homework_5

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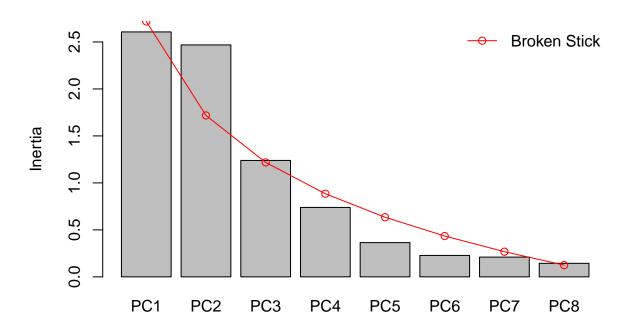
Homework 5

Ordination

Question 1) Conduct a Principal Component Analysis (PCA), Correspondence Analysis (CA), Principal Coordinate Analysis (PCoA), and Nonmetric Multidimensional Scaling Analysis (NMDS) using your dataset.

PCA - Environmental data

Broken Stick of PCs

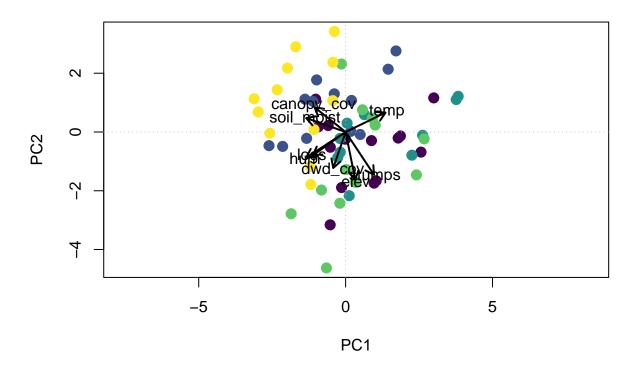


##

Eigenvectors:

```
PC1
                              PC3
                                            PC5
                                                   PC6
##
                       PC2
                                     PC4
                                  -0.396
## elev
              0.110 -0.542
                                          0.326
                                                0.516
              0.452 0.220 -0.390
                                   0.225
                                          0.224
## temp
             -0.445 -0.289
                            0.342
                                         -0.112 -0.342
## hum
## canopy_cov -0.352 0.279 -0.299 -0.648
                                          0.318
## dwd_cov
             -0.140 -0.408 -0.565 0.183
                                          0.349 -0.526
## soil_moist -0.446 0.158 0.230
                                   0.528
                                          0.582 0.318
## stumps
              0.322 -0.485 0.197
                                   0.101
                                          0.154
## logs
             -0.373 -0.265 -0.477 0.219 -0.495 0.475
```

PCA of Environmental Data



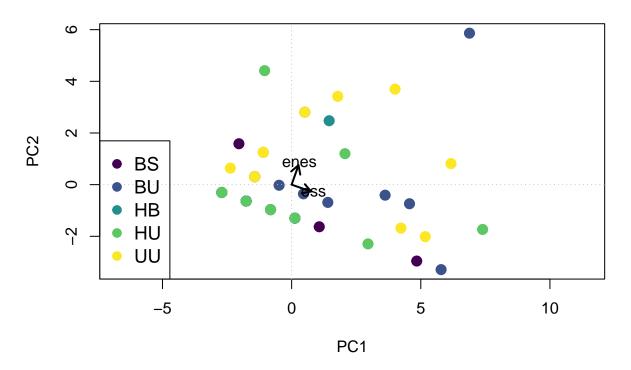
PCA - Sal data

This looks absolutely absurd, I would assume its mainly because I only have two species.

```
## Only 2 axes available
##
## Eigenvectors:

## PC1 PC2
## oss 0.943 -0.332
## enes 0.332 0.943
```

