

Advanced decision modelling in the context of Health Technology Assessment

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Contents

Introduction	5
Why reimbursement submissions fail?	5
Topics of the course	6
Statistical computing	6
Evaluation	6
Bibliography	8
1 What is HTA?	9
1.1 Pre-session readings	9
1.2 Definition and rationale	9
1.3 HTA process	10
1.4 Exercises	14
2 Introduction to decision-analytic models	15
2.1 Pre-session readings	15
2.2 Economic evaluation	16
2.3 Decision modelling	22
2.4 Exercises	24
3 Good practices in decision modelling and decision-tree models	25
3.1 Pre-session readings	25
3.2 Good practices in decision modelling	26
3.3 Decision-tree models	27

4	Markov models	31
4.1	Pre-session readings	31
4.2	Introduction	31
4.3	Best practices	32
5	Evidence synthesis	35
5.1	Pre-session readings	35
5.2	Identifying, aggregating and appraising the evidence	35
6	Uncertainty and decision-making	47
6.1	Pre-session readings	47
6.2	Introduction	47
6.3	Sources of uncertainty in cost-effectiveness models	48
6.4	Variability, heterogeneity and uncertainty	49
6.5	How to deal with uncertainty?	49
	Assignments	55
	Assignment 1	55
	Assignment 2	55
	Introduction to R	57
	Outline	57
	Babies Dataset	57
	Functions	63
	Working with more than one dataset	71
	Matrices	72
	Lists	73

Introduction

Bringing a new health technology to market and into the hands of a patient is a long process. Most of the times patients, who have a medical need, ask themselves why does it take so long to make the health technology available to everyone. When a health technology is in the market, it usually took between 5 to 10 years to make it available.

Depending on the country, governments usually are involved in the reimbursement process. They usually ask the next questions when a new health technology is available:

- How much does it cost?
- Will it save lives and/or improve quality of life?
- Do we have enough budget to fund it?
- If we have a pool of interventions for a specific disease, which one/ones should we reimburse?

Moreover, physicians, patients, insurance plans, and advocacy groups play an important role when new technologies are available in the market (why?). Even though a new technology see the light (i.e. it has proved to be safe and effective), insurance providers or the government will not necessarily cover it. Usually they argue that the new technology is “Not cost-effective” or “Not have good value for money”. *These notes aim to provide all the necessary tools to decide if a new intervention has a good value-for-money.* It is important to stress that value-for-money decision is only one of many questions that are asked by one of the users of a **health technology assessment (HTA)**: patients, healthcare workers, government, and others.

Why reimbursement submissions fail?

According to Goeree (2015), the reasons for rejection are:

1. Inappropriate comparator. Lack of proper statistical analysis.

2. Inappropriate outcome. Use of surrogates.
3. Inappropriate analysis. Lack of robust evidence for costs and quality of life.
4. High cost to the government.

Topics of the course

1. What is HTA?
2. Introduction to decision-analytic models
3. Good practices in decision modelling
4. Evidence-based medicine
5. Decision tree-models
6. State-transition models with the Markov assumption
7. Partitioned survival models
8. Microsimulation
9. Discrete-event simulation
10. Uncertainty and decision-making
11. Presentation of results

Statistical computing

The use of open-source programming languages, such as **R**, in health decision sciences is growing and has the potential to facilitate model transparency, reproducibility, and shareability. However, realizing this potential can be challenging. Models are complex and primarily built to answer a research question, with model sharing and transparency relegated to being secondary goals. Moreover, many decision modelers are not formally trained in computer programming and may lack good coding practices, further compounding the problem of model transparency. **Therefore, throughout this course, the programming language R will be used to show its potential for advanced modelling in the context of HTA.**

For this course, we will be using the book “R for Data Science”. To install **R** and **Rstudio**, instructions are provided in Chapter 1 of this book. We will also use Excel throughout this course.

Evaluation

Item	Percentage	Due date
Assignment 1	15%	Dec 5, 2021

Item	Percentage	Due date
Assignment 2	15%	jan 13, 2021
Take-home exam	30%	Jan 7, 2022
Project proposal	5%	Dec 2, 2021
Project presentation	5%	Jan 14, 2022
Final project	30%	Jan 17, 2022

The intent is to allow the students to demonstrate their mastery of this class through the following way. **Project proposal, presentation and final project will be done in pairs.**

Asssignments

The assignments are handed out approximately two weeks prior to the due date. Late work will not be marked, with the exception of an advance permission from the instructor.

Project proposal

(1 page)

The final deliverable for this course is a mini-HTA on a medical technology (preferably something topical), with a focus on the quantitative aspect of it. Given that the translation of a health policy question into a relevant research question is an essential first step in the conduct of HTA, students are required to formulate a research question and submit for grading purposes. This should include at least some of the following: an overview of the technology being assessed; a clear specification of the policy problem; and the research question(s) (including PICO) with objectives.

Project presentation

(20 minutes with extra 5 minutes for questions)

Students will be expected to present their final course paper and answer questions. Student will be graded on their presentations.

Final project

(20 pages double-spaced)

The main assignment will require students to produce a scaled down HTA, with a focus on the quantitative aspect of it. The objective of the final project is for the

student to show that they have obtained a clear understanding of the advanced methods in decision modelling in the context of HTA. More information will be provided throughout the course, but the paper should contain the following:

- a) Background and technology overview
- b) Formulation of the question you are trying to answer through your mini-HTA
- c) Review of the clinical literature
- d) Description of the structure of the model
- e) Description of the function of the model
- f) Results
- g) Conclusions

Bibliography

Briggs, A., Sculpher, M., & Claxton, K. (2006). Decision modelling for health economic evaluation. Oxford University Press.

Gray, A. M., Clarke, P. M., Wolstenholme, J. L., & Wordsworth, S. (2011). Applied methods of cost-effectiveness analysis in healthcare (Vol. 3). Oxford University Press.

Edlin, R., McCabe, C., Hulme, C., Hall, P., & Wright, J. (2015). Cost effectiveness modelling for health technology assessment: a practical course. Springer.

Chapter 1

What is HTA?

1.1 Pre-session readings

Goodman, C. S. (2004). Introduction to health technology assessment. The Lewin Group. virginia, USA. link. Chapters 1, 2, and 5.

Briggs, A., Sculpher, M., & Claxton, K. (2006). Decision modelling for health economic evaluation. Oxford University Press. Chapter 1.

Chapters 1 and 2 of *R for data science*.

1.2 Definition and rationale

The first thing that we need to know is the definition of a **health technology**. A health technology is any intervention that may be used to promote health, to prevent, diagnose or treat disease or for rehabilitation or long-term care.

Questions

1. List some examples of health technologies.

Depending on the agency, health technology assessment has a broad spectrum of definitions:

“HTA is a multidisciplinary process that uses explicit methods to determine the value of a health technology at different points in its lifecycle. The purpose is to inform decision-making in order to promote an equitable, efficient, and high-quality health system.” INAHTA

“Health technology assessment is a multidisciplinary process that uses explicit methods to determine the value of a health technology at different points in its

lifecycle. The purpose is to inform decision-making in order to promote an equitable, efficient, and high-quality health system.” EUnetHTA

“A comprehensive, objective, evidence-based analysis of the clinical effectiveness, cost-effectiveness and broader impact of drugs, medical technologies and health systems. HTA examines technologies at all stages of their life cycle, from development through to maturity and obsolescence.” CADTH

The purpose of HTA is to support/help decision makers by identifying technologies that will improve health outcomes and deliver value for every dollar invested.

- Does a new health technology offer a clinical advantage over the alternatives/standard approaches?
- Is it worth the investment?
- Can I pay for it?
- Who would benefit from it?
- Any ethical, social or legal issues

But, what are the reasons for conducting HTAs?

- Increased demand for healthcare (why?)
- Soaring healthcare costs
- Increased rate of diffusion of new technologies and associated evidence

Once we have seen the definition and rationale for conducting HTAs, it is important to talk about the potential users.

- Government
- Managers in hospitals
- Healthcare workers
- Researchers

1.3 HTA process

1. Identification and prioritization of technologies
2. Clear specification of the problem
3. Technology assessment and review
 - Evidence and systematic literature review
 - Aggregation and appraisal of evidence
 - Synthesize and consolidate
 - Collect primary data (if necessary)
 - Economic evaluation, budget and health system impact



Figure 1.1: Life expectancy in Mexico. Source: CONAPO

- Assessment of social, legal, and ethical consideration
 - Formulation of finding
4. Dissemination and implementation of recommendations
 5. Monitor the impact of assessment reports

1.3.1 Identification and prioritization of technologies

- Drugs seeking public or private reimbursement
- Variable for non-drug technologies. However candidates:
 - High potential to improve health outcomes, reduce harm or decrease costs with similar efficacy
 - Large numbers of individuals affected
 - Political pressure
 - Unmet needs—no current treatment

1.3.2 Clear specification of the problem

- Problem statements need to consider:
 - Patient population affected (indication; epidemiology)
 - Intervention being considered (drug, device, new/old)
 - Comparators
 - Outcome(s) or interest
 - Setting (e.g. hospital, community)
- Well formulated question

1.3.3 Technology assessment and review

1.3.3.1 Evidence and systematic literature review

- A comprehensive search of the literature based on systematic methods is essential
- 2 main types of resources relevant to HTA:
 - Published literature
 - Grey literature

1.3.3.2 Identification, aggregation & appraisal of evidence

- Objective, systematic process for screening and determine studies to be included in the synthesis
- Classify the studies

- Randomised, non-randomised and economic
- Critical appraisal of the quality of the evidence

1.3.3.3 Synthesize & consolidate

- Findings from multiple studies often combined to respond to the HTA question
- Techniques commonly used to synthesize data in HTA are:
 - Meta-analysis, meta-regression
 - Network meta-analysis

1.3.3.4 Economic evaluation

- Measures the incremental costs and benefits of the technology under review compared to one or more relevant technologies
- CEA, CUA and CBA
- Budget impact

1.3.3.5 Assessment of social, legal & ethical considerations

- Example: genetic information (why?)
- Any access or equity issues following the dissemination and implementation of technologies?

1.3.3.6 Formulation of findings

- Explicitly link quality of the evidence to the strength of findings and recommendations as well as any limitations
- Recommendations based on the findings that reflect the original question(s)

1.3.4 Dissemination of recommendations

- Findings translated into relevant and understandable information
- Knowledge translation

1.3.5 Monitoring the impact of reports

- Some recommendations are translated into policies with clear and quantifiable impacts (e.g. adoption of new technology, change in reimbursement)
- Others go ignored and are not readily adopted into general practice

1.4 Exercises

Read the following HTA published by NICE in the UK. Do the following:

- What is the population?
- What is the intervention and comparators?
- Is there a reproducible search strategy for the clinical evidence in the HTA?
- Was the clinical evidence critically appraised? How?
- Describe the evidence synthesis process
- What type of economic evaluation they used?
- What type of model was used in the economic evaluation?
- How was the uncertainty handled in the economic evaluation?
- Is there a budget impact in the HTA?
- What is the recommendation?

Chapter 2

Introduction to decision-analytic models

2.1 Pre-session readings

Chapter 3 of *R for data science*.

Economic evaluation

Gray, A. M., Clarke, P. M., Wolstenholme, J. L., & Wordsworth, S. (2011). Applied methods of cost-effectiveness analysis in healthcare (Vol. 3). Oxford University Press. Chapter 2.

Birch, S., & Gafni, A. (1992). Cost effectiveness/utility analyses: do current decision rules lead us to where we want to be?. Journal of health economics, 11(3), 279-296. Doubilet, P., Weinstein, M. C., & McNeil, B. J. (1986). Use and misuse of the term “cost effective” in medicine.

Decision modelling

Briggs, A., Sculpher, M., & Claxton, K. (2006). Decision modelling for health economic evaluation. Oxford University Press. Chapter 2. Sections 2.1 and 2.2

Gray, A. M., Clarke, P. M., Wolstenholme, J. L., & Wordsworth, S. (2011). Applied methods of cost-effectiveness analysis in healthcare (Vol. 3). Oxford University Press. Chapter 8. Sections 8.1 to 8.4

Buxton, M. J., Drummond, M. F., Van Hout, B. A., Prince, R. L., Sheldon, T. A., Szucs, T., & Vray, M. (1997). Modelling in economic evaluation: an unavoidable fact of life. Health economics, 6(3), 217-227.

2.2 Economic evaluation

Cost-effectiveness analysis (CEA) and cost-utility analysis (CUA) have been proposed as methods for comparing alternative uses of scarce health-care resources. The difference between CEA and CUA lies in the way outputs are measured.

We can find different objectives of CEA across the literature:

“The underlying premise of cost-effectiveness analysis in health problems is that for any given level of resources available, society (or the decision making jurisdiction involved) wishes to maximize the total aggregate health benefits conferred” Weinstein and Stason (1977).

“For any given rate of output [the combination of inputs] . that costs the decision maker least” Culyer (1980)

“A method of determining the most efficient way of dealing with a specified health problem” Green and Barker (1988)

The goal of CEA is to maximize health benefits produced from a given level of resources. Therefore, it is consistent with welfare economics concept of Pareto efficiency (Birch and Gafni, 1992) (Figure 2.1).

