

Write in Word,  
**Save in Markdown,**  
Publish in  $\text{\LaTeX}$

May take some pain out of scientific writing.

Katrin Leinweber

2015-Aug-31

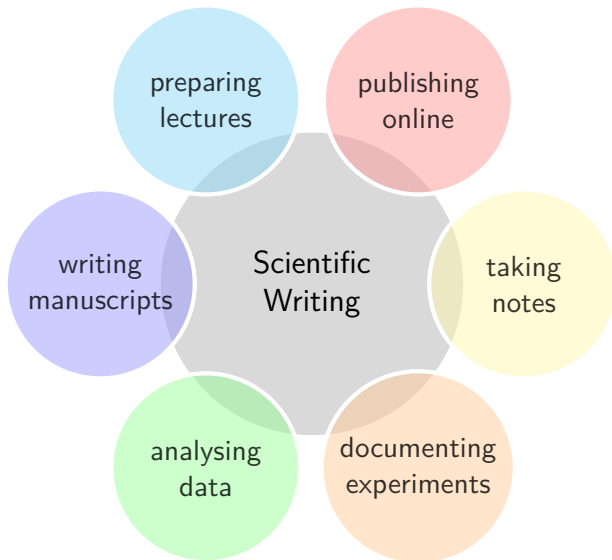
# Why not talk about a research project?

*Support research and training programs that transcend traditional disciplines.*<sup>1</sup>

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<sup>1</sup>[chembiol.uni.kn/statement.html](http://chembiol.uni.kn/statement.html)

# Markdown can help you with...



# Background: Markup languages



- ▶ design philosophy<sup>2</sup>: separate content from presentation
- ▶ most modern document formats are markup
- ▶ word processors just typeset continuously

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<sup>2</sup>[en.wikipedia.org/wiki/Separation\\_of\\_presentation\\_and\\_content](https://en.wikipedia.org/wiki/Separation_of_presentation_and_content)

# Background: Markup languages

LaTeX (1985)	HTML (1992)	
<code>\textbf{bold}</code>	<code>&lt;strong&gt;bold&lt;/strong&gt;</code>	<b>bold</b>
<code>\emph{Species name}</code>	<code>&lt;i&gt;Species name&lt;/i&gt;</code>	<i>Species name</i>
<code>\section{Heading 1}</code>	<code>&lt;h1&gt;Heading 1&lt;/h1&gt;</code>	<b>Heading 1</b>
<code>\subsection{Heading 2}</code>	<code>&lt;h2&gt;Heading 2&lt;/h2&gt;</code>	Heading 2
<code>\sout{strike out}</code>	<code>&lt;strike&gt;out&lt;/strike&gt;</code>	<del>strike out</del>

- ▶ a lot of formatting commands
- ▶ only really readable in typeset form (PDF, website)
- ▶ but machine-readable

# What is Markdown? Minimalistic markup language!

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<b>**bold**</b>	<b>bold</b>
<i>*Species name*</i>	<i>Species name</i>
<b># Heading 1</b>	<b>Heading 1</b>
<b>## Heading 2</b>	Heading 2
<del>~~strike out~~</del>	<del>strike out</del>

---

- ▶ fast to type & easy to read
- ▶ defined in 2004 by John Gruber<sup>3</sup> & Aaron Swartz
- ▶ designed for web publishing => converts to HTML
- ▶ has links, images, lists, quotes, etc.

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<sup>3</sup>[daringfireball.net/projects/markdown](http://daringfireball.net/projects/markdown)

## Science-related use-case examples for Markdown (MD)

## Words of caution: try with finished doc, or small new one!

- ▶ up-front time investment to install tools & get used to MD
- ▶ accept hand-over of styling & templating to others
- ▶ return to .docx possible in any case



Íshestar via [equitrekking.com](http://equitrekking.com)



