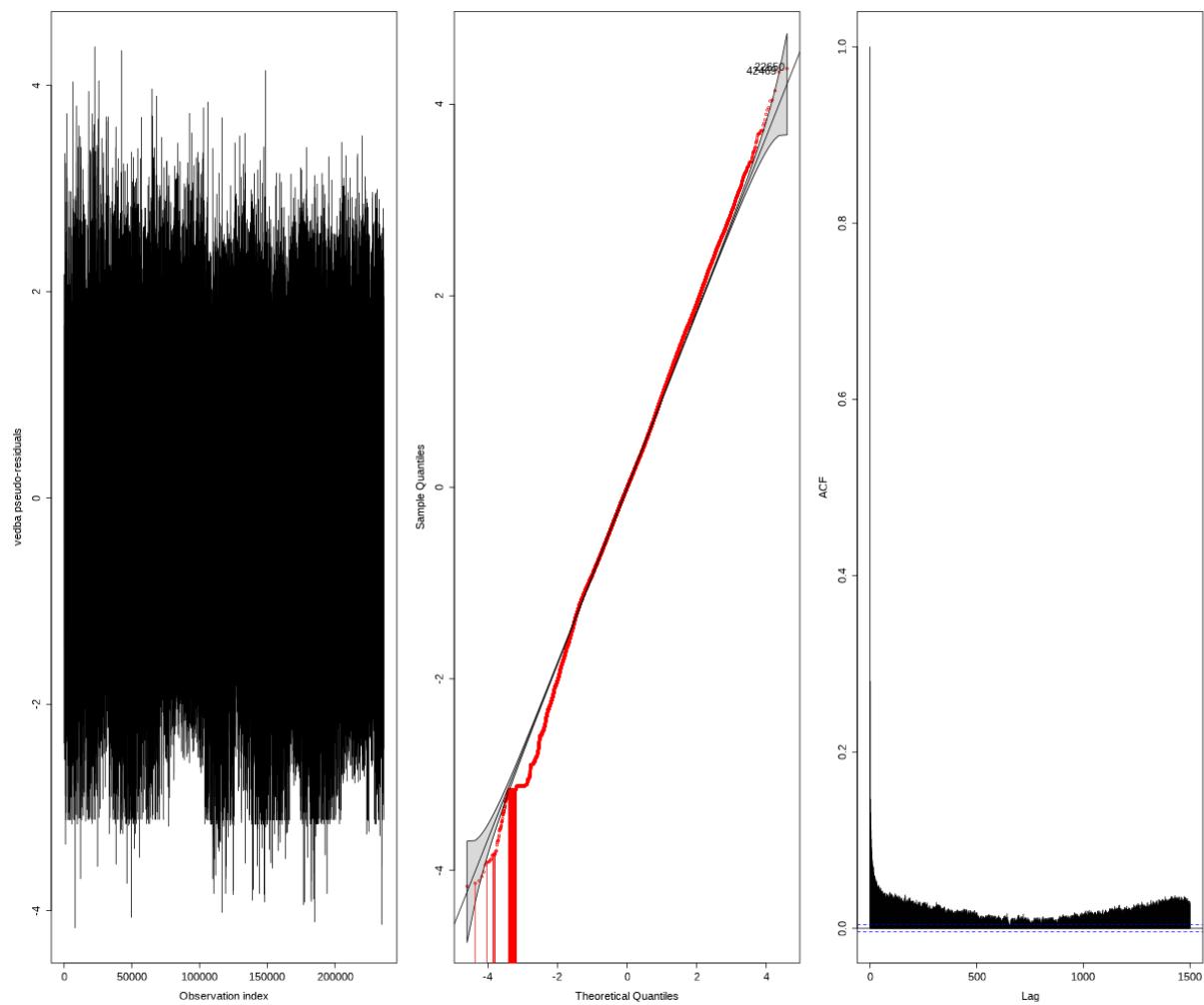
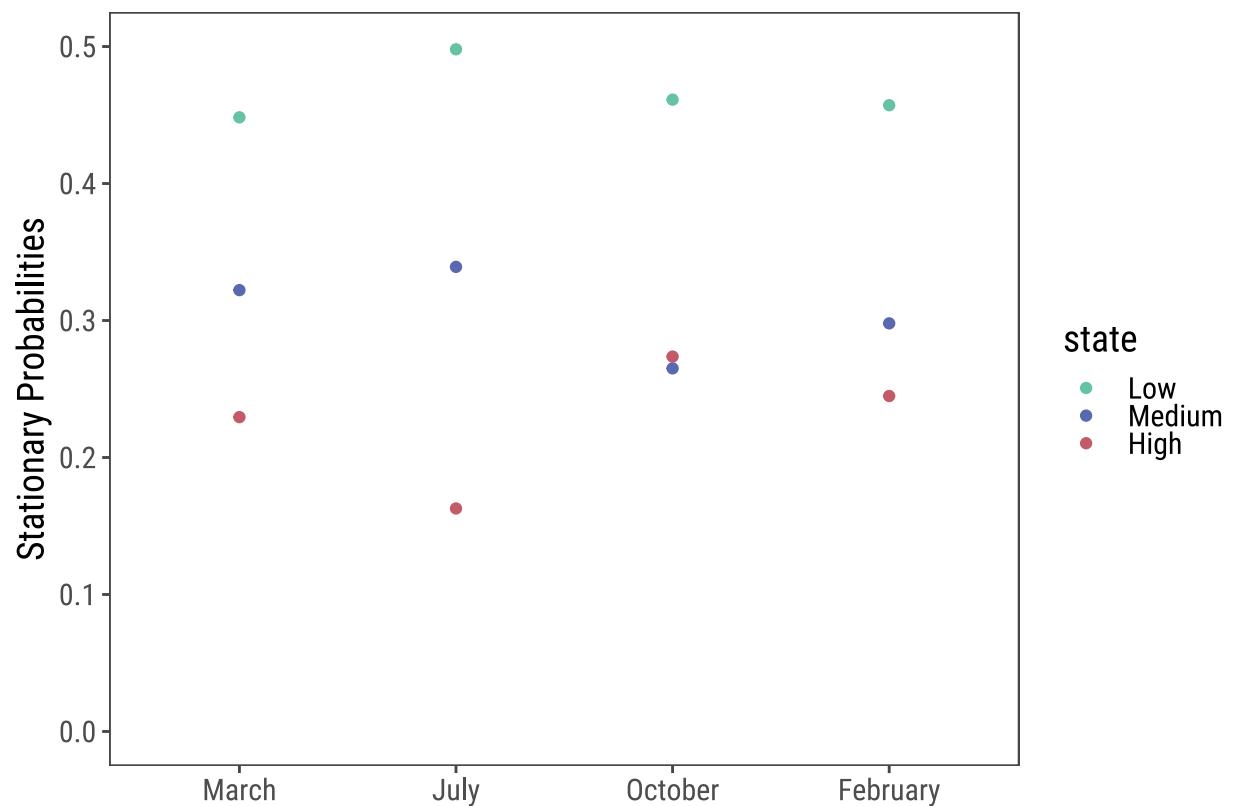