In practice, CEA **relaxes the constraint on available resources**. Because the focus of the evaluation is not a fixed resource pool, but a specific programme making demands on resources, the comparison of the programme under evaluation with an existing programme (e.g., services aimed at improving FS) must consider not only the inter-programme differences in outputs (incremental benefits), but also the inter-programme differences in resources used (incremental costs).

Weinstein and Stason (1977) state that: *“the criterion for cost-effectiveness is the ratio of the net increase of health-care costs to the net effectiveness. The lower the value of this ratio, the higher the priority in terms of maximizing benefits derived from a given health expenditure”*. Issue is that applying a ratio does not lead to the maximization of benefits from a fixed resource pool (movement from A to C in Figure). **The real problem of the application of CEA is the failure to adhere strictly to the notion of opportunity cost in the measurement of the (incremental) cost of a programme.** The existing programme represents a true opportunity cost for the entire resource requirements of the new programme, even though it does not absorb this level of resources.

2.2.1 Cost-effectiveness in practice

As mentioned before, CEA include both costs and effects, which are represented graphically in a plane:

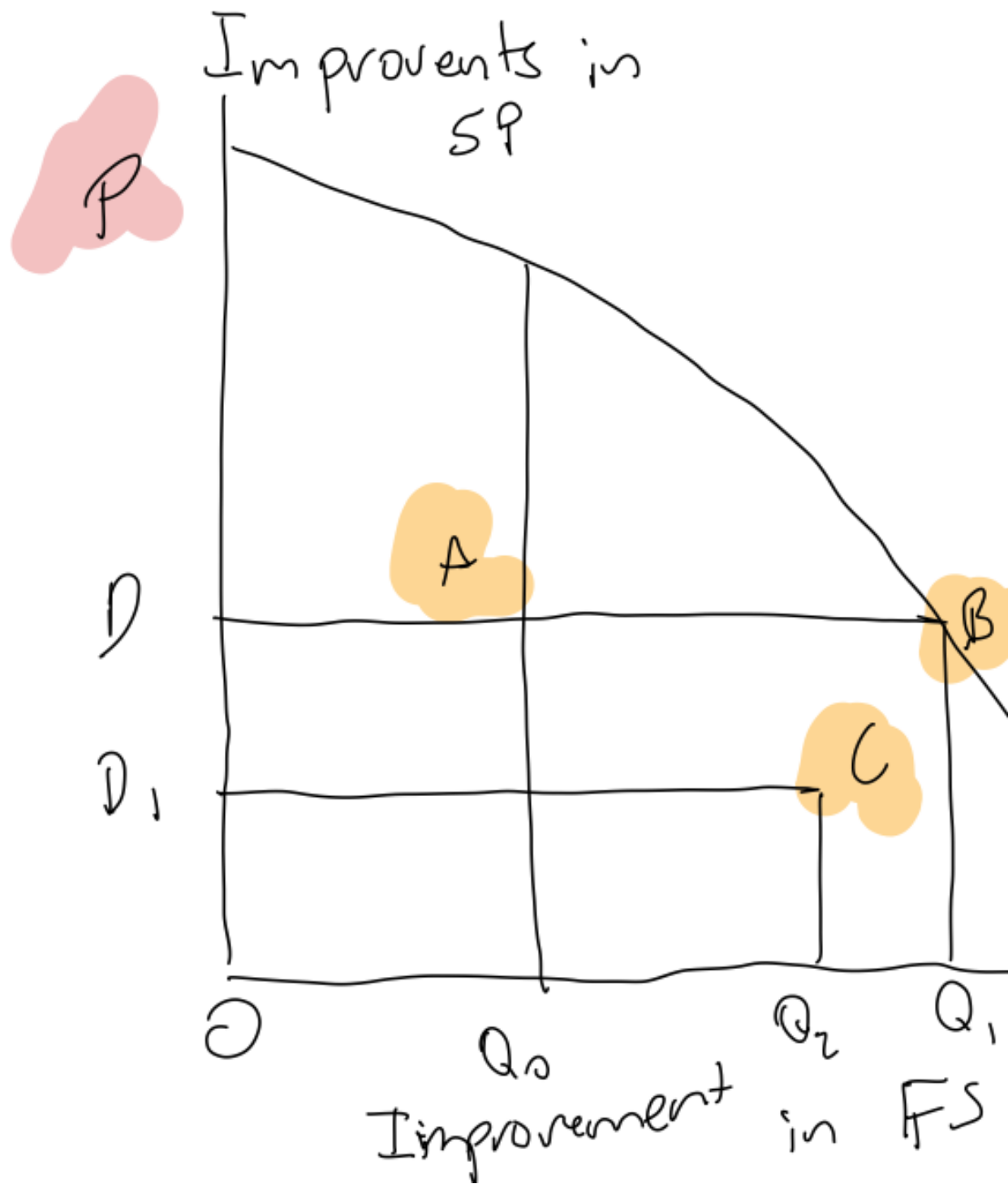


Figure 2.1: Production possibilities frontier

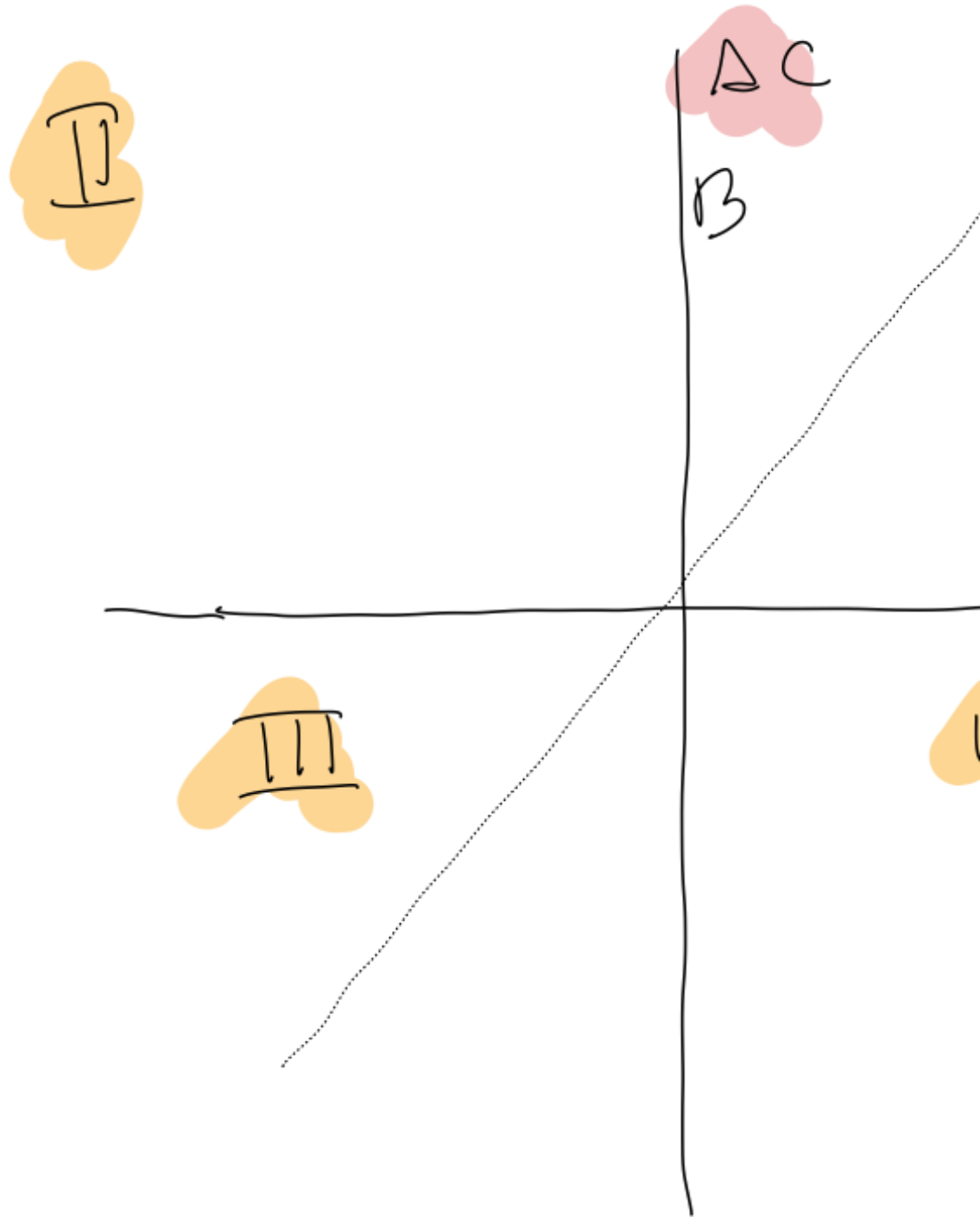


Figure 2.2: Incremental CE plane

Table 2.1: Calculating incremental cost-effectiveness

Option	Cost	QALYs	Incremental cost	Incremental QALY	ICER
A	0				
B	10,000	0.40	10,000	0.40	25,000
C	22,000	0.55	12,000	0.15	80,000
D	25,000	0.50	3,000	-0.05	-60,000
E	40,000	1.00	15,000	0.50	30,000

Questions

1. Describe the plane.
2. What is the willingness-to-pay in this plane?
3. What type of uncertainties are encountered in this plane?

Why incremental? We can see it using the example from Gray et al. (2011) applied to mutually exclusive options (see Figure 2.3):

- Diet and exercise (C): Reference
- Metformin (A): \$500k/250 life-years
- New drug (B): \$2500k/300 life-years

Clearly, as this example shows, it is quite misleading to calculate average cost-effectiveness ratios, as they ignore the alternatives available.

Questions

1. What is the difference between average cost-effectiveness ratios vs incremental?

The idea of CEA is to maximize health benefits with the available resources, which in terms of the CE plane represents pushing as far to the right as possible while moving up the vertical axis as little as possible. The next example from Gray et al. (2011) shows the ideas behind **cost-effectiveness frontier**, **dominance**, **extended dominance** (Table 2.1).

Once we have the ICERs for different independent programmes. How can we maximize health gains with this information? Note that now we are comparing different programmes as opposed to mutually exclusive options. Let's work in the next example:

Finally, we can ask ourselves. What is the maximum value of the incremental cost-effectiveness ratio (λ)?

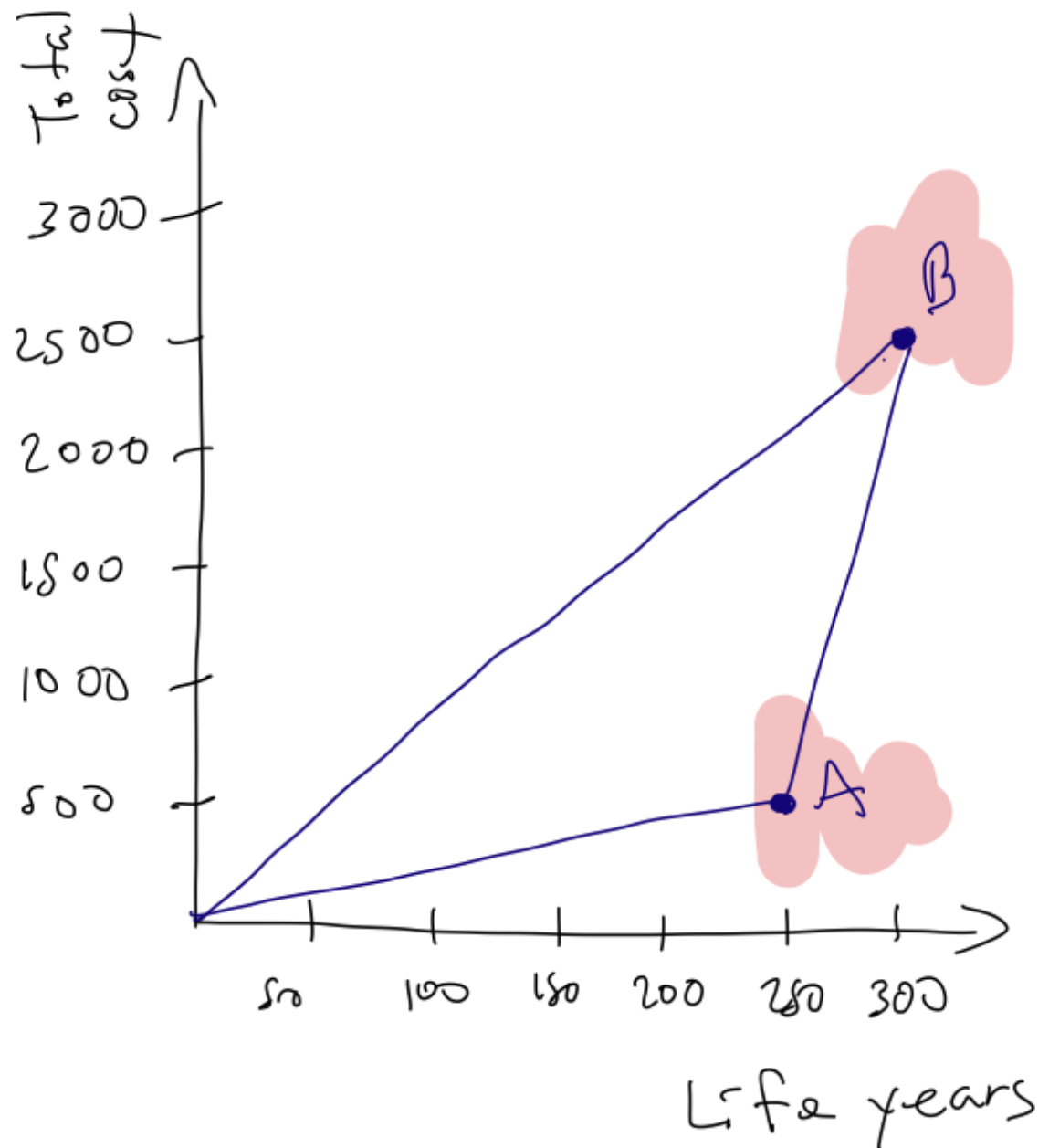


Figure 2.3: Average and incremental cost-effectiveness

Table 2.2: Using cost-effectiveness to maximize health gain

Intervention	Incremental cost (per k)	Incremental QALYs	ICER
1	1,300	165	7,879
2	600	28	21,429
3	750	110	6,818
4	750	13	57,692
5	2,200	75	29,333
6	400	85	4,706
Total	6,000	476	

- Rule-based approaches - Adopting arbitrary thresholds. For example, Laupacis et al. (1992) adopted identical cut-off points of CAN\$20,000 per QALY gained, up to which level they considered that there would be strong grounds for adoption, and CAN\$100,000 per QALY gained, above which they considered that evidence for adoption was weak. An alternative approach is to use the country's gross domestic product (GDP) per capita. Williams et al. (2004) advanced one line of reasoning in support of this, arguing that each person has an entitlement to a 'fair share' of the country's wealth.
- League table approach - If we had full information on the ICERs of all available interventions, it would be possible to rank them by ICER. See the example in Table 2.1.

