Class07: Machine Learning 1

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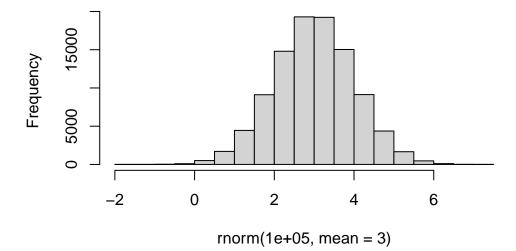
Today we will start our multi-part exploration of some keu machine learning methods. We will begin with clustering - finding groupings in data, and then dimensionallity reduction.

CLustering

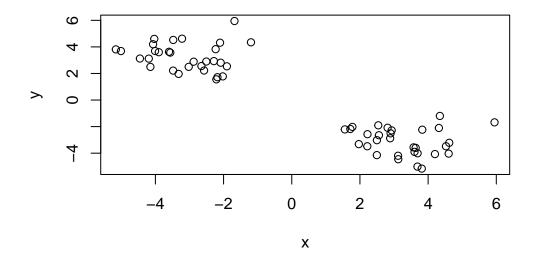
Let's start with "k-means" clustering. The main function in base R for this kmeans().

```
#Make up some data
hist(rnorm(100000, mean=3))
```

Histogram of rnorm(1e+05, mean = 3)



```
tmp <- c(rnorm(30, -3), rnorm(30, +3))
x <- cbind(x=tmp, y=rev(tmp))
plot(x)</pre>
```



Now let's try out kmeans()

```
km <- kmeans(x, centers=2)
km</pre>
```

K-means clustering with 2 clusters of sizes 30, 30

Cluster means:

```
x y
1 3.256896 -3.105346
2 -3.105346 3.256896
```

Clustering vector:

Within cluster sum of squares by cluster:

[1] 61.41485 61.41485 (between_SS / total_SS = 90.8 %)

Available components:

- [1] "cluster" "centers" "totss" "withinss" "tot.withinss"
- [6] "betweenss" "size" "iter" "ifault"

attributes(km)

\$names

[1] "cluster" "centers" "totss" "withinss" "tot.withinss"

[6] "betweenss" "size" "iter" "ifault"

\$class

- [1] "kmeans"
 - Q. How many points in each cluster?

km\$size

- [1] 30 30
 - Q. What component of your result object details cluster argument/membership?

km\$cluster

- - Q. What are centers/mean values of each cluster?

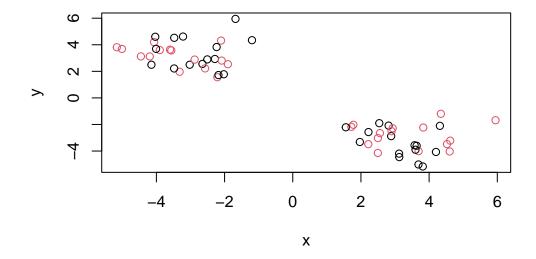
km\$centers

x y 1 3.256896 -3.105346

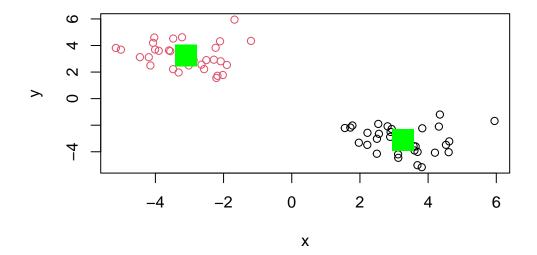
2 -3.105346 3.256896

Q. Make a plot of your data showing your clustering results.

plot(x, col=c(1,2))



#Selecting the cluster attribute vector from the the results of the kmeans() function we u
plot(x, col=km\$cluster)
points(km\$centers, col="green", pch=15, cex=3)



Q. Run kmeans() again and cluster in 4 groups and plot the results.

```
km4 <- kmeans(x, centers=4)
km4</pre>
```

K-means clustering with 4 clusters of sizes 12, 5, 30, 13

Cluster means:

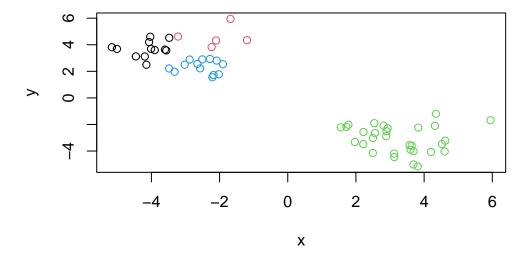
x y 1 -4.134665 3.672285 2 -2.086204 4.611556 3 3.256896 -3.105346 4 -2.547183 2.352437

Clustering vector:

Within cluster sum of squares by cluster:

[1] 6.951537 4.817664 61.414853 5.807639 (between SS / total SS = 94.1 %)

Available components:



Hierarchical Clustering

This form of clustering aims to reveal the structure in your data by progressively grouping points into an ever smaller number of clusters.

