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Genome-Wide Association Study of Alzheimer Disease Endophenotypes at Prediagnosis Stages

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Abstract

INTRODUCTION—Genetic associations for endophenotypes of Alzheimer's disease (AD) in cognitive stages preceding Alzheimer disease have not been thoroughly evaluated.

METHODS—We conducted GWAS for AD-related endophenotypes including hippocampal volume (HPV), logical memory scores (LMT), and cerebrospinal fluid (CSF) amyloid β_{42} and total/phosphorylated tau (t-/p-Tau) in cognitively normal (CN), mild cognitive impairment (MCI), and AD dementia subjects from the Alzheimer's Disease Neuroimaging Initiative study.

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RESULTS—In CN subjects, study-wide significant (SWS; $P < 8.3 \times 10^{-9}$) loci were identified for t-Tau near *SRRM4* and *C14orf79* and for HPV near *MTUS1*. In MCI subjects, SWS association was found with SNPs near *ZNF804B* for LMT-delayed recall. We found consistent expression patterns of *C14orf40* and *MTUS1* in carriers with risk alleles of eSNPs and in brains of AD patients, compared with in the non-carriers and in brains of controls.

DISCUSSION—Our findings for AD-related brain changes prior to AD provide insight about early AD-related biological processes.

Keywords

Alzheimer's disease; Genome-wide association; endophenotypes; cerebrospinal fluid; MRI; logical memory; biomarker; ADNI; tau; co-expression network

1. Introduction

Alzheimer's disease (AD) is the most common type of dementia and typically occurs after age 65 years. It is highly heritable, but the known genetic risk factors (currently numbering more than 25 loci including *APOE*) account for no more than 50% of the heritability of the disorder [1]. However, genetic association findings based on AD risk do not explain the whole genetic architecture of AD because the mechanistic complexity underlying AD is not captured entirely by disease status, especially in preclinical stages [2, 3]. To overcome this limitation and understand preclinical stages of AD, researchers have examined the genetic underpinnings of AD-related endophenotypes including cerebrospinal fluid (CSF) levels of beta amyloid peptide ($A\beta_{42}$) and tau proteins, structural brain changes quantified by magnetic resonance imaging (MRI), and neuropsychological test measures of cognitive functioning, including memory loss [4,5]. Genome-wide association (GWA) studies for AD-related endophenotypes have identified novel loci in the Alzheimer's Disease Neuroimaging Initiative (ADNI) study which enrolled appreciable numbers of subjects across three stages: AD dementia, mild cognitive impairment (MCI), and normal cognitive functioning (CN) [6]. Previous ADNI studies indicated the importance of delineating different stages of subjects [7, 8]. We hypothesized that some genes may contribute to AD-related processes specifically during stages prior to AD dementia onset. Genes and pathways that are strongly associated with AD-related endophenotypes in early disease stages may be promising targets for developing AD biomarkers and preventive medicines. To test this hypothesis, we conducted GWA analyses for AD-related endophenotypes in the ADNI sample stratified by stage. Here, we focused on the association tests in the CN and MCI subgroups because we were interested in identifying genes that may contribute to AD-related processes prior to AD dementia onset.

2. Subjects and Methods

2.1. Subjects

GWA and phenotype data for ADNI participants were downloaded from a public-access database (<http://www.loni.usc.edu>). A total of 1,189 subjects before QC were available with GWA data from two different chips (ADNI-1, n=757 and ADNI-GO/2, n=432). We stratified subjects by stage (CN, MCI, and AD dementia) based on diagnosis at the baseline

assessment as defined by the standard ADNI protocol. Demographic information and mean endophenotype values stratified by stage as well as for the entire sample are presented in Supplementary Table 1. Age is similarly distributed in each subgroup. Sample sizes for analyses of CSF biomarkers were considerably smaller than for those of other traits.

2.2. Phenotypic Evaluation

Previously suggested AD-related endophenotypes including CSF biomarkers [9], MRI brain imaging measures [10] and episodic memory tests [11] were selected for GWA analyses in this study. CSF biomarkers of A β 42, total tau (t-Tau), and phosphorylated tau (p-Tau), brain MRI measure for hippocampal volume (HPV), and scores for logical memory immediate (LMiT) and delayed (LMdT) recall tests which were all measured at baseline were analyzed in this study. Details about collection of CSF biomarkers, brain MRI scan data, and neuropsychological tests are reported elsewhere [12-15].

2.3. Genotyping, Quality Control, Imputation, and Population Substructure Analysis

Details of quality control, genotype imputation, and population substructure analyses are described in Supplementary Information. After QC, the ADNI-1 sample with genotype data consisted of 187 CN, 329 MCI, and 163 AD dementia subjects, and the ADNI-GO/2 sample contained 118 CN, 252 MCI, and 27 AD dementia subjects with genotype data.

2.4. Statistical Methods

2.4.1. Genome-wide association tests—Prior to the association tests, each of the six endophenotypes was adjusted for covariates using linear regression. Age and sex were used as covariates for the six endophenotypes. A term for education level was also included in the regression models for LMiT and LMdT, and the model for HPV was further adjusted for total intracranial volume. The residuals derived from the regression models were rank-transformed for normalization as previously described [16]. Analyses were conducted for all autosomal SNPs using the expected genotype dose, a quantitative measure between 0 and 2 of the number of effect alleles computed from the imputed genotype probabilities as the predictor. Association of the rank-normalized endophenotypes with each SNP was evaluated using a linear regression model including covariate terms for the first three principal components (PC) of population substructure using the R software package. The two ADNI datasets were analyzed independently for the CN and MCI subjects and the results from the two ADNI datasets were combined by meta-analysis using inverse variance weights as implemented in the METAL program [17]. AD cases from the two ADNI datasets were analyzed as one group because the ADNI-GO/2 sample included only 27 AD subjects, and regression models for this group included an extra covariate for ADNI dataset. The genome-wide significant (GWS) threshold was set at 5.0×10^{-8} . We determined a conservative study-wide significant (SWS) level of 8.3×10^{-9} which was calculated as the GWS level divided by the effective number of two independent endophenotypes and three clinical subgroups. The effective number of independent endophenotypes was computed by counting the number of eigenvalues greater than one from the PC analysis of all six endophenotypes. A threshold of $P < 10^{-6}$ was considered as suggestive evidence of association in the functional/pathway analysis.

2.4.2. Expression SNP Analysis—We examined association of the SWS SNPs (allele counts) with transcript-level expression, i.e., expression SNP (eSNP), using a publically available database via the GTEx Portal (<http://www.gtexportal.org>; [18]).