What are the issues with these approaches?

1. λ is a function of the budget. Therefore it is constantly changing
2. The dynamic nature of λ . As new programs are funded and others replaced, the identification of the last program funded changes.

2.2.2 Conclusions

- Irrespective of whether the problem face by decision-makers is simple (maximizing health gains from available resources) or complex (subject to considerations of equity, accessibility, etc.), if it is not to be considered in the context of a resource constraint there is little use for economics in the way the problem is considered.
- Other approaches have been considered, such as methods in linear programming, which is consistent with the objective of CEA.

$$\max_{x_1, \dots, x_n} \sum_{i=1}^n x_i E_i \text{ s.t. } \sum_{i=1}^n x_i c_i \leq C$$

where x_i is the health intervention presented as a binary outcome (1 or 0), E_i is the present value of the health benefits (measured using QALYs) generated by programme over the planning period, and c_i is the present value of the cost of providing programme i over the planning period.

2.3 Decision modelling

HTA is undertaken in order to inform decision-making regarding the appropriate use of particular healthcare programs and interventions, and involves the synthesis of a range of evidence. Broadly speaking, this implies two important components of this process:

1. Gathering evidence for the disease and technology of concern from a range of primary studies. This includes effectiveness, costs, epidemiology, natural history of the disease, quality of life, etc.
2. Synthesizing the evidence found in the first component in order to inform policy and decision-making.

Because of the nature of these two components, decision-analytic modelling has played an important role in HTA. This is because it represents an explicit approach to synthesizing currently available evidence regarding the effectiveness and costs of alternative (mutually exclusive) healthcare strategies (Philips et al., 2006). Therefore one of the main objectives of decision-analytic modelling is to address the relationship between the effectiveness and costs of alternative healthcare strategies in order to assess relative cost-effectiveness (CE) and to determine which options should be adopted given existing information (Philips et al., 2006). Consequently, modelling in the context of HTA is a typical problem of decision-making under uncertainty.

Traditionally randomised controlled trials (RCTs) have been a key component of many HTA process. This is because randomisation protects against selection bias and confounding. Nevertheless, the information produced by RCTs can be limited with respect to evaluating health care as delivered in the real world. This can be because:

1. Not all interventions to be compared are included in the trial.
2. Not enough follow-up.
3. Not enough flexibility (controlled trial).
4. Small sample sizes.
5. Patients in trial are not representative to the target population.

Consequently, it is advised that HTA submissions incorporate information from as many sources as possible to address some of these problems (Sculpher et al.,

2006). As mentioned before, decision analytical models allow for the synthesis of information across multiple sources and for the comparison of multiple options that might not have been included as part of an RCT.

Briefly, Dahabreh et al. (2016) consider some potential goals of modelling in a health-care context:

- To structure investigators' thinking and to facilitate the communication of data, assumptions, and results.
- To synthesize data from disparate sources.
- To make predictions.
- To support causal explanations.
- To inform decision making.

Moreover, they also distinguish different stages for the development of a decision analytic model

1. Define a question
2. Decide on the type of decision model most appropriate
3. Conceptualize the model (and its mathematical structure).
4. Gather all the evidence required for the model and synthesize it.
5. Implement and run the model.
6. Assess the model.

The development of models, especially those trying to explain complex phenomena and informing difficult decisions, is a demanding task. Choosing between alternative modeling approaches can be difficult because the correct choice would not be obvious at early stages in developing a decision analytical model. In general, modelling is most useful when data have limitations (e.g. non-randomised evidence, sparse evidence, etc.), when the research question is complex and when choices are preference laden (Dahabreh et al., 2016). Another important aspect when choosing between modelling approaches is whether the model in question is likely to show results that the intended audience will consider credible and useful. Multiple iterations are typically needed between the key activities outlined previously because at each activity the need for changes at earlier stages may become evident.

2.3.1 Importance of decision-analytic models in HTA

When doing a cost-effectiveness analysis in the context of HTA, one usually starts conceptualizing the model that will help answer the research question. But what is a model? A model is a simplified representation of reality (Roberts et al., 2012), where inputs from different sources inform it and its purpose, in the context of HTA, is to inform medical decisions and health-related resource allocation questions (Roberts et al., 2012).

Methods for the conduct of decision-analytic modelling have continued evolving to address the ever-increasing information needs of decision makers. The complexity and continued advances of the relevant methods have spurred the publication of recommendation statements on “best practices” for modeling in the context of HTA (Roberts et al., 2012; Briggs et al., 2012). Some of these modelling techniques (Caro et al., 2012) include:

- Decision-tree models.
- State-transition models.
- Micro-simulation models.
- Discrete event simulation (DES) models.
- Dynamic transmission models.

2.4 Exercises

1. Question 1 and 2 from Chapter 2 in *Applied methods of cost-effectiveness analysis in healthcare*.
2. Read the articles in our google classroom.

Chapter 3

Good practices in decision modelling and decision-tree models

3.1 Pre-session readings

Good practices

Roberts, M., Russell, L. B., Paltiel, A. D., Chambers, M., McEwan, P., & Krahn, M. (2012). Conceptualizing a model: a report of the ISPOR-SMDM modeling good research practices task force-2. Medical Decision Making, 32(5), 678-689.

Decision-tree models

Tarride, J. E., Blackhouse, G., Bischof, M., McCarron, E. C., Lim, M., Ferrusi, I. L., ... & Goeree, R. (2009). Approaches for economic evaluations of health care technologies. Journal of the American College of Radiology, 6(5), 307-316.

Briggs, A., Sculpher, M., & Claxton, K. (2006). Decision modelling for health economic evaluation. Oxford University Press. Chapter 2. Sections 2.2 and 2.3.1.

Gray, A. M., Clarke, P. M., Wolstenholme, J. L., & Wordsworth, S. (2011). Applied methods of cost-effectiveness analysis in healthcare (Vol. 3). Oxford University Press. Chapter 8. Sections 8.5 and 8.6.

3.2 Good practices in decision modelling

Best practices according to Roberts et al. (2012):

1. The modeling team should consult widely with subject experts and stakeholders to ensure that the model represents disease processes appropriately and adequately addresses the decision problem.
2. A clear, written statement of the decision problem, modeling objective, and scope should be developed. This should include disease spectrum considered, analytic perspective, target population, alternative interventions, health and other outcomes, and time horizon.
 - A model's scope and structure should be consistent with, and adequate to address, the decision problem and policy context.
 - The analytic perspective should be stated and defined. Outcomes modeled should be consistent with the perspective. Analyses that take a perspective narrower than societal should report which outcomes are included and excluded.
 - The target population should be defined in terms of features relevant to the decision (e.g., geography and patient characteristics, including comorbid conditions, disease prevalence, and stage).
 - Health outcomes, which may be events, cases of disease, deaths, life years gained, quality-adjusted life years, disability-adjusted life years, or other measures important to stakeholders, should be directly relevant to the question being asked.
 - Interventions modeled in the analysis should be clearly defined in terms of frequency, component services, dose or intensity, duration, and any variations required for subgroups and should include standard care and other strategies routinely considered and in use.
3. Although data are essential to a model, the conceptual structure should be driven by the decision problem or research question and not determined by data availability.
 - The choice of comparators crucially affects results and should be determined by the problem, not by data availability or quality. All feasible and practical strategies should be considered. Constraining the range of strategies should be justified
 - The time horizon of the model should be long enough to capture relevant differences in outcomes across strategies. A lifetime time horizon may be required.
4. The problem conceptualization should be used to identify key uncertainties in model structure where sensitivity analyses could inform their impact.
5. The policy context of the model should be clearly stated. This includes the funder, developer, whether the model is for single or multiple application, and the policy audience.

6. An explicit process (expert consultations, influence diagrams, concept mapping, or similar method) should be used to convert the problem conceptualization into an appropriate model structure, ensuring it reflects current disease knowledge and the process modeled.
7. Several model types may be suitable. Some problems are more naturally represented in some types than others.
 - For simple models or problems with special characteristics (e.g., very short time horizons, very few outcomes), a decision tree may be appropriate.
 - If the conceptualization involves representing the disease or treatment process as a series of health states, state-transition models are appropriate. Their primary disadvantage, the Markovian assumption that transition probabilities do not depend on history, can be addressed by increasing the number of states. Individual state-transition models, which do not require this assumption, are an alternative when the number of states grows too large.
 - When the disease or treatment process includes interactions between individuals, the methods should be able to represent those interactions and evaluate their effects.
 - When the problem involves resource constraints, the modeling method should be able to represent them and evaluate their effects.
 - For some problems, combinations of model types, hybrid models, and other modeling methodologies are appropriate.
8. Model simplicity is desirable for transparency, ease of analysis, validation, and description. However, the model must be complex enough to ensure that differences in value (e.g., health or cost) across the strategies considered are faithfully represented. Some degree of model complexity may be desirable to preserve face validity to clinical experts. Greater complexity may be necessary in policy models that are intended to be used for many problems.

3.3 Decision-tree models

A decision-tree, is a branching structure in which each branch represents an event that may take place in the future. Identifying alternatives and specifying the sequence and linkage of events are essential steps in constructing such a model, but are also in themselves of great value in clarifying complex decisions.

3.3.1 Structure and order

A hypothetical policy question concerning whether to introduce a breast cancer screening programme. It can be represented as:

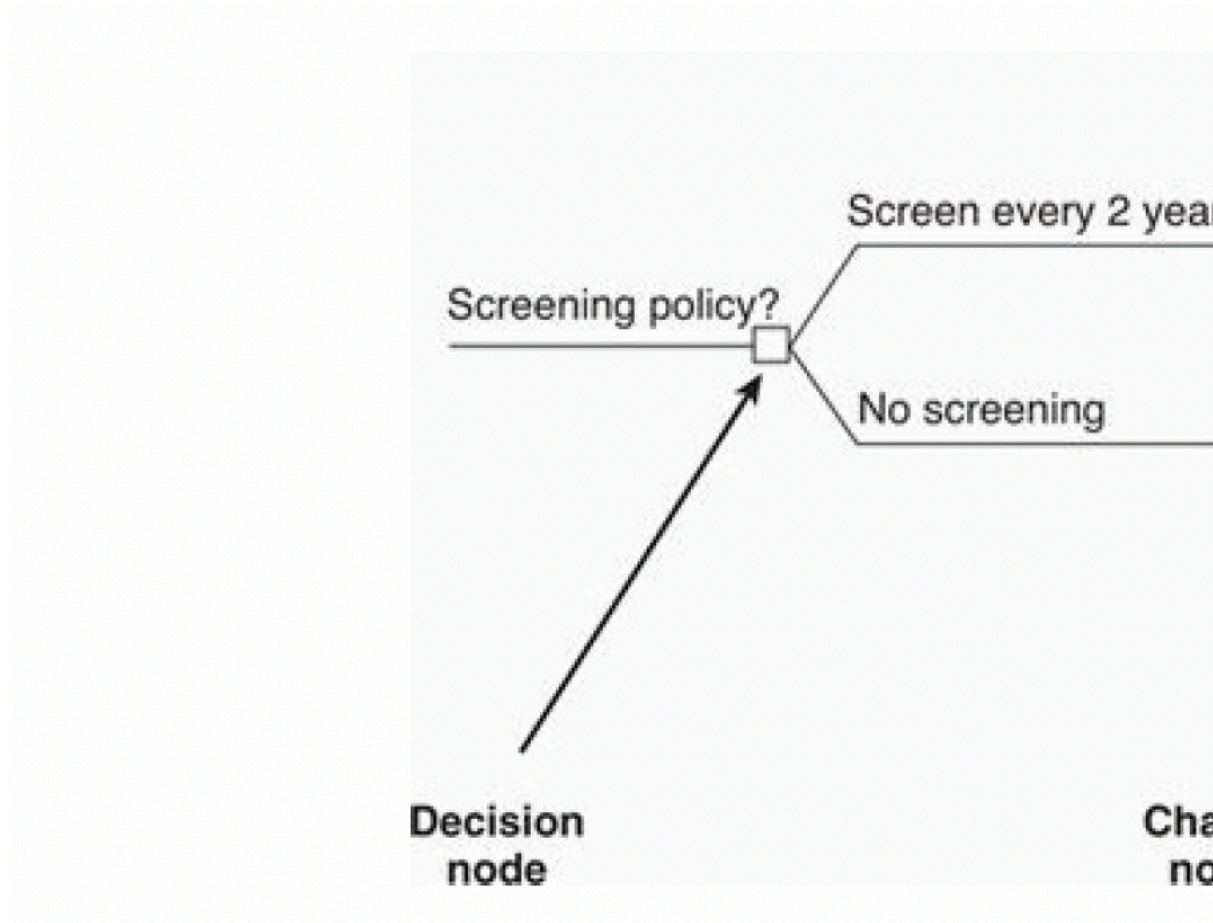


Figure 3.1: Decision problem, decision node, chance nodes, terminal nodes, and branches. Source: Gray et al., 2016

- Decision trees are by convention constructed from left to right, starting with the decision node and ending with the outcomes on the right, and follow the logical structure of the decision problem, usually following the sequence of events over time.
- Moving from left to right along the decision tree, the addition of chance nodes corresponds to the addition of subsequent uncertain events.

