## homework 6

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#### Homework 6

Constrained Ordination

Question 1: Constrained ordination is a type of ordination whereby the axes are constrained by environmental factors. For your dataset, which are the "response" variables and which are the "predictor" or "environmental" variables? Alternatively, do you think a symmetric analysis is more appropriate for your data (as opposed to an asymmetric or hypothesis-testing approach), and if so, why?

The response variables are the salamander data, and the predictor variables are my environmental matrix, i.e. downed wood, etc. I think symmetric could be more useful for my data for exploring structure and relationships.

# Question 2: Perform a Redundancy Analysis (RDA) on your dataset (you can use a simple, transformationbased,

or distance-based approach, but justify your choice!). Interpret the results by discussing the relationship between the response and predictor variables. What percentage of variance in the response data is explained by the predictor variables? Provide R output and visualizations (e.g., RDA triplots) to support your analysis.

I'm using tbRDA because it works well with data that has a lot of zeros.

```
sal.hel <- decostand(sals, "hellinger")

tbRDA <- rda(sal.hel ~ ., env_cont)

##

## Some constraints or conditions were aliased because they were redundant. This
## can happen if terms are linearly dependent (collinear): 'logs'

summary(tbRDA)</pre>
```

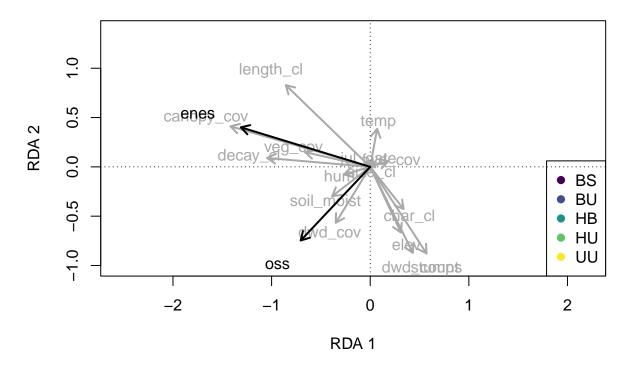
```
##
## Call:
## rda(formula = sal.hel ~ jul_date + elev + temp + hum + canopy_cov + veg_cov + dwd_cov + fwd_cov
##
## Partitioning of variance:
## Inertia Proportion
```

```
## Total
                 0.3394
                            1.0000
## Constrained
                 0.0677
                            0.1995
## Unconstrained 0.2717
                            0.8005
##
## Eigenvalues, and their contribution to the variance
##
## Importance of components:
##
                            RDA1
                                   RDA2
                                           PC1
                                                    PC2
## Eigenvalue
                         0.05114 0.01656 0.1775 0.09423
## Proportion Explained 0.15067 0.04879 0.5229 0.27764
## Cumulative Proportion 0.15067 0.19946 0.7224 1.00000
## Accumulated constrained eigenvalues
## Importance of components:
##
                            RDA1
## Eigenvalue
                         0.05114 0.01656
## Proportion Explained 0.75540 0.24460
## Cumulative Proportion 0.75540 1.00000
        anova(tbRDA, permutations = how(nperm = 999))
## Permutation test for rda under reduced model
## Permutation: free
## Number of permutations: 999
## Model: rda(formula = sal.hel ~ jul_date + elev + temp + hum + canopy_cov + veg_cov + dwd_cov + fwd_c
            Df Variance
                             F Pr(>F)
            15 0.067699 1.8438 0.011 *
## Model
## Residual 111 0.271709
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
        anova(tbRDA, by = "axis", permutations = how(nperm = 999))
## Permutation test for rda under reduced model
## Forward tests for axes
## Permutation: free
## Number of permutations: 999
## Model: rda(formula = sal.hel ~ jul_date + elev + temp + hum + canopy_cov + veg_cov + dwd_cov + fwd_c
##
            Df Variance
                              F Pr(>F)
## RDA1
             1 0.051140 23.3387 0.002 **
## RDA2
             1 0.016559 7.5572 0.988
## Residual 124 0.271709
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
