# STAT 656: Bayesian Data Analysis Fall 2024 Homework 1

Juanwu Lu\*

# Synthetic Data

The autoregressive model is frequently used to analyze time series data. The simplest autoregressive model has order 1, and is abbreviated as AR(1). This model assumes that an observation  $y_i$  at time point i (i = 1, ..., n) is generated according to

$$y_i = \rho y_{i-1} + \epsilon_i,$$

where  $\epsilon_i \sim \mathcal{N}(0, \sigma^2)$  independently, and rho and  $\sigma$  are unknown parameters. For simplicity, we shall assume that  $y_0$  is a fixed constant. We will also assume  $|\rho| < 1$ .

### 1. (5 points) Solution:

Given the formulation above, the log-likelihood function is calculated as follows:

$$\log L(\rho, \sigma^2 | y_0, y_1, \dots, y_n) = \log \prod_{i=1}^n (2\pi\sigma^2)^{-\frac{1}{2}} \cdot \exp\left\{-\frac{(y_i - \rho y_{i-1})^2}{2\sigma^2}\right\}$$
$$= -\frac{n}{2} \log(2\pi\sigma^2) - \frac{1}{2\sigma^2} \sum_{i=1}^n (y_i - \rho y_{i-1})^2$$
$$= -\frac{n}{2} \log(2\pi) - n \log(\sigma) - \frac{1}{2\sigma^2} \sum_{i=1}^n (y_i - \rho y_{i-1})^2.$$

#### 2. (10 points) Solution:

The code implementation is as follows:

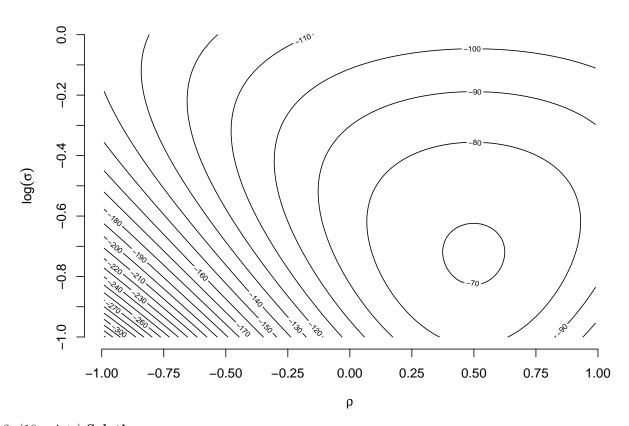
```
# Read data
if (file.exists("data/computation_data_hw_1.csv")) {
   data <- read.csv("data/computation_data_hw_1.csv")
   y <- data[["x"]]
} else {
   stop("Cannot find the file 'data/computation_data_hw_1.csv' at ", getwd())
}

# Define the log-likelihood function
ar_loglik <- function(rho, log_sig) {
   y <- .GlobalEnv$y
   n <- length(y)
   sig <- exp(log_sig)</pre>
```

<sup>\*</sup>College of Engineering, Purdue University, West Lafayette, IN, USA

```
rho <- rep(as.numeric(rho), times = n - 1)</pre>
  denom \leftarrow n * log(2 * pi) + 2 * n * log_sig
  log_lik <- -0.5 * (
    denom + (y[1]^2 + sum((y[2:n] - rho * y[1:(n - 1)])^2)) / sig^2
  )
  return(log_lik)
}
# Visualization
rho \leftarrow seq(-0.99, 0.99, length = 100)
log_sig \leftarrow seq(-1.0, 0.0, length = 100)
loglik <- outer(rho, log_sig, Vectorize(ar_loglik))</pre>
contour(
  x = rho,
  y = log_sig,
  z = loglik,
  xlab = expression(rho),
  ylab = expression(log(sigma)),
  main = "Log-Likelihood Function",
  nlevels = 20,
  axes = FALSE,
axis(side = 1, at = seq(-1.0, 1.0, by = 0.25))
axis(side = 2, at = seq(-1.0, 0.0, by = 0.10))
```

# Log-Likelihood Function



3. (10 points) Solution:

Since the prior is independent, we have  $p(\rho, \log(\sigma)) = p(\rho)p(\log(\sigma)) = \frac{1}{2} \cdot \frac{1}{10\sqrt{2\pi}} \exp\left\{-\frac{\log(\sigma)^2}{200}\right\}$ . Therefore, the posterior density is proportional to the product of the likelihood and the prior, and then the log of the posterior density is calculated up to a constant as follows:

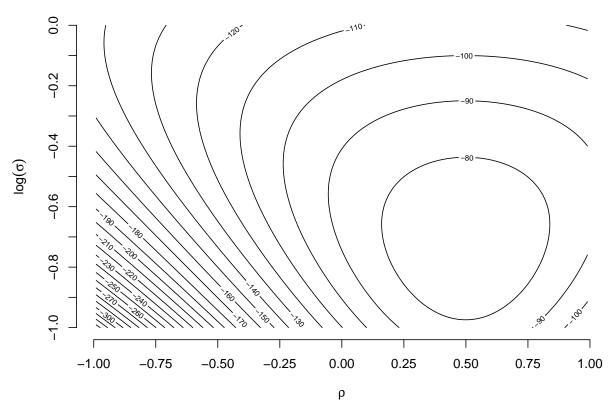
$$\log p(\rho, \log(\sigma)|y_0, y_1, \dots, y_n) = \log L(\rho, \log(\sigma)|y_0, y_1, \dots, y_n) + \log p(\rho, \log(\sigma)) + C$$

$$= -n\log(\sigma) - \frac{1}{2\exp(2*\log(\sigma))} \sum_{i=1}^{n} (y_i - \rho y_{i-1})^2 - \frac{\log(\sigma)^2}{200} + C'$$

where C is the constant log-normalizer and C' is a constant irrelevant to  $\rho$  and  $\log(\sigma)$ . The visualization of the log of the posterior density is as follows:

```
ar_logpost <- function(rho, log_sig) {</pre>
  \log_{prior} \leftarrow -\log(2) - 0.5 * (\log(2 * pi) + \log(100) + \log_{sig}^2 / 100)
  log_post <- ar_loglik(rho, log_sig) + log_prior</pre>
  return(log_post)
logpost <- outer(rho, log_sig, Vectorize(ar_logpost))</pre>
contour(
  x = rho,
  y = log_sig,
  z = logpost,
  main = "Unnormalized Log-Posterior Density",
  xlab = expression(rho),
  ylab = expression(log(sigma)),
  nlevels = 20,
  axes = FALSE,
axis(side = 1, at = seq(-1.0, 1.0, by = 0.25))
axis(side = 2, at = seq(-1.0, 0.0, by = 0.10))
```

### **Unnormalized Log-Posterior Density**



Compared to the log-likelihood function, the log posterior density is more concentrated around the maximum likelihood estimate of  $(\rho, \log(\sigma))^{\mathsf{T}}$ . The prior is not overly informative since the shape of the posterior density is still similar to the likelihood function, indicating that the posterior is **mainly** determined by the likelihood function.