In general, the order of the events in the decision tree usually follows the sequence of events over time according to the logical progression of the decision pathway. We can look at the next example:

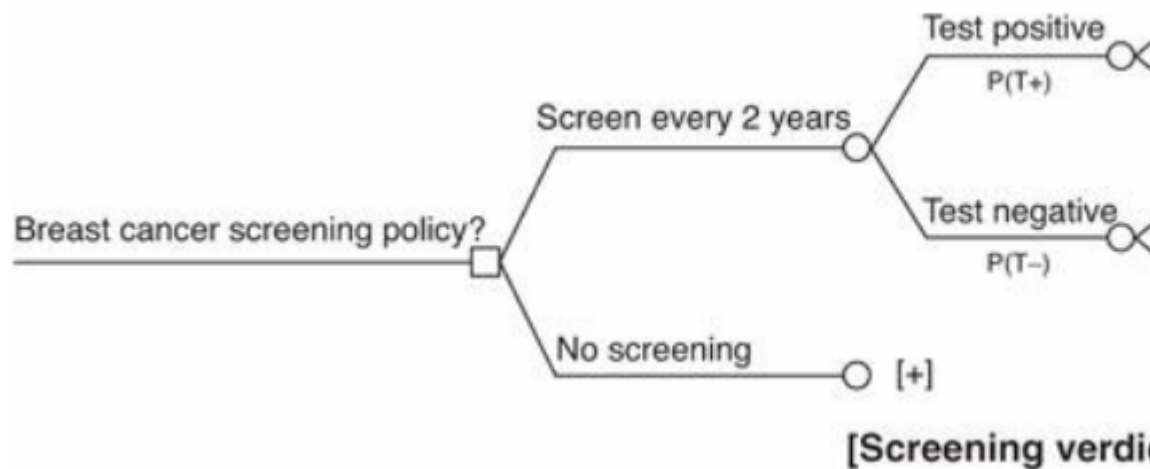


Figure 3.2: Structure of the tree: process ordered Source: Gray et al., 2016

3.3.2 Probabilities in a tree

- Once the structure of the model has been developed, the next step in the process is to start populating the model.
- Probabilities are usually derived from published studies. When there is more than one source of information on a probability, information will need to be synthesized.
- Probabilities (branch probabilities) are entered under the branches emanating from a chance node to represent the probability/likelihood of the uncertain event occurring. Since the events at a chance node must be mutually exclusive and exhaustive, the sum of the probabilities at each chance node must equal 1.

3.3.3 Payoffs

- Payoffs include the costs related to the events in the decision tree and the final outcomes (life-years, utilities, quality-adjusted life-years), and are entered at the terminal node.

3.3.4 Expected value

- The decision tree is ‘averaged out’ and ‘folded back’ (or ‘rolled back’). By folding back the tree, the expected values of each strategy are calculated. The folding-back process starts at the right-hand side of the tree and then averages back.
- The expected value is the sum of products of the estimates of the probability of events occurring and the consequence of the events, their outcomes, or their costs, i.e. the weighted average of the outcome or cost values. This process is conducted separately for costs and effects.

Chapter 4

Markov models

4.1 Pre-session readings

Siebert, U., Alagoz, O., Bayoumi, A. M., Jahn, B., Owens, D. K., Cohen, D. J., & Kuntz, K. M. (2012). State-transition modeling: a report of the ISPOR-SMDM modeling good research practices task force-3. Medical Decision Making, 32(5), 690-700. [link](#)

Briggs, A., Sculpher, M., & Claxton, K. (2006). Decision modelling for health economic evaluation. Oxford University Press. Chapter 2.3.2.

Gray, A. M., Clarke, P. M., Wolstenholme, J. L., & Wordsworth, S. (2011). Applied methods of cost-effectiveness analysis in healthcare (Vol. 3). Oxford University Press. Chapter 9.

4.2 Introduction

Many clinical situations can be described in terms of the conditions that individuals can be in (“states”), how they can move among such states (“transitions”), and how likely such moves are (“transition probabilities”). In these situations, state-transition models (STMs) are often well suited to the decision problem, as they conceptualize it in terms of a set of states and transitions among these states. **This chapter focuses in Markov Models.** An STM is a reasonable choice when the decision problem can be framed in terms of states, the interactions between individuals are not relevant, and the population of interest is a closed cohort.

4.3 Best practices

1. If the decision problem can be represented with a manageable number of health states that incorporate all characteristics relevant to the decision problem, including the relevant history, a cohort simulation should be chosen because of its transparency, efficiency, ease of debugging, and ability to conduct specific value of information analyses. If, however, a valid representation of any aspect of the decision problem would lead to an unmanageable number of states, then an individual-level state-transition model is recommended. Validity should not be sacrificed for simplicity.
2. The strategies being evaluated should be clearly defined. In particular, sequential decisions should not be modeled within the Markov cycle tree but rather be part of the specification of the alternative intervention strategies that precede the Markov tree
3. The starting cohort should be defined by the demographic and clinical characteristics that affect the transition probabilities or state values (e.g., quality of life and cost).
4. Specification of states and transitions should generally reflect the biological/theoretical understanding of the disease or condition being modeled.
5. States should adequately capture the type of intervention (i.e., prevention, screening, diagnostics, treatment) as well as the intervention's benefits and harms.
6. States need to be homogeneous with respect to both observed and unobserved (i.e., not known by the decision maker) characteristics that affect transition probabilities.
7. The time horizon for the model should be sufficiently large to capture all health effects and costs relevant to the decision problem.
8. Cycle length should be short enough to represent the frequency of clinical events and interventions.
9. Components of state-transition models that reflect similar clinical courses should not be recreated but rather should be incorporated once and linked to that structure throughout the model.
10. Transition probabilities and intervention effects should be derived from the most representative data sources for the decision problem.
11. All methods and assumptions used to derive transition probabilities and intervention effects should be described.

12. All parameters relating to the effectiveness of interventions derived from observational studies should be correctly controlled for confounding. Time-varying confounding is of particular concern in estimating intervention effects.
13. The valuation of intermediate outcomes/states should be justified.
14. A half-cycle correction should be applied to costs and effectiveness in the first cycle and in the final cycle if not using a lifetime horizon.
15. For certain decision problems, it may be important to report not only the expected value but also the distribution of the outcomes of interest.
16. The number of individuals simulated should be large enough to generate stable estimates of the expected values.
17. The report should use nontechnical language and clear figures and tables that enhance understanding of the STM to communicate its key structural elements, assumptions, and parameters.
18. In addition to final outcomes, intermediate outcomes that enhance the understanding and transparency of the model results should also be presented.

Chapter 5

Evidence synthesis

5.1 Pre-session readings

Murad, M. H., Montori, V. M., Ioannidis, J. P., Jaeschke, R., Devereaux, P. J., Prasad, K., ... & Guyatt, G. (2014). How to read a systematic review and meta-analysis and apply the results to patient care: users' guides to the medical literature. Jama, 312(2), 171-179. link

Schulz, K. F., Altman, D. G., Moher, D., & CONSORT Group. (2010). WITHDRAWN: CONSORT 2010 statement: Updated guidelines for reporting parallel group randomised trials. link

5.2 Identifying, aggregating and appraising the evidence

5.2.1 Identifying the evidence

- Comprehensive search following accepted search practices will help to:
 - avoid missing relevant studies
 - avoid other potential biases (publication, time lag, language)
 - provide detailed search documentation (aid transparency and increase confidence)

It is impossible to know if you have created the ‘perfect’ strategy and impossible to know if you have captured all the relevant evidence. The goal is to goal should instead be to search as comprehensively as is feasible, providing a

balance between sensitivity & specificity. One of the best sources for conducting a systematic review is the Cochrane Handbook for Systematic Reviews of Interventions.

Prior to the search process the following needs to be clarified:

- type of research question
- HTA components required
- type of studies required
- Databases

At protocol stage – scoping searching all occurs prior to protocol development so by protocol finalization, search shouldn't change after this. Search strategy should be described in report in such detail that it can be replicated and updated. You should include:

- Sources searched
- Search strategy used for at least one database
- Time periods of the search
- Any limits used

We can consider two types of literature: published and gray literature. It is important to know that an optimal search strategy will not necessarily identify all relevant items. The primary source of published literature are **bibliographic databases**, such as Medline and Embase. Other sources include:

- Conference abstracts & proceedings
- Electronically available full-text journals & report
- Reference lists & citation tracking
- Google

Hints

1. Check through bibliographic references of relevant review articles and/or trials to find references to other potentially relevant trials
2. Tracking the citations of key studies/papers.

5.2.2 How to search

Identify key topics and translate into a clearly focused question. Once this is we can create our PICO:

- P - Population

5.2. IDENTIFYING, AGGREGATING AND APPRAISING THE EVIDENCE³⁷

- I - Intervention
- C - Comparator
- O - Outcome
- S - Study type

Look at MeSH headings, words in title, abstract or other potentially relevant fields. Medical Subject Headings (MeSH) is a comprehensive controlled vocabulary for the purpose of indexing journal articles and books in the life sciences. Using only the controlled vocabulary that someone else deemed as right for an article is risky (e.g. MeSH). We need to use “free text” or keyword terms to ensure you find all your articles. Free text search words will find the word anywhere you tell it to. Moreover, consider combining body of search with study “filter” in order to restrict to a given study type.

It is crucial to **document your searches**, it allows other researchers to replicate your study. You need to provide details of which databases and interface you searched. Write down how many results you had from each database for inclusion in the Description of Studies or Results section, as well as a PRISMA flow chart.

Once you run your search, it is important to use bibliographic software (End-Note, Zotero, etc.)

5.2.3 Aggregating and appraising the evidence

We are generating new evidence, therefore we need to use systematic approaches which are documented, transparent and reproducible.

