

Class 9: Structural Bioinformatics 1.

Kyle Alvarez

The RCSB Protein Data Bank (PDB)

Protein structures by X-ray crystallography dominate this database. We are skipping Q1-3 as the website was too slow for us.

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

Hydrogen is too small to visualize, hence we have one atom per water molecule (shows the oxygen)

Q5: There is a critical "conserved" water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

We are able to identify this water molecule by turning out space fill for the ligand and the water molecule. We can then see where there is a single water molecule in the binding site and it's residue number which is HOH 308.

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain (we recommend "Ball & Stick" for these side-chains). Add this figure to your Quarto document. Discussion Topic: Can you think of a way in which indinavir, or even larger ligands and substrates, could enter the binding site?

In order to enter the binding site, the protein can undergo conformational change (through multiple causes such as another enzyme, pH change, etc) and allow larger ligands and substates to enter the binding site.

3. Introduction to Bio3D in R

Bio3D is an R package for structural bioinformatics. To use it we need to call it up with the `library()` function (just like any package).

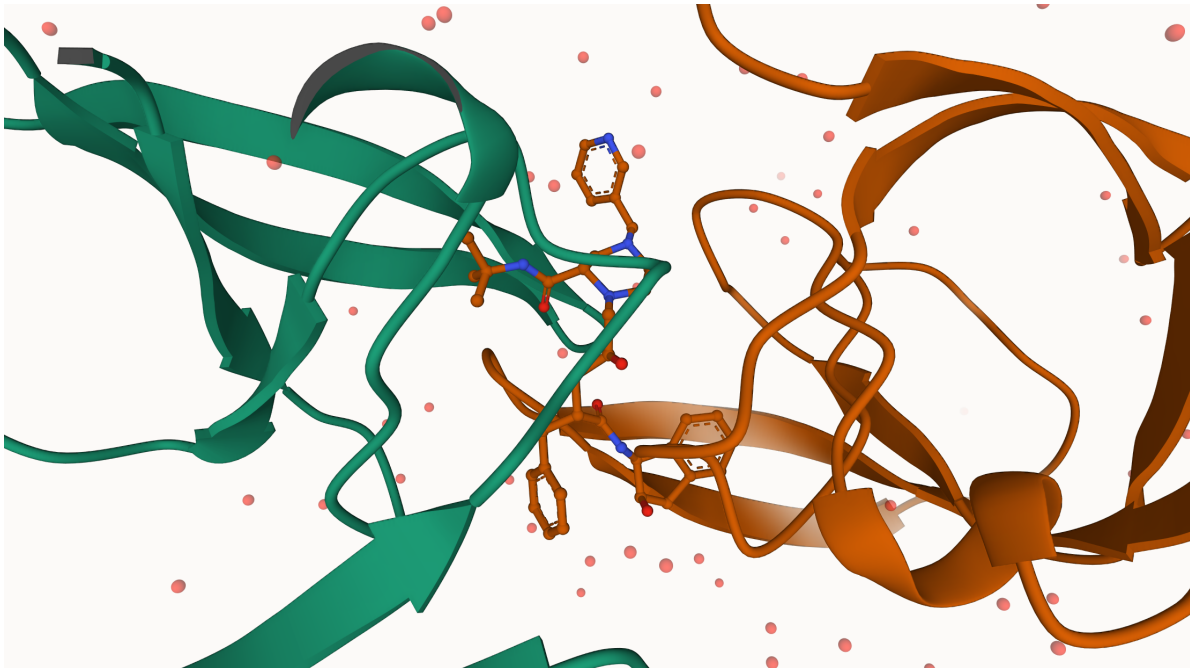


Figure 1: HIV-Pr Structure from 1HSG

```
library(bio3d)
```

To read a PDB file we can use `read.pdb()`

```
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
```

```
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

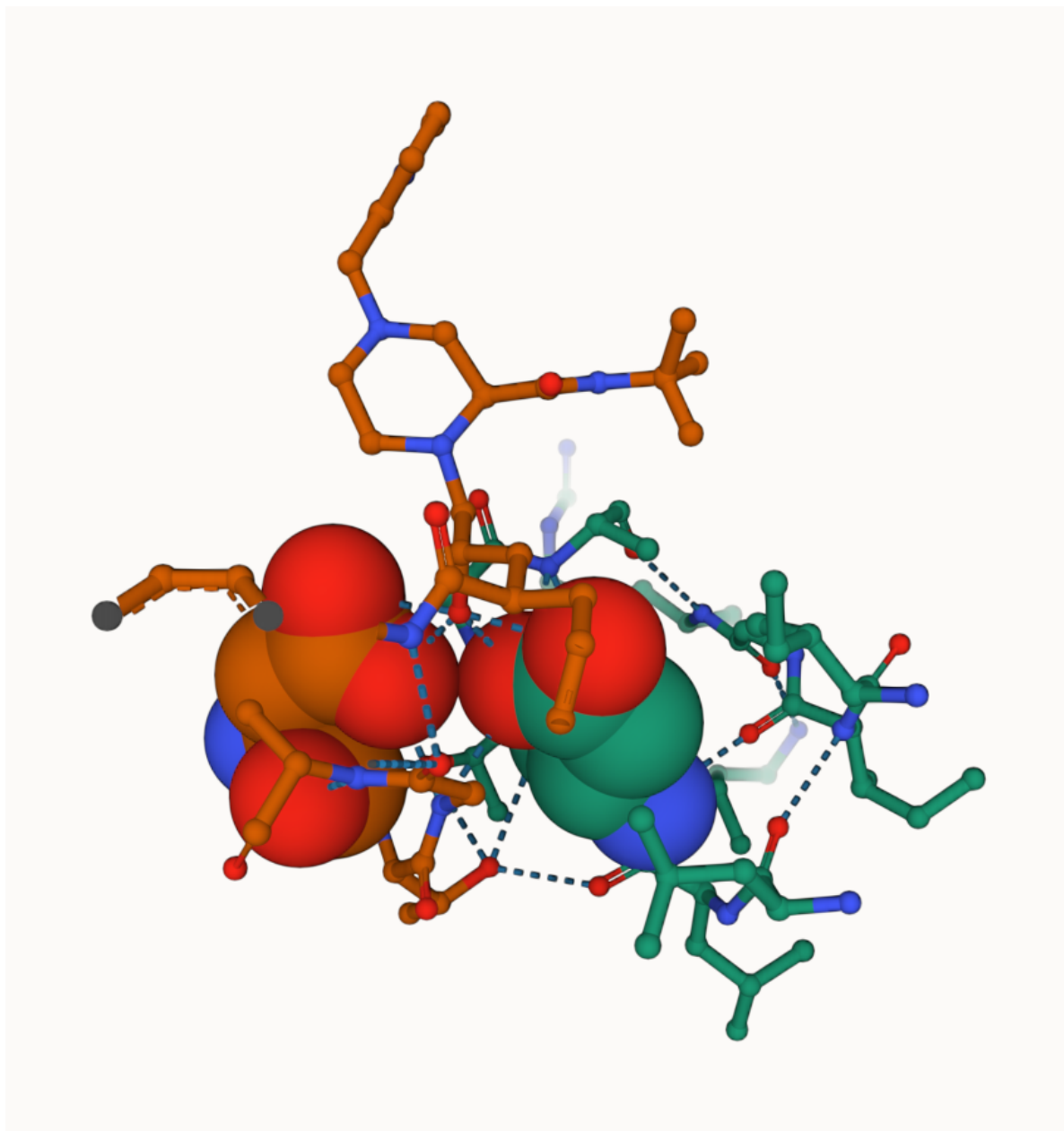


Figure 2: Spacefill of Protease ASP 25

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

Protein sequence:

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

The ATOM records of a PDB file are stored in `pdb$atom`

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elasy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

Comparative analysis of Aneylate kinase (ADK)

We will start our analysis with a single PDB id (code form the PDB database): 1AKE

First we get it's priamry sequence:

```
aa <- get.seq("1ake_a")
```

Warning in `get.seq("1ake_a")`: Removing existing file: seqs.fasta