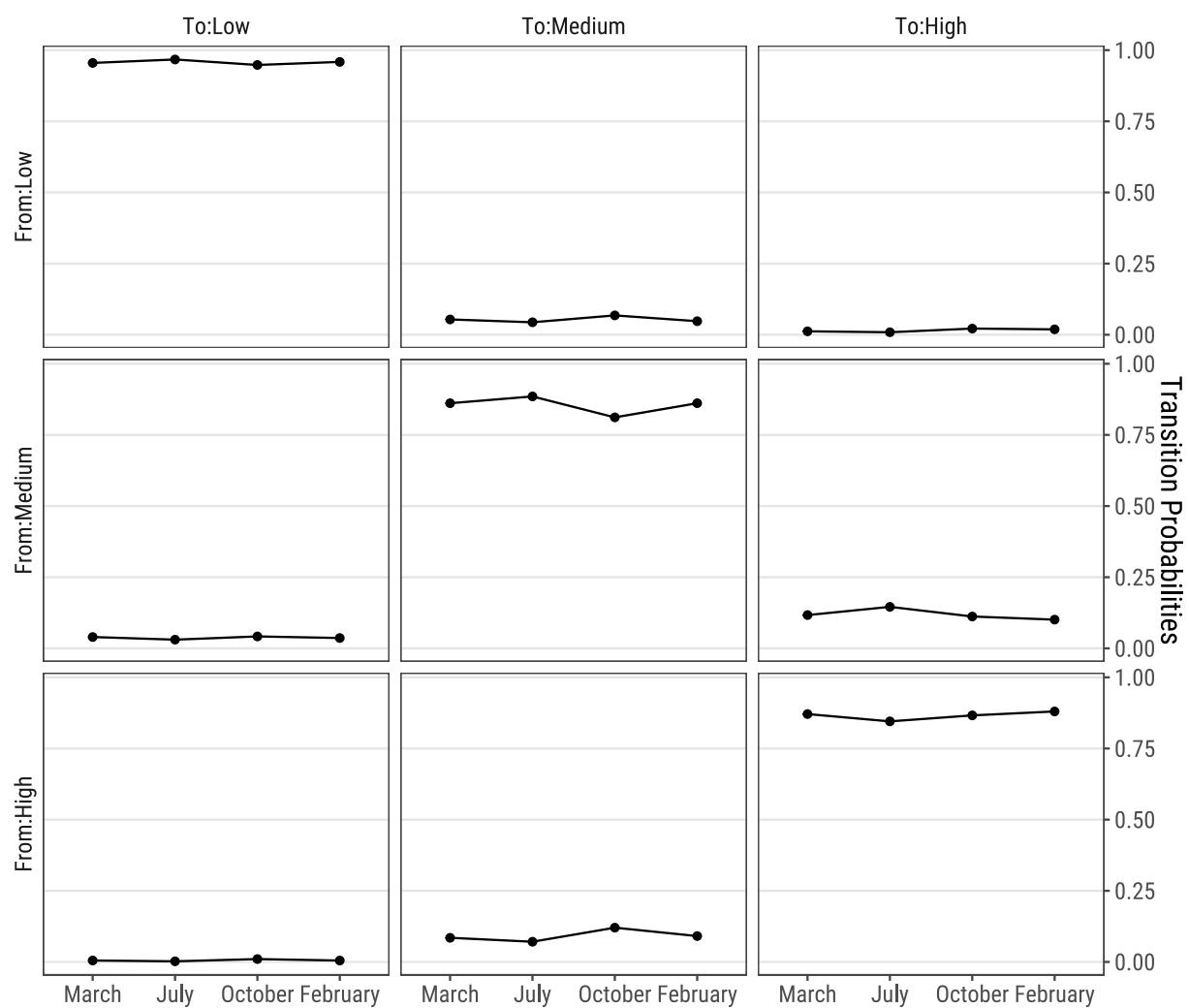


