

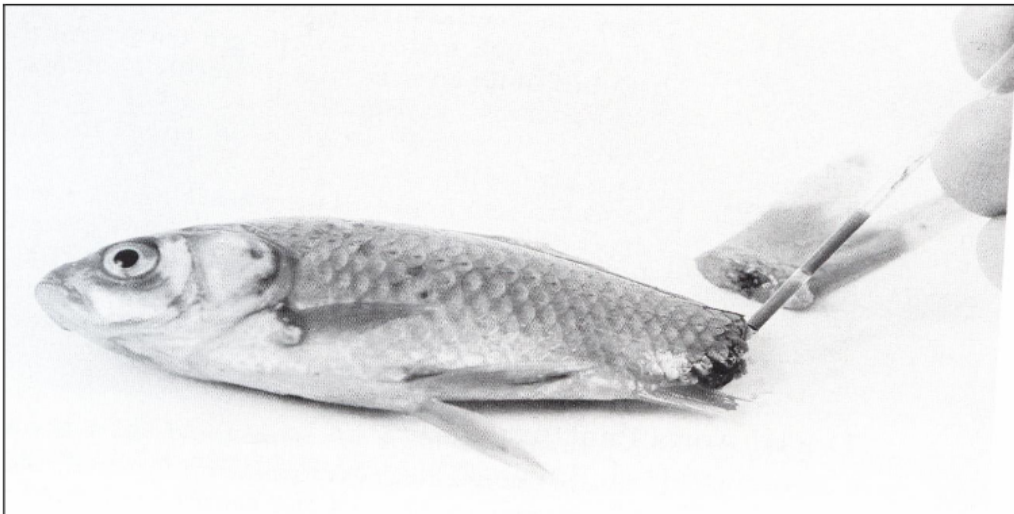
Juvenile Rockfish Blood Sampling SOP  
Logan Lab  
11-17-16

*Materials for Collection of Blood*

Heparinized glass hematocrit tubes (2-3 per fish)  
Scalpel and blades  
CritoSeal VWR#15407-103  
Centrifuge with hematocrit head rotor  
Hematocrit reader  
Black fine tip sharpie

**Tail Ablation**

1. Sever caudal peduncle with scalpel blade or sharp knife
2. Fill 3 replicate hematocrit tubes per fish with blood as it flows from caudal vein (fill tubes to  $\frac{2}{3}$  full or minimum  $\frac{1}{2}$  full). Duplicates are OK but aim for triplicate if possible
3. Plug one end of hematocrit tube with critoseal so that the seal is perpendicular to the tube



Blood collection from *Carassius auratus*. (Photo by R. Hebb)

4. Place 2 tubes for each fish in centrifuge (make sure that the centrifuge is equally balanced), and **make sure that the critoseal end in to the outside edge.**
5. Spin for 2-3 min (depends on centrifuge)
6. Remove from centrifuge carefully and place the tube on the hematocrit chart with the top of the plasma fraction on the 100% line and the bottom of the

red blood cells on line of the chart at 0%. - OR use digital calipers to do measurements

7. Read % red blood cells (red portion) to the bottom of the buffy coat (white blood cell layer) and record on data form.
8. The percent difference between values from the same fish should be within 5%.