#### 4. (10 points) Solution:

From the visualization in 3., we can see that the posterior density is concentrated around  $0.25 \le \rho \le 0.75$  and  $-0.9 \le \log(\sigma) \le -0.5$ . Therefore, we can choose a grid of  $\rho$  and  $\log(\sigma)$  as follows:

```
set.seed(42)
rho_grid <- seq(0.25, 0.75, length = 100)
log_sig_grid <- seq(-0.90, -0.50, length = 100)
grid_log_post <- outer(rho_grid, log_sig_grid, Vectorize(ar_logpost))
# Calculate the normalized probability density
probs <- exp(grid_log_post - max(grid_log_post))
probs <- probs / sum(probs)
# Randomly sample from the grid
indices <- sample(
    x = seq_len(length(as.vector(probs))),
    size = 1000,
    replace = TRUE,
    prob = probs
)
rho_sample <- rho_grid[((indices - 1) %% nrow(probs)) + 1]
log_sig_sample <- log_sig_grid[((indices - 1) %% nrow(probs)) + 1]</pre>
```

### 5. (5 points) Solution:

The code implementation is as follows:

```
library(moments)
  # Calculate summaries for rho
  print(quantile(rho_sample, probs = c(0.025, 0.25, 0.5, 0.75, 0.975)))
  ##
           2.5%
                      25%
                                 50%
                                           75%
                                                   97.5%
  ## 0.3357323 0.4419192 0.4974747 0.5542929 0.6691919
  sprintf("Mean: %.4f", mean(rho_sample))
  ## [1] "Mean: 0.5002"
  sprintf("Standard Deviation: %.4f", sd(rho_sample))
  ## [1] "Standard Deviation: 0.0847"
  sprintf("Skewnewss: %.4f", skewness(rho_sample))
  ## [1] "Skewnewss: 0.0490"
  sprintf("Kurtosis: %.4f", kurtosis(rho_sample))
  ## [1] "Kurtosis: 3.0670"
  # Calculate summaries for log(sigma)
  print(quantile(log_sig_sample, probs = c(0.025, 0.25, 0.5, 0.75, 0.975)))
            2.5%
                        25%
                                    50%
                                               75%
                                                         97.5%
  ## -0.8516162 -0.7707071 -0.7222222 -0.6696970 -0.5646465
  sprintf("Mean: %.4f", mean(log_sig_sample))
  ## [1] "Mean: -0.7199"
  sprintf("Standard Deviation: %.4f", sd(log_sig_sample))
  ## [1] "Standard Deviation: 0.0710"
  sprintf("Skewness: %.4f", skewness(log_sig_sample))
  ## [1] "Skewness: 0.1683"
  sprintf("Kurtosis: %.4f", kurtosis(log_sig_sample))
  ## [1] "Kurtosis: 2.7730"
6. (10 points) Solution:
  The code implementation is as follows:
  ar_post_predictive <- function(rho, log_sig) {</pre>
    # Sample from the posterior predictive distribution
    y <- .GlobalEnv$y
    new_y <- rep(0.0, times = length(y))</pre>
    new_y[1] \leftarrow y[1]
    sig <- exp(log_sig)</pre>
    for (i in 2:length(y)) {
      new_y[i] \leftarrow rnorm(n = 1, mean = rho * y[i - 1], sd = sig)
    }
    return(new_y)
```

```
params <- data.frame(rho = rho_sample, log_sig = log_sig_sample)</pre>
samples <- matrix(NA, nrow = nrow(params), ncol = length(y))</pre>
for (i in seq_len(nrow(params))) {
  samples[i, ] <- ar_post_predictive(params[i, "rho"], params[i, "log_sig"])</pre>
}
```

The above code makes use of the grid samples drawn in problem 4. For each pair of  $(\rho, \log(\sigma))$ , we use the AR(1) model to generate a new sample of sequences. The summary statistics of the posterior

```
predictive distribution are as follows:
print("Sequence means:")
## [1] "Sequence means:"
print(colMeans(samples))
##
     [1]
         0.00000000 -0.004768199 0.380467600 0.046329176
                                                            0.041871692
##
         0.230962145 -0.239262815 0.214538988 0.320248721
                                                            0.833946396
##
    [11] -0.047636240 0.154295216 0.091970943 0.491005558
                                                            0.167442727
##
    Г16Т
         0.448882392 0.221520741 0.374755026 -0.241297744 -0.400832166
##
    [21]
        0.027256619 -0.401138975 0.116944379 -0.011085951 0.170830053
##
    [31] -0.604599248 -0.828631029 -0.414124777 -0.153571316 -0.065036616
##
##
    [36]
         0.016159584 0.160684738 0.128898047 -0.165708086
                                                           0.295168094
##
    [41]
         0.400073542 - 0.100819868 \quad 0.057687704 - 0.194070728 - 0.274464118
    [46] -0.054924127 -0.393364772 -0.596120509 -0.254331298
        ##
    [51]
                                                            0.271008982
    [56]
         0.005264908 -0.169212597 0.181303334 0.197683160
                                                            0.174879318
##
        0.409731341 -0.031243178 -0.011589809 0.019277832 0.164153831
    [66] -0.070950225 -0.322370283 -0.174962173 -0.134343046 -0.2133333649
    [71] -0.360695027 0.253886889 -0.063299359 -0.218656995 -0.072649060
    [76]
        0.038109116 0.091598943 0.439842892 0.175992690 -0.205825087
##
   [81]
         0.249317439 -0.038973702 0.255908060 -0.029138104 -0.056703887
   [86] -0.183220258 -0.158864768 -0.146437883 -0.186715701 -0.252234505
    [91] -0.172413509 0.005136642 -0.111024282 0.241944648 0.249826061
    [96] 0.067545792 0.219785394 0.366799730 0.354160286 -0.187007079
print("Sequence standard deviations:")
## [1] "Sequence standard deviations:"
print(apply(samples, 2, sd))
##
     [1] 0.0000000 0.4875799 0.4952396 0.4853952 0.4985296 0.5092669 0.4943722
##
     [8] 0.5042714 0.4834231 0.4975461 0.4991716 0.4769155 0.4994349 0.4932221
##
    [15] 0.4738587 0.4824026 0.4765846 0.4818143 0.5206273 0.4930490 0.4913379
    [22] 0.4697823 0.5076087 0.4998257 0.4994883 0.4776279 0.5204864 0.4897410
    [29] 0.5075770 0.5083453 0.4970612 0.5102311 0.5127232 0.4965778 0.4776371
    [36] 0.4897456 0.4745159 0.4765058 0.4915986 0.4936407 0.4896082 0.5106595
    [43] 0.4947319 0.4906189 0.5111469 0.4905928 0.4780761 0.5285908 0.5027156
##
    [50] 0.4952306 0.4885762 0.4887156 0.4954808 0.4826266 0.5065151 0.4840795
    [57] \quad 0.4783016 \quad 0.4926563 \quad 0.5074513 \quad 0.4827778 \quad 0.4915453 \quad 0.4948413 \quad 0.4831472
##
    [64] 0.4882579 0.4893974 0.4978840 0.4923274 0.4964880 0.4812996 0.5041211
    [71] 0.4991301 0.4958067 0.5055281 0.4947765 0.4903025 0.4971520 0.4988327
##
    [78] 0.4844680 0.4888304 0.4860582 0.4848816 0.4869918 0.4963936 0.4814153
    [85] 0.4818288 0.5007236 0.4806716 0.5092220 0.4856871 0.4953563 0.4998521
```

