

# 07-thresholding

January 13, 2020

## 1 Thresholding

In this episode, we will learn how to use skimage functions to apply thresholding to an image. Thresholding is a type of *image segmentation*, where we change the pixels of an image to make the image easier to analyze. In thresholding, we convert an image from color or grayscale into a *binary image*, i.e., one that is simply black and white. Most frequently, we use thresholding as a way to select regions of interest of an image, while ignoring the parts we are not concerned with. It is therefore one way to automatically create masks.

We have already done some simple thresholding, in the "Manipulating pixels" section of the 3rd episode. In that case, we used a simple NumPy array manipulation to separate the pixels belonging to the root system of a plant from the black background. In this episode, we will learn how to use skimage functions to perform thresholding. Then, we will use the masks returned by these functions to select the parts of an image we are interested in.

### 1.1 Simple thresholding

Consider this image, with a series of crudely cut shapes set against a white background. The black outline around the image is not part of the image.

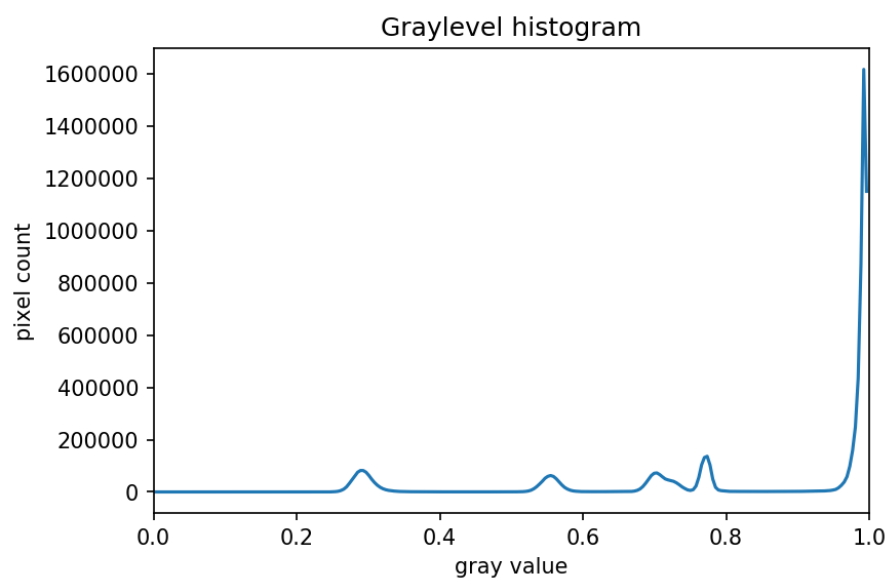
Now suppose we want to select only the shapes from the image. In other words, we want to leave the pixels belonging to the shapes "on," while turning the rest of the pixels "off," by setting their color channel values to zeros. The skimage library has several different methods of thresholding. We will start with the simplest version, which involves an important step of human input. Specifically, in this simple, *fixed-level thresholding*, we have to provide a threshold value,  $\tau$ .

Then, we will use the  $>$  operator to apply the threshold  $\tau$ , a number in the closed range  $[0.0, 1.0]$ . Pixels with color values on one side of  $\tau$  will be turned "on," while pixels with color values on the other side will be turned "off." In order to use this function, we have to determine a good value for  $\tau$ . How might we do that? Well, one way is to look at a grayscale histogram of the image. Here is the histogram produced by the code from the *Creating Histograms* episode, if we run it on the colored shapes image shown above.

Since the image has a white background, most of the pixels in the image are white. This corresponds nicely to what we see in the histogram: there is a spike near the value of 1.0. If we want to select the shapes and not the background, we want to turn off the white background pixels, while leaving the pixels for the shapes turned on. So, we should choose a value of  $\tau$  somewhere before the large peak and turn pixels above that value "off".



Original shapes image



Grayscale histogram

### 1.1.1 Exercise: Setting a fixed-level threshold

Now, it is your turn to apply a simple thresholding.