2.4.3. Differential Gene Expression Analysis—Differential gene expression (DGE) was evaluated for genes containing or near significantly associated SNPs in two independent human brain expression datasets from the Eisai Bio Bank (EBB) and Mt. Sinai Hospital (MSH) (which was downloaded from Gene Expression Omnibus [GEO]: GSE44772). The EBB has gene expression measures obtained from RNA sequencing from the hippocampus (HIPP) of samples collected from autopsied brains from 35 AD cases and 16 normal subjects ascertained at the University of Miami and McLean Hospital (Belmont, MA). A measure of neurofibrillary tangles (Braak stage) in the same samples was assessed following an established protocol [19]. Details about sample collection and preparation, and demographic characteristics are provided in Supplementary Information. The association of log₂-transformed transcript expression levels (outcome) with AD status (predictor) was evaluated using linear regression adjusting for site, age, sex, and RNA integrity number (RIN). A model testing the effect of Braak stage (0-6 stages) on transcript expression levels was also evaluated and included the same covariates. The MSH microarray expression data (GEO: GSE44772) were generated from autopsied brain tissue collected from dorsolateral prefrontal cortex (DLPFC), visual cortex (VCX), and cerebellum (CER) regions in 129 AD patients and 101 controls. Samples were profiled on a custom-made Agilent 44K array containing 40,638 probes. Gene expression data were normalized using Rosetta Resolver gene-expression analysis software as previously described [20]. The association of log₂-transformed gene expression level (outcome) and AD status (predictor) was tested using a linear regression model adjusting for RIN, postmortem interval (PMI), batch, preservation method, tissue pH, age, and sex. A significance threshold to correct for the number of eQTL analysis tests was applied which in this case was P=0.025 since only two genes (*C14orf40* and *MTUS1*) were tested.

2.4.4. Co-expression Network Analysis of Human Brain RNA-Sequence Data—The top-ranked genes at or near (< 50kb) loci that achieved suggestive significance (P<10⁻⁶) in GWA tests of any trait in CN or MCI subjects or in the total sample were further evaluated for gene co-expression networks. We built gene co-expression networks in the EBB HIPP RNA-Seq data (41,249 transcripts) by weighted gene co-expression network analysis (WGCNA) [21], an approach which defines modules (or subsets) of genes that are highly co-expressed (or co-regulated). Details of this approach are described in Supplementary Information. In the HIPP co-expression network, we selected modules which carry both the top-ranked genes from this study and previously known AD genes [22-25]. The selected modules were functionally annotated by two enrichment analyses of ‘gene ontology (GO)’ and ‘disease-associated genes’ with a hypergeometric test. The ‘disease-associated genes’, genes involved in risk of diseases, were downloaded from GWASdb2 [26]. A P-value of significance for each enrichment test was calculated along with a false discovery rate (FDR), estimated using the Benjamini-Hochberg procedure [27]. We used an FDR threshold of 0.05 to define associated GOs or diseases. The selected modules were further examined for

correlations with traits (AD status and Braak stages) using the EBB sample by calculating the Pearson's correlations between the module eigengene and the traits.

2.4.5. Functional Analysis and Brain Cell Type-Specific Expression Profiling—

We evaluated predicted functions of SNPs showing suggestive evidence for association ($P < 10^{-6}$) from the GWA tests using HaploReg (<http://www.broadinstitute.org/mammals/haploreg/>) [28]. ENCODE data [29] were used to evaluate potential regulatory function. We also investigated the expression profiles of the top-ranked gene from mouse and human brain from the cerebral cortex (http://web.stanford.edu/group/barres_lab/brainseqMariko/brainseq2.html) [30]. To identify shared functions among the top-ranked genes, we performed functional analysis using Ingenuity Pathway Analysis software (IPA, QIAGEN, Redwood, CA). IPA determines which molecular/cellular function terms (e.g., top-ranked genes in association tests) are statistically over represented, suggesting the GWA findings capture functional mechanisms underlying disease-related biological processes. We used a nominal P-value threshold of 0.05 to flag associated functions. We also examined neuronal cell type-specific expression for the top-ranked genes using single cell RNA sequencing data (which was downloaded from GEO: GSE67835). Further details of this analysis are described in Supplementary Information.

3. Results

3.1. Genome-wide Association Results

There was slight genomic inflation in GWA results for $\text{A}\beta_{42}$ and LMdT in CN subjects ($\lambda=1.02$ for both traits; Supplementary Figs. 1-12). Associations of the *APOE* ε4 allele were SWS with the CSF biomarkers and suggestive with the other traits in MCI subjects, whereas ε4 was significantly associated only with $\text{A}\beta_{42}$ level in CN subjects (Supplementary Table 2). Eleven of 25 other previously known AD loci [22-25] – *CR1*, *INPP5D*, *MEF2C*, *HBEGF*, *HLA region*, *ZCWPW1*, *USP6NL*, *MS4A region*, *PICALM*, *SLC2A4A*, and *CASS4* – were nominally significant ($P < 0.05$) with at least one trait in CN and/or MCI groups (Supplementary Table 3).

Novel SWS associations were observed for several endophenotypes in the CN and/or MCI groups (Table 1). Among CN subjects, t-Tau was associated with SNPs located 28 kb upstream of *SRRM4* (best SNP: rs10775009, $P=1.6 \times 10^{-9}$; Fig. 1A) and 66.8 kb upstream of *C14orf79* (rs2819438; $P=6.9 \times 10^{-9}$; Fig. 1B). In the same group, HPV was associated with rs4921790 in *PDGFRL* and near *MTUS1* ($P=4.6 \times 10^{-9}$; Fig. 1C). This finding was supported by associations with many SNPs in high LD ($r^2 > 0.8$ and $D' > 0.9$) with rs4921790 which span *MTUS1*. In the MCI subgroup, LMdT was associated with three SNPs in *ZNF804B* (best SNP: rs73705514; $P=2.9 \times 10^{-9}$; Fig. 1D). Association was also observed between HPV and a SNP near *LINC00271* and *PDE7B* ($P=1.76 \times 10^{-8}$) in AD subjects (Supplementary Fig. 13A). Several GWS associations are also noteworthy (Supplementary Table 4) including $\text{A}\beta_{42}$ level with *GRIN2B* SNP rs74442473 ($P=2.52 \times 10^{-8}$; Supplementary Fig. 13B) and rs2378873 near *BRIP1* and *NACA2* ($P=2.03 \times 10^{-8}$) in CN subjects, LMdT with *DAB1* SNP rs74834332 ($P=4.20 \times 10^{-8}$) and *PRKG1* SNP rs12268753 ($P=2.01 \times 10^{-8}$) in CN subjects, LMdT with *ARHGAP24* SNP rs111882035 ($P=2.74 \times 10^{-8}$) in MCI subjects, and LMiT with

NRG1 SNP rs118130881 ($P=1.72\times10^{-8}$; Supplementary Fig. 13C) in MCI subjects, and A β_{42} level with rs55644114 near *GFRA2* and *LZTS1* in the total sample.

3.2. Expression-SNP Association Results

According to the GTEx Portal database, several eSNPs under the SWS association peaks for the endophenotypes in the CN or MCI groups are significantly associated with expression of genes in those regions (Table 2). The major allele *C* of intergenic SNP rs2819438, which is associated with increasing CSF t-Tau, was significantly associated with lower expression of *C14orf79* in brain (P -values in HIPP = 3.8×10^{-4} and CER = 6.0×10^{-3} ; Supplementary Fig. 14A), but not with expression of *PLD4* and *AHNAK2* which are located between rs2819438 and *C14orf79*. Rs4921790 was not associated with expression of *MTUS1* ($P>0.05$), however the major allele *G* of rs55653268 (a proxy SNP for rs4921790; $r^2 = 0.7$; $D' = 1.0$), which is associated with decreasing HPV, was significantly associated with increased expression of *MTUS1* in the caudate ($P=9.7\times10^{-3}$; Supplementary Fig. 14B). *PDGFRL* expression was not tested, since its expression in brain is extremely low according to the GTEx database. *SRRM4* SNP rs119390525 and *ZNF804B* SNP rs73705514 were not significantly associated with expression in any brain regions ($P>0.05$).