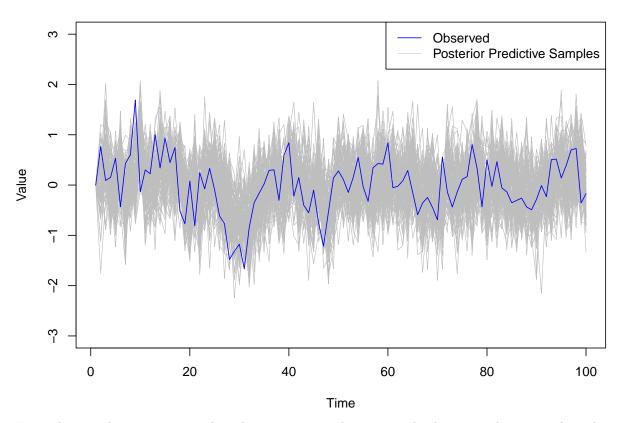
```
## [92] 0.4754400 0.4945970 0.4920551 0.4866788 0.4860907 0.4912455 0.4940571 ## [99] 0.4891187 0.4764067
```

#### 7. (10 points) Solution:

The visualization of the posterior predictive samples against the observed data is as follows:

```
x = seq_len(length(y)),
 y = samples[1, ],
 col = "gray",
 type = "1",
 1wd = 0.75,
 main = "Posterior Predictive Samples vs. Observed Data",
 xlab = "Time",
 ylab = "Value";
 ylim = c(-3.0, 3.0)
for (i in 2:100) {
  lines(x = seq_len(length(y)), y = samples[i, ], col = "gray", lwd = 0.75)
lines(x = seq_len(length(y)), y = y, col = "blue", lwd = 1.0)
legend(
  "topright",
 legend = c("Observed", "Posterior Predictive Samples"),
 col = c("blue", "gray"),
 lwd = c(1, 0.5),
  bg = "white",
```

### Posterior Predictive Samples vs. Observed Data



From the visualization, we see that the posterior predictive samples have a wider range than the observed data, indicating that the model has a higher uncertainty in prediction and may not be able to capture the true data distribution well. Therefore, the model is **not a good fit** for the observed data. My expectation for a good model is that the posterior predictive samples are close to the observed data with a similar tendency and range.

### Real Data

### 1. (5 points) Solution:

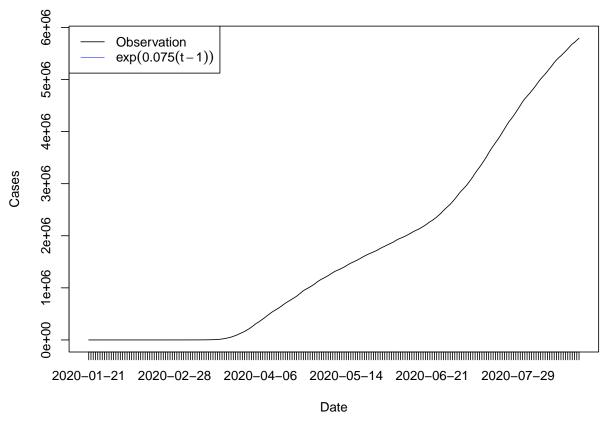
No, I do not believe that the information given is sufficient for analyzing the data. The GitHub page has only provided the dataset file, source of data, and definitions of different sorts of cases. All these pieces of information are related to the observation, i.e., COVID-19 cases, but none of them reflect any information to support the prior distribution of the parameters in the model. Other information such as demographic statistics, geofencing data, activity data, etc., can be useful for data analysis.

#### 2. (5 points) Solution:

Before establishing the model, the raw data is visualized as follows:

```
# Read data
if (file.exists("data/covid_us.txt")) {
 data <- read.table("data/covid_us.txt", header = TRUE, sep = ",")</pre>
 y <- data[["cases"]]</pre>
} else {
  stop("File not found: 'covid_us.txt' at ", getwd())
}
plot(
 x = seq_len(length(y)),
 y = y,
 type = "1",
 main = "COVID-19 Cases in the US",
 xlab = "Date",
 ylab = "Cases",
 xaxt = "n"
axis(side = 1, labels = data[["date"]], at = seq len(length(y)))
legend(
  "topleft",
 legend = c("Observation", expression(y=exp(0.075 * (t - 1)))),
  col = c("black", "blue"),
 lwd = c(1, 0.5),
 bg = "white",
```

#### COVID-19 Cases in the US



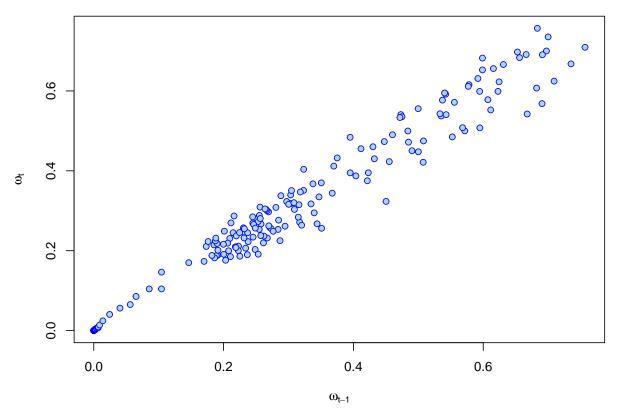
From the visualization, it is clear that the raw data has an exponential growth trend (blue line), which can not be directly captured by the AR(1) model. Therefore, the raw data needs to be transformed before being used for fitting an AR(1) model. The transformation consists of two steps: first, differentiating the transformed with lag = 1 to filter out the non-stationary trend that is not captured by the AR(1) model, and then, scale the differentiated data by a factor of 1/100000 to make the data lying in a reasonable range:

$$\omega_t = \frac{1}{100000} (y_t - y_{t-1}).$$

For consistency,  $\omega_1 = 0$ . The visualization of the transformed data is as follows:

```
omg <- c(0, diff(y, lag = 1) / 100000)
plot(
    x = omg[seq(1, length(omg) - 1)],
    y = omg[seq(2, length(omg))],
    type = "p",
    pch = 21,
    col = "blue",
    bg = "lightblue",
    main = "Visualization of Transformed Data",
    xlab = expression(omega[t - 1]),
    ylab = expression(omega[t]),
)</pre>
```

### **Visualization of Transformed Data**



As shown in the visualization, the transformed data has a more linear relationship between consecutive observations. Finally, the AR(1) model is constructed as:

$$\omega_i = \rho \omega_{i-1} + \epsilon_i, \quad \epsilon_i \sim \mathcal{N}(0, \sigma^2).$$

#### 3. (5 points) Solution:

Based on the visualization above, a majority of the point cloud is growing linearly with a slope around 1.0 and slopes are non-negative. Therefore, the prior distribution of  $\rho$  can be set as a Gamma distribution  $\rho \sim \text{Gamma}(1,1)$  with a mean of 1.0. For the log-variance  $\log(\sigma)$ , without losing generality, the prior can be set a normal distribution  $\log(\sigma) \sim \mathcal{N}(0,1^2)$ .