        anova(tbRDA, by = "terms", permutations = how(nperm = 999))
## Permutation test for rda under reduced model
## Terms added sequentially (first to last)
## Permutation: free
```

```
## Number of permutations: 999
##
## Model: rda(formula = sal.hel ~ jul_date + elev + temp + hum + canopy_cov + veg_cov + dwd_cov + fwd_c
              Df Variance
                           F Pr(>F)
## jul_date 1 0.000028 0.0115 0.990
## elev
             1 0.004956 2.0247 0.133
## temp
              1 0.000859 0.3510 0.672
         1 0.003031 1.2383 0.294
## hum
## canopy_cov 1 0.032173 13.1435 0.001 ***
## veg_cov 1 0.003876 1.5834 0.199
## dwd_cov
             1 0.002125 0.8681 0.422
## fwd_cov 1 0.004299 1.7563 0.187
## soil_moist 1 0.002106 0.8603 0.421
## dwd_count 1 0.001529 0.6245 0.520
## stumps 1 0.001473 0.6016 0.525
## size_cl 1 0.000373 0.1523 0.867
## decay_cl 1 0.004677 1.9105 0.145
## char cl
             1 0.000705 0.2880 0.745
## length_cl 1 0.005490 2.2429 0.115
## Residual 111 0.271709
## ---
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
        RsquareAdj(tbRDA)
## $r.squared
## [1] 0.1994623
## $adj.r.squared
## [1] 0.09128155
        goodness(tbRDA)
##
                       RDA2
             RDA1
## oss 0.05935405 0.1259271
## enes 0.27107790 0.2964183
       vif.cca(tbRDA)
##
     jul_date
                                          hum canopy_cov veg_cov
                                                                      dwd_cov
                   elev
                              temp
     2.193660
              2.400873
                          3.780248
                                     3.421559
                                                3.521891 1.931301
                                                                     2.245364
##
     fwd_cov soil_moist dwd_count
                                       stumps
                                                   logs
                                                           {\tt size\_cl}
                                                                     decay_cl
                                                                     2.359334
##
    2.059265
              1.938509
                         7.228652 14.209641
                                                     NΑ
                                                          1.151702
##
     char_cl length_cl
##
    1.930887
              7.079383
       groups <- levels(factor(env$trt))</pre>
       pt_col <- viridis(length(groups))</pre>
       site.sc <- summary(tbRDA)[2]</pre>
        spe.sc <- summary(tbRDA)[1]</pre>
```

```
env.sc <- summary(tbRDA)[4]</pre>
plot(tbRDA,
    choices=c(1,2),
    type="n",
    scaling=2,
    main="tbRDA of Hellinger-transformed Sal Count",
    xlab="RDA 1",
    ylab="RDA 2")
for (i in 1:length(groups)){
    dim_choice <- site.sc$sites[env$SMU==groups[i],]</pre>
    points(dim_choice[,1], dim_choice[,2],
    pch=19,
    cex=1.4,
    col=pt_col[i])
text(env.sc$biplot*2,
    row.names(env.sc$biplot),
    col="darkgray")
arrows(0, 0, env.sc$biplot[,1]*1.7, env.sc$biplot[,2]*1.7,
    lwd=2.
    length=0.1,
    col="darkgray")
text(spe.sc$species*2,
    row.names(spe.sc$species))
arrows(0, 0, spe.sc$species[,1]*1.5, spe.sc$species[,2]*1.5,
    lwd=2,
    length=0.1)
legend(x="bottomright",
    legend=groups,
    col=pt_col,
    pch=19)
```

## tbRDA of Hellinger-transformed Sal Count



The total constrained inertia from my predictors is about 20%, which isnt great. Of that, RDA1 accounts for about 91% and RDA2 accounts for about 8%.