3.3. Association of Gene Expression with AD Status and Braak Stage

Further examination of *C14orf79* and *MTUS1*, whose expression levels were associated with SWS SNPs (or their proxy SNPs), in the EBB RNA-Seq and the MSH microarray datasets revealed multiple significant associations with AD status and Braak stage in several brain regions (Table 3). *C14orf79* was not differentially expressed in HIPP ($P=0.76$), but its expression was significantly lower in AD cases than controls in CER ($P=7.5\times10^{-8}$), DLPFC ($P=3.3\times10^{-7}$, Fig 2A), and VCX ($P=2.1\times10^{-8}$). The association of *C14orf79* expression with decreasing Braak stage was nearly nominally significant (β [SE] = -0.07 [0.03], $P=0.06$) (Fig. 2A). *MTUS1* expression was nominally higher in AD cases than controls in HIPP ($P=0.02$), DLPFC ($P=0.01$, Fig. 2B) and VCX ($P=0.01$). *MTUS1* expression was also positively correlated with Braak stage (β [SE] = 0.16 [0.04], $P=3.9\times10^{-4}$; Fig. 2B). According to a publicly available murine brain expression profile data obtained from the cerebral cortex, *Mtus1* is highly expressed in microglia and myeloid cells. Moreover, expression is highest in activated microglia (Supplementary Figure 15A). However, *MTUS1* expression in mature human brain is ranked the second lowest in microglia/macrophage cells, whereas expression in adult mouse brain was ranked the second highest among glia, neurons, and vascular cells. This finding suggests that the role of *MTUS1* may differ in mouse and human myeloid cells (Supplementary Figure 15B).

3.4. Gene Co-expression Network Analysis in Hippocampus

Sixteen of 61 top-ranked genes (Supplementary Table 5) were clustered together with 17 known AD genes as co-expressed networks. Eight of 17 modules from WGCNA based on the human HIPP transcriptome (M1-M8) were enriched with the genes ($P<1.0\times10^{-6}$; Supplementary Table 5) identified in our GWAS analyses with AD-related endophenotypes (Supplementary Table 6). These eight modules were also significantly correlated with AD status or Braak stages (Fig. 3A). *C14orf79* was co-expressed with *MAPT-AS1* in M1. *MTUS1* was co-expressed with *ZCWPW1* in M4. *SRRM4* and *ZNF804B* were co-expressed

with previously known AD genes – *APP*, *BZRAPI*, *MAPT*, *MEF2C*, *PLXNA4*, *PTK2B*, and *PSEN2* – in M7. The M7 module is enriched for genes involved in neuronal processes including ‘chemical synaptic transmission’ ($FDR=1.8\times10^{-84}$), ‘dendrite’ ($FDR=1.7\times10^{-33}$), and ‘axon guidance’ ($FDR=1.6\times10^{-24}$). Modules M2, 4, 5, 6, and 8 are enriched for genes involved in ‘protein binding’, ‘signal transduction’, and ‘microtubule binding’ (Supplementary Table 6). Among the eight modules enriched for previously known AD risk genes and genes identified in the current study as associated with AD endophenotypes, modules 1, 2, 5, 7, and 8 were also highly enriched with genes for other neurodegenerative and neuropsychiatric disorders including Parkinson disease and schizophrenia, as well as for cognitive performance (Supplementary Table 6). The correlation matrix of expression levels among the genes in Modules 4 and 7 (Fig. 3B) is shown in Supplementary Table 7. Further evaluation revealed that M2 and M7 contained the largest number of known AD genes among all modules co-regulated in the hippocampal region (Supplementary Table 8).

3.5. Functional Analysis

The top-ranked genes from the GWA analyses in the CN and MCI groups (Supplementary Tables 5) are enriched in neuronal processes including synapse plasticity ($P=3.4\times10^{-5}$), axon quantity ($P=2.6\times10^{-4}$), microtubule dynamics ($P=2.8\times10^{-4}$), abnormal morphology of dentate gyrus ($P=3.4\times10^{-4}$), and neuronal development ($P=3.5\times10^{-4}$). These findings indicate that genes associated with AD-related endophenotypes among participants in clinical stages preceding AD dementia have particular roles in functioning of neuronal synapses (Fig. 3C). Single-cell transcriptome analysis of the human brain confirmed that the genes in Fig. 3C were highly expressed in neurons. Some genes were also expressed in other cell types: *APOE*, *MTUS1*, *ERBIN*, and *AKAP9* in astrocytes; *MTUS1*, *ERBIN*, and *AKAP9* in oligodendrocytes; and *AKAP9* and *RCAN1* in endothelial cells. Bioinformatic analysis using HaploReg and ENCODE suggests that GWS SNPs upstream of *NRG1* may be located in a transcriptional regulatory site and rs111969453, a SNP in high linkage disequilibrium ($r^2=0.94$ and $D'=0.97$) with the SNP (rs118130881) that is significantly associated with LMiT ($P=5.9\times10^{-8}$), is located on an enhancer histone mark in brain.

4. Discussion

A goal of this study was to identify genes that were both associated with AD-related endophenotypes in older, non-demented individuals and co-regulated with known AD genes. Using a GWA approach, we identified SWS associations in cognitively normal elders including CSF t-Tau level with *SRRM4* and *C14orf79*, HPV with *MTUS1*, and CSF A β_{42} level with *APOE*. In MCI subjects we detected SWS associations for LMdT with *ZNF804B*, and for CSF levels of A β_{42} , t-Tau, and p-Tau with *APOE*.

Angiotensin II interacting (AT2) protein (ATIP) isoforms encoded by *MTUS1* are highly expressed in most brain regions [31]. ATIP1 binds to AT2 proteins, mediating neuronal differentiation survival and regeneration in brain [31-33]. ATIP3 colocalizes with microtubules and regulates their polymerization, thereby regulating neuronal differentiation and neurite outgrowth [33]. Gene expression analysis revealed that the risk allele of eSNP near *MTUS1* was associated with increased expression of *MTUS1* in caudate. DGE analyses

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demonstrated that expression of *MTUS1* was higher in AD cases than controls in HIPP, DLPFC, and VCX. Also, expression of *MTUS1* was significantly greater in brains showing severe neurofibrillary tangle involvement. The ADNI GWAS findings together with these expression findings suggest that *MTUS1* expression may be related to changes in hippocampal volume prior to onset of cognitive impairment.