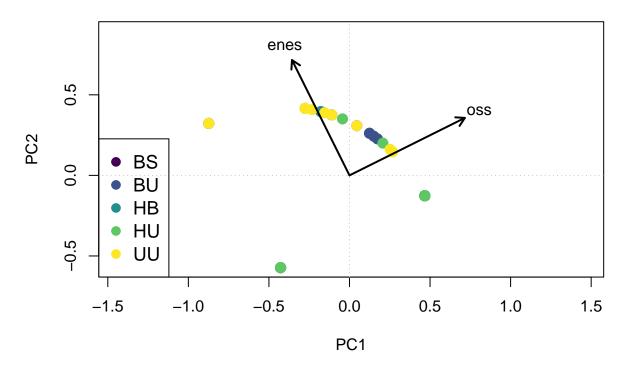
PCA of Sal Count Data (Untransformed)



```
## Only 2 axes available
##
## Eigenvectors:
## PC1 PC2
```

oss 0.895 0.446 ## enes -0.446 0.895

PCA of Sal Density Data (Hellinger)



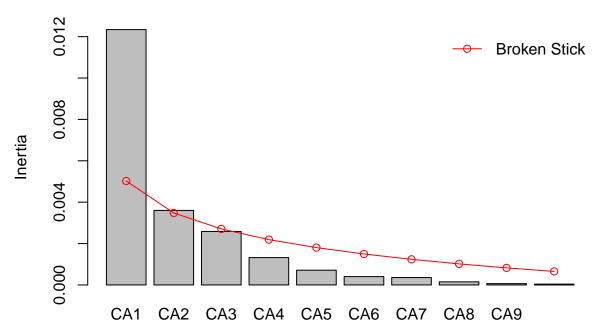
CA - Env data

##

```
## Call:
## cca(X = env_cont)
## Partitioning of scaled Chi-square:
##
                 Inertia Proportion
## Total
                 0.02164
                                   1
  Unconstrained 0.02164
                                   1
##
  Eigenvalues, and their contribution to the scaled Chi-square
##
##
  Importance of components:
##
                             CA1
                                                CA3
                                                         CA4
                                       CA2
                                                                    CA5
                                                                              CA6
## Eigenvalue
                         0.01234\ 0.003602\ 0.002585\ 0.001323\ 0.0007134\ 0.0004036
## Proportion Explained 0.57039 0.166478 0.119458 0.061140 0.0329730 0.0186530
## Cumulative Proportion 0.57039 0.736865 0.856323 0.917463 0.9504363 0.9690894
                                                    CA9
##
                                CA7
                                          CA8
                                                             CA10
                                                                        CA11
## Eigenvalue
                         0.0003582 0.0001505 6.427e-05 3.995e-05 2.533e-05
## Proportion Explained 0.0165561 0.0069542 2.971e-03 1.846e-03 1.171e-03
## Cumulative Proportion 0.9856454 0.9925996 9.956e-01 9.974e-01 9.986e-01
                              CA12
                                         CA13
## Eigenvalue
                         1.428e-05 1.094e-05 5.335e-06
## Proportion Explained 6.601e-04 5.059e-04 2.466e-04
```

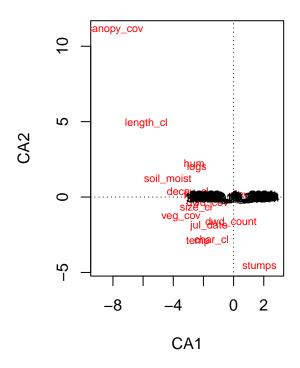
CA1 CA2 CA3 CA4 CA5 CA6 CA7 CA8 ## 57.03874 16.64779 11.94578 3.29730 1.86530 1.65561 0.69542 6.11402 ## CA9 CA10 CA11 CA12 CA13 **CA14** $0.29708 \quad 0.18464 \quad 0.11707 \quad 0.06601 \quad 0.05059$ 0.02466

