Sequence alignment and search A review on alignment and search tools

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How LARGE the data is

Bit

The bit is a basic unit of information in computing and digital communications. The bit represents a logical state with one of two possible values, 0 or 1.

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Magnitude

B -> KiB -> MiB -> GiB -> TiB -> PiB -> EiB ...

(Each -> represents a magnitude of 1024.)

Big O notation

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Formal definition

Let f be a real or complex valued function and g a real valued function. Let both functions be defined on some unbounded subset of the positive real numbers, and g(x) be strictly positive for all large enough values of x.

$$f(x) = O(g(x))(x \to \infty)$$

Application of Big O notation

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Example

Informally, especially in computer science, the big O notation often can be used to describe an asymptotic tight bound. For example, when considering a function $T(n)=73n^3+22n^2+58$, we can say:

$$T(n) = O(n^3)$$

Where n often represents the data amount of an input, and T(n) is the time for execution of an algorithm.

Comparing algorithms

Big O notation can compare the absolute speed of an algorithm as well as the relative speed.

- O(nlog(n))
- $O(n^k), (k > 1)$
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Example

Bubble sort has a speed of $O(n^2)$, while Quick sort has a speed of $O(n\log(n))$.

Therefore, on the same machine (with the same amount of input n), Quick sort is faster than Bubble sort. If we run Quick sort on a slow machine and Bubble sort on a fast machine, when n gets larger, Quick sort will eventually run faster than Bubble sort.

Traditional algorithms and Heuristic algorithms

For Traditional algorithms, their output is definite and accurate. While for Heuristic algorithms, especially when ramdomness is involved, their output is not stable.

Traditional algorithms and Heuristic algorithms

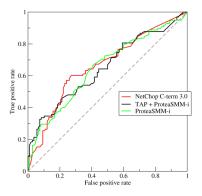
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Terminology

- TN, true negative
- FP, false positive
- FN, false negative
- TPR, true positive rate, TPR = TP/P = TP/(TP + FN)
- FPR, false positive rate, FPR = FP/N = FP/(FP + TN)
- ACC, accuracy, ACC = (TP + TN)/(P + N)

ROC curve

The ROC curve is created by plotting the true positive rate (TPR) against the false positive rate (FPR) at various threshold settings.



AUC

AUC is the area under the ROC curve.

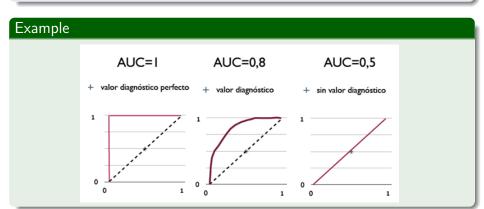


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Pairwise alignment

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Usage of Pairwise alignment

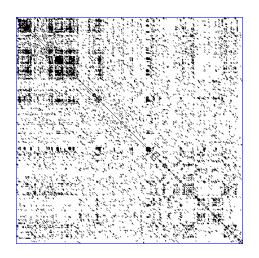
Pairwise alignments can only be used between two sequences at a time, but they are efficient to calculate and are often used for methods that do not require extreme precision (such as searching a database for sequences with high similarity to a query).

Algorithms for Pairwise alignment

- Dot-matrix methods
- Dynamic programming
- Word methods

Dot-matrix methods

The dot-matrix approach, which implicitly produces a family of alignments for individual sequence regions, is qualitative and conceptually simple, though time-consuming to analyze on a large scale. In the absence of noise, it can be easy to visually identify certain sequence features—such as insertions, deletions, repeats, or inverted repeats—from a dot-matrix plot. To construct a dot-matrix plot, the two sequences are written along the top row and leftmost column of a two-dimensional matrix and a dot is placed at any point where the characters in the appropriate columns match—this is a typical recurrence plot.



Dot-matrix tool

• Dotter: A dot-matrix program with interactive greyscale rendering for genomic DNA and Protein sequence analysis

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Dot-matrix method advantage

- Fairly easy to Implement.
- Easy to understand visually.
- It shows all possible alignment of pairs.
- Good overview of places fot good alignment.

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Dot-matrix method speed

For two sequences that have the length of n:

$$O(n^2)$$

Dynamic programming

The technique of dynamic programming can be applied to produce global alignments via the Needleman-Wunsch algorithm, and local alignments via the Smith-Waterman algorithm. In typical usage, protein alignments use a substitution matrix to assign scores to amino-acid matches or mismatches, and a gap penalty for matching an amino acid in one sequence to a gap in the other. DNA and RNA alignments may use a scoring matrix, but in practice often simply assign a positive match score, a negative mismatch score, and a negative gap penalty.

Input and output

The input of the algorithm is two sequences, and the output is a score representing how similar the sequences are. On most cases, a pattern showing the matches will be given.

For example:

TACGGGCCCGCTA-C

||---|-||-||-|

TA---G-CC-CTATC

Dynamic programming tool

- EMBOSS Water
- EMBOSS Needle
- http://emboss.sourceforge.net/

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Dynamic programming method speed

For two sequences that have the length of n:

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Pairwise alignment: Word methods

Word methods

Word methods, also known as k-tuple methods, are heuristic methods that are not guaranteed to find an optimal alignment solution, but are significantly more efficient than dynamic programming. These methods are especially useful in large-scale database searches where it is understood that a large proportion of the candidate sequences will have essentially no significant match with the query sequence.