Systematic literature review initial steps:

- Aggregate
- Abstract
- Appraise

5.2.3.1 Aggregate

- Multi-stage documented screening process using pre-defined selection criteria
- Recommended that this be completed by at least 2 individuals independently
- Using the a priori criteria

For reporting tools, follow PRISMA. Finally, the evidence can be categorized in different levels:

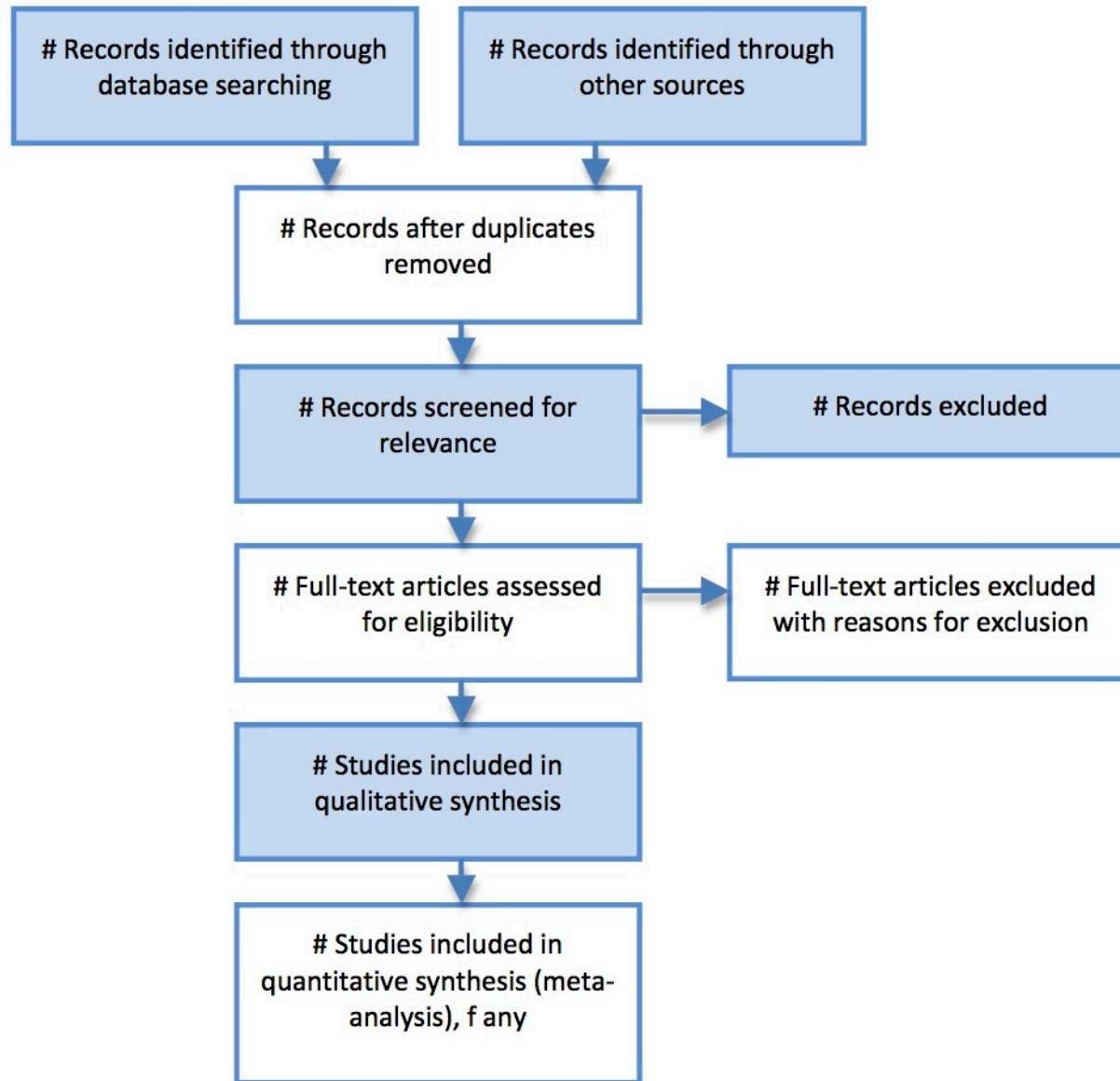


Figure 5.1: Flowchart PRISMA diagram

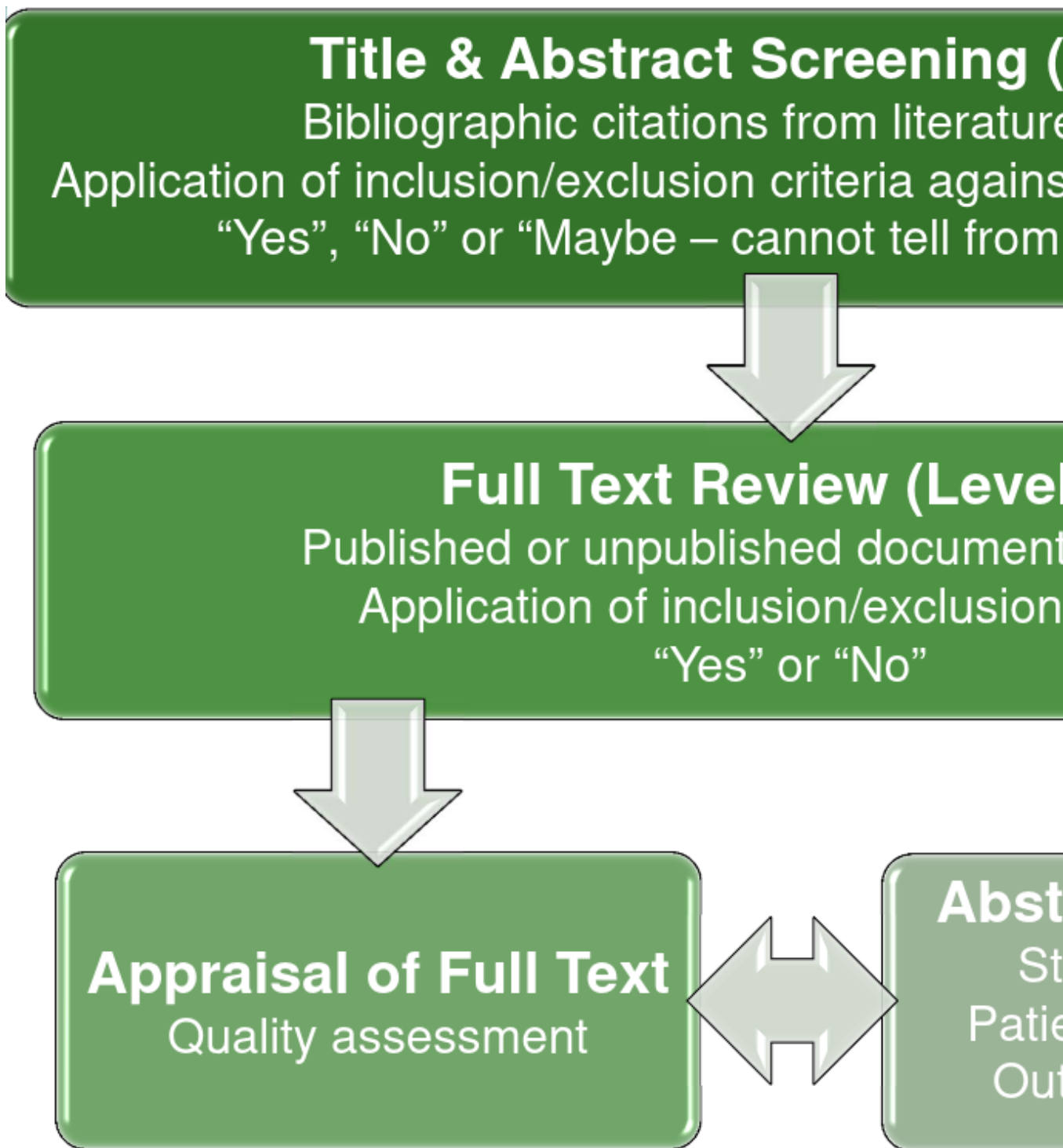


Figure 5.2: Aggregate process

1. Systematic review of RCTs.
2. RCTs.
3. Observational studies.
4. Case-series.
5. Experts opinion

Depending on age & type of health technology being evaluated, amount & quality of evidence may vary.

5.2.3.2 Abstract

Develop a data abstraction form to capture:

- Study characteristics
- Patient data
- Intervention information
- Outcome data

Data is entered into spreadsheet or database for reporting and synthesis.

5.2.3.3 Appraise

Many quality assessment tools exist depending on the type of literature:

- RCTs
- Observational studies
- Prognosis studies
- Economic evaluation

5.2.4 Evidence synthesis

5.2.4.1 Outcome measures

- Continuous: means
- Nominal data: proportions
- Nominal data: ranks

Before conducting a meta-analysis, it is important to define the outcome of interest.

Types of data	Outcome measures
Continuous	Mean difference. Standardized mean difference
Dichotomous	Risk difference. Relative Risk. Odds Ratio

- Use mean difference (MD) when studies have comparable outcome measures (i.e. same scale) $m_1 - m_2$
- Standardized mean difference (SMD) is used when studies use different outcome measurements which address the same clinical outcome (e.g. different instruments to measure pain intensity, anxiety...) $(m_1 - m_2)/sd_p$ where $sd_p = \sqrt{(sd_1 + sd_2)/2}$
- Risk difference (RD) $p_1 - p_2$
- Relative risk (RR) p_1/p_2
- Odds ratio (OR) $\frac{p_1/(1-p_1)}{p_2/(1-p_2)}$

In general, health practitioners are more familiar to the concept of relative risk than odds ratios. Remember that point estimates may be misleading. therefore uncertainty can be expressed by confidence intervals.

5.2.5 Meta-Analysis

A meta-analysis is a statistical analysis that combines the results of multiple scientific studies (more than 2 comparative studies). Studies may differ in terms of results or sample size.

- The results of a study (e.g. effect size) with 1,000 subjects is assumed to be a more precise estimate than an estimate derived from a study based on 10 subjects.
- Larger studies should carry more weight than smaller studies

5.2.5.1 Heterogeneity

Patients, interventions or outcomes may differ within and between studies. This is what is called heterogeneity. In order to take into account this heterogeneity, two models have been proposed: fixed effects and random effects models:

- Fixed effects model: Every study evaluates a common effect size. Variation between studies only due by chance.
- Random effects model: The effect sizes are different between studies and they follow a distribution.

In the presence of heterogeneity:

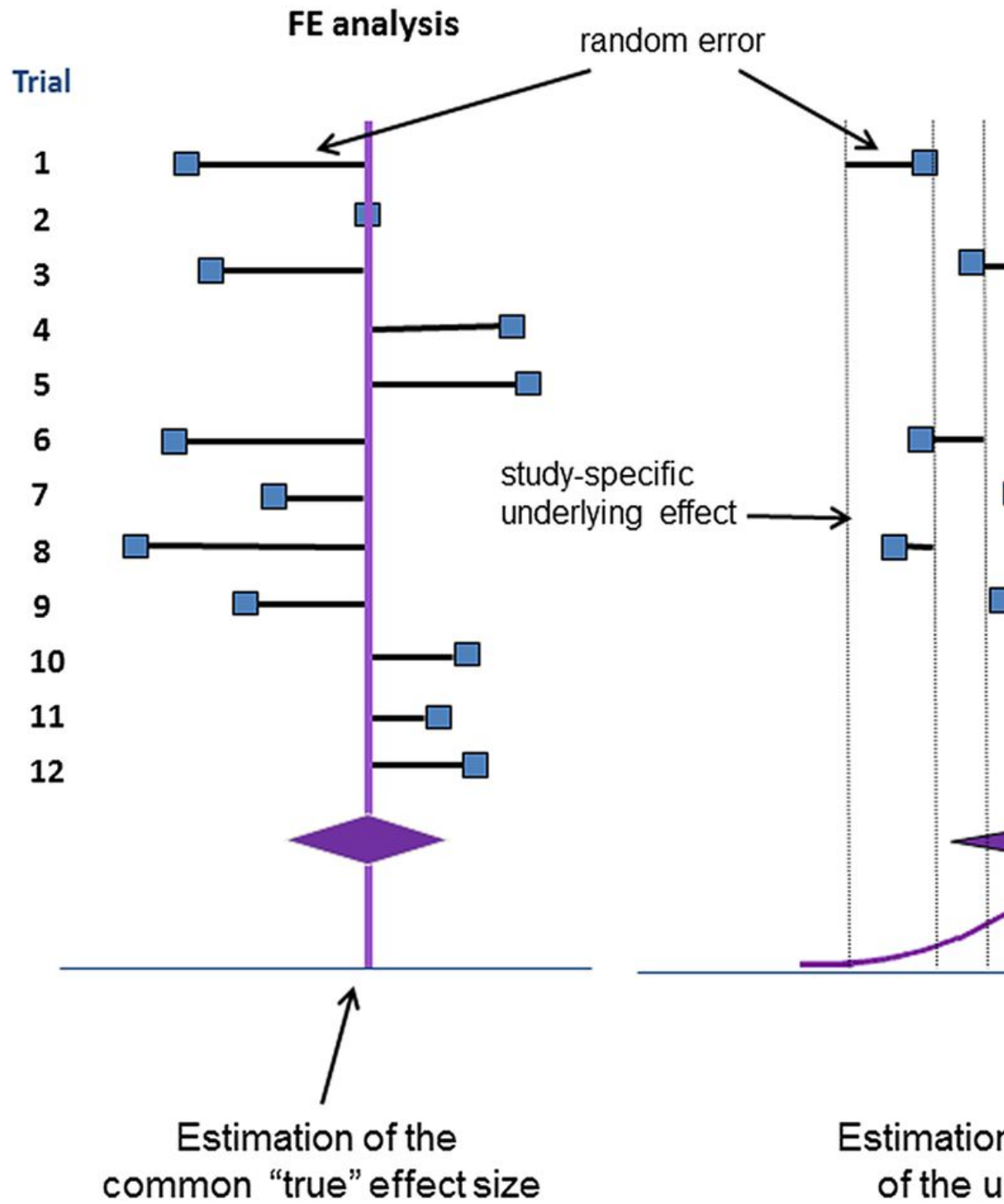


Figure 5.3: Fixed vs Random effects

5.2. IDENTIFYING, AGGREGATING AND APPRAISING THE EVIDENCE⁴³

- Ignore heterogeneity
- Incorporate heterogeneity into a random effects model
- Explain heterogeneity by sub-group analyses or meta-regressions

Presence of heterogeneity can be determined through a chi-squared test. Also, we can calculate a statistic (I^2) that describes the percentage of variation across studies that is due to heterogeneity rather than chance (i.e. how much heterogeneity).