Figure E.1: Model Residuals.

E.4 Stationary Probabilities



E.5 Transition Probabilities



Apêndice F

Individual Circadian Analysis

F.1 Individual Autocorrelation Plots

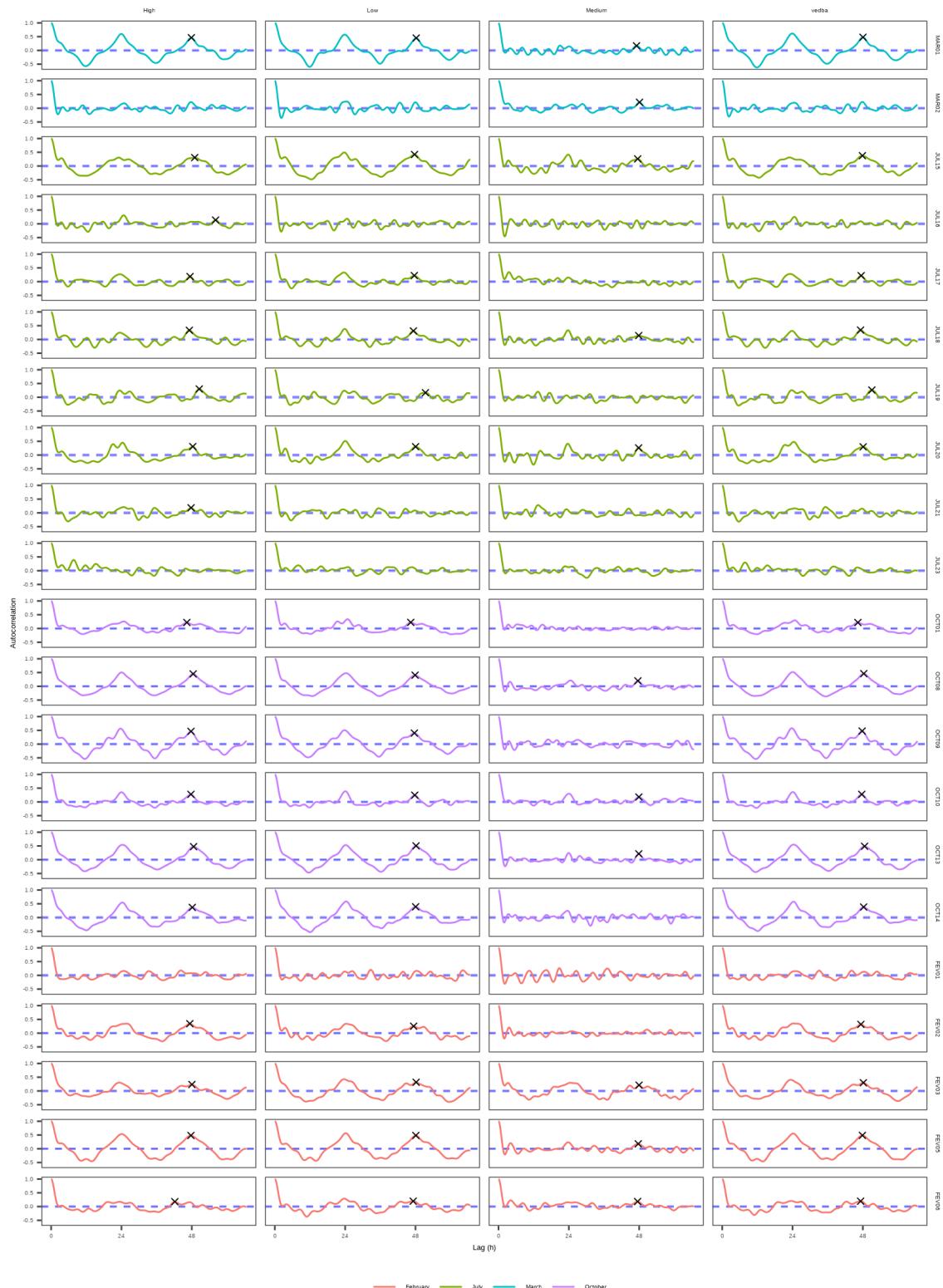
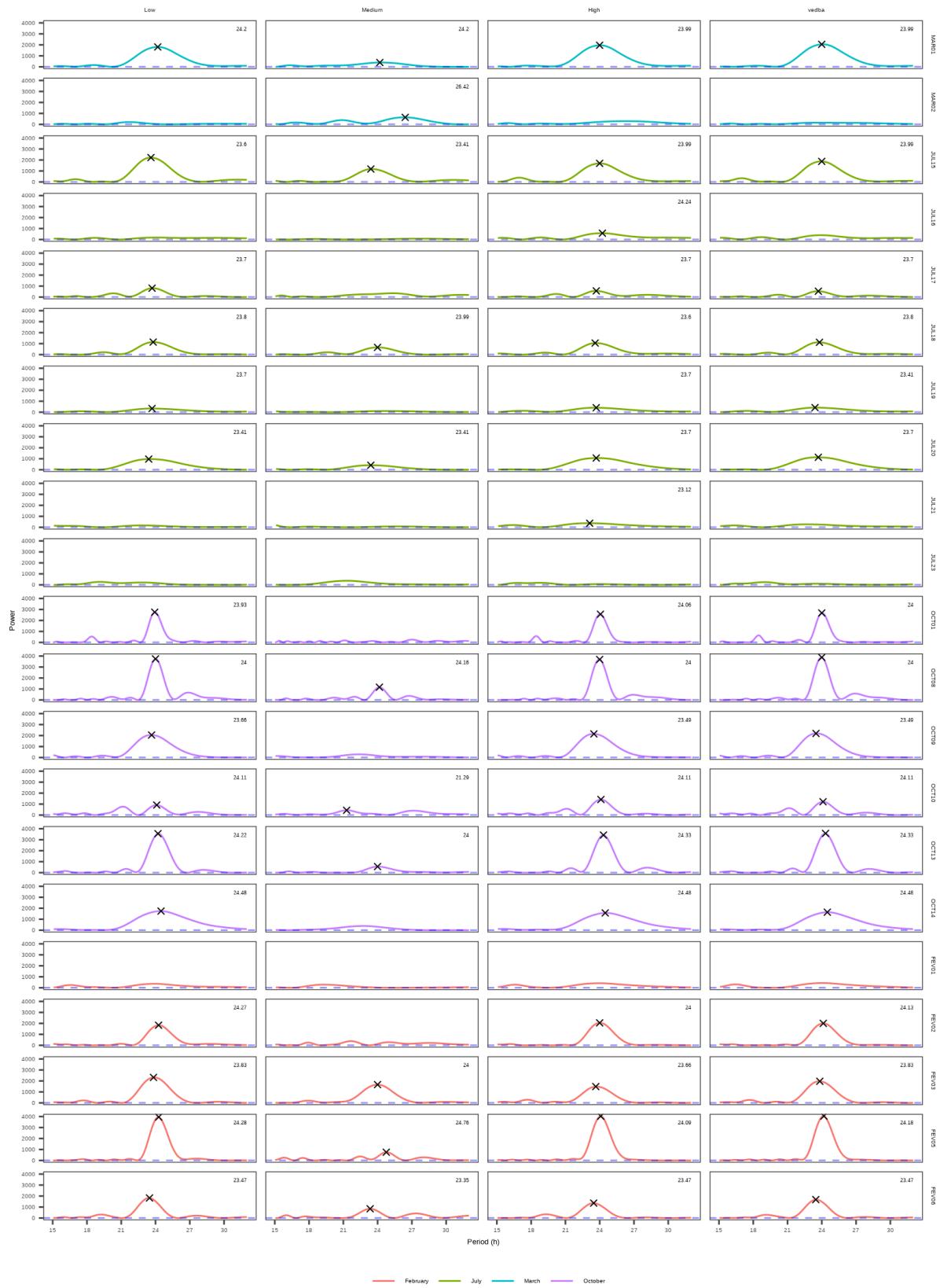


Figure F.1: Individual Autocorrelation plots.