SRRM4 encodes the neural-specific Ser/Arg repeat-related protein of 100kDa (nSR100) that promotes neurite outgrowth and alternative splicing and controls most neural microexons [34-36]. Down-regulated *SRRM4* alters splicing of microexons in autism brains [34]. Association of *SRRM4* SNP rs1997111 with t-Tau level in a subset of ADNI controls (i.e., from “ADNI 1” only) was previously identified at a significance level one order of magnitude less ($P=1.1\times10^{-8}$) than the result we obtained among CN subjects in our study ($P=1.76\times10^{-9}$) [37]. The functional relevance of *C14orf79* and *ZNF804B*, the two other SWS but poorly characterized genes, to AD is unclear.

GRIN2B encodes the GluN2B subunit of the N-methyl-D-aspartate (NMDA) receptor (GluN2B-NMDA) which is involved in synaptic plasticity and memory function. GluN2B-NMDA is the target of memantine, a drug that provides symptomatic relief in patients with AD by antagonizing the GluN2B-NMDA receptor [38]. A suggestive association of *GRIN2B* with temporal lobe volume ($P=1.3\times10^{-7}$) was reported in a previous GWA study in the ADNI-1 sample [39]. *NRG1* functions as an epidermal growth factor in the nervous system [40] and is involved in the neuregulin signal transduction pathway for synapse maturation and dendritic morphology [41, 42].

Among the known AD genes, we found suggestive evidence for association with several SNPs located 381 kb upstream of *AKAP9* with p-Tau level in MCI subjects (Supplementary Table 2, Supplementary Fig. 13D). We also found a suggestive association between rs149454736, located between exons 45 and 46 of *AKAP9*, and HPV in AD dementia subjects ($P=2.2\times10^{-7}$). Previously, we identified significant association of AD with two rare coding *AKAP9* missense mutations in exons 31 (rs144662445) and 46 (rs149979685) in African Americans [43], and our study is the first to report association with this gene for AD risk in European ancestry individuals. Rs149454736 is located 1.5 kb away from rs149979685. *AKAP9* has functional similarity with the tau protein in terms of microtubule stability and assembly [44].

With the exception of *APOE*, there is little overlap in GWS findings between our study and an investigation of genetic determinants of CSF A β ₄₂, t-Tau, and p-Tau levels in a much larger sample which included 787 ADNI subjects [45]. Some of the differences may be explained by our study design which conducted analyses within separate groups of CN, MCI, and AD subjects. The prior study combined cognitive groups and included multiple cohorts with highly variable ascertainment and distributions of subjects across cognitive groups. In addition, we extended our findings by incorporating them in analyses of co-regulated transcriptional networks enriched with previously established AD genes. Further comparison of results across the two studies revealed that four of the six GWS SNPs (excluding the *APOE* region) in the Deming *et al.* report [45] were nominally significant in specific cognitive groups in our study (Supplementary Table 9). For example, the association

of rs185031519 near *GLIS1* CSF A β 42 ($P=2.1\times10^{-8}$) in the prior study was nominally significant in the CN group ($P=8.3\times10^{-3}$) in our study. Several of our most significant findings (including *SRRM4*, *MTUS1*, and *GRIN2B*) are consistent with results of other studies of AD-related endophenotypes (Supplementary Table 10).

Our findings may provide important insights about the sequence of processes leading to AD. The SWS associations in CN subjects (*SRRM4* with t-Tau and *MTUS1* with HPV) implicate neuronal signaling, development and loss, but with the exception of *APOE*, they do not involve A β processing in the asymptomatic stage of AD. It is noteworthy that the *APOE* E4 allele was not associated with the CSF Tau biomarkers and other endophenotypes in cognitively normal subjects. The variants associated with CSF biomarkers and HPV could also be interpreted to be markers of cognitive reserve/resilience because they predict the extent of AD pathology in cognitively normal persons, but not in MCI or AD subjects. There is extensive evidence supporting the cognitive reserve hypothesis in AD [46]. Also, the variants identified for memory performance and HPV in cognitively normal individuals could be markers of inherent memory function and hippocampal volume, completely independent of AD.

Our study has several limitations. First, the sample sizes for analyses of all traits in each clinical group, especially AD cases, were relatively small. Thus, we had low power to identify variants having small effects. In addition, it is possible that our top findings are false positives. However, the significant SNPs were supported by evidence in both constituent datasets (ADNI-1 and -GO/2) and from expression data analysis. Second, in the transcriptomic analyses we did not consider differences in cellular composition between AD and control brain tissue. Therefore, our results may have resulted from the excessive neuronal death in brains from AD subjects compared to controls. However, this concern would not impact our finding of increased *MTSU1* expression in AD brains. Third, the two expression datasets generated using different platforms (RNA-seq versus microarray) from different brain regions (HIPP versus CER/DLPFC/VCX) limit direct comparisons between these datasets. Finally, it is necessary to repeat analyses in independent samples to confirm our findings and increase power to elevate the significance of true associations which did not attain study-wide significance.

In summary, we identified novel genes associated with AD-related endophenotypes in cognitively normal and MCI subjects. These genes had not been previously identified with AD risk and most of them are involved in neuronal development and signaling. Our findings suggest that genes influencing AD-related processes in individuals with normal cognition or with mild cognitive impairment may differ from those influencing these processes in individuals with AD dementia, but regulated together in the transcription level.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Research highlights

