

# Survival Analysis of Glioblastoma Patients Through Tumor Tissue Deconvolution



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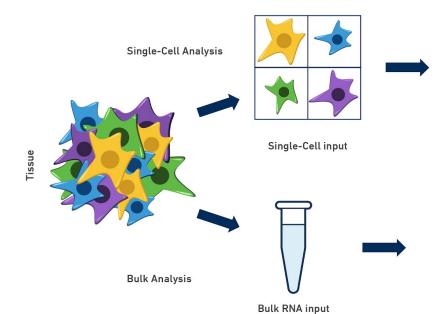
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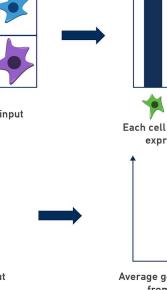


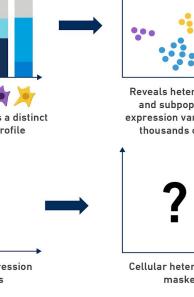


### **Background** ( (

- Glioblastoma multiforme (GBM) is the most aggressive type of brain cancer with a median survival time of 15-16 months and 14k cases in the U.S. each year
- Traditional treatments (surgery, radiation, and chemotherapy) and on-going research concentrates on understanding and estimating patient survival
- Immune cell presence in tumors has been found to be associated with patient survival estimated by the deconvolution of bulk RNA sequencing data
- Single Cell RNA Sequencing:
- Provides the gene expression profile of individual cells
- Reveals expression variability of different cell types subpopulations







Bulk RNA Sequencing:

Provides the average gene

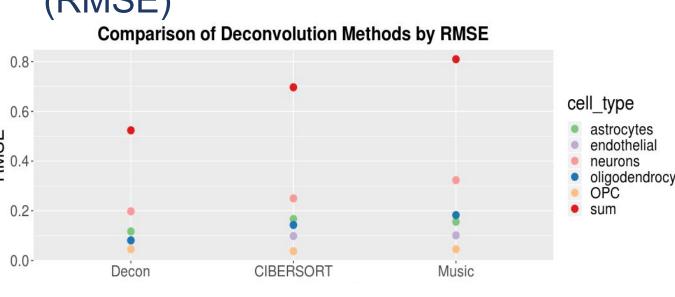
tissue; masks the cell type

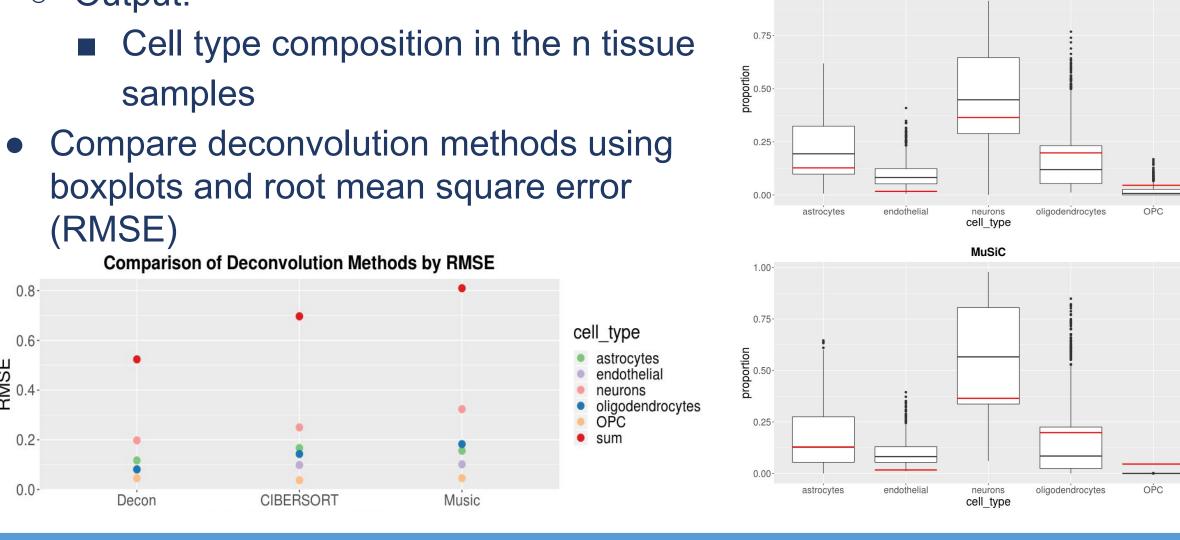
heterogeneity in the tissue

expression level of all cells in a

# Deconvolution Methods ( )

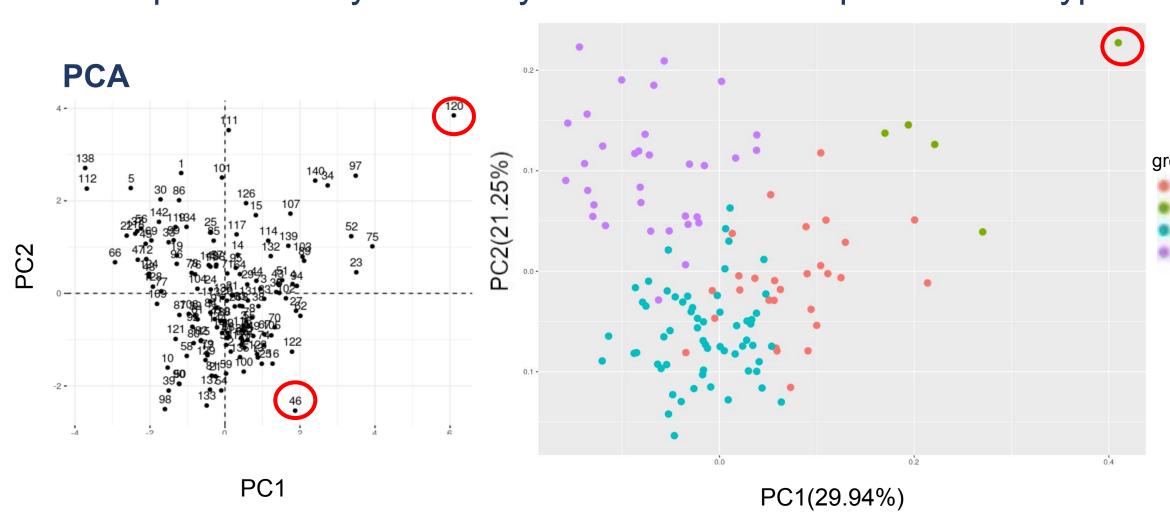
- General outline for deconvolution:
- o Input:
  - Gene signatures (single cell): Expression level of m genes in k cells
  - Mixture (bulk): Expression level of m genes in n tissue samples
- Output:
- samples
- boxplots and root mean square error (RMSE)





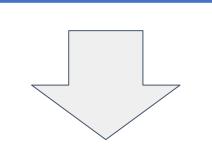
# Data Processing (

 Remove outlier samples in deconvolution results using Principal Component Analysis and by number of non-expressed cell types

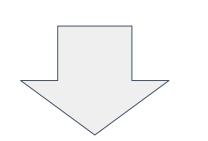


## Survival Analysis ( )

- K-means clustering:
- Use log rank and elbow method to decide the optimal number of clusters
- Use t-SNE dimensionality reduction to visualize clusters in two-dimensional space



- Survival Analysis:
  - Plot the Kaplan-Meier curve for each cluster to get the p-value for log rank test, which compares the survival curve for different clusters



- Fit Cox proportional hazards model to identify statistically significant variables
- Individual tests on group, B-cell, CD4+ T-cell, CD8+ T-cell, neutrophil, macrophage, dendritic cell (DC) to find which variables are statistically significant for predicting patient survival

#### Cox proportional hazards model:

 $\ln(\frac{H(t)}{H_0(t)}) = \beta_1 X_1 + \beta_2 X_2 + \beta_3 X_3 + \dots + \beta_m X_m$ 

Where H(t) is the hazard,  $H_0(t)$  is the baseline hazard at time t, which represents the hazard for a person with value 0 for all the predictor variables.  $X_1 \dots X_m$  are the set of covariates, and the beta coefficients measure their respective effect size. This model allows us to examine the effect of different factors on survival time simultaneously.