The main function in base R for this is called hclust(). This function does not take our input data directly but wants a "distance matrix" that details how (dis)similar all our input points are to each other.

```
#you can't just input x alone, hclust needs a distance matrix as it's input, which comes f
hc <- hclust(dist(x))
hc</pre>
```

```
Call:
hclust(d = dist(x))
```

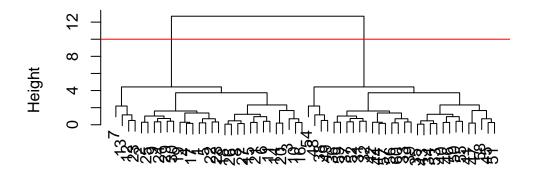
Cluster method : complete
Distance : euclidean

Number of objects: 60

The printout above is not very useful (unlike the one from kmeans) but there is a useful plot() method.

```
plot(hc)
abline(h=10, col="red")
```

Cluster Dendrogram

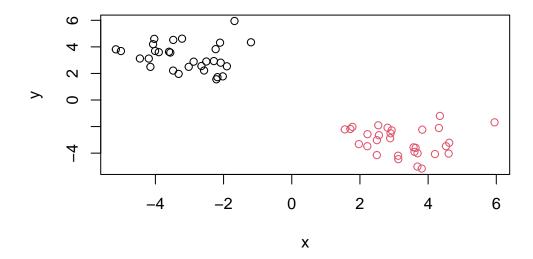


dist(x)
hclust (*, "complete")

To get my main result (my cluster membership vector) I need to "cut" my tree using the function cutree()

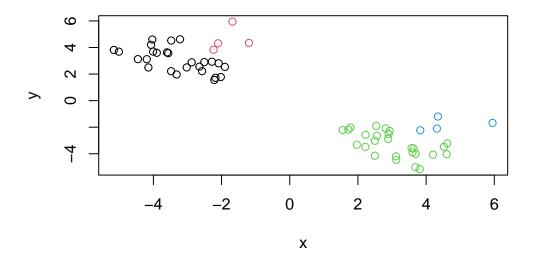
```
grps <- cutree(hc, h=10)
grps</pre>
```

plot(x, col=grps)



Cutting the tree a second time at h=4, splits the plotted data into more than two clusters.

```
plot(x, col=cutree(hc, h=4))
```



#Principal Component Analysis (PCA)

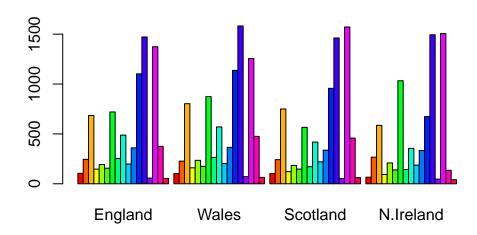
The goal of PCA is to reduce the dimensionality of a data set down to a smaller subset of new variables (called PCs) that are a useful basis for further analysis, like visualization, clustering, etc.

```
url <- "https://tinyurl.com/UK-foods"
x <- read.csv(url, row.names = 1)
head(x)</pre>
```

	England	Wales	Scotland	N.lreland
Cheese	105	103	103	66
Carcass_meat	245	227	242	267
Other_meat	685	803	750	586
Fish	147	160	122	93
Fats_and_oils	193	235	184	209
Sugars	156	175	147	139

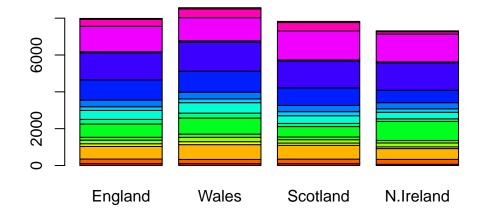
```
dim(x)
```

[1] 17 4



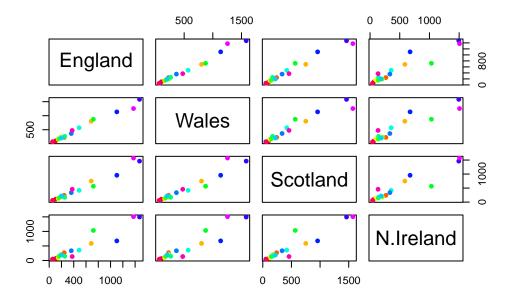
Changing the beside argument of the barplot() function changes the plot to be:

```
barplot(as.matrix(x), col=rainbow(nrow(x)))
```



The so-called "pairs" plot can be useful for small datasets:

```
pairs(x, col=rainbow(nrow(x)), pch=16)
```



These plots show each country in comparison to each other one - hence the title "pairs". You only need to look at half, the information is duplicated. Any departure from the straight diagnol line indicates a difference in the values between countries.