#### 4. (10 points) Solution:

With the above specified prior distributions, the log-posterior density is calculated as follows:

$$\log p(\rho, \log(\sigma)|\omega_{1}, \dots, \omega_{n}) = \log L(\rho, \log(\sigma)|\omega_{1}, \dots, \omega_{n}) + \log p(\rho) + \log p(\log(\sigma)) + C$$

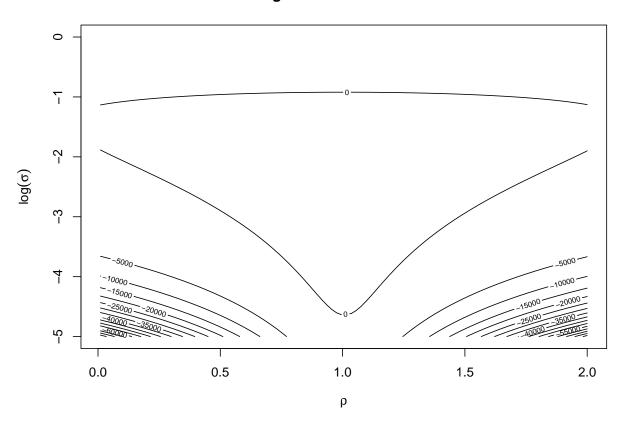
$$= -\frac{n}{2}\log(2\pi\sigma^{2}) - \frac{1}{2\sigma^{2}}\sum_{i=1}^{n}(\omega_{i} - \rho\omega_{i-1})^{2} + \log\left(\frac{1}{\Gamma(1)}\exp(-\rho)\right) - \frac{\log(\sigma)^{2}}{2} + C'$$

$$= -\frac{n}{2}\log(2\pi\sigma^{2}) - \frac{1}{2\sigma^{2}}\sum_{i=1}^{n}(\omega_{i} - \rho\omega_{i-1})^{2} - \rho - \frac{\log(\sigma)^{2}}{2} + C',$$

Therefore, the model fitted using data from the first time point until June 30 is as follows:

```
rho <- seq(0.01, 2.0, length = 100)
log_sig <- seq(-5.0, 0.0, length = 100)
log_lik <- outer(rho, log_sig, Vectorize(ar_loglik))
contour(
    x = rho,
    y = log_sig,
    z = log_lik,
    main = "Log-Likelihood Function",
    xlab = expression(rho),
    ylab = expression(log(sigma)),
    nlevels = 20,
)
</pre>
```

# Log-Likelihood Function

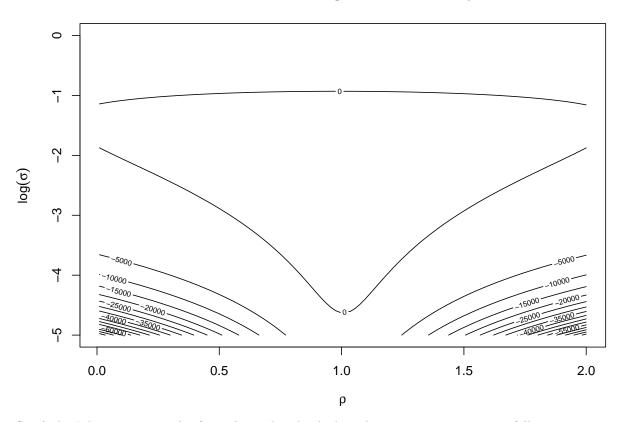


The visualization of the logarithm of unnormalized posterior density is shown below:

```
ar_logpost <- function(rho, log_sig) {
  log_prior <- -rho - 0.5 * (log_sig * log_sig)
  log_post <- ar_loglik(rho, log_sig) + log_prior
  return(log_post)
}
log_post <- outer(rho, log_sig, Vectorize(ar_logpost))
contour(
  x = rho,
  y = log_sig,
  z = log_post,
  main = "Unnormalized Log-Posterior Density",</pre>
```

```
xlab = expression(rho),
ylab = expression(log(sigma)),
nlevels = 20,
)
```

### **Unnormalized Log-Posterior Density**



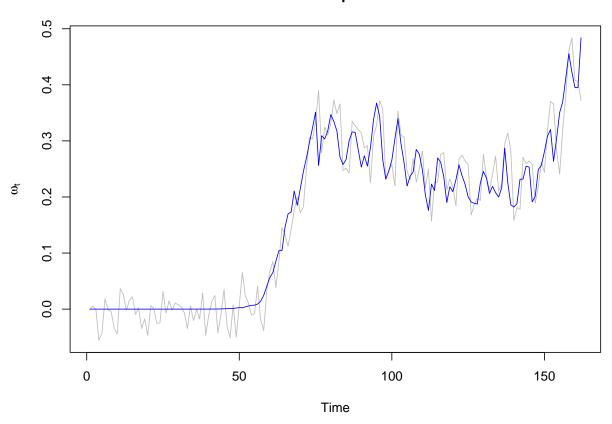
Similarly, I draw 1000 samples from the grid and calculate the summary statistics as follows:

```
library(moments)
grid_log_post <- outer(rho, log_sig, Vectorize(ar_logpost))</pre>
# Calculate the normalized probability density
probs <- exp(grid_log_post - max(grid_log_post))</pre>
probs <- probs / sum(probs)</pre>
# Randomly sample from the grid
indices <- sample(</pre>
  x = seq_len(length(as.vector(probs))),
  size = 1000,
  replace = TRUE,
  prob = probs
rho_sample <- rho[((indices - 1) %% nrow(probs)) + 1]</pre>
log_sig_sample <- log_sig[((indices - 1) %/% nrow(probs)) + 1]</pre>
# Calculate summaries for rho
print(quantile(rho_sample, probs = c(0.025, 0.25, 0.5, 0.75, 0.975)))
##
        2.5%
                                                  97.5%
                    25%
                               50%
                                          75%
```

```
## 0.9949495 0.9949495 1.0150505 1.0150505 1.0150505
  sprintf("Mean: %.4f", mean(rho_sample))
  ## [1] "Mean: 1.0088"
  sprintf("Standard Deviation: %.4f", sd(rho_sample))
  ## [1] "Standard Deviation: 0.0102"
  sprintf("Skewnewss: %.4f", skewness(rho_sample))
  ## [1] "Skewnewss: -0.3685"
  sprintf("Kurtosis: %.4f", kurtosis(rho_sample))
  ## [1] "Kurtosis: 2.2449"
  # Calculate summaries for log(sigma)
  print(quantile(log_sig_sample, probs = c(0.025, 0.25, 0.5, 0.75, 0.975)))
  ##
          2.5%
                      25%
                                50%
                                           75%
                                                   97.5%
  ## -3.737374 -3.636364 -3.636364 -3.585859 -3.484848
  sprintf("Mean: %.4f", mean(log_sig_sample))
  ## [1] "Mean: -3.6194"
  sprintf("Standard Deviation: %.4f", sd(log_sig_sample))
  ## [1] "Standard Deviation: 0.0571"
  sprintf("Skewness: %.4f", skewness(log_sig_sample))
  ## [1] "Skewness: 0.1094"
  sprintf("Kurtosis: %.4f", kurtosis(log_sig_sample))
  ## [1] "Kurtosis: 3.3055"
5. (10 points) Solution:
  Similarly, the posterior predictive samples are compared to the observed transformed data. The code
  implementation is as follows:
  params <- data.frame(rho = rho_sample, log_sig = log_sig_sample)</pre>
  samples <- matrix(NA, nrow = nrow(params), ncol = length(y))</pre>
  for (i in seq_len(nrow(params))) {
    samples[i, ] <- ar_post_predictive(params[i, "rho"], params[i, "log_sig"])</pre>
  }
  plot(
    x = seq along(samples[1, seq(1, length(y))]),
    y = samples[1, seq(1, length(y))],
    type = "l",
    col = "gray";
    xlab = "Time",
    ylab = expression(omega[t]),
    main = "Posterior Predictive Samples vs. Observed Data",
  for (i in 2:1000) {
    lines(x = seq_along(samples[i]), y = samples[i], col = "gray")
```

```
}
lines(x = seq_along(y), y = y, col = "blue")
```