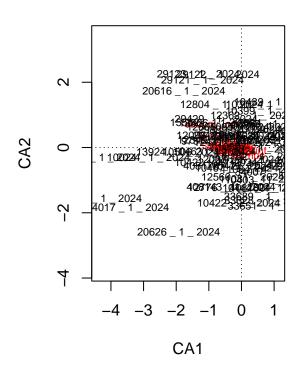
Inertia, CA vs. Random



Env CA, Scaling 1

Env CA, Scaling 2





null device
1

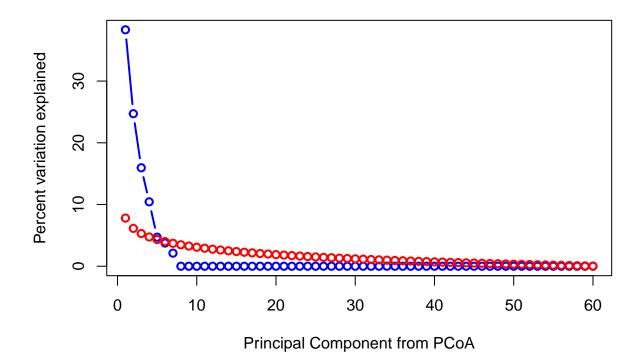
CA - Sal data

I believe this warning is happening because I have a lot of zeros in my salamander data, but I can't figure out what to do about that. I think this just isnt a great analysis for my data.

```
#sal.ca <- cca(sals)
#Error in cca.default(sals) :
# all row sums must be >0 in the community data matrix
```

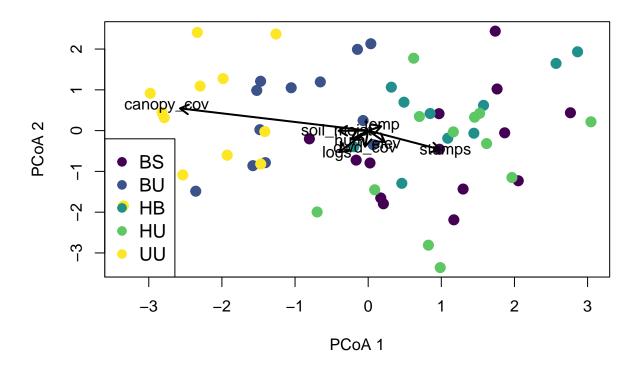
PCoA - Env data

Broken Stick Model



```
ylab = "PCoA 2",
     pch = 19)
for (i in 1:length(groups))
    dim_choice <- site.sc[dat2$trt==groups[i],]</pre>
    points(dim_choice[,1], dim_choice[,2],
        pch=19,
        cex=1.4,
        col=pt_col[i])
text(spe.sc*1.5, row.names(spe.sc))
arrows(0, 0, spe.sc[,1]*1.4, spe.sc[,2]*1.4,
    lwd=2,
    length=0.1)
legend(x="bottomleft",
    legend=levels(factor(dat2$trt)),
    col=pt_col[1:6],
    pch=19,
    cex=1.2)
```

Env PCoA



```
# Fit environmental variables to PCoA space for interpretation
envfit_results <- envfit(env.pcoa, env_std_subset)
print(envfit_results)</pre>
```

```
## ***VECTORS
##
##
                  Dim1
                           Dim2
                                    r2 Pr(>r)
              0.73756  0.67528  0.3514  0.001 ***
## temp
## dwd_cov
             -0.06858 -0.99765 0.7659 0.001 ***
## soil moist -0.99983 -0.01842 0.3091 0.001 ***
## stumps
              0.80849 -0.58851 0.7629 0.001 ***
## logs
              -0.43885 -0.89856 0.8121 0.001 ***
## decay_cl -0.98900 0.14792 0.6418 0.001 ***
## canopy_cov -0.95012  0.31189  0.7685  0.001 ***
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
## Permutation: free
## Number of permutations: 999
PCOA - Sal data
Again, I don't think this is useful for my data since I have a lot of zeros.
sal.bray <- vegdist(log_sal_dens, "bray")</pre>
## Warning in vegdist(log_sal_dens, "bray"): results may be meaningless because data have negative entr
                    in method "bray"
sal.pcoa <- cmdscale(sal.bray,</pre>
            k=5,
            eig=TRUE,
            add=T)
## Warning in cmdscale(sal.bray, k = 5, eig = TRUE, add = T): only 0 of the first
## 5 eigenvalues are > 0
NMDS - Env data only
## 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## Run 0 stress 0.1740743
## Run 1 stress 0.174237
## ... Procrustes: rmse 0.005032674 max resid 0.02553826
## Run 2 stress 0.1742868
## ... Procrustes: rmse 0.006032315 max resid 0.03043585
## Run 3 stress 0.1740887
## ... Procrustes: rmse 0.001649388 max resid 0.009675061
## ... Similar to previous best
## Run 4 stress 0.1740766
## ... Procrustes: rmse 0.002066923 max resid 0.009792351
## ... Similar to previous best
## Run 5 stress 0.174237
## ... Procrustes: rmse 0.005028381 max resid 0.02553346
```