Pairwise alignment: Word methods

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Word methods speed

For two sequences that have the length of n:

$$O(n^k), 1 < k < 2$$

Pairwise alignment: Word methods

Word methods tool

- EMBL FASTA http://www.ebi.ac.uk/fasta33/
- NCBI BLAST https://blast.ncbi.nlm.nih.gov/Blast.cgi

Multiple sequence alignment

Modified pairwise alignment algorithms

Most Pairwise alignment algorithms can be easily modified for Multiple sequence alignment. However, such algorithms are too slow to handle massive amount of data.

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Specialized agorithms and tools

Some algorithms are designed for fast multiple sequence alignment, but most of them are not sensitive compared with Pairwise alignment algorithms.

Below are some tools.

- Clustal
- T-Coffee
- LINCLUST
- MMseqs2
- UCLUST

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MMseqs2 Publications

- Steinegger M and Soeding J. MMseqs2 enables sensitive protein sequence searching for the analysis of massive data sets. Nature Biotechnology, doi: 10.1038/nbt.3988 (2017).
- Steinegger M and Soeding J. Clustering huge protein sequence sets in linear time. Nature Communications, doi: 10.1038/s41467-018-04964-5 (2018).
- Mirdita M, Steinegger M and Soeding J. MMseqs2 desktop and local web server app for fast, interactive sequence searches. Bioinformatics, doi: 10.1093/bioinformatics/bty1057 (2019).
- Mirdita M, Steinegger M, Breitwieser F, Soding J, Levy Karin E: Fast and sensitive taxonomic assignment to metagenomic contigs. bioRxiv, doi: 10.1101/2020.11.27.401018 (2020).

Install MMseqs2

Before installation, read the documents of the source code carefully, and make sure you understand all the instructions in the doc.

Install gcc, cmake and libz

To compile MMseqs2 git, g++ (4.9 or higher) and

cmake (2.8.12 or higher) are needed. Zlib and Bzlib are optional.

apt-get install build-essential cmake zlib1g zlib1g-devel libbz2-dev

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Download source code

git clone https://github.com/soedinglab/MMseqs2.git && cd MMseqs2

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```

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Download source code

git clone https://github.com/soedinglab/MMseqs2.git && cd MMseqs2

Compile and Install

```
mkdir build && cd build/
cmake -DCMAKE_BUILD_TYPE=RELEASE -DCMAKE_INSTALL_PREFIX=/usr ...
```

make -j && make install

Use MMseqs2: Search

Create a database

You can create a database from a FASTA file.

mmseqs createdb examples/DB.fasta targetDB

mmseqs createindex targetDB tmp

Or you can download a database online.

mmseqs databases UniProtKB/Swiss-Prot swissprot tmp

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Search

You can search some sequences in a fasta file.

mmseqs easy-search examples/QUERY.fasta examples/DB.fasta alnRes.m8 tmp
Or search some sequences in a pre-built database, which is much faster.

mmseqs easy-search examples/QUERY.fasta targetDB alnRes.m8 tmp

Use MMseqs2: Search

Search Results

The results of a search is in m8 format, which is a tab-separated ascii file with lines like this:

A0A061HXN7 P13020 0.953 780 37 0 1 778 1 780 0.000E+00 1521 Each column has the definition listed below:

- Query id
- Subject id
- Identity
- Alignment length
- Mismatches
- Gap Openings
- q. start
- a. end
- s start
- s. end
- e-value
- bit score

Clustering FASTA files

For clustering, MMseqs2 easy-cluster and easy-linclust are available.

mmseqs easy-cluster examples/DB.fasta clusterRes tmp

mmseqs easy-linclust examples/DB.fasta clusterRes tmp

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mmseqs easy-linclust examples/DB.fasta clusterRes tmp

Cluster results

The commands above produces three files: clusterRes_cluster.tsv, clusterRes_all_seqs.fasta and clusterRes_rep_seq.fasta.

Cluster results

The clusterRes_cluster.tsv file follows the following format:

```
#cluster-representative cluster-member
QOKJ32 QOKJ32
QOKJ32 COW539
QOKJ32 D6KVP9
...
```

Cluster results

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```
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QOKJ32 QOKJ32
QOKJ32 COW539
QOKJ32 D6KVP9
...
```

Cluster results

The clusterRes_all_seqs.fasta is FASTA-like format file, with a new cluster being marked by two identical name lines of the representative sequence.

The clusterRes_rep_seq.fasta contains all representative sequences.

Computing environment

- Hardware: Xeon Gold 5117 @2.00GHz 28C56T, 32GB RAM, SSD
- Software: Ubuntu 18.04, MMseqs2 Release 13 compiled with gcc 7.5.0

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- Software: Ubuntu 18.04, MMseqs2 Release 13 compiled with gcc 7.5.0

Data size

The input file is swiss.fasta, which contains 1129276 seqs from UniProtKB. The size of the file is 264MB

Performance

Using command

time mmseqs easy-cluster examples/swiss.fasta clusterRes tmp to do the clustering, it took: 1m21.693s.

Performance

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time mmseqs easy-cluster examples/swiss.fasta clusterRes tmp to do the clustering, it took: 1m21.693s.

Performance

Using command

time mmseqs easy-linclust examples/swiss.fasta clusterRes tmp to do the clustering, it took: Om16.869s.

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Algorithm?

- Parallel computing?
- Machine learning based methods?
- Quantum computing?

Hardware?

- GPU?
- CPU Clusters?
- FPGA?
- Playstation?
- Wirawan A, Kwoh C K, Hieu N T, et al. CBESW: sequence alignment on the playstation 3

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Q & A

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