 Final model fitted with T\_cell.CD8 and Macrophage

 $\ln(\frac{H(t)}{H_0(t)}) = 2.19 \text{ T_cell. CD8} + 1.19 \text{Macrophage}$ 

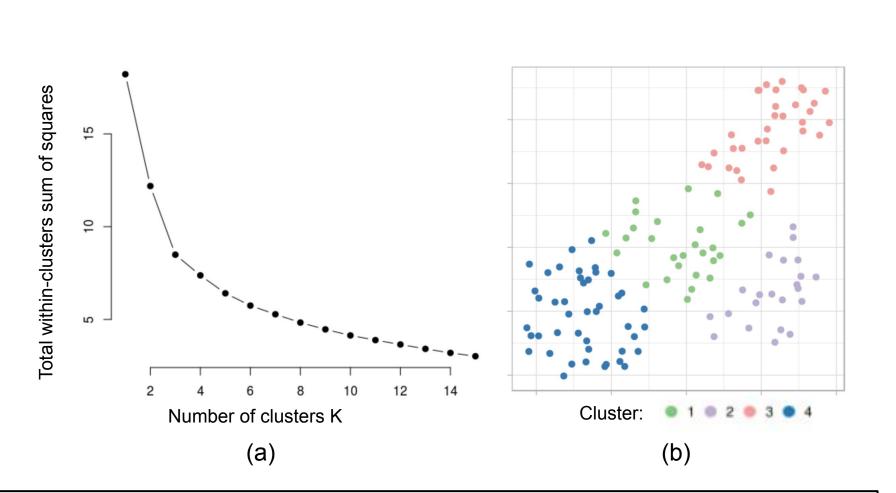


Figure 1: (a) Within-clusters sum of squares for 2 to 14 clusters. (b) Visualization of 4 clusters after t-SNE dimensionality reduction and k-means

#### **Kaplan-Meier Curves for Patients in Different Clusters** Strata - unlist(group)=1 - unlist(group)=2 - unlist(group)=3 - unlist(group)=4

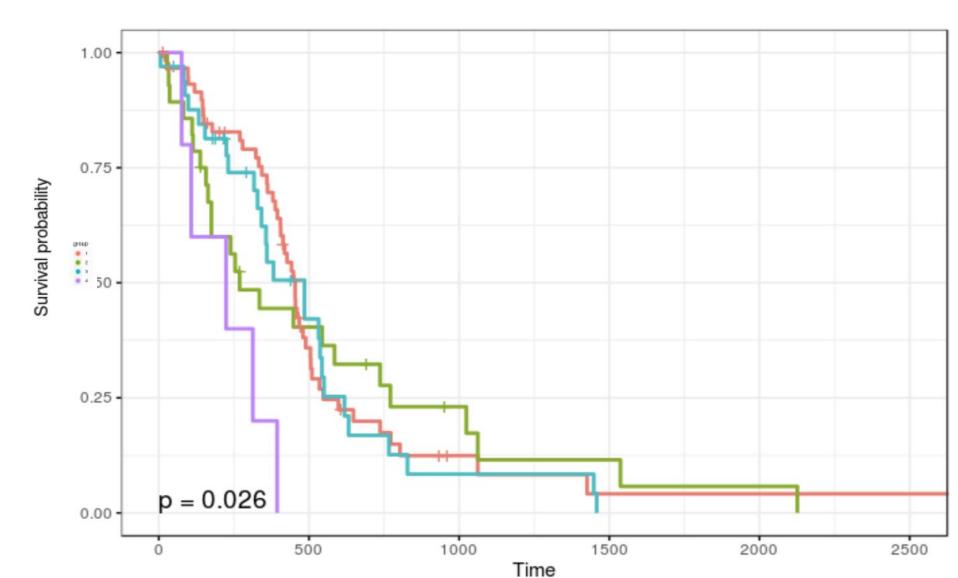


Figure 2: Kaplan-Meier Curves for different clusters of glioblastoma patients. Patients with similar tumor tissue immune cell type compositions are grouped into the same cluster represented by a unique color. A p-value of 0.026 for the log rank test comparison of different Kaplan-Meier curves indicates a significant difference among the survival curves.

#### **Summary of Beta Coefficients for Different Cell Types**

Variable	group	B_cell	T_cell.C D4	T_cell.CD 8	Neutrophil	Macroph age	DC
beta	-0.14	-0.57	0.32	2.7	1.7	2.5	0.44
Hazard ratio (95% CI)	0.87 (0.7-1.1)	0.57 (0.19-1.7)	1.4 (0.41-4.6)	15 (2.8-86)	5.3 (0.38-76)	13 (1.7-94)	1.6 (0.69-3.5)
p - value	0.2	0.31	0.6	0.0018	0.22	0.012	0.28