### Posterior Predictive Samples vs. Observed Data

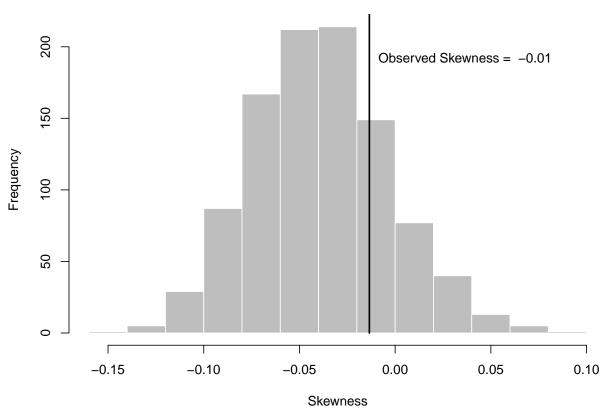


Following the posterior predictive check covered in the lecture, the visualization of the skewness and kurtosis of the posterior predictive samples against the observed data is as follows:

```
skewness_obs <- skewness(y[seq(2, length(y))])</pre>
sample_skewness <- apply(samples, 1, function(x) skewness(x[seq(2, length(y))]))</pre>
kurtosis_obs <- kurtosis(y[seq(2, length(y))])</pre>
sample_kurtosis <- apply(samples, 1, function(x) kurtosis(x[seq(2, length(y))]))</pre>
hist(
  x = sample_skewness,
  main = "Skewness of Posterior Predictive Samples",
  xlab = "Skewness",
  col = "gray",
  border = "white",
lines(
  x = c(skewness_obs, skewness_obs),
  y = c(0, 300),
  col = "black",
  lwd = 2,
)
text(
  x = skewness_obs + 0.05,
 y = 200,
```

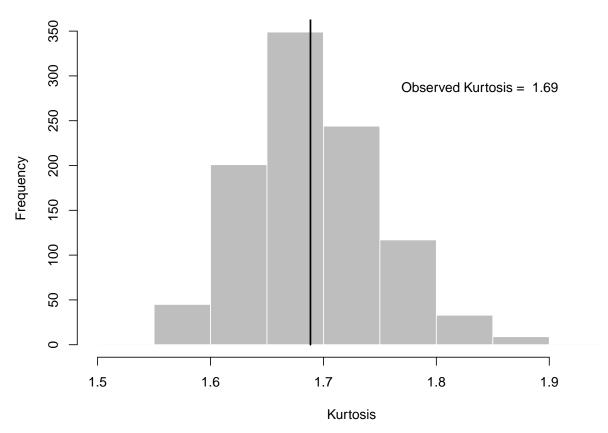
```
labels = paste("Observed Skewness = ", round(skewness_obs, 2)),
pos = 1
)
```

# **Skewness of Posterior Predictive Samples**



```
hist(
    x = sample_kurtosis,
    main = "Kurtosis of Posterior Predictive Samples",
    xlab = "Kurtosis",
    col = "gray",
    border = "white",
)
lines(
    x = c(kurtosis_obs, kurtosis_obs),
    y = c(0, 500),
    col = "black",
    lwd = 2,
)
text(
    x = kurtosis_obs + 0.15,
    y = 300,
    labels = paste("Observed Kurtosis = ", round(kurtosis_obs, 2)),
    pos = 1
)
```

### **Kurtosis of Posterior Predictive Samples**



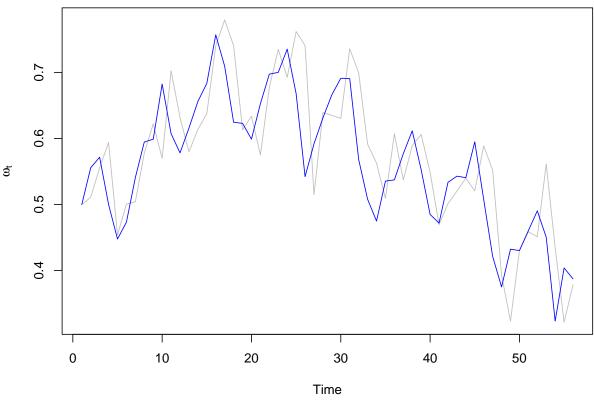
Based on the posterior predictive checks, the model provides good predictions for the data until June 30. The skewness and kurtosis of the observed data are close to the expected values from the posterior predictive samples, and the time-series visualization shows that the posterior predictive samples are close to the observed data.