Which one to use? Heterogeneity can be explained either by:

- Sub-group analyses
- Meta regression

5.2.5.2 Example

- Type 2 Diabetes mellitus management
 - P: Type II DM
 - I: Glitazones
 - C: Sulfonylurea
 - O: HbA1c

Study or Subgroup	Pioglitazones			Sulfonylur	
	Mean	SD	Total	Mean	SD
Tan (2)	-0.5	1.37	83	-0.4	1.18
Lawrence	-0.81	0.63	20	-1.21	0.82
Tan (1)	-0.78	1.69	109	-0.68	1.68
Yamanouchi	-2.3	0.68	35	-2.1	0.87
Forst	-0.81	0.74	89	-0.61	0.74
Agarwal	-0.1	1.2	22	-0.4	1.8
Basu	0.6	0.65	10	0.4	0.65
Pfutzner	-0.81	0.74	89	-0.61	0.74
Perriello	-0.79	0.97	140	-0.79	1.22
Nakamura	-1.6	1.12	17	-1.6	1.06
Mazzone	-0.32	0.04	203	0	0.04
Perez (a)	-2.34	2.08	323	-1.85	2.07
Al Majali	-0.09	0.8	10	-0.01	0.75
Taramato	-0.8	1.14	46	-1.43	1.09
Heliovaara	-0.6	0.16	29	-0.6	0.16
Total (95% CI)			1225		
Heterogeneity: $\text{Chi}^2 = 708.97$, $\text{df} = 14$ ($P < 0.00001$); $I^2 = 9$					
Test for overall effect: $Z = 6.24$ ($P < 0.00001$)					

Figure 5.4: Fixed effects

Study or Subgroup	Pioglitazones			Sulfonylurea			Weight
	Mean	SD	Total	Mean	SD	Total	
Tan (2)	-0.5	1.37	83	-0.4	1.18	96	6.9
Lawrence	-0.81	0.63	20	-1.21	0.82	20	6.9
Tan (1)	-0.78	1.69	109	-0.68	1.68	99	6.9
Yamanouchi	-2.3	0.68	35	-2.1	0.87	34	6.7
Forst	-0.81	0.74	89	-0.61	0.74	84	6.9
Agarwal	-0.1	1.2	22	-0.4	1.8	22	6.0
Basu	0.6	0.65	10	0.4	0.65	11	6.7
Pfutzner	-0.81	0.74	89	-0.61	0.74	84	6.9
Perriello	-0.79	0.97	140	-0.79	1.22	135	6.9
Nakamura	-1.6	1.12	17	-1.6	1.06	18	6.9
Mazzone	-0.32	0.04	203	0	0.04	206	6.0
Perez (a)	-2.34	2.08	323	-1.85	2.07	333	6.9
Al Majali	-0.09	0.8	10	-0.01	0.75	11	6.7
Taramato	-0.8	1.14	46	-1.43	1.09	45	6.9
Heliovaara	-0.6	0.16	29	-0.6	0.16	30	6.7
Total (95% CI)			1225			1228	100.0
Heterogeneity: $\tau^2 = 1.58$; $\chi^2 = 708.97$, $df = 14$ ($P < 0.00001$); $I^2 = 98.0\%$							
Test for overall effect: $Z = 1.53$ ($P = 0.13$)							

Not

Figure 5.5: Random effects

Chapter 6

Uncertainty and decision-making

6.1 Pre-session readings

Edlin, R., McCabe, C., Hulme, C., Hall, P., & Wright, J. (2015). Cost effectiveness modelling for health technology assessment: a practical course. Springer. Chapters 4, 6 and 7.

Briggs, A., Sculpher, M., & Claxton, K. (2006). Decision modelling for health economic evaluation. Oxford University Press.. Chapters 4.1, 4.2 and 4.3.

6.2 Introduction

The ultimate purpose of any economic evaluation is to inform decision-making; it is inherently concerned with estimating the expected future costs and outcomes of alternative courses of action; and because it is concerned with the future, uncertainty is a central concern for the analyst. In the context of decision-making, we always are interested to quantify the risk of making a wrong decision.

Questions

1. What do you think I mean of making a wrong decision in the context of HTA?

Decision uncertainty can be thought of as the probability that the decision maker will refuse to pay for a therapy that is good value or agree to pay for a therapy that is not good value. Uncertainty comes in different forms, so it is

important to understand the different sources of uncertainty that can impact upon the results of an economic evaluation.

6.3 Sources of uncertainty in cost-effectiveness models

6.3.1 Sampling variation

RCTs (or even observational studies) come from a subset of the population of interest. Therefore, a different sample from the same population will result in different estimates of efficacy in an experimental study.

6.3.2 Generalisability

Generalisability is concerned with the degree of confidence with which data obtained in one setting can be used to inform a model of what should be expected in a different setting. Generalisability is a particularly acute concern when evaluating technologies close to launch, when almost all evidence is derived from the clinical trial development programme.

Questions

2. Think about COVID-19 vaccines. What issues arise with this type of uncertainty?
3. In CEA, what other components are prone to generalisability issues?

6.3.3 Structural uncertainty

A model structure attempts to describe the relevant clinical pathways for patients treated with the interventions being evaluated. Usually, the simpler the model the better. The question is whether the simplifications exclude important characteristics of the potential care process that, if included in the model, would be likely to lead to a different decision?

Methodological choices, and different methods will often lead to different predictions and very different estimates of the uncertainty in those predictions. It is important that the analyst be explicit about the rationale for the modelling strategy adopted, ensuring that the decision maker is aware of any uncertainty that this introduces.

6.4 Variability, heterogeneity and uncertainty

We must distinguish between *variability* (the differences that occur between patients by chance) and *heterogeneity* (differences that occur between patients that can be explained) from decision *uncertainty* – the fundamental quantity that we wish to capture from our decision models.

The best way to explain the difference between these 3 concepts is with a traditional linear regression model, i.e.,

$$y_i = \beta_0 + \beta_p x_{i,p} + \epsilon_i, \epsilon_i \sim \mathcal{N}(0, \sigma)$$

6.5 How to deal with uncertainty?

Different methods have been developed to address uncertainty in CEA. These methods fall under the umbrella term *sensitivity analysis*. Sensitivity analysis is the process of varying model input values and recording the impact of those changes on the model outputs. The next table shows the different ways to handle uncertainty in CEA.

Type of sensitivity analysis	Definition
One way	The variation of one parameter whilst all other parameters are held constant to observe the impact on the predicted costs and outcomes
Multiway	The variation of more than one parameter whilst all other parameters are held constant to observe the impact on predicted costs and outcomes
Probabilistic sensitivity analysis	Using probability distributions to characterise the credible range and the likelihood of any given value being observed, combined with simulation to promulgate the uncertainty in the parameters to uncertainty in the predicted costs and outcomes

6.5.1 One-way (deterministic) sensitivity analysis

One-way sensitivity analysis examines how the ICER changes in response to changes in a single parameter whilst holding the value of all other model parameters constant. It is helpful to identify which parameters in a model have the strongest direct effect on the model results.

One way to represent this variation is by using tornado diagrams (see Figure 6.1).

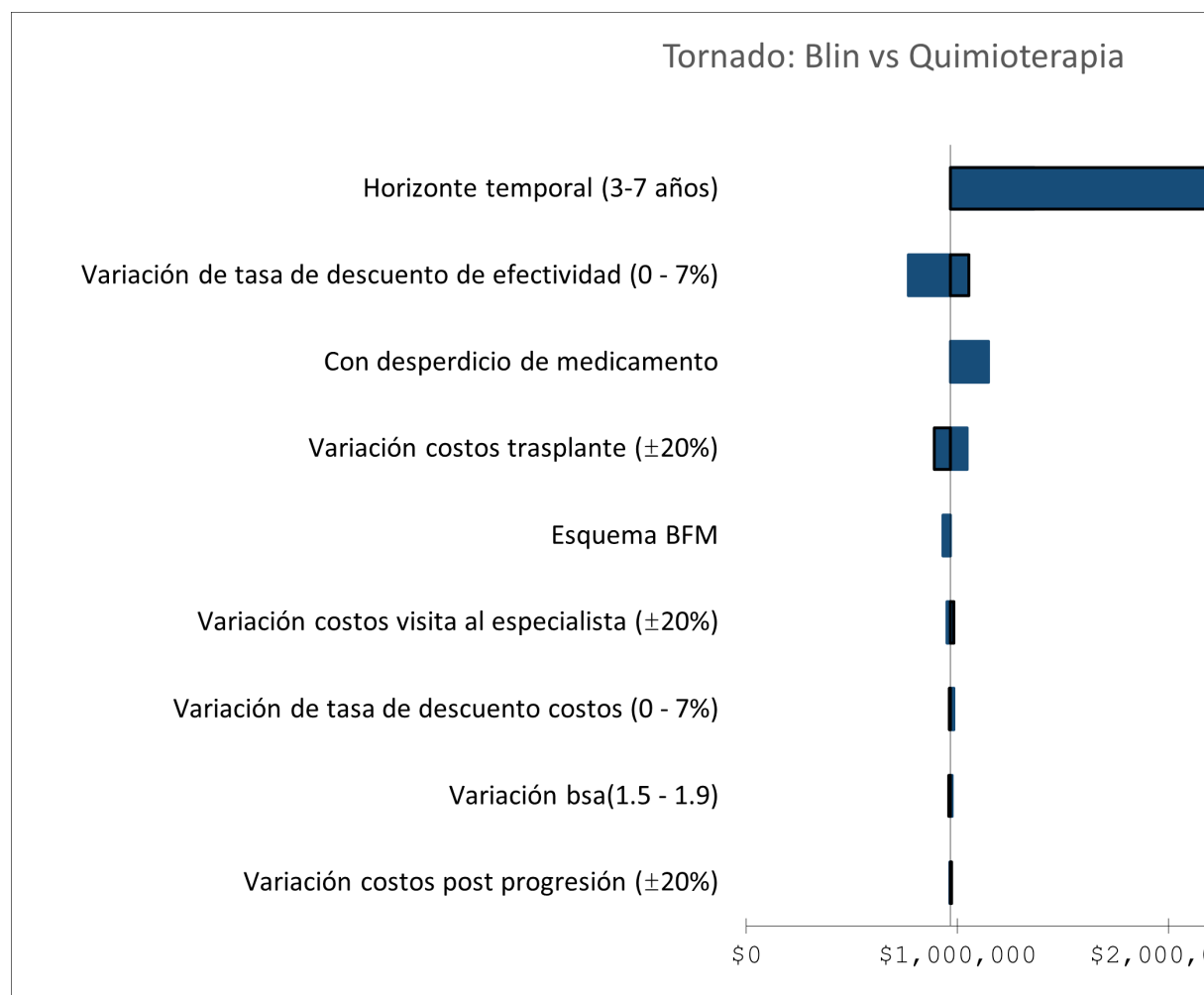


Figure 6.1: Tornado diagram

6.5.2 Probabilistic Sensitivity Analysis (PSA)

PSA requires that each parameter is expressed as a probability distribution. Each input to the model is known with a certain degree of uncertainty. This uncertainty can be characterised as a probability density function with an associated probability distribution.

Once a probability distribution has been described for each parameter in the cost effectiveness model, PSA allows the analyst to vary the value of each parameter simultaneously. Values are drawn from the probability distributions, and the outputs of the model for each draw are recorded.

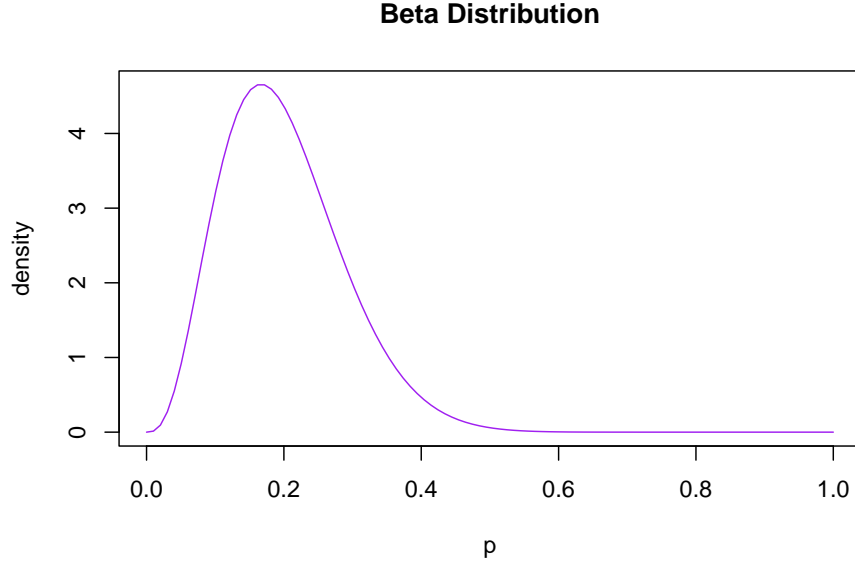
6.5.2.1 Distributions for probability parameters

Probability parameters have an important constraint – all probabilities can only take values between the range of zero and one. Furthermore, probabilities of mutually exclusive events must sum to one.