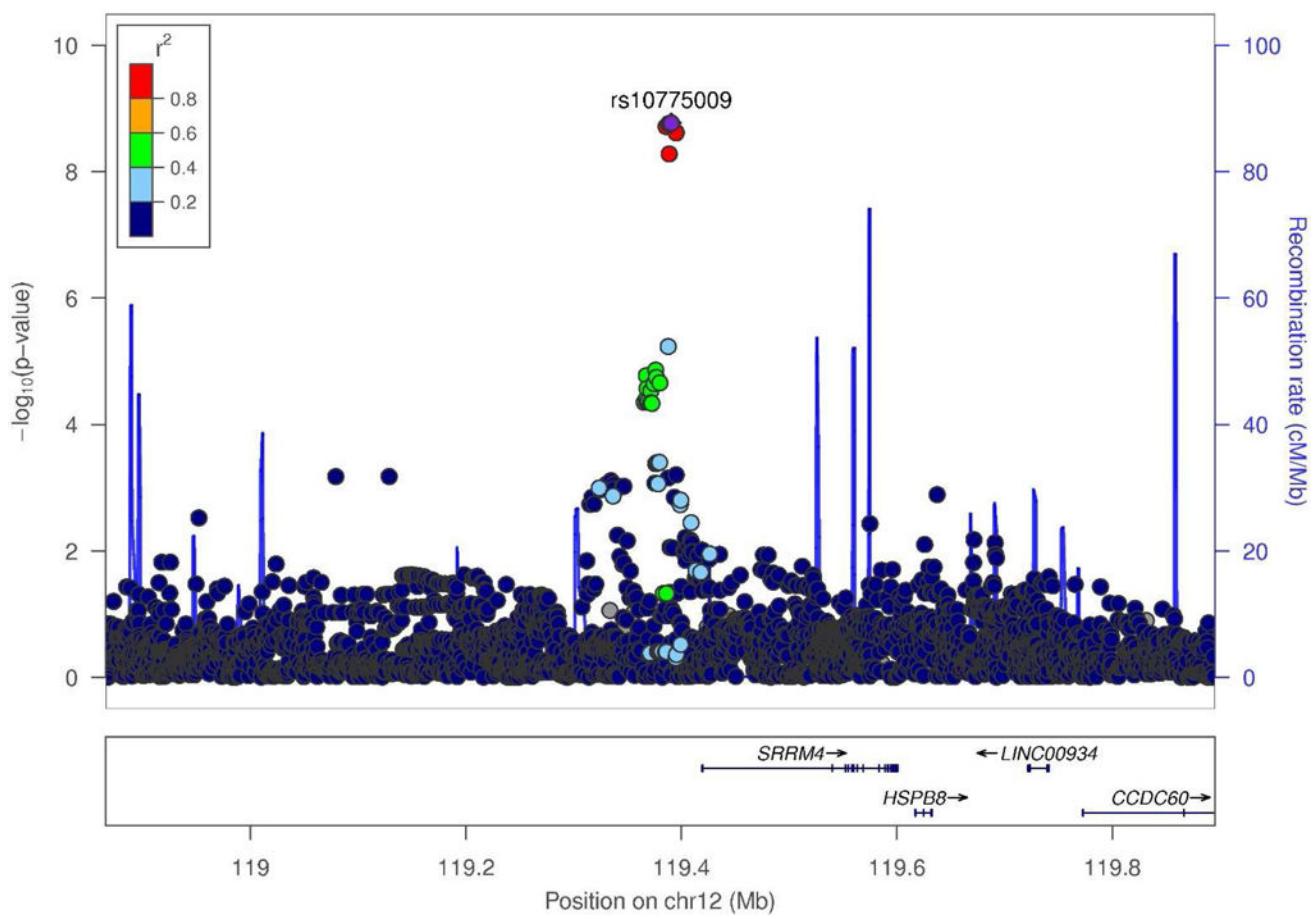
- Genetic factors for Alzheimer disease related endophenotypes influencing prediagnosis stages is evaluated
- Genetic factors affecting early changes for Alzheimer disease via expression are proposed.
- Co-regulated gene expression network containing genetic signatures associated with early changes in Alzheimer disease related endophenotypes that are interconnected with known AD risk genes.
- Genes and pathways in early biological processes in Alzheimer disease are enriched in neuronal or synaptic function.