Run 6 stress 0.174237

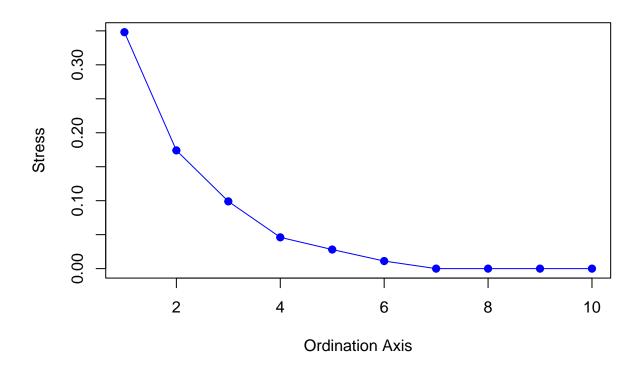
... Procrustes: rmse 0.005021948 max resid 0.02553619

```
## Run 7 stress 0.1742519
## ... Procrustes: rmse 0.005282821 max resid 0.0253712
## Run 8 stress 0.1742511
## ... Procrustes: rmse 0.004875527 max resid 0.02536426
## Run 9 stress 0.1740624
## ... New best solution
## ... Procrustes: rmse 0.00122971 max resid 0.007034305
## ... Similar to previous best
## Run 10 stress 0.1740766
## ... Procrustes: rmse 0.00165056 max resid 0.009695893
## ... Similar to previous best
## Run 11 stress 0.1742519
## ... Procrustes: rmse 0.005162522 max resid 0.02543396
## Run 12 stress 0.1742519
## ... Procrustes: rmse 0.005171929 max resid 0.02543997
## Run 13 stress 0.1740895
## ... Procrustes: rmse 0.002294135 max resid 0.009898852
## ... Similar to previous best
## Run 14 stress 0.1742868
## ... Procrustes: rmse 0.005907962 max resid 0.03022985
## Run 15 stress 0.174237
## ... Procrustes: rmse 0.004912636 max resid 0.02564774
## Run 16 stress 0.174252
## ... Procrustes: rmse 0.005199122 max resid 0.0254382
## Run 17 stress 0.1740766
## ... Procrustes: rmse 0.001652255 max resid 0.009705327
## ... Similar to previous best
## Run 18 stress 0.174237
## ... Procrustes: rmse 0.004907842 max resid 0.02564286
## Run 19 stress 0.1742425
## ... Procrustes: rmse 0.004871095 max resid 0.02530995
## Run 20 stress 0.1740766
## ... Procrustes: rmse 0.001651455 max resid 0.0097003
## ... Similar to previous best
## *** Best solution repeated 5 times
## 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## Run 0 stress 0.09896962
## Run 1 stress 0.09896971
## ... Procrustes: rmse 9.016584e-05 max resid 0.0004376369
## ... Similar to previous best
## Run 2 stress 0.09896972
## ... Procrustes: rmse 8.577676e-05 max resid 0.0004127917
## ... Similar to previous best
## Run 3 stress 0.09896977
## ... Procrustes: rmse 0.0001223393 max resid 0.0005867479
## ... Similar to previous best
## Run 4 stress 0.09896965
## ... Procrustes: rmse 5.866884e-05 max resid 0.0002897183
## ... Similar to previous best
## Run 5 stress 0.09896962
## ... New best solution
## ... Procrustes: rmse 3.259576e-05 max resid 0.0001404313
```