F.2 Individual Period Estimation



Referências

- Ables, Ernest D. 1969. "Activity Studies of Red Foxes in Southern Wisconsin." *The Journal of Wildlife Management* 33 (1): 145. <https://doi.org/10.2307/3799662>.
- Abraham, E., H.F. del Valle, F. Roig, L. Torres, J.O. Ares, F. Coronato, and R. Godagnone. 2009. "Overview of the Geography of the Monte Desert Biome (Argentina)." *Journal of Arid Environments* 73 (2): 144–53. <https://doi.org/10.1016/j.jaridenv.2008.09.028>.
- Amaya, Juan Pablo, Juan I. Areta, Veronica S. Valentiniuzzi, and Emmanuel Zufiaurre. 2016. "Form and Function of Long-Range Vocalizations in a Neotropical Fossorial Rodent: The Anillaco Tuco-Tuco (*Ctenomys* Sp.)." *PeerJ* 4 (October): e2559. <https://doi.org/10.7717/peerj.2559>.
- Antinuchi, C. Daniel, Roxana R. Zenuto, Facunda Luna, Ana Paula Cutrera, Paula P. Perissinotti, and Cristina Busch. 2007. "Energy Budget in Subterranean Rodents: Insights from the Tuco-Tuco *Ctenomys Talarum* (Rodentia: Ctenomyidae)." In *The Quintessential Naturalist: Honoring the Life and Legacy of Oliver P. Pearson*. University of California Press. <http://california.universitypressscholarship.com/view/10.1525/california/9780520098596.001.0001/upso-9780520098596>.
- Aranda-Rickert, Adriana, Patricia Diez, and Brigitte Marazzi. 2014. "Extrafloral Nectar Fuels Ant Life in Deserts." *AoB PLANTS* 6 (January). <https://doi.org/10.1093/aobpla/plu068>.

- Aranda-Rickert, Adriana, and Sebastián Fracchia. 2011. "Pogonomyrmexcunicularius as the Keystone Disperser of Elaiosome-Bearing *Jatropha Excisa* Seeds in Semi-Arid Argentina: Pogonomyrmex Cunicularius Ants as Keystone Seed Dispersers." *Entomologia Experimentalis Et Applicata* 139 (2): 91–102. <https://doi.org/10.1111/j.1570-7458.2011.01111.x>.
- Bisigato, A. J., P. E. Villagra, J. O. Ares, and B. E. Rossi. 2009. "Vegetation Heterogeneity in Monte Desert Ecosystems: A Multi-Scale Approach Linking Patterns and Processes." *Journal of Arid Environments* 73 (2): 182–91. <https://doi.org/10.1016/j.jaridenv.2008.09.001>.
- Bivand, Roger, and Nicholas Lewin-Koh. 2020. "Maptools: Tools for Handling Spatial Objects." <https://CRAN.R-project.org/package=maptools>.
- Blanchong, J. A., T. L. McElhinny, M. M. Mahoney, and L. Smale. 1999. "Nocturnal and Diurnal Rhythms in the Unstriped Nile Rat, *Arvicanthis Niloticus*." *Journal of Biological Rhythms* 14 (5): 364–77. <https://doi.org/10.1177/074873099129000777>.
- Bloch, Guy, Brian M. Barnes, Menno P. Gerkema, and Barbara Helm. 2013. "Animal Activity Around the Clock with No Overt Circadian Rhythms: Patterns, Mechanisms and Adaptive Value." *Proceedings of the Royal Society B: Biological Sciences* 280 (1765): 20130019. <https://doi.org/10.1098/rspb.2013.0019>.
- Brown, Danielle D., Roland Kays, Martin Wikelski, Rory Wilson, and A. Peter Klimley. 2013. "Observing the Unwatchable Through Acceleration Logging of Animal Behavior." *Animal Biotelemetry* 1 (1): 20. <https://doi.org/10.1186/2050-3385-1-20>.
- Bunn, Andrew G. 2008. "A Dendrochronology Program Library in R (dplR)." *Dendrochronologia* 26 (2): 115–24. <https://doi.org/10.1016/j.dendro.2008.01.002>.
- Calisi, Rebecca M., and George E. Bentley. 2009. "Lab and Field Experiments: Are They the Same Animal?" *Hormones and Behavior* 56 (1): 1–10. <https://doi.org/10.1016/j.yhbeh.2008.11.014>.

<https://doi.org/10.1016/j.yhbeh.2009.02.010>.

Chmura, Helen E., Thomas W. Glass, and Cory T. Williams. 2018. "Biologging Physiological and Ecological Responses to Climatic Variation: New Tools for the Climate Change Era." *Frontiers in Ecology and Evolution* 6 (July): 92. <https://doi.org/10.3389/fevo.2018.00092>.

Daan, Serge. 1981. "Adaptive Daily Strategies in Behavior." In *Handbook of Behavioral Neurobiology*, 4:563. Plenum Press.

Daan, Serge, and Steven Slopsema. 1978. "Short-Term Rhythms in Foraging Behaviour of the Common Vole, *Microtus Arvalis*." *Journal of Comparative Physiology* 127 (3): 215–27. <https://doi.org/10.1007/BF01350112>.

de Freitas, Thales Renato Ochotorena, Gislene Lopes Gonçalves, and Renan Maestri, eds. 2021. *Tuco-Tucos: An Evolutionary Approach to the Diversity of a Neotropical Subterranean Rodent*. Cham: Springer International Publishing. <https://doi.org/10.1007/978-3-030-61679-3>.

Dominoni, Davide M., Susanne Åkesson, Raymond Klaassen, Kamiel Spoelstra, and Martin Bulla. 2017. "Methods in Field Chronobiology." *Philosophical Transactions of the Royal Society B: Biological Sciences* 372 (1734): 20160247. <https://doi.org/10.1098/rstb.2016.0247>.

Dowse, Harold B. 2009. "Chapter 6 Analyses for Physiological and Behavioral Rhythmicity." In, 454:141–74. Elsevier. [https://doi.org/10.1016/S0076-6879\(08\)03806-8](https://doi.org/10.1016/S0076-6879(08)03806-8).

Duda, Niklas, Thorsten Nowak, Markus Hartmann, Michael Schadhauser, Björn Cassens, Peter Wägemann, Muhammad Nabeel, et al. 2018. "BATS: Adaptive Ultra Low Power Sensor Network for Animal Tracking." Article. *Sensors* 18 (10, 10): 3343. <https://doi.org/10.3390/s18103343>.

Enright, J T. 1970. "Ecological Aspects of Endogenous Rhythmicity." *Annual Review of Ecology and Systematics* 1 (1): 221–38. <https://doi.org/10.1146/annurev.>

es.01.110170.001253.