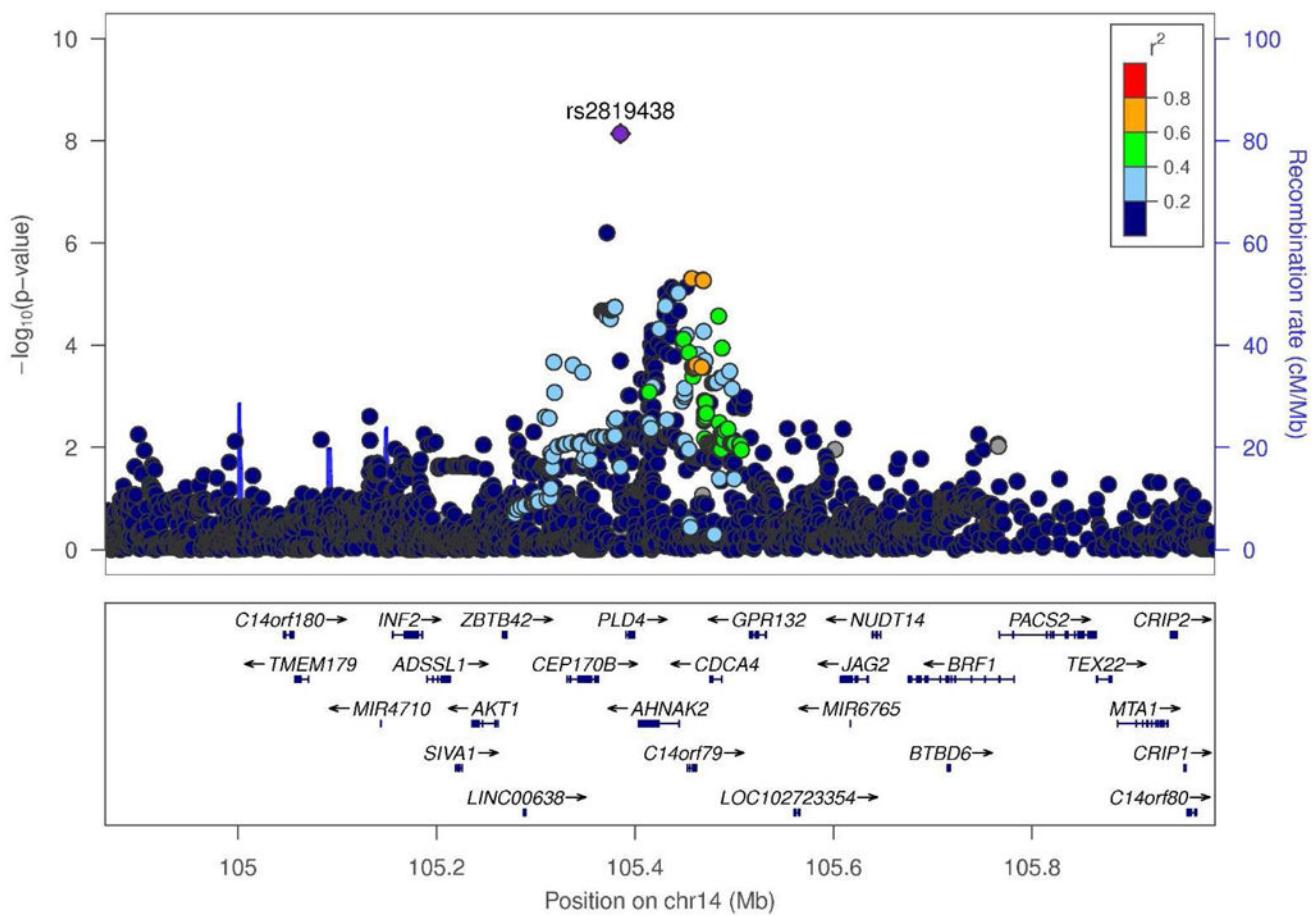
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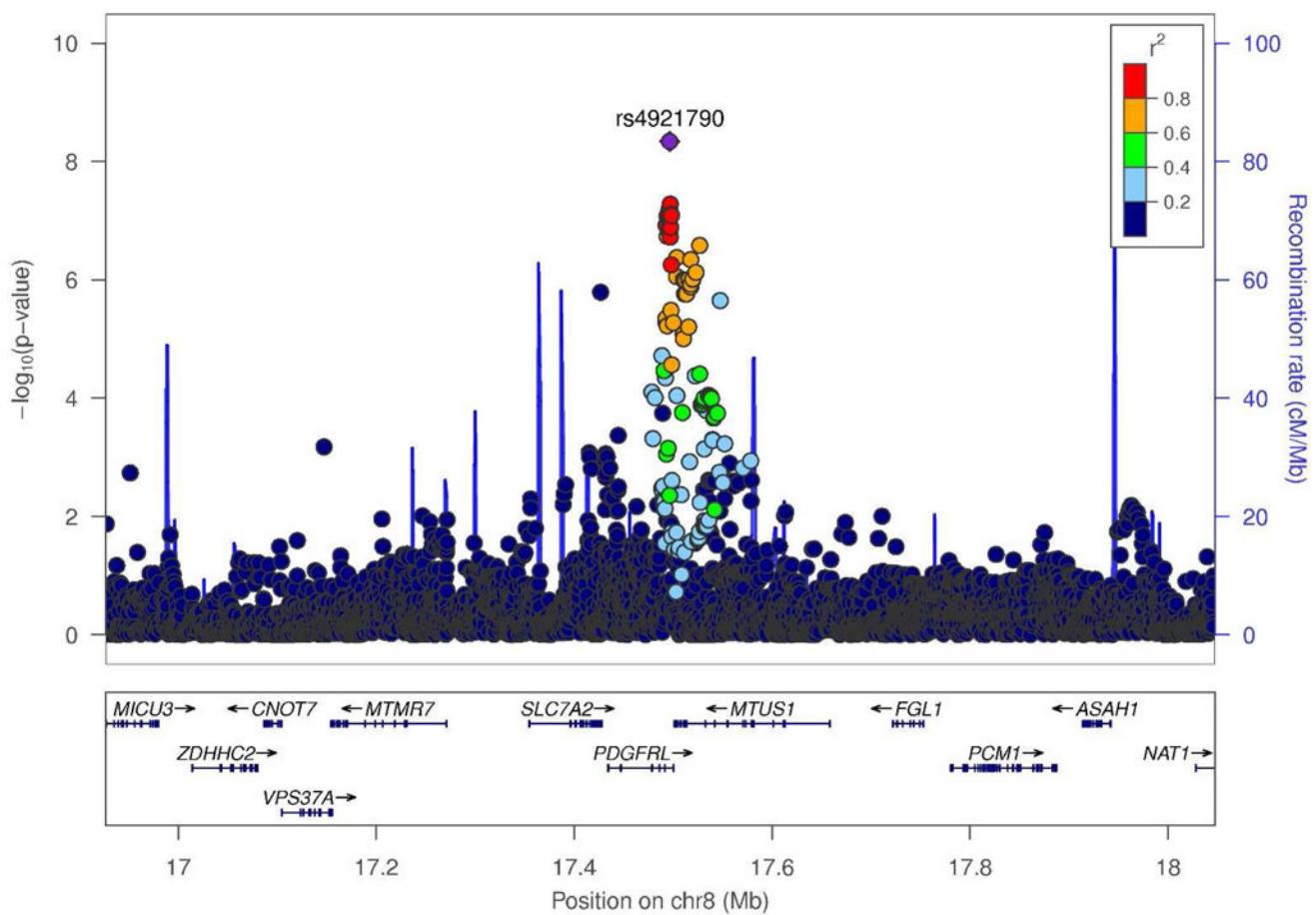
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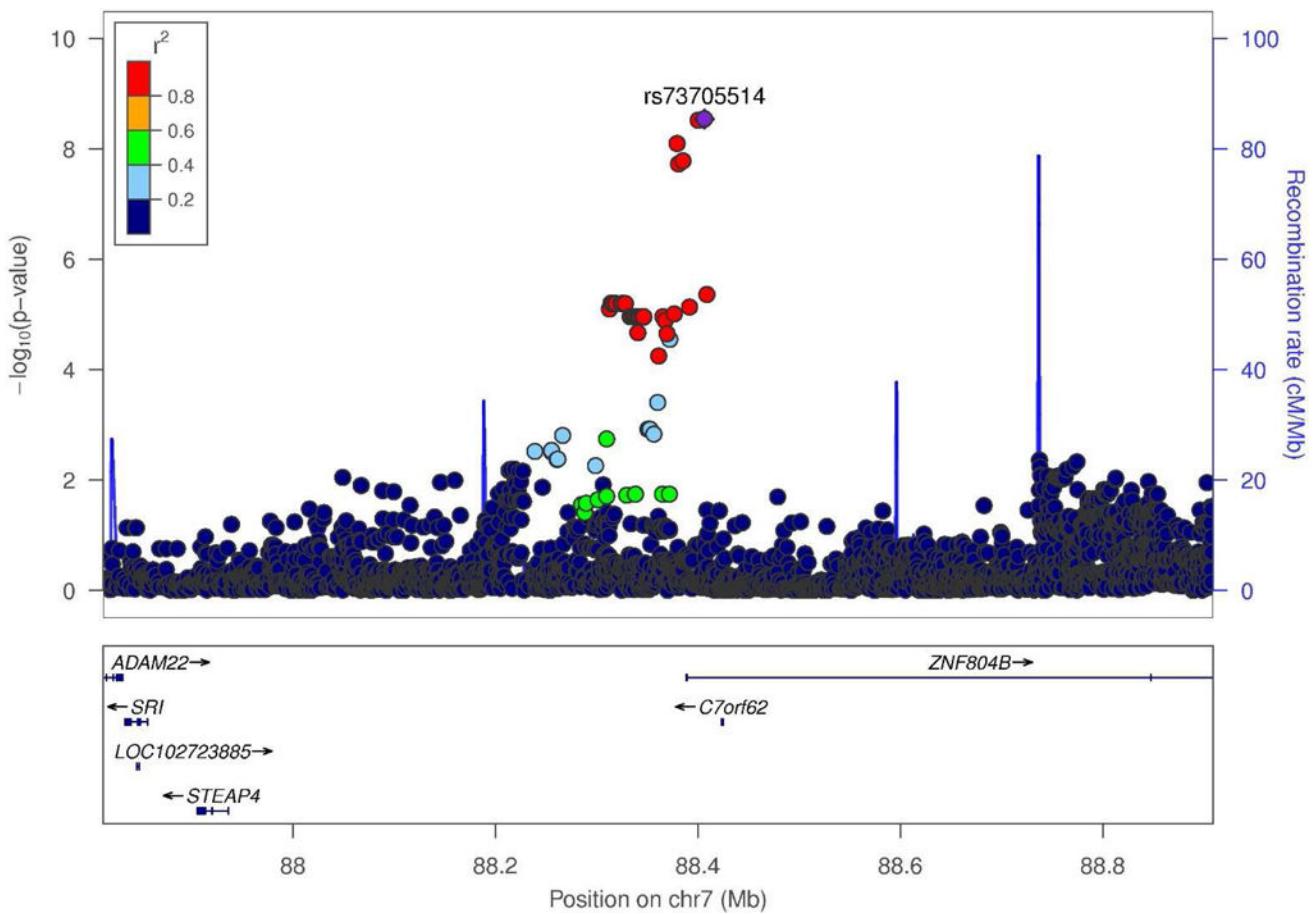