#### 6. (10 points) Solution:

The code implementation is as follows:

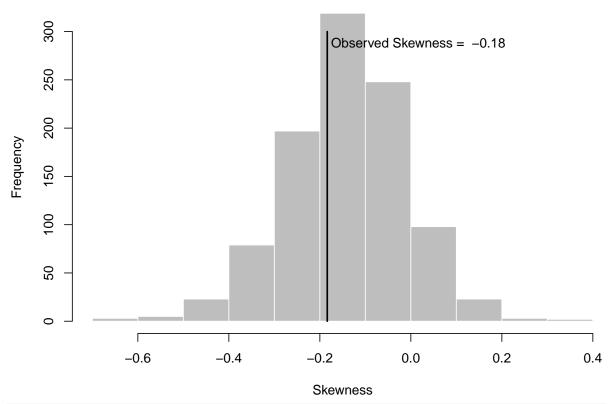
```
lines(x = seq_along(samples[i]), y = samples[i], col = "gray")
}
lines(x = seq_along(y), y = y, col = "blue")
```

### **Posterior Predictive Samples vs. Observed Data**



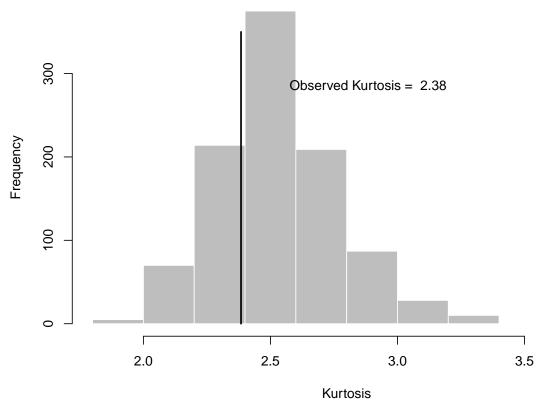
```
skewness_obs <- skewness(y[seq(2, length(y))])</pre>
sample_skewness <- apply(samples, 1, function(x) skewness(x[seq(2, length(y))]))</pre>
hist(
  x = sample_skewness,
  main = "Skewness of Posterior Predictive Samples",
 xlab = "Skewness",
 col = "gray",
  border = "white",
lines(
  x = c(skewness_obs, skewness_obs),
 y = c(0, 300),
 col = "black",
  lwd = 2,
text(
  x = skewness_obs + 0.2,
  y = 300,
  labels = paste("Observed Skewness = ", round(skewness_obs, 2)),
```

# **Skewness of Posterior Predictive Samples**



```
kurtosis_obs <- kurtosis(y[seq(2, length(y))])</pre>
sample_kurtosis <- apply(samples, 1, function(x) kurtosis(x[seq(2, length(y))]))</pre>
hist(
  x = sample_kurtosis,
 main = "Kurtosis of Posterior Predictive Samples",
 xlab = "Kurtosis",
 col = "gray",
  border = "white",
)
lines(
  x = c(kurtosis_obs, kurtosis_obs),
  y = c(0, 350),
 col = "black",
  lwd = 2,
text(
  x = kurtosis_obs + 0.5,
 y = 300,
  labels = paste("Observed Kurtosis = ", round(kurtosis_obs, 2)),
 pos = 1
```

### **Kurtosis of Posterior Predictive Samples**



Based on the posterior predictive checks, the model provides a good prediction for the future data from June 30 to August 25. The skewness and kurtosis of the observed data are close to the expected values from the posterior predictive samples, and the time-series visualization shows that the posterior predictive samples are close to the observed data.

#### 7. (10 points) Solution:

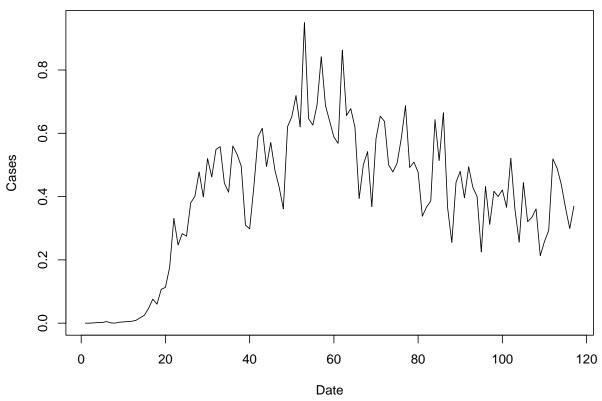
The code implementation is as follows:

```
# Read data
if (file.exists("data/covid_us.txt")) {
    data <- read.table("data/covid_us-states.txt", header = TRUE, sep = ",")
    data <- data[data["state"] == "Indiana", ] # Filter data for Indiana
    y <- data[["cases"]]
} else {
    stop("File not found: 'covid_us-states.txt' at ", getwd())
}

y <- c(
    0,
    diff(y[as.Date(data$date) <= as.Date("2020-06-30")], lag = 1) / 1000
)
plot(
    x = seq_len(length(y)),
    y = y,
    type = "l",
    main = "Daily COVID-19 Case Increase in Indiana",</pre>
```

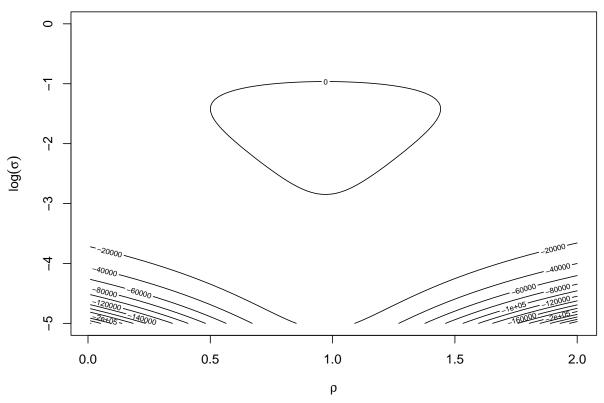
```
xlab = "Date",
ylab = "Cases",
)
```

# Daily COVID-19 Case Increase in Indiana



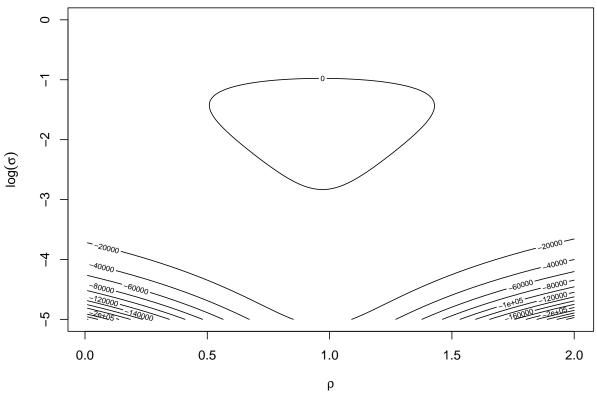
```
rho <- seq(0.01, 2.0, length = 100)
log_sig <- seq(-5.0, 0.0, length = 100)
log_lik <- outer(rho, log_sig, Vectorize(ar_loglik))
contour(
    x = rho,
    y = log_sig,
    z = log_lik,
    main = "Log-Likelihood Function",
    xlab = expression(rho),
    ylab = expression(log(sigma)),
    nlevels = 20,
)</pre>
```