- Everts, Lammina G., Arjen M. Strijkstra, Roelof A. Hut, Ilse E. Hoffmann, and Eva Millesi. 2004. "Seasonal Variation in Daily Activity Patterns of Free-Ranging European Ground Squirrels (*Spermophilus Citellus*)."*Chronobiology International* 21 (1): 57–71. <https://doi.org/10.1081/cbi-120027982>.
- Fanjul, M. Sol, Roxana R. Zenuto, and Cristina Busch. 2006. "Seasonality of Breeding in Wild Tuco-tucos *Ctenomys Talarum* in Relation to Climate and Food Availability."*Acta Theriologica* 51 (3): 283–93. <https://doi.org/10.1007/BF03192680>.
- Flôres, Danilo E. F. L., Milene G. Jannetti, Giovane C. Imrota, Patricia Tachinardi, Veronica S. Valentinuzzi, and Gisele A. Oda. 2021. "Telling the Seasons Underground: The Circadian Clock and Ambient Temperature Shape Light Exposure and Photoperiodism in a Subterranean Rodent."*Frontiers in Physiology* 12: 1690. <https://doi.org/10.3389/fphys.2021.738471>.
- Flôres, Danilo E. F. L., Milene G. Jannetti, Veronica S. Valentinuzzi, and Gisele A. Oda. 2016. "Entrainment of Circadian Rhythms to Irregular Light/Dark Cycles: A Subterranean Perspective."*Scientific Reports* 6 (1): 34264. <https://doi.org/10.1038/srep34264>.
- Flôres, Danilo E. F. L., Barbara M. Tomotani, Patricia Tachinardi, Gisele A. Oda, and Veronica S. Valentinuzzi. 2013. "Modeling Natural Photic Entrainment in a Subterranean Rodent (*Ctenomys Aff. Knighti*), the Tuco-Tuco." Edited by Gianluca Tosini. *PLoS ONE* 8 (7): e68243. <https://doi.org/10.1371/journal.pone.0068243>.
- Fracchia, S., L. Krapovickas, A. Aranda-Rickert, and V.S. Valentinuzzi. 2011. "Dispersal of Arbuscular Mycorrhizal Fungi and Dark Septate Endophytes by *Ctenomys Cf. Knighti* (Rodentia) in the Northern Monte Desert of Argentina."*Journal of Arid Environments* 75 (11): 1016–23. <https://doi.org/10.1016/j.jaridenv.2011.04.034>.

- Frey, Sandra, Jason T. Fisher, A. Cole Burton, and John P. Volpe. 2017. "Investigating Animal Activity Patterns and Temporal Niche Partitioning Using Camera-Trap Data: Challenges and Opportunities." *Remote Sensing in Ecology and Conservation* 3 (3): 123–32. <https://doi.org/10.1002/rse2.60>.
- Geissmann, Quentin, Luis Garcia Rodriguez, Esteban J. Beckwith, and Giorgio F. Gilestro. 2019. "Rethomics: An R Framework to Analyse High-Throughput Behavioural Data." *PLOS ONE* 14 (1): e0209331. <https://doi.org/10.1371/journal.pone.0209331>.
- Gerkema, Menno P., Serge Daan, Marieke Wilbrink, Martina W. Hop, and Floris van der Leest. 1993. "Phase Control of Ultradian Feeding Rhythms in the Common Vole (*Microtus Arvalis*): The Roles of Light and the Circadian System." *Journal of Biological Rhythms* 8 (2): 151–71. <https://doi.org/10.1177/074873049300800205>.
- Gerkema, Menno, and Serge Daan. 1985. "Ultradian Rhythms in Behavior The Case of the Common Vole (*Microtus Arvalis*)."https://doi.org/10.1007/978-3-642-70483-3_3.
- Halle, S., and Nils Chr Stenseth, eds. 2000. *Activity Patterns in Small Mammals: An Ecological Approach*. Ecological Studies, v. 141. Berlin ; New York: Springer.
- Hans W. Borchers. 2019. "Pracma: Practical Numerical Math Functions." <https://CRAN.R-project.org/package=pracma>.
- Helm, Barbara, Marcel E. Visser, William Schwartz, Noga Kronfeld-Schor, Menno Gerkema, Theunis Piersma, and Guy Bloch. 2017. "Two Sides of a Coin: Ecological and Chronobiological Perspectives of Timing in the Wild." *Philosophical Transactions of the Royal Society B: Biological Sciences* 372 (1734): 20160246. <https://doi.org/10.1098/rstb.2016.0246>.
- Hut, Roelof A., Noga Kronfeld-Schor, Vincent van der Vinne, and Horacio De la Iglesia. 2012. "In Search of a Temporal Niche: Environmental Factors." In *Progress*

in Brain Research, 199:281–304. Elsevier. <https://doi.org/10.1016/B978-0-444-59427-3.00017-4>.

Hut, Roelof A., Bob E. H. van Oort, and Serge Daan. 1999. “Natural Entrainment Without Dawn and Dusk: The Case of the European Ground Squirrel (*Spermophilus Citellus*).” *Journal of Biological Rhythms* 14 (4): 290–99. <https://doi.org/10.1177/074873099129000704>.

Jabbur, Maria Lu’isa, Chi Zhao, and Carl Hirschie Johnson. 2021. “Insights into the Evolution of Circadian Clocks Gleaned from Bacteria.” In *Circadian Rhythms in Bacteria and Microbiomes*, edited by Carl Hirschie Johnson and Michael Joseph Rust, 111–35. Cham: Springer International Publishing. <https://doi.org/10.1007/978-3-030-72158-9>.

Jannetti, Milene G., C. Loren Buck, Veronica S. Valentinuzzi, and Gisele A. Oda. 2019. “Day and Night in the Subterranean: Measuring Daily Activity Patterns of Subterranean Rodents (*Ctenomys* Aff. *Knighti*) Using Bio-Logging.” *Conservation Physiology* 7 (1). <https://doi.org/10.1093/conphys/coz044>.

Jansen, Heiko T., Tanya Leise, Gordon Stenhouse, Karine Pigeon, Wayne Kasworm, Justin Teisberg, Thomas Radandt, Robert Dallmann, Steven Brown, and Charles T. Robbins. 2016. “The Bear Circadian Clock Doesn’t ‘Sleep’ During Winter Dormancy.” *Frontiers in Zoology* 13 (1): 42. <https://doi.org/10.1186/s12983-016-0173-x>.

Kays, Roland, Sameer Tilak, Margaret Crofoot, Tony Fountain, Daniel Obando, Alejandro Ortega, Franz Kuemmeth, et al. 2011. “Tracking Animal Location and Activity with an Automated Radio Telemetry System in a Tropical Rainforest.” *The Computer Journal* 54 (12): 1931–48. <https://doi.org/10.1093/comjnl/bxr072>.

Kenagy, G. J. 1973. “Daily and Seasonal Patterns of Activity and Energetics in a Heteromyid Rodent Community.” *Ecology* 54 (6): 1201–19. <https://doi.org/10.2307/1934184>.

