Lab #8

By the beginning of the next lab (March 26th), send what you have to [afodor@uncc.edu](mailto:afodor@uncc.edu) with “Lab #8” in the subject line. As usual, show all of your code.

1. We return to our RNA seq dataset of E. Coli genes from mice. The URL is here:

http://afodor.github.io/classes/stats2015/longitdunalRNASeqData.zip

Read and normalize the counts table ( “nc101\_scaff\_dataCounts.txt “ into R). For example:

setwd("C:\\somewhere")

myT<-read.table("nc101\_scaff\_dataCounts.txt",sep="\t",header=TRUE,row.names=1)

# remove rare genes

myT <- myT[ apply( myT,1, median)> 5,]

myTNorm <- myT

for ( i in 1:ncol(myT))

{

colSum = sum(myT[,i])

myTNorm[,i] =myTNorm[,i]/colSum

}

(The first 3 columns are “day 2”, the next 3 columns are “week 12” and the last 5 are “week 18”). Remember, that day 2 is before the mice have inflammation symptoms, week 12 is associated with inflammation and week 18 is associated with cancer.

1. For every row in the normalized spreadsheet, run three t-tests ( “day 2” vs. “week 12”, “day 2” vs. “week 18” and “week 12” vs. “week 18”. At a p < .05 threshold fill in the following table:

|  |  |  |  |
| --- | --- | --- | --- |
|  | # of genes significant at p <0.05 uncorrected | # genes significant at p <0.05 BH FDR corrected | # genes significant at p <0.05 Bonferroni corrected |
| “day 2” vs. “week 12” |  |  |  |
| “day 2” vs. “week 18” |  |  |  |
| “week 12” vs. “week 18” |  |  |  |

1. Make histograms of all the uncorrected p-values for each of the three companions. Are any of the distributions uniform?
2. Based on these data, when is the biggest shift in the transcriptome? Which samples are most different from one another?