# Use-case: digital lab journalling

## ##### Preparation

- [x] Glucose standards ("4/2/13", [150304a](https://docs.google.com/spreadsheets/d/1z1411v1qddJ3-jqSSM1V340X9w0-gmmEzDZJ0wgDEdg/edit#gid=0) & [141015a](https://trello.com/c/h07txK0a/104-141015a-achmi-sugar-standard-curves) mix)
- [x] solution of 5% crystalline phenole (not Roti-Phenol) in MQ-H<sub>2</sub>O
- [x] shaker(s) at room temperature
- [ ] Multipette with 5mL- & 10mL tips
- [ ] PMMA cuvettes

## ##### Procedure

- 2 1mL aliquots taken from Erli for non-concentrated measurements:
  - centrifuged down at 18k\*g\* for 3min => SN transferred into "oSN" sample & centrifuged again => V\_oSN = 978.4μL
- 184.8mL cell suspension centrifuged down at 5k\*g\* & 20°C for 3min
  - slightly lower recovery of supernatant for concentration ("cSN") due to disturbances of pellet with 25mL pipettes => V\_cSN = 175mL
  - lyophilisation at [Spitellers'](https://trello.com/c/j7bmrNW2/135-spitellers-lyophilisator) at 0°C,

## ##### Conclusions



- \*\*high salt complicates assay procedure due to overboiling & degrades standard curve\*\*
- \*\*conc. supernatant only 2-5x\*\*

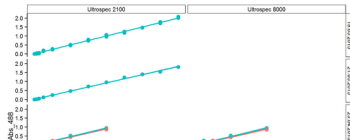
## Preparation

- ☒ Glucose standards ("4/2/13", 150304a & 141015a\_mix)
- ☒ solution of 5% crystalline phenole (not Roti-Phenol) in MQ-H<sub>2</sub>O
- ☒ shaker(s) at room temperature
- ☐ Multipette with 5mL- & 10mL tips
- ☐ PMMA cuvettes

## Procedure

- 2 1mL aliquots taken from Erli for non-concentrated measurements:
  - centrifuged down at 18kg for 3min => SN transferred into "oSN" sample centrifuged again => V\_oSN = 978.4μL
- 184.8mL cell suspension centrifuged down at 5kg & 20°C for 3min
  - slightly lower recovery of supernatant for concentration ("cSN") due to disturbances of pellet with 25mL pipettes => V\_cSN = 175mL
  - lyophilisation at Spitellers' at 0°C,

## Conclusions



editors with live preview:



MarkdownPad,



MacDown, etc.

# Use-case: documenting data analysis

```
## Distribution of the number of bacteria cells adherent to diatom valve faces of
## different surface types (frustule or capsule) in xenic A. minutissimum biofilms
## incubated for 11 to 31 days.

Katrín Leinweber, Uni Konstanz -- 5. Oct. 2014; revised in Feb. 2015

```{r ggplot2, fig.width = 9, fig.height = 3}
library(ggplot2)

data_raw <- read.csv("I41005a_fig_attachment.csv")

subset_by_celltype <- function(celltype){return(subset(data_raw, diatom_valve == celltype
))}

frustules <- subset_by_celltype("frustule")
capsules <- subset_by_celltype("capsule")

N_frustules <- dim(frustules)[1]
N_capsules <- dim(capsules)[1]

plot <- ggplot(data_raw, aes(x = diatom_valve, y = N_bacteria)) +
  geom_boxplot(fill = "darkgrey", size = 1) + # thicker outlines
  coord_flip() +
  labs(title = NULL, x = NULL, y = "bacteria cells per diatom") +
  scale_x_discrete(breaks = c("capsule", "frustule"), # original category names / tick
  labels =
    labels = c("adherent to ncapsules", "adherent to nfrustules") #
  ) + # learned from http://stat.ethz.ch/R-manual/R-patched/library/base/html/sprintf.html
  9/#setting-tick-mark-labels
  stat_summary(fun.y = mean, geom = "point", shape = 5, size = 4) + # adds symbol for me
  theme(title = element_text(size = 16),
    axis.title.y = element_blank(),
    axis.text = element_text(size = 16, color = "black"),
    axis.ticks = element_blank(),
    panel.grid.major = element_line(color = "white", size = 1)
  ) # learned from http://docs.ggplot2.org/0.9.3/theme.html

plot
....