# **Log-Likelihood Function**



```
ar_logpost <- function(rho, log_sig) {
  log_prior <- -rho - 0.5 * (log_sig * log_sig)
  log_post <- ar_loglik(rho, log_sig) + log_prior
  return(log_post)
}
log_post <- outer(rho, log_sig, Vectorize(ar_logpost))
contour(
  x = rho,
  y = log_sig,
  z = log_post,
  main = "Unnormalized Log-Posterior Density",
  xlab = expression(rho),
  ylab = expression(log(sigma)),
  nlevels = 20,
)</pre>
```

### **Unnormalized Log-Posterior Density**

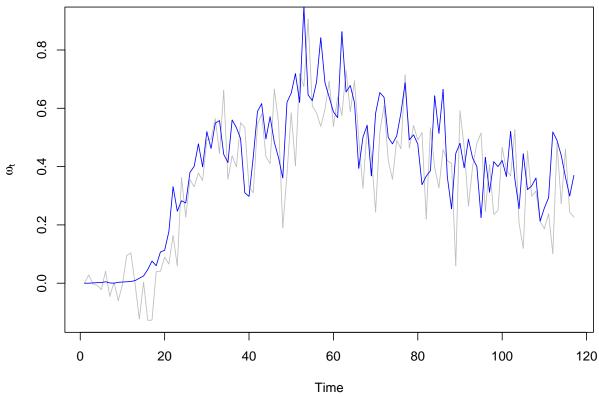


```
grid_log_post <- outer(rho, log_sig, Vectorize(ar_logpost))</pre>
# Calculate the normalized probability density
probs <- exp(grid_log_post - max(grid_log_post))</pre>
probs <- probs / sum(probs)</pre>
# Randomly sample from the grid
indices <- sample(</pre>
 x = seq_len(length(as.vector(probs))),
 size = 1000,
 replace = TRUE,
 prob = probs
rho_sample <- rho[((indices - 1) %% nrow(probs)) + 1]</pre>
log_sig_sample <- log_sig[((indices - 1) %/% nrow(probs)) + 1]</pre>
# Calculate summaries for rho
print(quantile(rho_sample, probs = c(0.025, 0.25, 0.5, 0.75, 0.975)))
##
        2.5%
                    25%
                              50%
                                         75%
                                                  97.5%
## 0.9145455 0.9547475 0.9748485 0.9949495 1.0150505
sprintf("Mean: %.4f", mean(rho_sample))
## [1] "Mean: 0.9712"
sprintf("Standard Deviation: %.4f", sd(rho_sample))
```

## [1] "Standard Deviation: 0.0248"

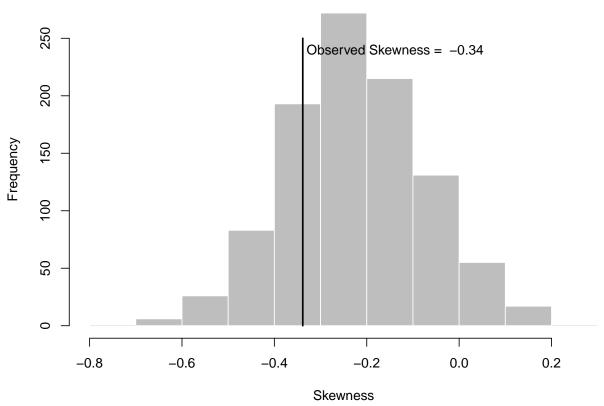
```
sprintf("Skewnewss: %.4f", skewness(rho_sample))
## [1] "Skewnewss: -0.0953"
sprintf("Kurtosis: %.4f", kurtosis(rho_sample))
## [1] "Kurtosis: 3.0754"
# Calculate summaries for log(sigma)
print(quantile(log_sig_sample, probs = c(0.025, 0.25, 0.5, 0.75, 0.975)))
##
                                        75%
        2.5%
                   25%
                              50%
                                                97.5%
## -2.272727 -2.171717 -2.171717 -2.121212 -2.020202
sprintf("Mean: %.4f", mean(log_sig_sample))
## [1] "Mean: -2.1523"
sprintf("Standard Deviation: %.4f", sd(log_sig_sample))
## [1] "Standard Deviation: 0.0664"
sprintf("Skewness: %.4f", skewness(log_sig_sample))
## [1] "Skewness: 0.0944"
sprintf("Kurtosis: %.4f", kurtosis(log_sig_sample))
## [1] "Kurtosis: 3.1947"
params <- data.frame(rho = rho_sample, log_sig = log_sig_sample)</pre>
samples <- matrix(NA, nrow = nrow(params), ncol = length(y))</pre>
for (i in seq_len(nrow(params))) {
  samples[i, ] <- ar_post_predictive(params[i, "rho"], params[i, "log_sig"])</pre>
plot(
  x = seq_along(samples[1, seq(1, length(y))]),
  y = samples[1, seq(1, length(y))],
  type = "1",
  col = "gray",
  xlab = "Time",
 ylab = expression(omega[t]),
 main = "Posterior Predictive Samples vs. Observed Data",
for (i in 2:1000) {
  lines(x = seq_along(samples[i]), y = samples[i], col = "gray")
lines(x = seq_along(y), y = y, col = "blue")
```

### Posterior Predictive Samples vs. Observed Data



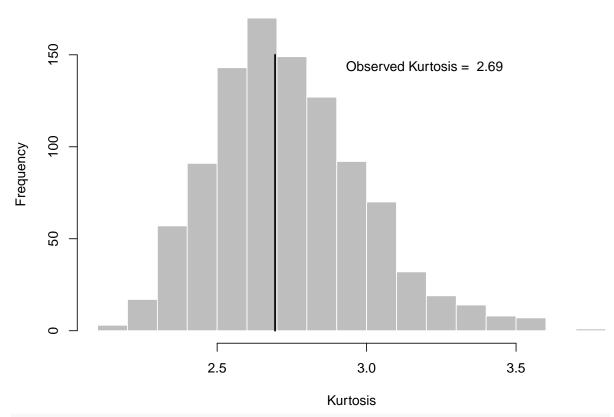
```
skewness_obs <- skewness(y[seq(2, length(y))])</pre>
sample_skewness <- apply(samples, 1, function(x) skewness(x[seq(2, length(y))]))</pre>
hist(
 x = sample_skewness,
 main = "Skewness of Posterior Predictive Samples",
 xlab = "Skewness",
  col = "gray",
  border = "white",
)
lines(
  x = c(skewness_obs, skewness_obs),
 y = c(0, 250),
  col = "black",
  lwd = 2,
)
text(
  x = skewness_obs + 0.2,
  y = 250,
  labels = paste("Observed Skewness = ", round(skewness_obs, 2)),
 pos = 1
)
```