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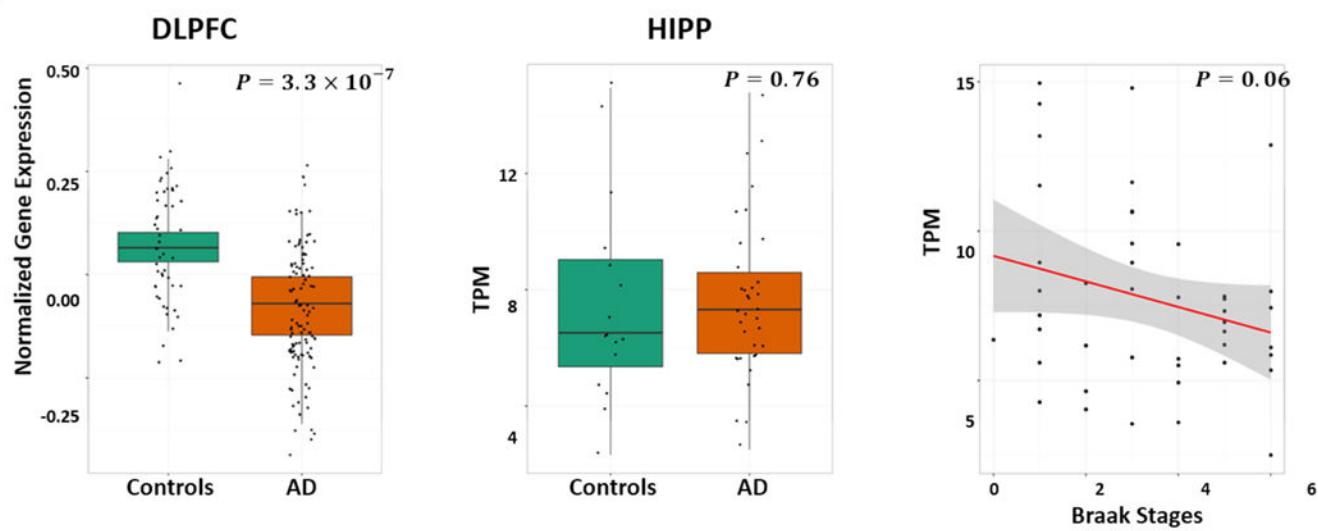
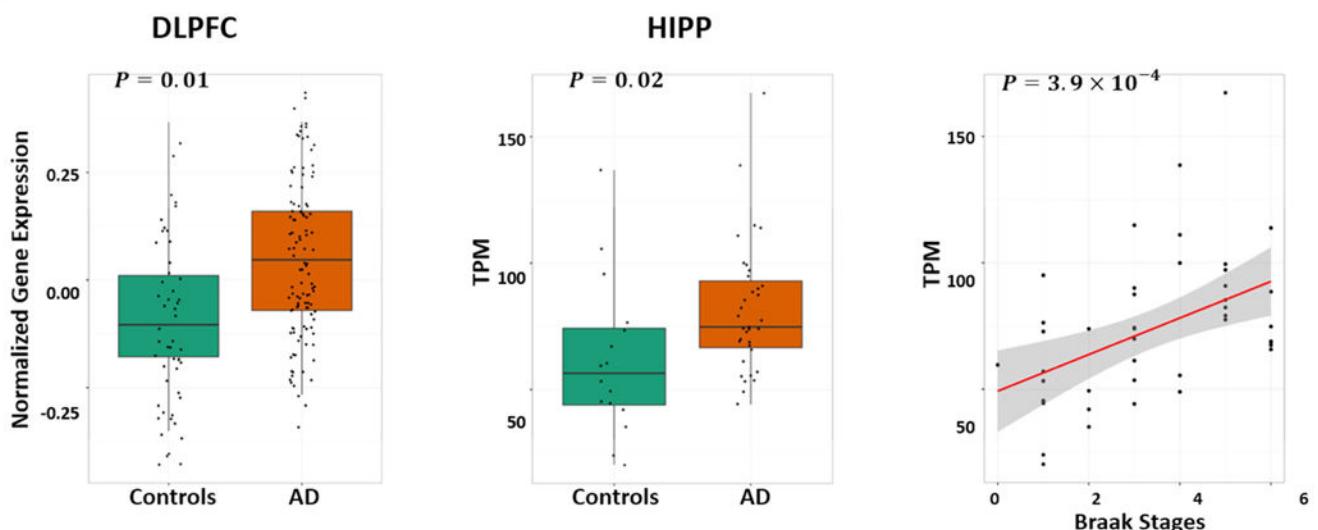
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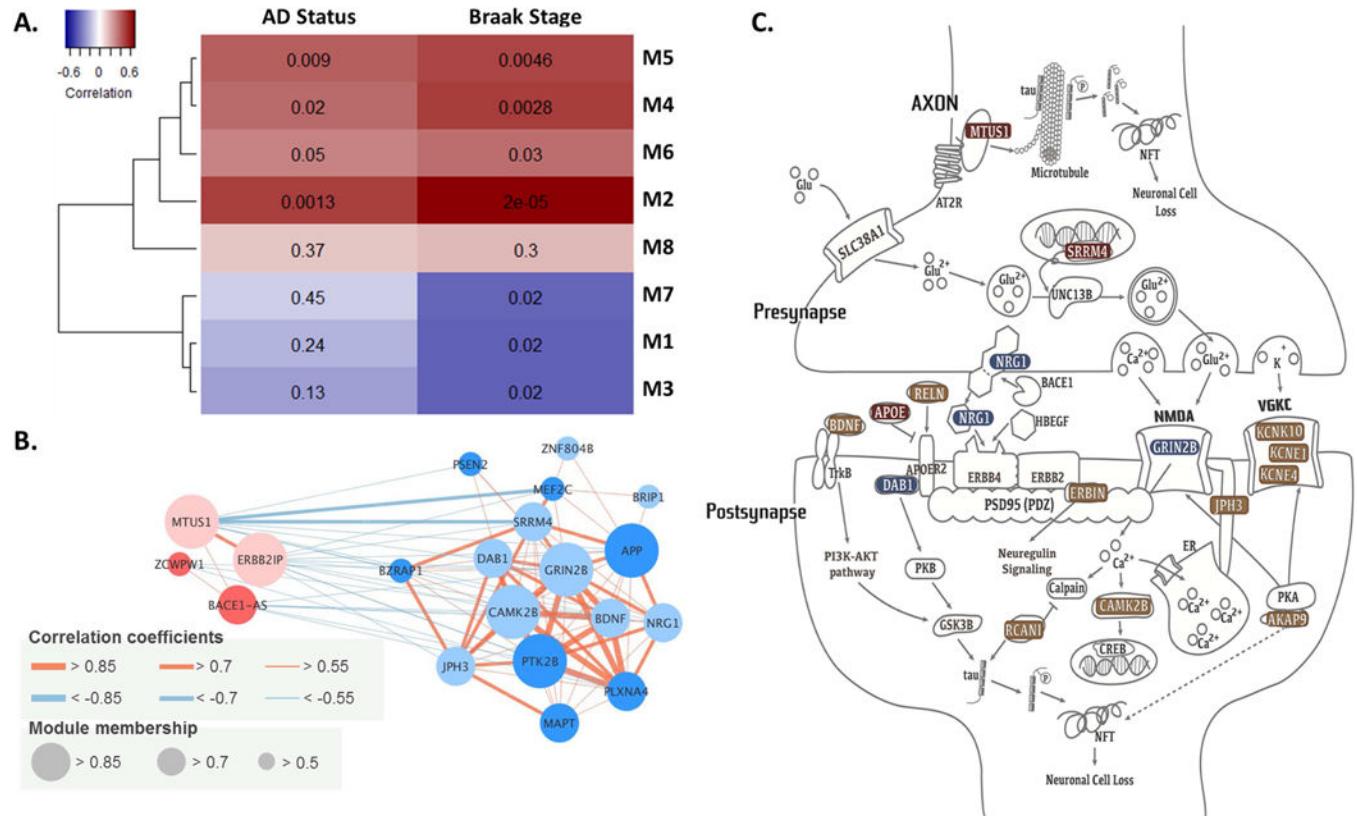
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**Fig. 1.**