So the pairs plot is useful for small data sets but it can be a lot of work to interpret and gets untraceable for larger datasets.

PCA to the rescue...

The main function to do PCA in base R is called prcomp(). This functions wants the transpose of our data in this case.

```
#need to switch the data, we want foods as the columns for PCA
pca <- prcomp(t(x))
summary(pca)</pre>
```

Importance of components:

	PC1	PC2	PC3	PC4
Standard deviation	324.1502	212.7478	73.87622	3.176e-14
Proportion of Variance	0.6744	0.2905	0.03503	0.000e+00
Cumulative Proportion	0.6744	0.9650	1.00000	1.000e+00

```
attributes(pca)
```

```
$names
```

```
[1] "sdev" "rotation" "center" "scale" "x"
```

\$class

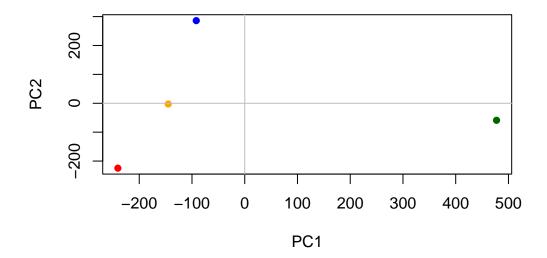
```
[1] "prcomp"
```

x shows how the data lies on the new axes

```
pca$x
```

```
PC1 PC2 PC3 PC4
England -144.99315 -2.532999 105.768945 -4.894696e-14
Wales -240.52915 -224.646925 -56.475555 5.700024e-13
Scotland -91.86934 286.081786 -44.415495 -7.460785e-13
N.Ireland 477.39164 -58.901862 -4.877895 2.321303e-13
```

A major PCA result viz is called a "PCA plot" (a.k.a. a score plot, bi-plot, PC1 vs PC2 plot, ordination plot)



Another important output from PCA is called the "loadings" vector or "rotation" component - this tells us how much the original variables (the foods in this case) contribute to the new PCs.

pca\$rotation

	PC1	PC2	PC3	PC4
Cheese	-0.056955380	0.016012850	0.02394295	-0.694538519
Carcass_meat	0.047927628	0.013915823	0.06367111	0.489884628
Other_meat	-0.258916658	-0.015331138	-0.55384854	0.279023718
Fish	-0.084414983	-0.050754947	0.03906481	-0.008483145
Fats_and_oils	-0.005193623	-0.095388656	-0.12522257	0.076097502
Sugars	-0.037620983	-0.043021699	-0.03605745	0.034101334
Fresh_potatoes	0.401402060	-0.715017078	-0.20668248	-0.090972715
Fresh_Veg	-0.151849942	-0.144900268	0.21382237	-0.039901917
Other_Veg	-0.243593729	-0.225450923	-0.05332841	0.016719075
Processed_potatoes	-0.026886233	0.042850761	-0.07364902	0.030125166
Processed_Veg	-0.036488269	-0.045451802	0.05289191	-0.013969507
Fresh_fruit	-0.632640898	-0.177740743	0.40012865	0.184072217
Cereals	-0.047702858	-0.212599678	-0.35884921	0.191926714
Beverages	-0.026187756	-0.030560542	-0.04135860	0.004831876
Soft_drinks	0.232244140	0.555124311	-0.16942648	0.103508492

PCA looks to be a super useful method for gaining some insight into high dimensional data that is difficult to examine in other ways.

PCA of RNAseq Data

```
url2 <- "https://tinyurl.com/expression-CSV"</pre>
  rna.data <- read.csv(url2, row.names=1)</pre>
  head(rna.data)
       wt1 wt2 wt3 wt4 wt5 ko1 ko2 ko3 ko4 ko5
gene1 439 458 408 429 420 90 88 86 90
gene2 219 200
                204 210 187 427 423 434 433 426
gene3 1006 989 1030 1017 973 252 237 238 226 210
      783 792 829 856 760 849 856 835 885 894
gene4
      181 249
                204 244 225 277 305 272 270 279
gene5
      460 502 491 491 493 612 594 577 618 638
gene6
  ## Again we have to take the transpose of our data
  pca <- prcomp(t(rna.data), scale=TRUE)</pre>
  summary(pca)
Importance of components:
                          PC1
                                 PC2
                                         PC3
                                                 PC4
                                                          PC5
                                                                  PC6
                                                                          PC7
Standard deviation
                       9.6237 1.5198 1.05787 1.05203 0.88062 0.82545 0.80111
Proportion of Variance 0.9262 0.0231 0.01119 0.01107 0.00775 0.00681 0.00642
Cumulative Proportion 0.9262 0.9493 0.96045 0.97152 0.97928 0.98609 0.99251
                           PC8
                                   PC9
                                            PC10
Standard deviation
                       0.62065 0.60342 3.457e-15
Proportion of Variance 0.00385 0.00364 0.000e+00
Cumulative Proportion 0.99636 1.00000 1.000e+00
```