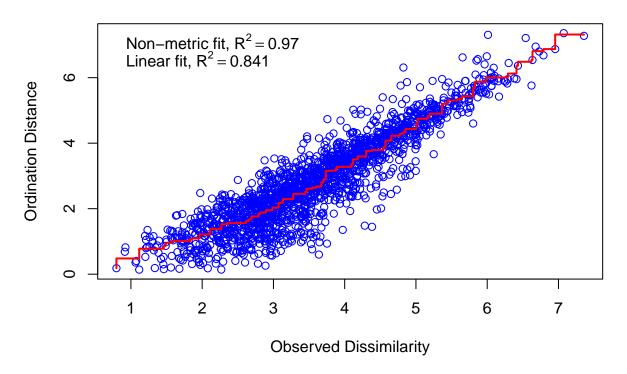
```
## ... Similar to previous best
## Run 6 stress 0.09896965
## ... Procrustes: rmse 8.313361e-05 max resid 0.0003665721
## ... Similar to previous best
## Run 7 stress 0.09896987
## ... Procrustes: rmse 0.000185915 max resid 0.0008367507
## ... Similar to previous best
## Run 8 stress 0.09896993
## ... Procrustes: rmse 0.0001501375 max resid 0.0008206557
## ... Similar to previous best
## Run 9 stress 0.09896962
## ... Procrustes: rmse 2.501235e-05 max resid 0.0001413601
## ... Similar to previous best
## Run 10 stress 0.09896968
## ... Procrustes: rmse 0.0001033617 max resid 0.0004440732
## ... Similar to previous best
## Run 11 stress 0.09896971
## ... Procrustes: rmse 9.430648e-05 max resid 0.0005388081
## ... Similar to previous best
## Run 12 stress 0.09896979
## ... Procrustes: rmse 0.0001556599 max resid 0.000696825
## ... Similar to previous best
## Run 13 stress 0.09896967
## ... Procrustes: rmse 7.68342e-05 max resid 0.0004590056
## ... Similar to previous best
## Run 14 stress 0.09896978
## ... Procrustes: rmse 0.0001239758 max resid 0.0006894128
## ... Similar to previous best
## Run 15 stress 0.09896974
## ... Procrustes: rmse 0.0001364889 max resid 0.0006982714
## ... Similar to previous best
## Run 16 stress 0.09896965
## ... Procrustes: rmse 5.763074e-05 max resid 0.0003457259
## ... Similar to previous best
## Run 17 stress 0.09896967
## ... Procrustes: rmse 7.211913e-05 max resid 0.0004325678
## ... Similar to previous best
## Run 18 stress 0.09896978
## ... Procrustes: rmse 0.0001229242 max resid 0.0007154321
## ... Similar to previous best
## Run 19 stress 0.09896974
## ... Procrustes: rmse 0.0001018879 max resid 0.0005856811
## ... Similar to previous best
## Run 20 stress 0.09896977
## ... Procrustes: rmse 0.0001102717 max resid 0.0006202152
## ... Similar to previous best
## *** Best solution repeated 16 times
## Attaching package: 'MASS'
## The following object is masked from 'package:dplyr':
##
##
      select
```

```
\#\# 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## Warning in metaMDS(x, distance = distance, k = i, trymax = trymax,
## autotransform = autotransform, : stress is (nearly) zero: you may have
## insufficient data
\mbox{\tt \#\#} 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## Warning in metaMDS(x, distance = distance, k = i, trymax = trymax,
## autotransform = autotransform, : stress is (nearly) zero: you may have
## insufficient data
## 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## Warning in metaMDS(x, distance = distance, k = i, trymax = trymax,
## autotransform = autotransform, : stress is (nearly) zero: you may have
## insufficient data
## 'comm' has negative data: 'autotransform', 'noshare' and 'wascores' set to FALSE
## Warning in metaMDS(x, distance = distance, k = i, trymax = trymax,
## autotransform = autotransform, : stress is (nearly) zero: you may have
## insufficient data
```