Bacteria were counted in SEM images, if they were in direct, visible contact with the
valve face of either a frustule (N = r N_frustules) or a completely encapsulated diatom
cell (N = r N_capsules; see figures 2B and 3A for illustration). Boxes represent 1st and
3rd quartile, black center lines represent medians, whiskers extend to 1.5-fold of the
inter-quartile range. Diamond symbols represent means. Black dots are outliers.

```{r}
# basic statistics
```

Distribution of the number of bacteria cells adherent to diatom valve faces of different surface types (frustule or capsule) in xenic *A. minutissimum* biofilms incubated for 11 to 31 days.

Katrín Leinweber, Uni Konstanz -- 5. Oct. 2014; revised in Feb. 2015

```
library(ggplot2)

data_raw <- read.csv("I41005a_fig_attachment.csv")

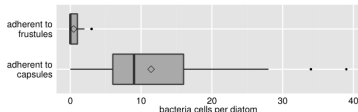
subset_by_celltype <- function(celltype){return(subset(data_raw, diatom_valve == celltype)
)}

frustules <- subset_by_celltype("frustule")
capsules <- subset_by_celltype("capsule")

N_frustules <- dim(frustules)[1]
N_capsules <- dim(capsules)[1]

plot <- ggplot(data_raw, aes(x = diatom_valve, y = N_bacteria)) +
  geom_boxplot(fill = "darkgrey", size = 1) + # thicker outlines
  coord_flip() +
  labs(title = NULL, x = NULL, y = "bacteria cells per diatom") +
  scale_x_discrete(breaks = c("capsule", "frustule"), # original category names / tick la
  labels =
    labels = c("adherent to ncapsules", "adherent to nfrustules") # learned
    ) + # learned from http://www.cookbook-r.com/Graphs/Axes_228ggplot222
  stat_summary(fun.y = mean, geom = "point", shape = 5, size = 4) + # adds symbol for me
  theme(title = element_text(size = 16),
    axis.title.y = element_blank(),
    axis.text = element_text(size = 16, color = "black"),
    axis.ticks = element_blank(),
    panel.grid.major = element_line(color = "white", size = 1)
  ) # learned from http://docs.ggplot2.org/0.9.3/theme.html

plot
```

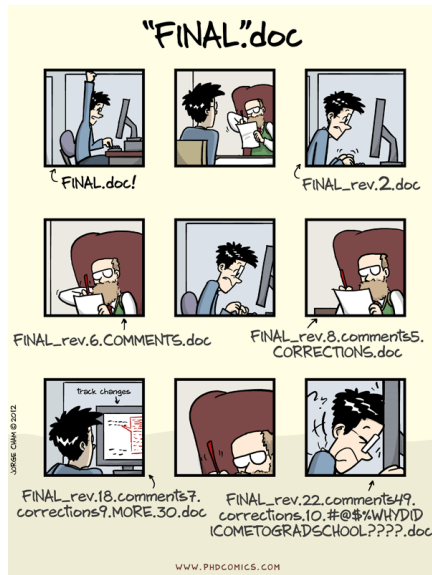


Bacteria were counted in SEM images, if they were in direct, visible contact with the valve face of either a frustule (N = 54) or a completely encapsulated diatom cell (N = 71; see main figures 2B and 3A for illustration). Boxes represent 1st and 3rd quartile. Black center lines represent medians. Whiskers extend to 1.5-fold of the inter-quartile range. Diamond symbols represent means. Black dots are outliers.