The beta distribution is a natural choice for representing uncertainty in a probability parameter where the data informing that parameter are binomial. The beta distribution is constrained on the interval 0–1 and is characterized by two parameters, *alpha* and *beta*. When data are binomially distributed, fitting the beta distribution turns out to be extremely straightforward. If the data are represented by a number of events of interest r , observed from a given sample size n , the proportion of events to the total sample gives the point estimate of the probability. Uncertainty in this probability can be represented by a beta (α, β) distribution, simply by setting $\alpha = r$ and $\beta = n - r$. Let's say that in a RCT, 4 adverse events were present in 20 patients. Therefore, shows the result of fitting a beta(4,16) distribution to the data of the previous example.

```
#define range
p = seq(0,1, length=100)

#create custom plot of Beta distribution
plot(p, dbeta(p, 4, 16), ylab='density',
     type='l', col='purple', main='Beta Distribution')
```



Questions

4. Why not using the normal distribution

Most of the when times when fitting beta distributions to secondary data or meta-analysis results, it may only be the mean/proportion and standard error/variance that are reported. If this is the case then it is still possible to fit the beta distribution using an approach known as method of moments. Let \hat{p} represents the sample proportion reported for example in a meta-analysis and $\text{se}(\hat{p})$ the standard error associated to it. Then

$$\alpha = \frac{\hat{p}^2(1 - \hat{p})}{\text{se}(\hat{p})^2} - \hat{p} \quad \beta = \alpha \frac{1 - \hat{p}}{\hat{p}}$$

6.5.2.2 Distributions for relative risks (or odd ratios) parameters

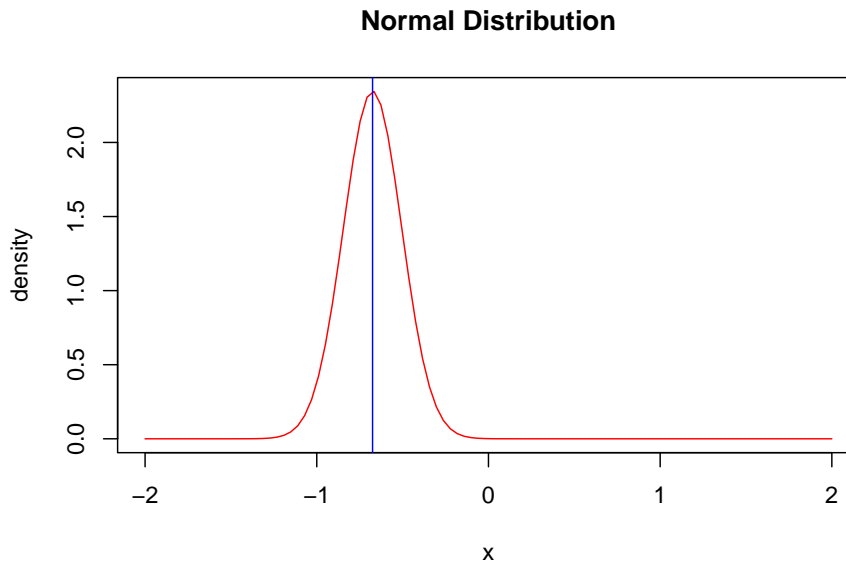
We will use the normal distribution for the log relative risks (odd ratios) (why?). Let's say that we employ data again from a published meta-analysis where the authors employ a relative risk estimate of 0.51 with a quoted 95% confidence interval of 0.365–0.710. Taking the natural logs of the point and interval estimates generates the following log scale estimates: -0.675 (-1.008 , -0.342). Dividing the range through by 2×1.96 recovers the estimate of log scale standard error:

$$\text{se}(\ln(RR)) = \frac{UL - LL}{2 * 1.96}$$

where UL and LL denote the upper and lower limits of the confidence interval. Now we simply take a random draw from a $\mathcal{N}(-0.675, 0.170)$ distribution and exponentiate the result.

```
#define range
x = seq(-2,2, length=100)

#create custom plot of Beta distribution
plot(x, dnorm(x, -.675, .17), ylab='density',
      type='l', col='red', main='Normal Distribution')
abline(v=-.675, col="blue")
```



6.5.2.3 Distributions for costs

Just as our choice of distribution for probability data was based upon the range of the data, so it should be noted that cost data are constrained to be non-negative and are made up of counts of resource use weighted by unit costs.

A natural choice is the gamma distribution. The reason is because this distribution is constrained on the interval 0 to positive infinity. To fit a gamma distribution to cost data we can again make use of the method of moments approach. The gamma distribution is parameterized as $\text{gamma}(\alpha, \beta)$. Let $\hat{\mu}$ represents the mean cost and $\text{se}(\hat{\mu})$ the standard error associated to it. Then

$$\alpha = \frac{\hat{\mu}^2}{\text{se}(\hat{\mu})^2} \quad \beta = \frac{\text{se}(\hat{\mu})^2}{\hat{\mu}}$$

As an example, we can consider the direct medical costs associated to a particular state of the model, which will be \$6948 for the purposes of this example. Unfortunately, although this estimate seems to have been taken from a patient-level cost data set, no standard error was reported. For the purposes of this example, suppose that the standard error is the same value as the mean. Therefore

$$\alpha = 6948/6948 = 1 \quad \beta = 6948/6948 = 6948,$$

6.5.2.4 Distributions for utilities

A pragmatic approach, often employed when health state utilities are far from zero, is to use a beta distribution. However, this is not appropriate for states close to death where values less than one are possible.

A simple transformation of $D = 1 - U$, such that D is a utility decrement or a disutility provides the solution. This utility decrement is now constrained on the interval 0 to positive infinity and the previous methods of fitting a gamma distribution can be applied.

Assignments

Assignment 1

1. Section 3.5 in *Edlin, R., McCabe, C., Hulme, C., Hall, P., & Wright, J. (2015). Cost effectiveness modelling for health technology assessment: a practical course. Springer.*
2. Build the precious exercise in R.
3. From Google Classroom week 2, fill the CHEERS table 1 checklist using the next CE article.

Assignment 2

1. Chapter 9 in *Edlin, R., McCabe, C., Hulme, C., Hall, P., & Wright, J. (2015). Cost effectiveness modelling for health technology assessment: a practical course. Springer.*
2. To the Markov model built in class (i.e. `Markov.R`), add the half-cycle correction as seen in the excel file from class (Chapter 9 from Gray et al.)

Introduction to R

The goal of this tutorial is to orient the learner to R Studio and the R programming language.

Please complete the following:

1. Create a folder somewhere you can easily find it (e.g., on your desktop) called 'R Course'.
2. Open R Studio
3. Session -> Set Working Directory -> to source file location

Outline

1. Read in the 'babies' data set
2. Basic data manipulation
3. Functions
4. Saving and viewing results

Babies Dataset

The `babies` dataset will be used throughout this session to illustrate basic R concepts.

The dataset is a collection of variables taken for each new mother in a Child and Health Development Study. It has 1,236 observations on male live births for the following 23 variables.

Variables in data file

<code>id</code>	identification number
<code>date</code>	birth date as character string (mon-dd-yyyy)
<code>ddate</code>	day of birth
<code>mdate</code>	month of birth

ydate	year of birth
gestation	length of gestation in days
wt	birth weight in ounces (999 unknown)
parity	total number of previous pregnancies including fetal deaths and still births
age	mother's age in years at end of pregnancy 999=unknown
ed	mother's education 0=less than 8th grade 1=8th-12th grade - did not graduate 2=HS graduate???no other schooling 3=HS+trade 4=HS+some college 5=College graduate, 6=Trade school HS unclear 9=unknown
ht	mother's height in inches to the last completed inch 999=unknown
wt1	mother prepregnancy wt in pounds 999=unknown
dage	father's age, coding same as mother's age.
ded	father's education, coding same as mother's education.
dht	father's height, coding same as for mother's height
dwt	father's weight coding same as for mother's weight
inc	family yearly income in \$2500 increments 0=under 2500 1=2500-4999 ..., 8= 12,500-14,999 9=15000+ 998=unknown 999=not asked
smoke	does mother smoke? 0=never 1=smokes now 2=until current pregnancy 3=once did, not now 9=unknown
time	If mother quit smoking, how long ago? 0=never smoked 1=still smokes 2=during current preg 3=within 1 yr 4=1 to 2 years ago 5=2 to 3 years ago 6=3 to 4 years ago

```

7=5 to 9 years ago
8=10+years ago
9=quit and don't know,
998=unknown
999=not asked
number      number of cigs smoked per day for past and current smokers
0=never
1=1-4
2=5-9
3=10-14
4=15-19
5=20-29
6=30-39
7=40-60
8=60+
9=smoke but don't know
998=unknown
999=not asked
race        mother's race
marital     marital status of mother
drace       father's race, coding same as mother's race

```

Loading babies dataset

We can load the data from our working directory as shown below. It is a .csv file, so can be read in with `read.csv`. specifying that

- the file has a “header” row (`header=T`) with variable names
- values are separated by commas.
- values of ‘’, ‘998’ or ‘999’ represent missing data

We need to be confident that 998 and 999 are not legitimate values for any variables, so that R does not interpret legitimate values as missing. We will not distinguish between ‘unknown’ and ‘not asked’ in the data, so the code below will read two consecutive commas (“,”), 998, and 999 as missing values. We will have to deal separately with the values ‘9’ that mean a value is missing.

```

babies <- read.csv('babies.csv',header=T, sep=",",
                  na.strings=c("", "998", "999"))

```

Data manipulation

What are the dimensions of the data set?

```
dim(babies)
```

```
[1] 1236 23
```

What are the names of the variables?

```
names(babies)
```

```
## [1] "id"      "date"    "gestation" "wt"      "parity"  "age"
## [7] "ed"      "ht"      "wt1"       "dage"    "ded"     "dht"
## [13] "dwt"     "inc"     "smoke"     "time"    "number"  "race"
## [19] "marital" "drace"   "ddate"     "mdate"   "ydate"
```

```
#head(babies)
```

```
#tail(babies)
```

Isolating variables

```
# babies$id
# babies[, 1]
# babies[, 'id']
# babies[1:10, c('id', 'date', 'gestation')]
```

Assignment and subsetting

```
under.30<- babies$age<30
b<- babies[under.30, ]
summary(b$age)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.   NA's
##  15.00   22.00   24.00   24.03   27.00   29.00     2
```

```
b<- babies[which(under.30), ]
summary(b$age)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##  15.00   22.00   24.00   24.03   27.00   29.00
```

Exercises (1)

1. Create a new data set consisting of only the rows where the father's age is 40 or over (use '>=')

2. Include only the mother's age, father's age and marital status in the new dataset
3. Display the dimensions of this data set in the console.
4. Locate the new dataset in the Global Environment and view it

NB: Be sure that you have handled missing father's ages properly

Exercises (1) solutions

Creating some new variables

A baby weight more than 4kg is defined as macrosomia. We can create this variable (converting ounces to kg) as:

```
babies$wtKg <- babies$wt/(16*2.2)
babies$macrosomia <- babies$wtKg > 4
```

A baby weighing less than 2.5kg is defined as "small". We can create this variable as:

```
babies$smallBaby <- babies$wtKg < 2.5
```

We can make a single character variable with three categories of weight:

```
babies$birthWeightCat <- ifelse(babies$wtKg < 2.5, "small",
                                ifelse(babies$wtKg > 4, "large", "normal"))
```

Notice that when we tabulate these, they appear in alphabetical order.

```
table(babies$birthWeightCat)
```

```
##
##  large normal  small
##    145   1033    58
```

To impose an order, we can convert to a factor and specify the order:

```
babies$birthWeightCat <- factor(babies$birthWeightCat,
                                levels=c("small", "normal", "large"),
                                labels=c("Small baby", "Normal weight baby", "Large baby"))
table(babies$birthWeightCat)
```

```
##
##           Small baby Normal weight baby           Large baby
##                58                1033                145
```

A baby born before 37 completed weeks of pregnancy is defined as preterm. We can create this logical variable as:

```
babies$preterm <- babies$gestation < 37*7
table(babies$preterm)
```

```
##
## FALSE TRUE
## 1126   97
```

This is just a logical variable (TRUE/FALSE or T/F) so we need to make a factor to have it appear more user-friendly:

```
babies$whenBorn <- factor(babies$preterm,
                          levels=c(TRUE,FALSE),
                          labels=c("Preterm","Full-term"))
```

We can make a binary variable for the mother's and father's races:

```
babies$whiteRace <- ifelse(babies$race=="white","white","other")
babies$dwhiteRace <- ifelse(babies$drace=="white","white","other")
```

Exercises (2)

1. Starting with the numeric variable `smoke`, in the babies dataset, create a new variable `smokeCat` in the data set that has three levels: `never smoker`, `past smoker`, `current smoker`. Use the `factor` function and ensure that the levels appear in that order.
2. Tabulate the numerical variable against the new one to make sure you have not made an error.
3. Make a binary variable in the babies dataset `smokeNow` that is 'Yes' when the mother smokes and 'No' otherwise (i.e., it is not yes (1) or it is missing (9).)
4. [optional] Make a factor variable `eduCat` from the mother's education variable `ed` using the information at the top of this file about the meaning of 1,2,3,...

Exercises (2) solutions

1. Make the two variables:
2. Check the coding was correct
3. Make a binary variable
4. [optional] make the education variable and check it

Functions

Two types:

1. Built in functions
2. User defined functions

Built in Functions

When you open R, there are many functions available to you: Here, we will review a few useful built in functions.