Question 3: Next, perform a Canonical Correspondence Analysis (CCA) on your dataset. Compare and contrast the results of the RDA with the CCA. Discuss the assumptions made in RDA and CCA and whether they are met by your dataset. How do the results differ in terms of species-environment (or other response-predictor) relationships?

I cant get this to work with my data, as it doesn't want any zeros in the community data matrix.

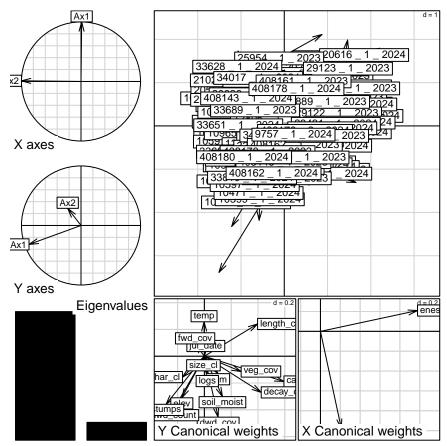
Question 4: Conduct a Co-inertia Analysis (CoIA), even if your "Y1" and "Y2" matrices are better suited for a directional hypothesis. How do the two datasets relate to one another? Use the RV coefficient to assess the strength of the shared structure between the datasets.

```
dudi.sal <- dudi.pca(sal.hel, scale=FALSE, scannf=FALSE)
  dudi.env <- dudi.pca(env_cont, scale=TRUE, scannf = FALSE)</pre>
```

```
coia <- coinertia(dudi.sal, dudi.env, scannf = FALSE, nf=2)</pre>
            summary(coia)
## Coinertia analysis
## Class: coinertia dudi
## Call: coinertia(dudiX = dudi.sal, dudiY = dudi.env, scannf = FALSE,
##
       nf = 2)
## Total inertia: 0.1152
## Eigenvalues:
       Ax1
               Ax2
## 0.10116 0.01403
##
## Projected inertia (%):
      Ax1
               Ax2
           12.18
##
    87.82
##
## Cumulative projected inertia (%):
            Ax1:2
##
      Ax1
    87.82 100.00
##
##
## Eigenvalues decomposition:
                    covar
                                sdX
            eig
                                         sdY
## 1 0.10115803 0.3180535 0.3778140 1.797890 0.4682298
## 2 0.01403038 0.1184499 0.4404449 1.240336 0.2168223
## Inertia & coinertia X (dudi.sal):
        inertia
                      max
## 1 0.1427434 0.1939982 0.7357975
## 12 0.3367351 0.3367351 1.0000000
## Inertia & coinertia Y (dudi.env):
       inertia
                  max
## 1 3.232408 3.838730 0.8420516
## 12 4.770841 6.341899 0.7522733
##
## RV:
## 0.08229505
       randtest(coia, nrepet = 999)
## Monte-Carlo test
## Call: randtest.coinertia(xtest = coia, nrepet = 999)
## Observation: 0.08229505
## Based on 999 replicates
## Simulated p-value: 0.001
## Alternative hypothesis: greater
```

##

### plot(coia)



The total inertia is 0.11, which is pretty low, and means that the variance in the two datasets is not explained by their common structure. The RV is 0.08, which again is very low. This reinforces that there is not a strong relationship between the environmental variables and the salamander data. I dont know why and I dont like it.

Question 5: Based on the analyses you conducted (RDA, CCA, CoIA), summarize the main ecological insights gained from your data. What limitations did you encounter in your analysis and how might they affect your interpretations? Which method do you feel is most appropriate for your data and why?

These all felt like a bit of a bust for my data. Not sure if its user error, but none of them reported notable relationships between the matrices, and I couldnt get the CCA to work at all. Maybe it just means that my environmental data actually doesnt have any strong relationships with my salamander data, which would probably mean I made mistakes in my study design somehow, because we know that environmental variables should be related to salamander occupancy and abundance.

The tbRDA plot does associate OSS with soil moisture and downed wood cover, which tracks what we know about the species, and ENES with canopy cover, which means its occurring in our less disturbed sites.