RMarkdown.RStudio.com

# Use-case: preventing this...







# ...by plain text version control with Git

## Unsynced changes

-  remove meta-info  
2 days ago by Katrin Leinweber
-  proof-read  
16 days ago by Katrin Leinweber
-  fixed judgemental "outlier" designati...  
26 days ago by Katrin Leinweber
-  reverted SF2 scale bar variant  
1 month ago by Katrin Leinweber
-  Peter's corrections & scale bar varian...  
1 month ago by Katrin Leinweber
-  moved incubation times to legends  
1 month ago by Katrin Leinweber
-  150213 PJ requests coherent label siz...  
1 month ago by Katrin Leinweber
-  renamed  
1 month ago by Katrin Leinweber
-  initial commit: as in 150211 PJ AchMi...  
1 month ago by Katrin Leinweber
-   Added .gitattributes  
1 month ago by Katrin Leinweber

## proof-read

	Katrin Leinweber		 
12	12		
13		- Identification of *A. minutissimum* capsules (asterisks) by subsequent observation of cell clusters by both bright-field and scanning electron microscopy of xenic biofilm (scale bars: 5 µm).	
	13	+ Identification of *A. minutissimum* capsules (asterisks) by successive observation of cell clusters by first bright-field and then scanning electron microscopy of xenic biofilm (scale bars: 5 µm).	
14	14		
15	15	**A:** Bright-field micrograph of crystal violet (CV) stained, 31 days old culture. Encapsulated cells (asterisks) are strongly stained, while weak staining indicates few extracellular polymeric substances (EPS) on the frustule surfaces. **B:** Scanning electron micrograph of the the same cell cluster. Encapsulated cells (asterisks) are surrounded by an opaque material. Frustule pores are visible on cells that did not possess a capsule in the hydrated biofilm. Note also the unequal distribution of bacteria ...(line truncated)...	
16	16		
...	...	@@ -30,7 +30,7 @@ Comparison of microstructures on *A. minutissimum* cell surfaces in a xenic biof	
30	30		
31	31		
32	32		
33		- Scanning electron micrographs of terminal parts of *A. minutissimum* cells at potentially different encapsulation stages of xenic biofilms (scale bars: 1 µm).	
	33	+ Scanning electron micrographs of terminal parts of *A. minutissimum* cells at potentially different encapsulation stages within xenic biofilms (scale bars: 1 µm).	
34	34		

# Use-case: easier collaboration on manuscripts



Paper Now: [github.com/PeerJ/paper-now](https://github.com/PeerJ/paper-now)

- ▶ Git-based template & generator for article websites
- ▶ no submission options (yet)



[Authorea.com](https://authorea.com)

- ▶ academic text editor with citations, formulas, figures, commenting, etc.
- ▶ 1-click-formatting & journal submission

# Use-case: offline Scientific Markdown<sup>4</sup>

In contrast, axenic *\achi* cells did not form biofilms, so that even careful rinsing left much fewer cells attached to the disks and thus available for SEM analysis.

This observation is in agreement with studies that utilised other growth substrates to compare biofilm formation by axenic and xenic diatom cultures. By measuring chl concentrations, the possibility that axenic cells might simply be less proliferate was excluded [Windler\_biofile\_2015]. Xenic *\achi* cultures on the other hand have also been found to develop biofilms on glass beads as well as in plastic multi-well plates [Lubarsky\_stabilisation\_2010; Windler\_biofile\_2015].

Our results demonstrate, that xenic biofilms of *\achi* can also be grown on Thermanox disks, enabling direct preparation for electron microscopy of native biofilm samples.

## \*\*\* Identification of *\achi* capsule microstructures

!["Identification of *\achi* capsules (asterisks) by successive observation of cell clusters by first bright-field and then scanning electron microscopy of xenic biofilm (scale bars: 5  $\mu$ m)."]

""A"" Bright-field micrograph of CV stained, 31 days old culture.

Encapsulated cells (asterisks) are strongly stained, while weak staining indicates few extracellular polymeric substances (EPS) on the frustule surfaces.

""B"" Scanning electron micrograph of the same cell cluster.

Encapsulated cells (asterisks) are surrounded by an opaque material. Frustule pores are visible on cells that did not possess a capsule in the hydrated biofilm.