nrow(rna.data)

```
[1] 100
```

```
attributes(pca)
$names
[1] "sdev"
              "rotation" "center"
                                  "scale"
$class
[1] "prcomp"
  head(pca$x)
         PC1
                   PC2
                              PC3
                                        PC4
                                                  PC5
                                                             PC6
wt1 -9.697374 1.5233313 -0.2753567 0.7322391 -0.6749398 1.1823860
wt2 -9.138950  0.3748504  1.0867958 -1.9461655  0.7571209 -0.4369228
wt4 -8.731483 -0.7468371 0.5875748 0.2268129 -1.5404775 -1.2723618
wt5 -9.006312 -0.2945307 -1.8498101 -0.4303812 0.8666124 -0.2496025
ko1 8.846999 2.2345475 -0.1462750 -1.1544333 -0.6947862 0.7128021
           PC7
                      PC8
                                  PC9
                                             PC10
wt1 -0.24446614 1.03519396 0.07010231 3.073930e-15
wt2 -0.03275370  0.26622249  0.72780448  1.963707e-15
wt3 -0.03578383 -1.05851494 0.52979799 2.893519e-15
wt4 -0.52795595 -0.20995085 -0.50325679 2.872702e-15
wt5 0.83227047 -0.05891489 -0.81258430 1.693090e-15
ko1 -0.07864392 -0.94652648 -0.24613776 4.052314e-15
  kmeans(pca$x[,1], centers=2)
K-means clustering with 2 clusters of sizes 5, 5
Cluster means:
       [,1]
1 -9.125676
2 9.125676
Clustering vector:
wt1 wt2 wt3 wt4 wt5 ko1 ko2 ko3 ko4 ko5
```

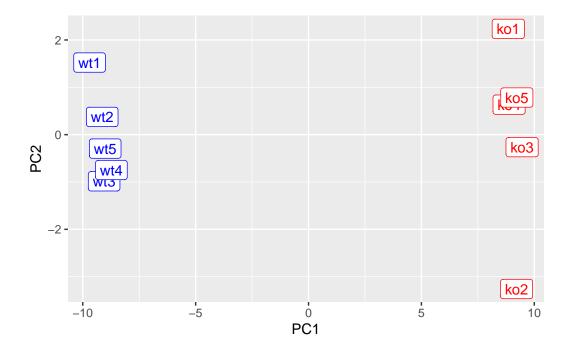
```
1 1 1 1 1 2 2 2 2 2
Within cluster sum of squares by cluster:
[1] 0.5017505 0.2648467
(between_SS / total_SS = 99.9 %)
Available components:
[1] "cluster"
                 "centers"
                               "totss"
                                             "withinss"
                                                           "tot.withinss"
[6] "betweenss"
                 "size"
                               "iter"
                                             "ifault"
I will make a main result figure using ggplot.
  library(ggplot2)
  res <- as.data.frame(pca$x)
  head(res)
         PC1
                                        PC4
                   PC2
                             PC3
                                                  PC5
                                                            PC6
wt1 -9.697374 1.5233313 -0.2753567 0.7322391 -0.6749398 1.1823860
wt2 -9.138950 0.3748504 1.0867958 -1.9461655 0.7571209 -0.4369228
wt4 -8.731483 -0.7468371 0.5875748 0.2268129 -1.5404775 -1.2723618
wt5 -9.006312 -0.2945307 -1.8498101 -0.4303812 0.8666124 -0.2496025
ko1 8.846999 2.2345475 -0.1462750 -1.1544333 -0.6947862 0.7128021
                                 PC9
                                            PC10
           PC7
                      PC8
wt1 -0.24446614 1.03519396 0.07010231 3.073930e-15
wt2 -0.03275370 0.26622249 0.72780448 1.963707e-15
wt3 -0.03578383 -1.05851494 0.52979799 2.893519e-15
wt4 -0.52795595 -0.20995085 -0.50325679 2.872702e-15
wt5 0.83227047 -0.05891489 -0.81258430 1.693090e-15
ko1 -0.07864392 -0.94652648 -0.24613776 4.052314e-15
```

[1] "blue" "blue" "blue" "blue" "red" "red" "red" "red" "red"

mycols <- c(rep("blue", 5), rep("red", 5))</pre>

mycols

```
ggplot(res) +
  aes(x=PC1, y=PC2, label=row.names(res)) +
  geom_point(col=mycols) +
  geom_label(col=mycols)
```



colnames(rna.data)

[1] "wt1" "wt2" "wt3" "wt4" "wt5" "ko1" "ko2" "ko3" "ko4" "ko5"