- . 1976. "The Periodicity of Daily Activity and Its Seasonal Changes in Free-Ranging and Captive Kangaroo Rats." *Oecologia* 24 (2): 105–40. <https://doi.org/10.1007/BF00572754>.
- Kronfeld-Schor, Noga, Guy Bloch, and William J. Schwartz. 2013. "Animal Clocks: When Science Meets Nature." *Proceedings of the Royal Society B: Biological Sciences* 280 (1765): 20131354. <https://doi.org/10.1098/rspb.2013.1354>.
- Kronfeld-Schor, Noga, and Tamar Dayan. 2003. "Partitioning of Time as an Ecological Resource." *Annual Review of Ecology, Evolution, and Systematics* 34 (1): 153–81. <https://doi.org/10.1146/annurev.ecolsys.34.011802.132435>.
- Kronfeld-Schor, Noga, Marcel E. Visser, Lucia Salis, and Jan A. van Gils. 2017. "Chronobiology of Interspecific Interactions in a Changing World." *Philosophical Transactions of the Royal Society B: Biological Sciences* 372 (1734): 20160248. <https://doi.org/10.1098/rstb.2016.0248>.
- Lacey, E. A., J. L. Patton, and G. N. Cameron. 2000. *Life Underground: The Biology of Subterranean Rodents*. Life Underground: The Biology of Subterranean Rodents. University of Chicago Press. <https://books.google.com.br/books?id=sxmPx07rWscC>.
- Langrock, Roland, Ruth King, Jason Matthiopoulos, Len Thomas, Daniel Fortin, and Juan M. Morales. 2012. "Flexible and Practical Modeling of Animal Telemetry Data: Hidden Markov Models and Extensions." *Ecology* 93 (11): 2336–42. <https://doi.org/10.1890/11-2241.1>.
- Leise, Tanya L. 2017. "Analysis of Nonstationary Time Series for Biological Rhythms Research." *Journal of Biological Rhythms* 32 (3): 187–94. <https://doi.org/10.1177/0748730417709105>.
- Leos-Barajas, Vianey, Theoni Photopoulou, Roland Langrock, Toby A. Patterson, Yuuki Y. Watanabe, Megan Murgatroyd, and Yannis P. Papastamatio. 2017. "Analysis of Animal Accelerometer Data Using Hidden Markov Models." Edited

by Robert B. O'Hara. *Methods in Ecology and Evolution* 8 (2): 161–73. <https://doi.org/10.1111/2041-210X.12657>.

Levine, Joel D, Pablo Funes, Harold B Dowse, and Jeffrey C Hall. 2002. "Signal Analysis of Behavioral and Molecular Cycles." *BMC Neuroscience*, 25.

Levy, Ofir, Tamar Dayan, and Noga Kronfeld-Schor. 2007. "The Relationship Between the Golden Spiny Mouse Circadian System and Its Diurnal Activity: An Experimental Field Enclosures and Laboratory Study." *Chronobiology International* 24 (4): 599–613. <https://doi.org/10.1080/07420520701534640>.

Malizia, Ana I., and Cristina Busch. 2009. "Breeding Biology of the Fossorial Rodent *Ctenomys Talarum* (Rodentia: Octodontidae)." *Journal of Zoology* 242 (3): 463–71. <https://doi.org/10.1111/j.1469-7998.1997.tb03849.x>.

Massot, Bertrand, S'ebastien Arthaud, Baptiste Barrillot, Johanna Roux, Gianina Ungurean, Pierre-Herv'e Luppi, Niels C. Rattenborg, and Paul-Antoine Libourel. 2019. "ONEIROS, a New Miniature Standalone Device for Recording Sleep Electrophysiology, Physiology, Temperatures and Behavior in the Lab and Field." *Journal of Neuroscience Methods*, Methods and models in sleep research: A Tribute to Vincenzo Crunelli, 316 (March): 103–16. <https://doi.org/10.1016/j.jneumeth.2018.08.030>.

McClintock, Brett T., Roland Langrock, Olivier Gimenez, Emmanuelle Cam, David L. Borchers, Richard Glennie, and Toby A. Patterson. 2020. "Uncovering Ecological State Dynamics with Hidden Markov Models." Edited by Tim Coulson. *Ecology Letters* 23 (12): 1878–1903. <https://doi.org/10.1111/ele.13610>.

McClintock, Brett T, and Theo Michelot. 2021. "momentuHMM: R Package for Analysis of Telemetry Data Using Generalized Multivariate Hidden Markov Models of Animal Movement," 155.

Mendes, Calebe P., Daiane Carreira, Felipe Pedrosa, Gabrielle Beca, La'is Lautenschlager, Paula Akkawi, William Bercê, Katia M. P. M. B. Ferraz, and Mauro Galetti.

2020. "Landscape of Human Fear in Neotropical Rainforest Mammals." *Biological Conservation* 241 (January): 108257. <https://doi.org/10.1016/j.biocon.2019.108257>.

Michelot, Theo, and Roland Langrock. 2019. "A Short Guide to Choosing Initial Parameter Values for the Estimation in moveHMM." <https://cran.r-project.org/web/packages/moveHMM/vignettes/moveHMM-starting-values.pdf>.

Moore-Ede, M. C., F. M. Sulzman, and C. A. Fuller. 1982. *The Clocks That Time Us: Physiology of the Circadian Timing System*. A Commonwealth Fund Book. Harvard University Press. <https://books.google.com.br/books?id=W3RZPgAACAAJ>.

Morgan, Elfed. 2004. "Ecological Significance of Biological Clocks." *Biological Rhythm Research* 35 (1-2): 3–12. <https://doi.org/10.1080/09291010412331313205>.

Naef-Daenzer, B. 2005. "Miniaturization (0.2 g) and Evaluation of Attachment Techniques of Telemetry Transmitters." *Journal of Experimental Biology* 208 (21): 4063–68. <https://doi.org/10.1242/jeb.01870>.

Papastamatiou, Yannis P., Yuuki Y. Watanabe, Urška Demšar, Vianey Leos-Barajas, Darcy Bradley, Roland Langrock, Kevin Weng, Christopher G. Lowe, Alan M. Friedlander, and Jennifer E. Caselle. 2018. "Activity Seascapes Highlight Central Place Foraging Strategies in Marine Predators That Never Stop Swimming." *Movement Ecology* 6 (1): 9. <https://doi.org/10.1186/s40462-018-0127-3>.

Paruelo, Jos'e M., Esteban G. Jobb'agy, and Osvaldo E. Sala. 2001. "Current Distribution of Ecosystem Functional Types in Temperate South America." *Ecosystems* 4 (7): 683–98. <https://doi.org/10.1007/s10021-001-0037-9>.