- By inspecting the histogram above, chose a threshold value  $t$  which separates the white background from the objects
- Create a mask by using the  $<$  operator and the chosen threshold value  $t$
- Plot the mask using matplotlib's `imshow`
- Apply the mask to the image
- Use the histogram function provided in the code cell below to show how the pixel distribution has changed after thresholding

Here are the lines of a Python program to apply simple thresholding to the image. For convenience a function plotting a grayscale histogram is also given. As usual, we first have to import the modules needed and open the image:

```
In [ ]: # %load ../exercises/07-simple-thresholding.py
import numpy as np
import skimage
import matplotlib.pyplot as plt
# 'magic' to display plots in the jupyter notebook
%matplotlib inline

# we want to look at grayscale histograms
def plot_histogram(image):

    histogram, bin_edges = np.histogram(image, bins = 256, range = (0,1))
    plt.plot(bin_edges[:-1], histogram, color = 'k')

    plt.xlabel('Pixel value')
    plt.ylabel('Counts')
    plt.title('Grayscale Histogram')

    image = skimage.io.imread('../data/07-shapes.tif', as_gray = True)

    plt.figure()
    plt.imshow(image, cmap = 'gray')

    # YOUR CODE HERE

In [ ]: # 0.9 should be fine
t = 0.9
mask = image < t
plt.imshow(mask, cmap = 'gray')

In [ ]: # create masked image
sel = np.zeros_like(image)
sel[mask] = image[mask]
plt.imshow(sel, cmap = 'gray')

In [ ]: plot_histogram(image[mask])
```

### 1.1.2 Exercise: Blurring + Thresholding

In many real life applications, applying a threshold directly to an acquired image may not be optimal. Consider the following example where noise was added to our collection of shapes:

```
In [ ]: %load ../exercises/07-blurring-simple-thresh.py
```

As you can see, the *segmentation* into foreground and background is not so good. However, we can use blurring to suppress most of the noise!

- Again plot a histogram, this time of the noisy image already opened above. Note how the pixel distribution makes direct thresholding difficult.
- From what you learned in the *Gaussian Blur* episode blur the image before thresholding. Choose a sigma which suppresses the noise but keeps the shapes mostly intact.
- Look at the histogram of the blurred image and compare it to the pixel distribution of the raw image. Now choose a threshold  $t$ .
- Repeat the steps of the preceding exercise to create a mask and hence segment the blurred image

*Hint: The gaussian filter is available as `skimage.filters.gaussian`*

The final result should be much better segmented compared to thresholding without blurring first.

```
In [ ]: # histogram of the noisy image
        plot_histogram(image)
```

```
In [ ]: # apply some gaussian blur
        import skimage.filters

        blurred = skimage.filters.gaussian(image, 3)
        plt.imshow(blurred, cmap = 'gray')
```

```
In [ ]: plot_histogram(blurred)
```

```
In [ ]: # create the mask
        t = 0.8
        mask = blurred < t
        plt.imshow(mask, cmap = 'gray')
```

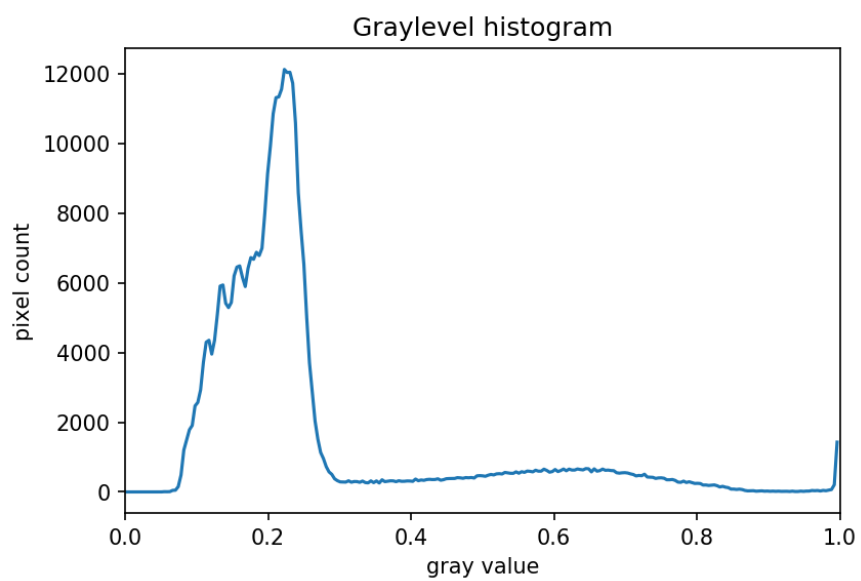
```
In [ ]: # create masked image
        sel = np.zeros_like(blurred)
        sel[mask] = blurred[mask]
        plt.imshow(sel, cmap = 'gray')
```

### 1.1.3 Adaptive thresholding

There are also `skimage` methods to perform *adaptive thresholding*. The chief advantage of adaptive thresholding is that the value of the threshold,  $t$ , is determined automatically for us. One such method, *Otsu's method*, is particularly useful for situations where the grayscale histogram of an



Maize root system



Maize root histogram

image has two peaks. Consider this maize root system image, which we have seen before in the *Skimage Images* episode.