If you need help using a function, execute `?` followed by the function name, with or without the parentheses.

```
? table
```

The following functions are widely used in descriptive statistics

table(), prop.table()

```
t<- with(babies, table(marital))
t
```

```
## marital
##          divorced legally separated          married          never married
##              5              15              1208              6
##          unknown
##              2
```

```
p<- prop.table(t)
p
```

```
## marital
##          divorced legally separated          married      never married
##      0.004045307      0.012135922      0.977346278      0.004854369
##          unknown
##      0.001618123
```

```
p*100
```

```
## marital
##          divorced legally separated          married      never married
##      0.4045307      1.2135922      97.7346278      0.4854369
##          unknown
##      0.1618123
```

```
ifelse()
```

```
babies$first.preg<- with(babies, ifelse(parity==0, 'first','not first'))
table(babies$first.preg)
```

```
##
##      first not first
##      315      921
```

```
summary(), mean(), median(), sd(), quantile()
```

```
summary(babies)
```

```
##          id          date          gestation          wt
## Min.    : 15  Length:1236  Min.    :148.0  Min.    : 55.0
## 1st Qu.:5286  Class :character 1st Qu.:272.0 1st Qu.:108.8
## Median :6730  Mode  :character Median :280.0 Median :120.0
## Mean   :6001          Mean   :279.3 Mean   :119.6
## 3rd Qu.:7583          3rd Qu.:288.0 3rd Qu.:131.0
## Max.   :9263          Max.   :353.0 Max.   :176.0
##                                     NA's   :13
##      parity          age          ed          ht
```



```

## Min. : 0.000 Min. :15.00 Min. :0.000 Min. :53.00
## 1st Qu.: 0.000 1st Qu.:23.00 1st Qu.:2.000 1st Qu.:62.00
## Median : 1.000 Median :26.00 Median :2.000 Median :64.00
## Mean : 1.932 Mean :27.26 Mean :2.916 Mean :64.05
## 3rd Qu.: 3.000 3rd Qu.:31.00 3rd Qu.:4.000 3rd Qu.:66.00
## Max. :13.000 Max. :45.00 Max. :9.000 Max. :72.00
## NA's :2 NA's :22
## wt1 dage ded dht dwt
## Min. : 87.0 Min. :18.00 Min. :0.000 Min. :60.0 Min. :110.0
## 1st Qu.:114.8 1st Qu.:25.00 1st Qu.:2.000 1st Qu.:68.0 1st Qu.:155.0
## Median :125.0 Median :29.00 Median :4.000 Median :71.0 Median :170.0
## Mean :128.6 Mean :30.35 Mean :3.189 Mean :70.2 Mean :171.2
## 3rd Qu.:139.0 3rd Qu.:34.00 3rd Qu.:5.000 3rd Qu.:72.0 3rd Qu.:185.0
## Max. :250.0 Max. :62.00 Max. :9.000 Max. :78.0 Max. :260.0
## NA's :36 NA's :7 NA's :492 NA's :499
## inc smoke time number
## Min. :0.000 Min. :0.0000 Min. :0.0000 Min. :0.000
## 1st Qu.:2.000 1st Qu.:0.0000 1st Qu.:0.0000 1st Qu.:0.000
## Median :3.000 Median :1.0000 Median :1.0000 Median :1.000
## Mean :3.701 Mean :0.8681 Mean :0.9625 Mean :1.825
## 3rd Qu.:5.000 3rd Qu.:1.0000 3rd Qu.:1.0000 3rd Qu.:3.000
## Max. :9.000 Max. :9.0000 Max. :9.0000 Max. :9.000
## NA's :124 NA's :10 NA's :10
## race marital drace ddate
## Length:1236 Length:1236 Length:1236 Min. : 1.00
## Class :character Class :character Class :character 1st Qu.: 8.00
## Mode :character Mode :character Mode :character Median :15.00
## Mean :15.37
## 3rd Qu.:23.00
## Max. :31.00
## mdate ydate wtKg macrosomia
## Min. : 1.000 Min. :1961 Min. :1.562 Mode :logical
## 1st Qu.: 4.000 1st Qu.:1961 1st Qu.:3.089 FALSE:1091
## Median : 7.000 Median :1962 Median :3.409 TRUE :145
## Mean : 6.617 Mean :1962 Mean :3.397
## 3rd Qu.: 9.000 3rd Qu.:1962 3rd Qu.:3.722
## Max. :12.000 Max. :1962 Max. :5.000
## smallBaby birthWeightCat preterm whenBorn
## Mode :logical Small baby : 58 Mode :logical Preterm : 97
## FALSE:1178 Normal weight baby:1033 FALSE:1126 Full-term:1126
## TRUE :58 Large baby : 145 TRUE :97 NA's : 13
## NA's :13
##
##

```

```
##
##   whiteRace          dwhiteRace          smokeCat      smokeNow
## Length:1236          Length:1236          Never smoker :544 Length:1236
## Class :character     Class :character     Past smoker   :198 Class :character
## Mode  :character     Mode  :character     Current smoker:484 Mode  :character
##                                     NA's           : 10
##
##
##
##           eduCat      first.preg
## HS graduate      :444 Length:1236
## HS+some college :298 Class :character
## College graduate:219 Mode  :character
## 8th-12th grade  :183
## HS+trade        : 65
## (Other)         : 26
## NA's            : 1
```

```
gestAge<- babies$gestation
mean(gestAge)
```

```
## [1] NA
```

```
mean(gestAge, na.rm=T)
```

```
## [1] 279.3385
```

```
gestAge<- gestAge[complete.cases(gestAge)]
summary(gestAge)
```

```
##   Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##  148.0   272.0   280.0   279.3   288.0   353.0
```

```
mean(gestAge)
```

```
## [1] 279.3385
```

```
median(gestAge)
```

```
## [1] 280
```

```
sd(gestAge)
```

```
## [1] 16.02769
```

```
range(gestAge)
```

```
## [1] 148 353
```

```
min(gestAge)
```

```
## [1] 148
```

```
max(gestAge)
```

```
## [1] 353
```

```
quantile(gestAge)
```

```
##    0%   25%   50%   75%  100%
##   148   272   280   288   353
```

```
quantile(gestAge, seq(0, 1, by = 0.2))
```

```
##    0%   20%   40%   60%   80%  100%
##   148   270   277   283   290   353
```

```
round()
```

```
x<- 1121.933384
round(x, 3)
```

```
## [1] 1121.933
```

```
round(x, -2)
```

```
## [1] 1100
```

```
paste()
```

We can build up complex quoted strings with `paste`:

```
paste("Mean (days) =", mean(gestAge))
```

```
## [1] "Mean (days) = 279.338511856092"
```

```
paste("Mean (days) =", round(mean(gestAge)))
```

```
## [1] "Mean (days) = 279"
```

```
paste('mean day (sd) = ',
      round(mean(gestAge)),
      ' (',
      round(sd(gestAge), 1),
      ')',
      sep='')
```

```
## [1] "mean day (sd) = 279 (16)"
```

apply()

Carries out an operation on the rows or columns of a dataset. We pick the `MARGIN` to specify whether we want this to be rows or columns

- 1 = rows
- 2 = columns

```
apply(babies[, c('age', 'dage', 'wtKg')], MARGIN = 2, FUN = mean, na.rm=T)
```

```
##      age      dage      wtKg
## 27.25527 30.34825  3.39707
```

tapply()

Carries out an operation on one variable, split by a second variable (or group of variables) - e.g., means by group:

```
wt.by.smoke.term<- with(babies,
                        tapply(wt, list(smokeCat, whenBorn), mean, na.rm=T))
rw<- round(wt.by.smoke.term, 1)
rw
```

```
##               Preterm Full-term
## Never smoker    107.0    124.1
## Past smoker     103.4    125.9
## Current smoker   91.7     116.2
```

User Defined Functions

You can write your own functions, `text(arguments){ body }`

```
custom.summary<- function(x){
  out<- paste(round(mean(x, na.rm=T), 2),
               ' (SD=',
               round(sd(x, na.rm=T), 2),
               ')',
               sep='')
  return(out)
}
custom.summary(babies$age)
```

```
## [1] "27.26 (SD=5.78)"
```

Saving output and opening in word

```
write.csv(rw, 'test.csv', quote= F)
```

Now open 'test.csv' with word. Highlight and click Table -> convert text to table -> ok.

Exercises (3)

1. Create a function to summarize a binary (0/1 or FALSE/TRUE)) variable and return a character string that looks like, for example "18/54 (33.3%)"

Framework

```
bin.sum <- function(x){
  t <- # tabulate x
  n <- # how many nonmissing observations?
  x <- # how many 1's?
  pct <- round()
  paste()

}
```

2. Use `tapply` to apply your function to the macrosomia using `smokeCat` as the grouping variable

Hint

```
result <- tapply(X =
                 INDEX=
                 FUN=bin.sum)
```

3. View your results as a Word table using by using write.csv to save to a CSV file.
4. [Advanced] Create a function to summarize a continuous variable in the following format: “Median (IQR.low, IQR.high), n”. Use **apply** to apply this function to the variables **age**, **dage**, ‘ht’, ‘dht’. The result should be a table that you can save in the same way as in question 3.
5. [More advanced] Use the function from question 4. to summarize a the variables **age**, **dage**, ‘ht’, ‘dht’ in groups formed by ‘preterm’. Hint: Look at the help for the function **aggregate**.

Hint:

```
aggregate(x=DATASET,
          by=list(BY VARIABLES HERE),
          FUN=FUNCTION.FROM.4)
```

Exercises (3) solutions

- 1.
- 2.
- 3.
- 4.
- 5.

Working with more than one dataset

Suppose you want to know how far from the average for the ethnic group each baby's birth weight is as a Z-score:

$$Z = (\text{wt} - \text{average})/\text{SD}$$

You need to

1. find the average and SD for each group
2. merge this data with the full babies dataset
3. calculate each baby's Z score

The R code for these steps is shown below

1. find the average and SD for each group and save it in a data.frame

```
average.by.group <- tapply(X = babies$wtKg,
                           INDEX=babies$race,
                           mean,na.rm=T)

sd.by.group <- tapply(X = babies$wtKg,
                      INDEX=babies$race,
                      sd,na.rm=T)
stats <- data.frame(race=names(average.by.group),
                   average=average.by.group,
                   SD=sd.by.group)
print(stats)
```

```
##           race average      SD
## asian      asian 3.137268 0.4543554
## black      black 3.216980 0.5422872
## mexican    mexican 3.526989 0.4017935
## mixed      mixed 3.360719 0.5363923
## unknown    unknown 3.892045      NA
## white      white 3.455721 0.5027514
```

2. merge this data with the full babies dataset

```
babies <- merge(babies,
                stats,
                by="race")

head(babies[, c("wtKg", "average", "SD", "race")])
```

```
##          wtKg average          SD race
## 1 3.409091 3.137268 0.4543554 asian
## 2 3.125000 3.137268 0.4543554 asian
## 3 2.642045 3.137268 0.4543554 asian
## 4 3.352273 3.137268 0.4543554 asian
## 5 2.982955 3.137268 0.4543554 asian
## 6 2.840909 3.137268 0.4543554 asian
```

3. calculate each baby's Z score

```
babies$Z.wt <- (babies$wtKg - babies$average)/babies$SD
```

Finally, please save the alterations to the babies data set for the next session:

```
write.csv(babies, 'babiesAugmented.csv', quote=F, row.names = F)
```

Matrices

A data structure useful in statistical programming and for more advanced data analysis is a matrix. A matrix is a 2-dimensional array, with rows and columns, that all contains items of the same type (as it is with vectors). We access their content using two indices (given the two dimensions).

```
M1 <- matrix(1:12,nrow=3,byrow = F)
M1a <- matrix(1:12,nrow=3,byrow = T)
M2 <- matrix(letters[1:25],nrow=5,ncol = 5)
M3 <- matrix(c(letters,LETTERS[1:4]),nrow=5,ncol = 6)
```

```
M3[1,1]
```

```
## [1] "a"
```

```
M3[1:3,1:3]
```

```
##          [,1] [,2] [,3]
## [1,] "a"   "f"   "k"
## [2,] "b"   "g"   "l"
## [3,] "c"   "h"   "m"
```


Lists

The most versatile data structure is the `list`. It can be used as a collection of different, even heterogeneous, objects. It can have one or many dimensions (e.g. list array). We need to access a list's items using double square brackets `[[]]`.

```
some.stuff <- list(first.matrix=M1,
                  second.matrix=M2,
                  my.data.frame=babies,
                  OldDads=babies$dage > 50)
```

```
class(some.stuff)
```

```
## [1] "list"
```

```
length(some.stuff)
```

```
## [1] 4
```

```
names(some.stuff)
```

```
## [1] "first.matrix" "second.matrix" "my.data.frame" "OldDads"
```

```
some.stuff[[1]]
```

```
##      [,1] [,2] [,3] [,4]
## [1,]    1    4    7   10
## [2,]    2    5    8   11
## [3,]    3    6    9   12
```

```
table(some.stuff[["OldDads"]])
```

```
##
## FALSE  TRUE
## 1217    12
```

```
some.stuff$my.vector[3]
```

```
## NULL
```

```
some.stuff$my.data.frame$gestation[41:45]
```

```
## [1] 278 280 280 280 275
```

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