Note also the unequal distribution of bacteria cells on capsules versus non-encapsulated frustules. \label{CLEM}][capsule-microstructure=figures/CLEM.png]

In order to correlate the hydrated *\achi* capsules visible in light microscopy to their dehydrated appearance in SEM, areas were marked by scratches on the CV stained disks and cells of interest were identified by BM. Subsequently, the same areas and cells were found again in the SEM (Fig. \ref{CLEM}).

The same technique was successfully applied to axenic cultures, despite the lower prevalence of adherent cells (Suppl. Fig. \ref{CLEM-ax}, \ref{CLEM-ax}).

after removal from the medium. Staining with the dye CV and subsequent bright-field microscopy showed that large portions of the diatom cells were surrounded by capsules.

In contrast, axenic *A. minutissimum* cells did not form biofilms, so that even careful rinsing left much fewer cells attached to the disks and thus available for SEM analysis. This observation is in agreement with studies that utilised other growth substrates to compare biofilm formation by axenic and xenic diatom cultures. By measuring chl concentrations, the possibility that axenic cells might simply be less proliferate was excluded [Windler et al., 2015]. Xenic *A. minutissimum* cultures on the other hand have also been found to develop biofilms on glass beads as well as in plastic multi-well plates [Lubarsky et al., 2010; Windler et al., 2015]. Our results demonstrate, that xenic biofilms of *A. minutissimum* can also be grown on Thermanox disks, enabling direct preparation for electron microscopy of native biofilm samples.

## Identification of *A. minutissimum* capsule microstructures

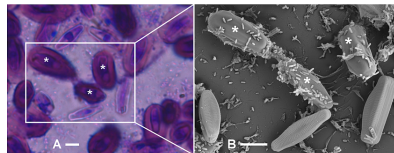


Figure 4.8: Identification of *A. minutissimum* capsules (asterisks) by successive observation of cell clusters by first bright-field and then scanning electron microscopy of xenic biofilm (scale bars: 5  $\mu$ m). A: Bright-field micrograph of CV

toolset for bridging Markdown to  $\text{\LaTeX}$  (and anything else)

# Write in Word? Save in Markdown! Publish in L<sup>A</sup>T<sub>E</sub>X!

- ▶ Writage.com adds Markdown support in Word
- ▶ messy plain text, renamed image files & lost figure captions

Capsules of the diatom *Achnantheidum minutissimum* arise from fibrillar precursors and foster attachment of bacteria

## Abstract

Please note: This is an experimental Paper Now version of this PeerJ article based on [this source repository](#). No guarantees are given for the correctness or completeness of this experimental version.

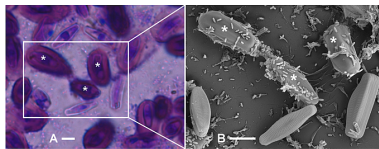


Figure 1: Crystal violet (CV) stained capsules (grey ovals) in xenic *A. minutissimum* biofilm (scale bar: 20  $\mu$ m).

Capsules of the diatom *Achnantheidum minutissimum* arise from fibrillar precursors and foster attachment of bacteria

## Abstract

**Please note:** This is an experimental **Paper Now** (<https://github.com/PeerJ/paper-now>) **version** of **this PeerJ article** (<https://peerj.com/articles/858/>) **based on** **this source repository** (<https://github.com/katrinleinweber/paper-now/>). No guarantees are given for the correctness or completeness of this experimental version.



Figure 1: Crystal violet (CV) stained capsules (grey ovals) in xenic *A. minutissimum* biofilm (scale bar: 20  $\mu$ m).

Write in Word,  
Save in Markdown,  
Publish in L<sup>A</sup>T<sub>E</sub>X



~~Write in Word,~~  
Use Markdown wherever possible,  
Convert to whatever is necessary.

# Thanks for your attention! Questions?

*katrin.leinweber@uni-konstanz.de*

*notes, links & slides on [konsense.de/md](https://konsense.de/md)*

## Acknowledgements

- ▶ retreat organisers
- ▶ [github.com/JensErat/scientific-markdown](https://github.com/JensErat/scientific-markdown)

## Funding



Actual lab work and thesis writing happened as well ;-)