Now, look at the grayscale histogram of this image, as produced by our grayscale histogram code from the *Creating Histograms* episode.

The histogram has a significant peak around 0.2, and a second, albeit smaller peak very near 1.0. Thus, this image is a good candidate for thresholding with Otsu's method. The mathematical details of how this work are complicated (see the [skimage documentation](#) if you are interested), but the outcome is that Otsu's method finds a threshold value between the two peaks of a grayscale histogram.

The `skimage.filters.threshold_otsu()` function can be used to determine the adaptive threshold via Otsu's method. Then numpy comparison operators can be used to apply it as before.

```
In [ ]: import skimage
import numpy as np
image = skimage.io.imread('../data/07-roots-original.tif', as_gray = True)

In [ ]: # blurring
blurred = skimage.filters.gaussian(image, 1)

# Otsu threshold
t = skimage.filters.threshold_otsu(image)
print(f'Otsu threshold: {t:.2f}')

# create the mask
mask = blurred > t
plt.imshow(mask)

In [ ]: # create masked image
sel = np.zeros_like(blurred)
sel[mask] = image[mask]
plt.imshow(sel)
```

#### 1.1.4 Exercise: Thresholding a bacteria colony image (10 min)

In the `/data` directory, you will find an image named `colonies01.tif`; this is one of the images you will be working with in the morphometric challenge at the end of the workshop. First, create a grayscale histogram of the image, and determine a threshold value for the image. Then, write a Python program to threshold a grayscale version of the image, leaving the pixels in the bacteria colonies "on," while turning the rest of the pixels in the image "off."

*Hint: Try both manual and automatic thresholding.*

```
In [ ]: import skimage
import skimage.io
import skimage.filters
import numpy as np
import matplotlib.pyplot as plt
# 'magic' to display plots in the jupyter notebook
%matplotlib inline
```

```

In [ ]: image = skimage.io.imread('../data/colonies01.tif', as_gray=True)
        image = skimage.filters.gaussian(image, 0.5)
        plt.figure()
        plot_histogram(image)
        plt.figure()
        plt.imshow(image, cmap = 'gray')

In [ ]: # the colonies are at the far left end of the histogram
        mask = image < 0.1
        colonies = np.zeros_like(image)
        colonies[mask] = image[mask]
        plot_histogram(image[mask])

In [ ]: plt.imshow(mask, cmap = 'gray')

```

### 1.1.5 Bonus Exercise: Quantification of Combined Colony Sizes

We want to measure the fraction of the dish which is covered by bacteria colonies.

- Write a Python function which takes an image of a colony as input, and returns the ratio of dish area vs. overgrown area.
- Compute the area ratio for all three colonies.tif in the /data directory.
- Make a bar plot displaying the measured values for all three dishes

*Hint: Here one binary segmentation into background and foreground is not enough: try segmenting the dish + colonies, and as before the colonies alone. You can use `np.sum(mask)` to compute the number of pixels which are 'on'. A simple bar plot can be produced with `plt.bar(...)`, use `plt.bar?` in a code cell to learn how to use it.*

```

In [ ]: def measure_colonies(image):

        image = skimage.filters.gaussian(image, 0.5)

        dish_mask = image < 0.8
        colonies_mask = image < 0.1

        dish_area = np.sum(dish_mask)
        c_area = np.sum(colonies_mask)
        fraction = c_area/dish_area
        print(f'Area fraction: {fraction}')

        return fraction

In [ ]: from glob import glob

        fnames = glob('../data/colo*')

        fractions = []
        for name in fnames:

```

```
image = skimage.io.imread(name, as_gray = True)
plt.figure()
plt.imshow(image)
plt.title(name)
frac = measure_colonies(image)
fractions.append(frac)

print(len(fractions), fnames)
plt.figure()
plt.bar(np.arange(1,4), fractions, tick_label = fnames)
plt.ylabel('area fraction')
```