Adapting Tavtigian et al.'s Bayesian System for LLRs

Background

We recently developed a method to translate variant effect scores from VE maps into measures of log likelihood of pathogenicity based on the densities of positive and negative reference variants across the VE map's score spectrum. The goal behind these efforts was to provide a more useful metric for variant classification. These classifications however are classically based on the ACMG guidelines which only use simple categories.

Tavtigian et al. 2018 previously introduced the idea of representing the ACMG variant classification guidelines as a Bayesian framework. They based this on equating the different evidence levels (Pathogenic-Very Strong, Pathogenic-Strong, Pathogenic-Moderate, Pathogenic-Supporting, Benign-Supporting, and Benign-Strong) to different thresholds of what they call Odds of Pathogenicity or OddsPath (OP).

They state a number of modeling goals, which are based on the ACMG's loose definition of the 'Pathogenic' classification as corresponding to a greater than 99% chance of being pathogenic, and 'Likely pathogenic' as corresponding to a greater than 90% chance of being pathogenic. This means a successful model should fulfill the following criteria:

- (A) Evidence combination rules with the same classification outcome (e.g. likely pathogenic) should ideally yield the same lower bound on posterior probability.
- (B) Evidence combination rules for 'Likely Pathogenic' should yield posteriors greater than 90%.
- (C) Evidence combination rules for 'Pathogenic' should yield posteriors greater than 99%.

A quick examination of Tavtigian et al.'s underlying methods shows that what they refer to as 'OddsPath' (OP) is just another name for the likelihood ratio

$$\frac{P(Data|Pathogenic)}{P(Data|Benign)}$$

, often also referred to as the Bayes Factor, or Λ . This makes it easy for us to adopt to our existing LLR metric, since the log Likelihood Ratio (LLR) is simply $\log(\Lambda)$.

The authors make a number of modeling decisions:

- 1. Adjacent evidence levels should scale in OP exponentially by a power of X. That means our equivalent LLRs scale multiplicatively by X.
- 2. ACMG rules for evidence combination work by multiplying OPs. That means our equivalent LLRs will be combined by addition.

Next they have to pick a number of modeling parameters. The parameters to choose are:

- 1. The scaling factor X.
- 2. An underlying prior probability of pathogenicity P_{prior} .
- 3. A starting point for their OP scale from which all other OP thresholds follow. For this they pick highest pathogenic evidence level Pathogenic-Very strong OP_{PVSt} .

For the scaling factor, they choose X=2. This is because it does best at fulfilling criterion (A).

For the prior, they more or less arbitrarily pick $P_{prior} = 0.1$. The authors claim that they roughly base this on the prevalence of pathogenic variants in BRCA1 and BRCA2.

Finally, they have to choose a starting point for their OP scale, which they do by picking an arbitrary OP for the Pathogenic-Very strong $(OP_{PVSt} = 350)$. This number was picked because it fulfilled criteria (B) and (C) given their choice of X and the prior.

Re-implementation with LLRs

We can quickly test that by choosing the same parameters, our LLR-based implementation yields the exact same results for the combination rules:

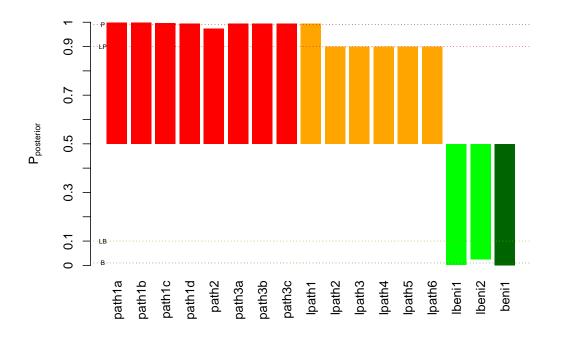
```
> #Modeling parameters
> prior <- 0.1
> LLRpvst <- log10(350)
> X <- 2
> #function to calculate posterior given prior and LLR
> llr2post <- function(llr,prior=0.1) {</pre>
    #log prior odds
+
          lpo <- log10(prior/(1-prior))</pre>
          #posterior odds
          posto <- 10^(llr+lpo)</pre>
+
          #posterior prob
          (posto) / (1+posto)
+
> #here we derive the LLR evidence thresholds based on our parameters
> calcThresholds <- function(LLRpvst,X) {</pre>
    list(
+
+
      #pathogenic very strong
            LLRpvst=LLRpvst,
             #pathogenic strong
            LLRpst=LLRpvst/X,
             #pathogenic moderate
            LLRpm=LLRpvst/(X^2),
             #pathogenic supporting
            LLRpsu=LLRpvst/(X^3),
```

```
#benign supporting
             LLRbsu=-LLRpvst/(X^3),
             #benign strong
             LLRbst = -LLRpvst/(X^1)
+
    )
+ }
> thresholds <- calcThresholds(LLRpvst,X)</pre>
> #print thresholds
> round(unlist(thresholds),digits=3)
LLRpvst LLRpst
                   LLRpm LLRpsu LLRbsu LLRbst
  2.544
          1.272
                   0.636
                           0.318 - 0.318 - 1.272
> #test evidence combination rules
> calcComboLLRs <- function(thresholds) {</pre>
    with(thresholds,c(
+
      #pathogenic combinations
             path1a=LLRpvst+LLRpst,
             path1b=LLRpvst+2*LLRpm,
+
             path1c=LLRpvst+LLRpm+LLRpsu,
+
             path1d=LLRpvst+2*LLRpsu,
             path2 =2*LLRpst,
             path3a=LLRpst+3*LLRpm,
             path3b=LLRpst+2*LLRpm+2*LLRpsu,
             path3c=LLRpst+LLRpm+4*LLRpsu,
             #likely pathogenic combinations
             lpath1=LLRpvst+LLRpm,
             lpath2=LLRpst+LLRpm,
             lpath3=LLRpst+2*LLRpsu,
             lpath4=3*LLRpm,
             lpath5=2*LLRpm+2*LLRpsu,
             lpath6=LLRpm+4*LLRpsu,
             #likely benign combinations
             lbeni1=LLRbst+LLRbsu,
+
             1beni2=2*LLRbsu,
             #benign combinations
             beni1 =2*LLRbst
    ))
+
+ }
> #calculate resulting posteriors
> comboPosts <- llr2post(calcComboLLRs(thresholds),prior)</pre>
> #and print
> print(round(comboPosts,digits=3))
path1a path1b path1c path1d path2 path3a path3b path3c lpath1 lpath2 lpath3
 0.999 \quad 0.999 \quad 0.997 \quad 0.994 \quad 0.975 \quad 0.994 \quad 0.994 \quad 0.994 \quad 0.994 \quad 0.900 \quad 0.900
```

```
lpath4 lpath5 lpath6 lbeni1 lbeni2 beni1
0.900 0.900 0.900 0.003 0.025 0.000
```