Patterson, Allison, Hugh Grant Gilchrist, Lorraine Chivers, Scott Hatch, and Kyle Elliott. 2019. "A Comparison of Techniques for Classifying Behavior from Accelerometers for Two Species of Seabird." *Ecology and Evolution* 9 (6): 3030–45. <https://doi.org/10.1002/ece3.4740>.

Patterson, Toby A., Marinelle Basson, Mark V. Bravington, and John S. Gunn. 2009. "Classifying Movement Behaviour in Relation to Environmental Conditions Using Hidden Markov Models." *Journal of Animal Ecology* 78 (6): 1113–23. <https://doi.org/10.1111/j.1365-2656.2009.01583.x>.

Pittendrigh, C S. 1993. "Temporal Organization: Reflections of a Darwinian Clock-Watcher." *Annual Review of Physiology* 55 (1): 17–54. <https://doi.org/10.1146/annurev.ph.55.030193.000313>.

Pohle, Jennifer, Roland Langrock, Floris M. van Beest, and Niels Martin Schmidt. 2017. "Selecting the Number of States in Hidden Markov Models: Pragmatic Solutions Illustrated Using Animal Movement." *Journal of Agricultural, Biological and Environmental Statistics* 22 (3): 270–93. <https://doi.org/10.1007/s13253-017-0283-8>.

Qasem, Lama, Antonia Cardew, Alexis Wilson, Iwan Griffiths, Lewis G. Halsey, Emily L. C. Shepard, Adrian C. Gleiss, and Rory Wilson. 2012. "Tri-Axial Dynamic Acceleration as a Proxy for Animal Energy Expenditure; Should We Be Summing Values or Calculating the Vector?" *PLOS ONE* 7 (2): e31187. <https://doi.org/10.1371/journal.pone.0031187>.

R Core Team. 2020. "R: A Language and Environment for Statistical Computing." <https://www.R-project.org/>.

Rattenborg, Niels C., Bryson Voirin, Sebastian M. Cruz, Ryan Tisdale, Giacomo Dell'NAOMO, Hans-Peter Lipp, Martin Wikelski, and Alexei L. Vyssotski. 2016. "Evidence That Birds Sleep in Mid-Flight." OriginalPaper. *Nature Communications* 7 (1, 1): 1–9. <https://doi.org/10.1038/ncomms12468>.

Rattenborg, Niels C., Bryson Voirin, Alexei L. Vyssotski, Roland W. Kays, Kamiel Spoelstra, Franz Kuemmeth, Wolfgang Heidrich, and Martin Wikelski. 2008. "Sleeping Outside the Box: Electroencephalographic Measures of Sleep in Sloths Inhabiting a Rainforest." *Biology Letters* 4 (4): 402–5. <https://doi.org/10.1098/rbl.2008.0530>.

rsbl.2008.0203.

Ripperger, Simon P., Gerald G. Carter, Niklas Duda, Alexander Koelpin, Björn Cassens, Rüdiger Kapitza, Darija Josic, Jineth Berr'io-Mart'inez, Rachel A. Page, and Frieder Mayer. 2019. "Vampire Bats That Cooperate in the Lab Maintain Their Social Networks in the Wild." *Current Biology* 29 (23): 4139–4144.e4. <https://doi.org/10.1016/j.cub.2019.10.024>.

Ripperger, Simon P., Gerald G. Carter, Rachel A. Page, Niklas Duda, Alexander Koelpin, Robert Weigel, Markus Hartmann, et al. 2019. "Thinking Small: Next-Generation Sensor Networks Close the Size Gap in Vertebrate Biologging." *bioRxiv*, September, 767749. <https://doi.org/10.1101/767749>.

Ripperger, Simon, Linus Günther, Hanna Wieser, Niklas Duda, Martin Hierold, Björn Cassens, Rüdiger Kapitza, Alexander Koelpin, and Frieder Mayer. 2019. "Proximity Sensors on Common Noctule Bats Reveal Evidence That Mothers Guide Juveniles to Roosts but Not Food." *Biology Letters* 15 (2): 20180884. <https://doi.org/10.1098/rsbl.2018.0884>.

Ropert-Coudert, Yan, and Rory P. Wilson. 2005. "Trends and Perspectives in Animal-Attached Remote Sensing." *Frontiers in Ecology and the Environment* 3 (8): 437–44. [https://doi.org/10.1890/1540-9295\(2005\)003%5B0437:TAPIAR%5D2.0.CO;2](https://doi.org/10.1890/1540-9295(2005)003%5B0437:TAPIAR%5D2.0.CO;2).

Rowcliffe, J. Marcus, Roland Kays, Bart Kranstauber, Chris Carbone, and Patrick A. Jansen. 2014. "Quantifying Levels of Animal Activity Using Camera Trap Data." *Methods in Ecology and Evolution* 5 (11): 1170–79. <https://doi.org/10.1111/2041-210X.12278>.

Ruf, T. 1999. "The Lomb-Scargle Periodogram in Biological Rhythm Research: Analysis of Incomplete and Unequally Spaced Time-Series." *Biological Rhythm Research* 30 (2): 178–201. <https://doi.org/10.1076/brrm.30.2.178.1422>.

Rutz, Christian, and Graeme C. Hays. 2009. "New Frontiers in Biologging Science." *Biology Letters* 5 (3): 289–92. <https://doi.org/10.1098/rsbl.2009.0089>.

Shepard, Elc, Rp Wilson, Lg Halsey, F Quintana, A Gómez Laich, Ac Gleiss, N Liebsch, Ae Myers, and B Norman. 2008. "Derivation of Body Motion via Appropriate Smoothing of Acceleration Data." *Aquatic Biology* 4 (December): 235–41. <https://doi.org/10.3354/ab00104>.

Shkolnik, A. 1971. "Diurnal Activity in a Small Desert Rodent." *International Journal of Biometeorology* 15 (2-4): 115–20. <https://doi.org/10.1007/BF01803884>.

Signer, Claudio, Thomas Ruf, Franz Schober, Gerhard Fluch, Thomas Paumann, and Walter Arnold. 2010. "A Versatile Telemetry System for Continuous Measurement of Heart Rate, Body Temperature and Locomotor Activity in Free-Ranging Ruminants." *Methods in Ecology and Evolution / British Ecological Society* 1 (1): 75–85. <https://doi.org/10.1111/j.2041-210X.2009.00010.x>.

Silvério, Jefferson T., and Patricia Tachinardi. 2020. "Chronobiology in the Wild: Toolkit to Study Daily Rhythms in Free-Living Animals." *Sleep Science*. <https://doi.org/10.5935/1984-0063.20200020>.