# **Skewness of Posterior Predictive Samples**



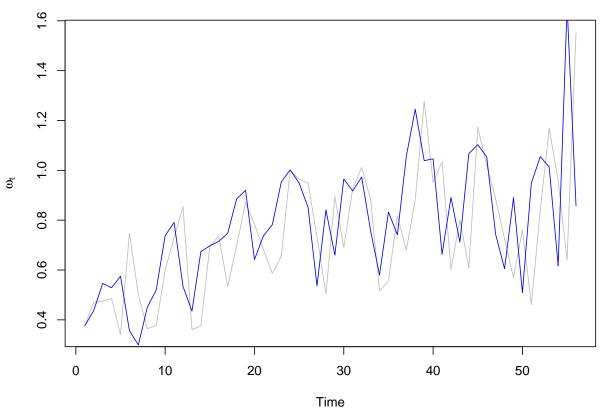
```
kurtosis_obs <- kurtosis(y[seq(2, length(y))])</pre>
sample_kurtosis <- apply(samples, 1, function(x) kurtosis(x[seq(2, length(y))]))</pre>
hist(
 x = sample_kurtosis,
 main = "Kurtosis of Posterior Predictive Samples",
 xlab = "Kurtosis",
  col = "gray",
  border = "white",
)
lines(
  x = c(kurtosis_obs, kurtosis_obs),
  y = c(0, 150),
  col = "black",
  lwd = 2,
text(
  x = kurtosis_obs + 0.5,
  y = 150,
  labels = paste("Observed Kurtosis = ", round(kurtosis_obs, 2)),
```

# **Kurtosis of Posterior Predictive Samples**



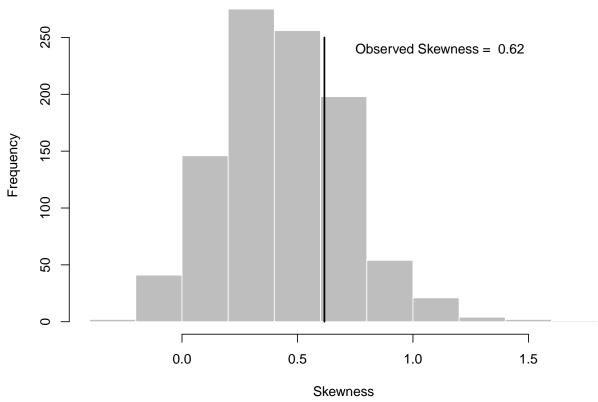
```
y <- data[["cases"]]</pre>
y \leftarrow diff(y[as.Date(data$date) >= as.Date("2020-06-30") &
              as.Date(data$date) <= as.Date("2020-08-25")], lag = 1) / 1000
samples <- matrix(NA, nrow = nrow(params), ncol = length(y))</pre>
for (i in seq_len(nrow(params))) {
  samples[i, ] <- ar_post_predictive(params[i, "rho"], params[i, "log_sig"])</pre>
}
plot(
 x = seq_along(samples[1, seq(1, length(y))]),
  y = samples[1, seq(1, length(y))],
  type = "1",
  col = "gray",
  xlab = "Time",
 ylab = expression(omega[t]),
  main = "Posterior Predictive Samples vs. Observed Data",
for (i in 2:1000) {
  lines(x = seq_along(samples[i]), y = samples[i], col = "gray")
lines(x = seq_along(y), y = y, col = "blue")
```

### Posterior Predictive Samples vs. Observed Data



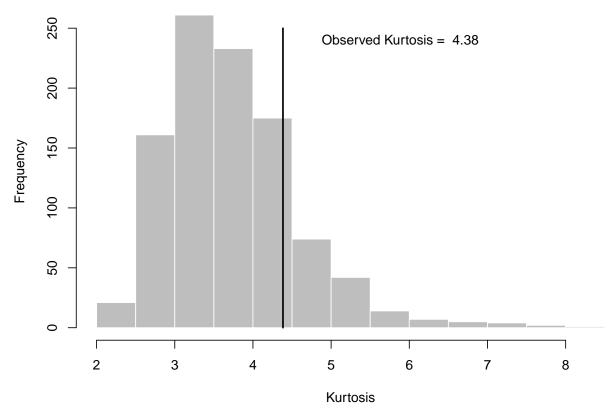
```
skewness_obs <- skewness(y[seq(2, length(y))])</pre>
sample_skewness <- apply(samples, 1, function(x) skewness(x[seq(2, length(y))]))</pre>
hist(
 x = sample_skewness,
 main = "Skewness of Posterior Predictive Samples",
 xlab = "Skewness",
  col = "gray",
  border = "white",
lines(
  x = c(skewness_obs, skewness_obs),
 y = c(0, 250),
 col = "black",
  lwd = 2,
)
text(
  x = skewness_obs + 0.5,
 y = 250,
  labels = paste("Observed Skewness = ", round(skewness_obs, 2)),
 pos = 1
)
```

# **Skewness of Posterior Predictive Samples**



```
kurtosis_obs <- kurtosis(y[seq(2, length(y))])</pre>
sample_kurtosis <- apply(samples, 1, function(x) kurtosis(x[seq(2, length(y))]))</pre>
hist(
 x = sample_kurtosis,
 main = "Kurtosis of Posterior Predictive Samples",
 xlab = "Kurtosis",
 col = "gray",
  border = "white",
)
lines(
 x = c(kurtosis_obs, kurtosis_obs),
 y = c(0, 250),
 col = "black",
  lwd = 2,
text(
  x = kurtosis_obs + 1.5,
 y = 250,
 labels = paste("Observed Kurtosis = ", round(kurtosis_obs, 2)),
```

### **Kurtosis of Posterior Predictive Samples**



From the results, the model is capable of providing good predictions for the COVID-19 cases in Indiana. By comparing the time-series plot of cases in Indiana and in the US, the trend of the cases in Indiana is similar to the trend of the cases in the US. Therefore, the AR(1) model with the same prior can retain its predictive power transferring from the entire US to just Indiana.

#### 8. (5 points) Solution:

In summary, a AR(1) model on daily COVID-19 case increase is established in this problem. Specifically, the number of increased COVID-19 cases at the t-th day  $\omega_t$  is linearly correlated with the number of increased cases at the (t-1)-th day  $\omega_{t-1}$ , with some random noise.

$$\omega_t = \rho \omega_{t-1} + \epsilon_t, \quad \epsilon_t \sim \mathcal{N}(0, \sigma^2).$$

The model is fitted to the data both in the entire US and in the Indiana state. We conclude that the model provides good predictions in both cases by drawing samples from its predictive distribution, and these samples have the distribution similar to the observed data. The success can be intuitively attributed to two main reasons: first, the similar trend of the COVID-19 cases in Indiana and the US, which makes the AR(1) model transferable from the entire US to Indiana; meanwhile, the differentiating data preprocess helps to remove the non-stationary trend in the data, which can not be captured by the AR(1) model.

# Feedback

- a. Yes, the instructor present the material in a clear and understandable way.
- b. Yes, the instructor well-addressed the questions.
- c. I would like to hear more about how to properly select the appropriate prior under different circumstances.
- d. Yes, quite representative.
- e. None that I can think of now.
- f. I believe the instructor can provide more examples to illustrate posterior predictive checks for time-series data, which is more related to the homework.
- g. No, I am satisfied with the course so far.