Indeed that matches the numbers in Tavtigian et al. 2018 perfectly. Let's plot these numbers, so we can inspect the consistency of the rules:

```
> plotRules <- function(comboPosts,...) {</pre>
    op \leftarrow par(las=3,mar=c(10,5,4,1))
    barplot(comboPosts-.5, ylim=c(-0.5,.5), border=NA, ylab=expression(P[posterior]),
            col=c(rep("red",8),rep("orange",6),rep("green",2),"darkgreen"),
            axes=FALSE,...
    )
+
    axis(2,at=seq(-.5,.5,.1),seq(0,1,.1))
+
    hs <-c(.99,.9,.1,.01)-.5
    abline(h=hs,lty="dotted",
+
      col=c("firebrick4", "firebrick3", "chartreuse3", "chartreuse4")
    text(0,hs,c("P","LP","LB","B"),cex=.5)
+
+ }
> plotRules(comboPosts)
```



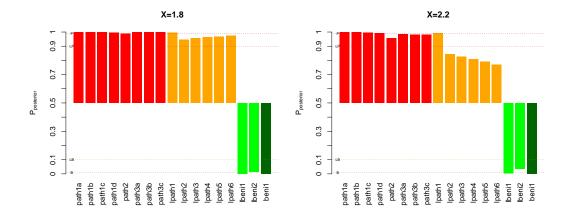
The same observations that Tavtigian et al. make stick out here as well: Rule 'path2' as defined in the ACMG guidelines is not strong enough; rule 'lpath1' is too strong; and rule 'lbeni1' is too strong.

A deeper exploration of parameter choice

Now, let's explore the effects of the different modeling parameters ourselves.

Indeed, choosing any other X than 2 results in violation of criterion (A), by skewing the values of equivalent rules further from each other.

- > layout(cbind(1,2))
- > thresholds <- calcThresholds(LLRpvst,X=1.8)</pre>
- > comboPosts <- llr2post(calcComboLLRs(thresholds),prior)</pre>
- > plotRules(comboPosts,main="X=1.8")
- > thresholds <- calcThresholds(LLRpvst,X=2.2)</pre>
- > comboPosts <- llr2post(calcComboLLRs(thresholds),prior)</pre>
- > plotRules(comboPosts,main="X=2.2")



The more we stray from X=2 in either direction, the more the 'Likely pathogenic' combination rules fall out of sync with each other. So we must conclude that X isn't really much of a free parameter at all. Only X=2 is a choice that can fulfill our criteria.

As for OP_{PVSt} and P_{prior} , we can see that both primarily affect overall scale of the rule posteriors. Thus, only certain combinations of the two can be expected to fulfill criteria (B) and (C). We can actually derive a precise relationship between the two that allows us to strike this balance by making using the likely pathogenic combination rules as a guide. Since they must achieve a posterior of $P_{post} = 90\%$, the following must hold:

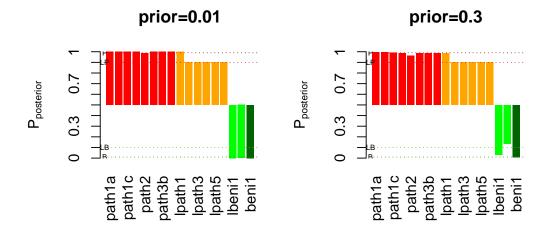
$$\frac{LLR_{PVst}}{2} + \frac{LLR_{PVst}}{4} + \log(\frac{P_{prior}}{1 - P_{prior}}) = \log(\frac{P_{post}}{1 - P_{post}})$$