Tachinardi, Patricia, Jos'e Eduardo Wilken Bicudo, Gisele Akemi Oda, and Ver'onica Sandra Valentinuzzi. 2014. "Rhythmic 24 h Variation of Core Body Temperature and Locomotor Activity in a Subterranean Rodent (*Ctenomys* Aff. *Knighti*), the Tuco-Tuco." Edited by Ralph E. Mistlberger. *PLoS ONE* 9 (1): e85674. <https://doi.org/10.1371/journal.pone.0085674>.

Tachinardi, Patricia, Øivind Tøien, Veronica S. Valentinuzzi, C. Loren Buck, and Gisele A. Oda. 2015. "Nocturnal to Diurnal Switches with Spontaneous Suppression of Wheel-Running Behavior in a Subterranean Rodent." Edited by Eric M Mintz. *PLOS ONE* 10 (10): e0140500. <https://doi.org/10.1371/journal.pone.0140500>.

- Tachinardi, Patricia, Verónica S. Valentiniuzzi, Gisele A. Oda, and C. Loren Buck. 2017. "The Interplay of Energy Balance and Daily Timing of Activity in a Subterranean Rodent: A Laboratory and Field Approach." *Physiological and Biochemical Zoology* 90 (5): 546–52. <https://doi.org/10.1086/693003>.
- Tøien, Øivind, John Blake, Dale M. Edgar, Dennis A. Grahn, H. Craig Heller, and Brian M. Barnes. 2011. "Hibernation in Black Bears: Independence of Metabolic Suppression from Body Temperature." *Science* 331 (6019): 906–9. <https://doi.org/10.1126/science.1199435>.
- Tomotani, Barbara M., Danilo E. F. L. Flores, Patrícia Tachinardi, José D. Paliza, Gisele A. Oda, and Verônica S. Valentiniuzzi. 2012. "Field and Laboratory Studies Provide Insights into the Meaning of Day-Time Activity in a Subterranean Rodent (*Ctenomys Aff. Knighti*), the Tuco-Tuco." Edited by Ralph E. Mistlberger. *PLoS ONE* 7 (5): e37918. <https://doi.org/10.1371/journal.pone.0037918>.
- Valentiniuzzi, Verónica Sandra, Gisele Akemi Oda, John Fontenele Araújo, and Martin Roland Ralph. 2009. "Circadian Pattern of Wheel-Running Activity of a South American Subterranean Rodent (*Ctenomys Cf Knightii*)."*Chronobiology International* 26 (1): 14–27. <https://doi.org/10.1080/07420520802686331>.
- van de Kerk, Madelon, David P. Onorato, Marc A. Criffield, Benjamin M. Bolker, Ben C. Augustine, Scott A. McKinley, and Madan K. Oli. 2015. "Hidden Semi-Markov Models Reveal Multiphasic Movement of the Endangered Florida Panther." Edited by Tim Coulson. *Journal of Animal Ecology* 84 (2): 576–85. <https://doi.org/10.1111/1365-2656.12290>.
- van der Vinne, Vincent, Patricia Tachinardi, Sjaak J. Riede, Jildert Akkerman, Jamey Scheepe, Serge Daan, and Roelof A. Hut. 2019. "Maximising Survival by Shifting the Daily Timing of Activity." *Ecology Letters* 22 (12): 2097–2102. <https://doi.org/10.1111/ele.13404>.

- van Rosmalen, Laura, and Roelof A. Hut. 2021. "Negative Energy Balance Enhances Ultradian Rhythmicity in Spring-Programmed Voles." *Journal of Biological Rhythms* 36 (4): 359–68. <https://doi.org/10.1177/07487304211005640>.
- Vyssotski, Alexei L., Andrei N. Serkov, Pavel M. Itskov, Giacomo Dell'Omo, Alexander V. Latanov, David P. Wolfer, and Hans-Peter Lipp. 2006. "Miniature Neurologgers for Flying Pigeons: Multichannel EEG and Action and Field Potentials in Combination With GPS Recording." *Journal of Neurophysiology* 95 (2): 1263–73. <https://doi.org/10.1152/jn.00879.2005>.
- Weinert, D., and J. Waterhouse. 1998. "Diurnally Changing Effects of Locomotor Activity on Body Temperature in Laboratory Mice." *Physiology & Behavior* 63 (5): 837–43. [https://doi.org/10.1016/s0031-9384\(97\)00546-5](https://doi.org/10.1016/s0031-9384(97)00546-5).
- Wikelski, Martin, and Steven J. Cooke. 2006. "Conservation Physiology." *Trends in Ecology & Evolution* 21 (1): 38–46. <https://doi.org/10.1016/j.tree.2005.10.018>.
- Williams, Cory T., Michael J. Sheriff, Joel A. Schmutz, Franziska Kohl, Øivind Tøien, C. Loren Buck, and Brian M. Barnes. 2011. "Data Logging of Body Temperatures Provides Precise Information on Phenology of Reproductive Events in a Free-Living Arctic Hibernator." *Journal of Comparative Physiology B* 181 (8): 1101–9. <https://doi.org/10.1007/s00360-011-0593-z>.
- Williams, Cory T., Kathryn Wilsterman, Amanda D. Kelley, André R. Breton, Herbert Stark, Murray M. Humphries, Andrew G. McAdam, Brian M. Barnes, Stan Boutin, and C. Loren Buck. 2014. "Light Loggers Reveal Weather-Driven Changes in the Daily Activity Patterns of Arboreal and Semifossorial Rodents." *Journal of Mammalogy* 95 (6): 1230–39. <https://doi.org/10.1644/14-MAMM-A-062>.
- Williams, Cory T., Kathryn Wilsterman, Victor Zhang, Jeanette Moore, Brian M. Barnes, and C. Loren Buck. 2016. "The Secret Life of Ground Squirrels: Accelerometry Reveals Sex-Dependent Plasticity in Above-Ground Activity." *Royal*

Society Open Science 3 (9): 160404. <https://doi.org/10.1098/rsos.160404>.

Willmer, Pat, G. Stone, and Ian A. Johnston. 2005. *Environmental Physiology of Animals*. 2nd ed. Malden, Mass: Blackwell Pub.

Wilmers, Christopher C., Barry Nickel, Caleb M. Bryce, Justine A. Smith, Rachel E. Wheat, and Veronica Yovovich. 2015. "The Golden Age of Bio-Logging: How Animal-Borne Sensors Are Advancing the Frontiers of Ecology." *Ecology* 96 (7): 1741–53. <https://doi.org/10.1890/14-1401.1>.

Zucchini, Walter, Iain MacDonald, and Roland Langrock. 2016. *Hidden Markov Models for Time Series - An Introduction Using R*. Vol. 43.