Regional association plots of (A) *SRRM4* and (B) *C14orf79* for CSF total tau in the CN subjects, (C) *MTUS1* for hippocampal volume in the CN subjects, and (D) *ZNF804B* for logical memory delayed recall test in the MCI subjects.

A.**B.****Fig. 2.**

Expression studies of (A) *C14orf79* and (B) *MTUS1* including boxplots for differentially expressed genes in the brain dorsolateral prefrontal cortex (DLPFC) in the microarray data (GEO: GSE44772; left column) and in the hippocampus (HIPP) in the RNA-Seq data (Eisai Bio Bank data; middle column), and regression plots of gene expression by Braak stage in the RNA-Seq data (Eisai Bio Bank data; right column). P-values in plots were computed from linear regression models after adjusting for covariates (details in the Methods).

**Fig. 3.**

(A) A heat-map of correlations between AD traits and the first principal component values of genes in modules. Deeper colors indicate higher correlation with the traits (red: positive and blue: negative). Values in the heat-map are correlation *P*-values. (B) Connectivity plot of genes in modules M4 and M7, which were identified in this study or in previously reported in AD genome-wide association studies (GWAS). Light colored circles indicate the genes identified in this study, and deep colored circles indicate previously identified AD genes. (C) The role of genes identified in GWAS of AD-related endophenotypes among cognitively normal and mild cognitively impaired subjects in the top-ranked canonical pathways; colors indicate the level of association significance of the genes identified in this study (red = study-wide significance; blue = genome-wide significance; brown = $P < 1.0 \times 10^{-6}$ (except for *BDNF* in Supplementary Fig. 13H).

Table 1

Genome-wide significant association ($P < 5.0 \times 10^{-8}$) of novel genes in cognitively normal (CN), mild cognitive impairment (MCI), or AD dementia subjects with CSF protein levels ($\text{A}\beta_{42}$ and total and phosphorylated taur), hippocampal volume (HPV), and logical memory tests of immediate (LMdT) and delayed (LMdT) recall.

Group	Traits	CHR	BP	SNP	MA	MAF	BETA	SE	P	Closest Genes
ALL	$\text{A}\beta_{42}$	8	20647323	rs5644114	A	0.16	0.41	0.07	2.54×10^{-8}	<i>GFRA2, LZTS1</i>
		12	13870464	rs7442473	G	0.07	-1.02	0.18	2.53×10^{-8}	<i>GRIN2B</i>
CN	$t\text{-Tau}$	17	59687842	rs2378873	T	0.44	-0.53	0.10	2.03×10^{-8}	<i>BRIP1, NACA2</i>
		12	119390525	rs10775009	T	0.34	0.51	0.09	1.59×10^{-9}	<i>SRRM4</i>
CN	HPV	14	105385352	rs2819438	A	0.13	-0.80	0.14	6.94×10^{-9}	<i>PLD4, C14orf79</i>
		8	17496561	rs4921790	C	0.12	0.61	0.10	4.58×10^{-9}	<i>PDGFRL, MTUS1</i>
LMdT	LMdT	1	57739164	rs74834332	A	0.03	0.79	0.14	4.30×10^{-8}	<i>DAB1</i>
		10	53818149	rs12268753	C	0.21	0.30	0.05	2.01×10^{-8}	<i>PRKG1</i>
MCI	LMdT	4	86416554	rs111882035	G	0.02	-0.93	0.17	2.74×10^{-8}	<i>ARHgap24</i>
		7	88406552	rs73705514	C	0.02	-0.84	0.14	2.86×10^{-9}	<i>ZNF804B</i>
AD	HPV	8	31228770	rs118130881	G	0.04	-0.61	0.11	1.72×10^{-8}	<i>NRG1</i>
		6	136077929	rs79846291	T	0.02	1.85	0.31	1.76×10^{-8}	<i>LINC00271, PDE7B</i>

MA is minor allele, and MAF is the minor allele frequency. Bold SNPs denote the study-wide significant association ($P < 8.33 \times 10^{-9}$) with a trait.

Table 2

Genotype specific effect of the expression level among study-wide significant (8.33×10^{-9}) SNPs using the GTEx Portal database.

eQTL	EA	RA	Gene	eSNP Association Summary		
				<u>Hippocampus</u>		Other brain region
				P-value (β)	P-value (β)	
rs10775009	T	C	<i>SRM4</i>	0.09 (0.09)	0.16 (0.10)	Frontal Cortex
rs2819438	A	C	<i>PLD4</i>	0.47 (0.12)	0.10 (0.42)	Anterior cingulate cortex
			<i>AHNAK2</i>	0.15 (0.23)	0.06 (-0.28)	Cerebellar Hemisphere
			<i>C14orf79</i>	3.8×10^{-4} (0.44)	3.9×10^{-3} (0.31)	Cortex
						Cerebellum
rs4921790	C	A	<i>MTUS1</i>	1.0 (0.0)	0.09 (-0.16)	Caudate
rs55653268	T	G				Caudate
						Nucleus accumbens

EA and RA indicate effect allele and reference allele, respectively. Positive effect (β) in eSNP association summary means that carriers with effect alleles of a SNP tend to have higher expression level of a gene. Rs55653268 is in high LD with rs4921790 ($r^2 = 0.7$; $D = 1.0$) and is significantly associated with the hippocampal volume (effective allele: T; β [SE]: 0.6 [0.1], P-value: 4.2×10^{-7})

Table 3

Summary statistics of association between expression levels of the genes (*MTUS1* and *C14orf79*) and AD status as well as Braak Stage.

Source	Brain region	Predictor	<i>MTUS1</i>			<i>C14orf79</i>		
			β	SE	P	β	SE	P
Eisai Bio Bank (RNA-Seq)	HIPP	AD Status	0.38	0.16	0.02	0.04	0.14	0.76
	HIPP	Braak Stages	0.16	0.04	3.9×10 ⁻⁴	-0.07	0.03	0.06
Mt. Sinai Hospital (Microarray)	CER	AD Status	0.00	0.02	0.78	-0.13	0.02	7.5×10 ⁻⁸
	DLPFC	AD Status	0.09	0.04	0.01	-0.13	0.02	3.3×10 ⁻⁷
	VCX	AD Status	0.09	0.03	0.01	-0.15	0.03	2.1×10 ⁻⁸

Brain regions tested were hippocampus (HIPP), cerebellum (CER), dorsolateral prefrontal cortex (DLPFC), visual cortex (VCX). Positive effect (β) for AD status means that a gene is up-regulated in AD cases vs. in controls. Positive effect for Braak stages means that the expression of a gene is positively correlated with the Braak stages.