If we solve for LLR_{PVst} :

$$LLR_{PVst} = \frac{4}{3} \log \left(\frac{P_{post}(1 - P_{prior})}{P_{prior}(1 - P_{post})} \right)$$

Let's test this:

```
> #function to calculate the optimal LLRpvst from the prior
> optiLLR <- function(prior,posterior=0.9) {
+ log10(posterior*(1-prior)/(prior*(1-posterior)))*4/3
+ }
> layout(cbind(1,2))
> prior <- 0.01
> LLRpvst <- optiLLR(prior)
> thresholds <- calcThresholds(LLRpvst,X)
> comboPosts <- llr2post(calcComboLLRs(thresholds),prior)
> plotRules(comboPosts,main="prior=0.01")
> prior <- 0.3
> LLRpvst <- optiLLR(prior)
> thresholds <- calcThresholds(LLRpvst,X)
> comboPosts <- llr2post(calcComboLLRs(thresholds),prior)
> plotRules(comboPosts,main="prior=0.3")
```



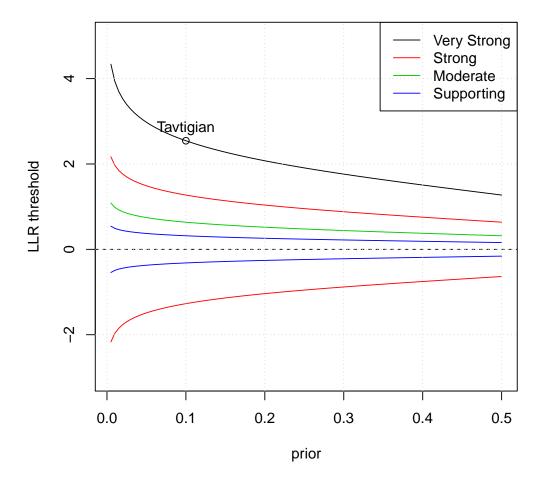
Indeed it looks like all rules except for the problematic ones already identified by Tavtigian et al themselves remain consistent.

Having derived this relationship also lets us figure out why Tavtigian et al. settled on $OP_{PVSt} = 350$. It is numerically very close to the optimal solution for a prior of 0.1:

```
> 10^optiLLR(0.1)
[1] 350.4666
```

Now, lets see what the relationship between the prior and the LLR evidence thresholds looks like in graphical form. I am highlighting Tavtigian's parameter choice in the plot as well.

```
> priors <- seq(0.005, 0.5, 0.005)
> LLRpvst <- optiLLR(priors)</pre>
> plot(priors,LLRpvst,type="1",
          xlab="prior", ylab="LLR threshold",
          vlim=c(-3,5)
+ )
> lines(priors,LLRpvst/2,col=2)
> lines(priors,LLRpvst/4,col=3)
> lines(priors,LLRpvst/8,col=4)
> lines(priors,-LLRpvst/8,col=4)
> lines(priors,-LLRpvst/2,col=2)
> abline(h=0,lty="dashed")
> grid()
> legend("topright",
          c("Very Strong", "Strong",
                   "Moderate", "Supporting"),
          col = c(1:4), lty = 1
+ )
> #Highlight Tavtigian's parameter set
> points(0.1,log10(350))
> text(0.1,log10(350),"Tavtigian",pos=3)
```



Finally let's look at this in the form of a table

```
> priors <- seq(0.05,0.5,0.05)
> LLRpvst <- optiLLR(priors)
> llrThresholds <- data.frame(prior=priors,patho.Vstr=LLRpvst,patho.str=LLRpvst/2,
+ patho.mod=LLRpvst/4,patho.sup=LLRpvst/8
+ )
> llrThresholds
```

```
prior patho.Vstr patho.str patho.mod patho.sup
1
    0.05
           2.977328 1.4886641 0.7443320 0.3721660
2
    0.10
           2.544647 1.2723233 0.6361617 0.3180808
3
    0.15
           2.276760 1.1383801 0.5691901 0.2845950
4
    0.20
           2.075070 1.0375350 0.5187675 0.2593838
5
    0.25
           1.908485 0.9542425 0.4771213 0.2385606
6
    0.30
           1.762959 0.8814795 0.4407398 0.2203699
7
    0.35
           1.630784 0.8153919 0.4076959 0.2038480
```

```
8 0.40 1.507112 0.7535558 0.3767779 0.1883890
9 0.45 1.388524 0.6942618 0.3471309 0.1735654
10 0.50 1.272323 0.6361617 0.3180808 0.1590404
```

Conclusions

Of the three modeling parameters Tavtigian et al use, only one is really a free parameter: The prior probability of pathogenicity. From it the optimal LLR thresholds of all the evidence levels can be derived such as to yield the best compatibility with the ACMG ruleset.

This raises the question whether prior should probably be chosen separately for each gene, based on the best guess of the incidence rate of pathogenic variants in the gene.