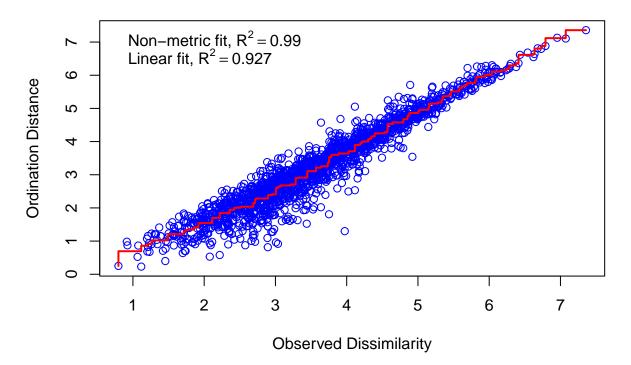
Scree Plot of Stress vs. Dimension



Shepard Plot

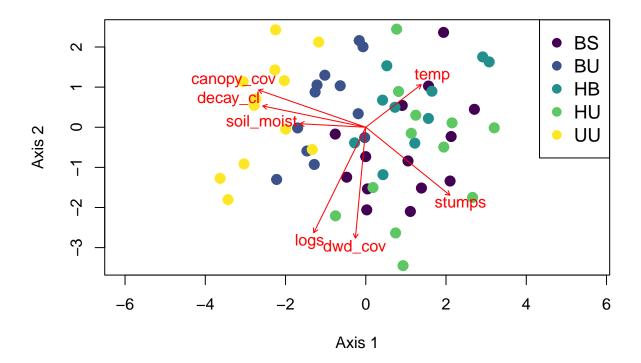


Shepard Plot



```
##
   ***VECTORS
##
##
##
                  Dim1
## temp
               0.73756  0.67528  0.3514  0.001 ***
## dwd cov
              -0.06858 -0.99765 0.7659
                                        0.001 ***
## soil_moist -0.99983 -0.01842 0.3091
                                       0.001 ***
## stumps
               0.80849 -0.58851 0.7629
                                       0.001 ***
## logs
              -0.43885 -0.89856 0.8121
                                        0.001 ***
             -0.98900 0.14792 0.6418
## decay_cl
                                        0.001 ***
## canopy_cov -0.95012  0.31189  0.7685  0.001 ***
##
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
## Permutation: free
## Number of permutations: 999
## species scores not available
```

NMDS2



1a. Were you able to successfuly apply each of these analyses to your dataset? If not, explain why.

Most of these worked with my environmental data, but I dont think I can do species ordinations. I have many zeros, and only two species.

1b. Which transformation or standardization procedures, if any, did you use prior to each analysis?

PCA: z-scored environmental data, hellinger salamander data

CA: raw environmental data

PCoA: euclidean distance matrix from z-scored subset of env data

NMDS: z-scored environmental data

1c. Did you remove any outliers prior to running the ordination procedure? Are there any apparent

outliers in the biplot(s) that appear to warrant further examination?

No, I dont think so.

1d. Which ordination procedure is the most appropriate for your dataset? Defend your answer based on what you know about the strengths and limitations of each procedure.

I think the PCoA is the best option. None of them are showing very clear clusters, but this one does show a grouping of the control sites aside from the rest.

Since euclidean distance is appropriate for my data, I wouldve expected PCA to be more helpful. I also expected the PCoA using euclidean distance to be similar to the PCA, but it was not.

1e. How does the choice of ordination method influence the interpretation of your data?

Different ordination methods can give varying interpretations of your data and need to match your data structure. They can focus on different aspects of the data- for example, CA is more sensitive to rare species. Choosing one that shows absolute (PCA, CA) or relative (NMDS) distances impacts your interpretation as well.

Question 2) For the PCA, PCoA, and/or CA, interpret the meaning of the principal components in the context of your dataset. What do the first few principal components explain about the variation in your data? How do these components relate to the original variables? Are there any variables that are strongly correlated with the principal components?

My PCoA plot of environmental data shows a high loading of canopy cover on axis 1, which may explain the grouping of UU (control) plots on the left side of the plot because they are all unlogged. Im having a harder time understanding the results because apparently every variable is showing as having a significant relationship with each vector. Downed wood is strongly correlated with axis 2, but it doesnt look that way on the plot, so that's odd. Still some workshopping to do.

Question 3) Do a little digging and find at least one creative way of presenting your ordination output that we did not discuss in lab or in class. How does this approach allow you to interpret your data more effectively?

Struggling to make a plot that shows a gradient of downed wood density, so that's in the works. Here's one that shows salamander count by point size. It is interesting to see that the sites in the top right have lower counts than on the left side of the plot.

```
oss_scale <- 1 + log(dat2$oss + 1)
plot(site.sc[, 1:2], # First two dimensions
    main = "Env PCoA, points sized to OSS count",
    xlab = "PCoA 1",
    ylab = "PCoA 2",
    pch = 19)
for (i in 1:length(groups))
{
    dim_choice <- site.sc[dat2$trt==groups[i],]
}
text(spe.sc*1.5, row.names(spe.sc))
points(site.sc,
    pch=20,
    cex=oss_scale,
    col="purple")
arrows(0, 0, spe.sc[,1]*1.4, spe.sc[,2]*1.4,</pre>
```

Env PCoA, points sized to OSS count