#### **Comparison of Beta Coefficient for Different Cell Types**

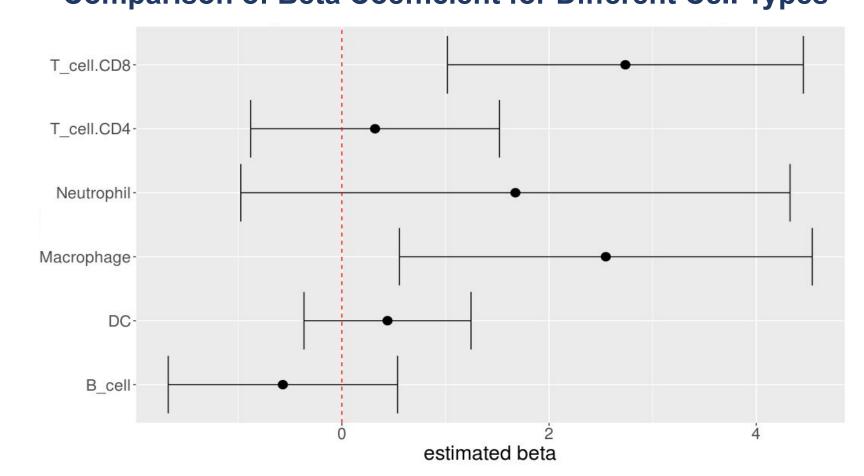


Figure 3: 95% confidence intervals of beta coefficients obtained from univariate Cox proportional hazards model on different immune cell types. The red dash line is 0. A negative beta indicates positive effect on survival time; a positive beta indicates negative effects on survival time.

# Results (

- A p-value of 0.026 for the log rank test to compare different Kaplan-Meier Curves indicates a significant difference among the survival curves.
- The p-values for the likelihood ratio test, wald test, and score test for the final model are all significant, indicating that the final model is significant. \_cell.CD8 is still significant in the final model while Macrophage fails to be significant in the presence of T\_cell.CD8 in the model.
- The hazard ratios of covariates are interpretable as multiplicative effects on the hazard. The hazard ratios for T\_cell.CD8(8.939) and Macrophage(3.288) indicate a strong relationship between higher proportion of these cell types in the patient's tumor tissue and increased risk of death. Holding the other covariates constant, a higher proportion of \_cell.CD8 and Macrophage in the patient's tumor tissue is associated with shorter survival time.

# **Discussion**

- Our current results are based on deconvolution output from TIMER due to technical difficulties we incurred while trying to run CIBERSORT and DeconRNASeq. TIMER's distinct output format impedes comparison with the other deconvolution methods, so its accuracy could not be determined.
- A possible explanation for inconsistencies with existing literature: More CD8+ T-cells may be sent to attack tumor cells, but their influence to prolong patient survival time may be limited for aggressive GBM tumors for patients that are severely ill
- Future work:
  - Explore interactions between T-cell subtypes in terms of effect on GBM patient survival time
  - Run CIBERSORT using the phenotype matrix with combined gene expression level of T\_cell.CD4 and T\_cell.CD8

#### Comparison of Beta Coefficient for T\_cell.CD8 and Macrophage

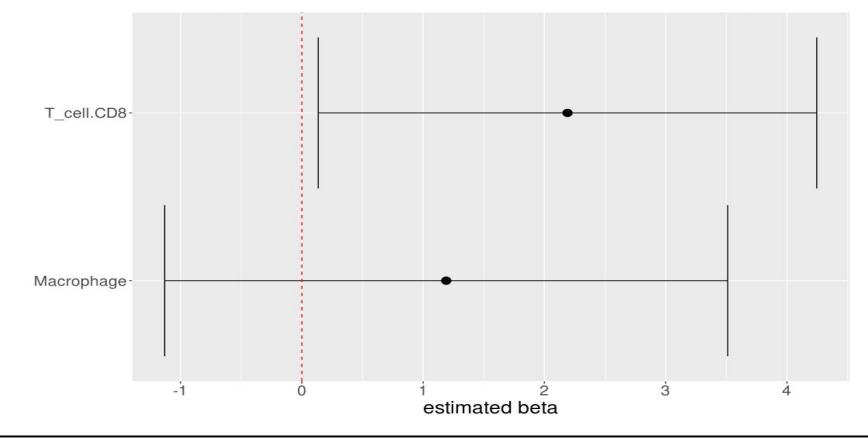


Figure 4: 95% confidence intervals of beta coefficients for T\_cell.CD8 and Macrophage obtained from final model. The red dash line is 0. A negative beta indicates positive effect on survival time; a positive beta indicates negative effects on survival time

#### **References:**

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