Fetching... Please wait. Done.

aa

```

      1      .      .      .      .      .      .      60
pdb|1AKE|A  MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLV
      1      .      .      .      .      .      .      60

      61      .      .      .      .      .      .      120
pdb|1AKE|A  DELVIALVKERIAQEDCRNGFLLDGFPRITIPQADAMKEAGINVDYVLEFDVPDELIVDRI
      61      .      .      .      .      .      .      120

     121      .      .      .      .      .      .      180
pdb|1AKE|A  VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
     121      .      .      .      .      .      .      180

     181      .      .      .      214
pdb|1AKE|A  YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
     181      .      .      .      214
```

Call:

```
read.fasta(file = outfile)
```

Class:

```
fasta
```

Alignment dimensions:

```
1 sequence rows; 214 position columns (214 non-gap, 0 gap)
```

+ attr: id, ali, call

```
# Blast or hmmer search
#b <- blast.pdb(aa)
```

```
# Plot a summary of search results
#hits <- plot(b)
# List out some 'top hits'
#head(hits$ pdb.id)
```

Use these ADK structures for analysis:

```
hits <- NULL
hits$ pdb.id <- c('1AKE_A', '6S36_A', '6RZE_A', '3HPR_A', '1E4V_A', '5EJE_A', '1E4Y_A', '3X2S_A', '1E4Z_A')
```

Download all these PDB files from the online database...

```
# Download related PDB files
files <- get.pdb(hits$ pdb.id, path="pdbc", split=TRUE, gzip=TRUE)
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbc", split = TRUE, gzip = TRUE): pdbc/
1AKE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbc", split = TRUE, gzip = TRUE): pdbc/
6S36.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbc", split = TRUE, gzip = TRUE): pdbc/
6RZE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbc", split = TRUE, gzip = TRUE): pdbc/
3HPR.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbc", split = TRUE, gzip = TRUE): pdbc/
1E4V.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbc", split = TRUE, gzip = TRUE): pdbc/
5EJE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbc", split = TRUE, gzip = TRUE): pdbc/
1E4Y.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbc", split = TRUE, gzip = TRUE): pdbc/
3X2S.pdb.gz exists. Skipping download
```

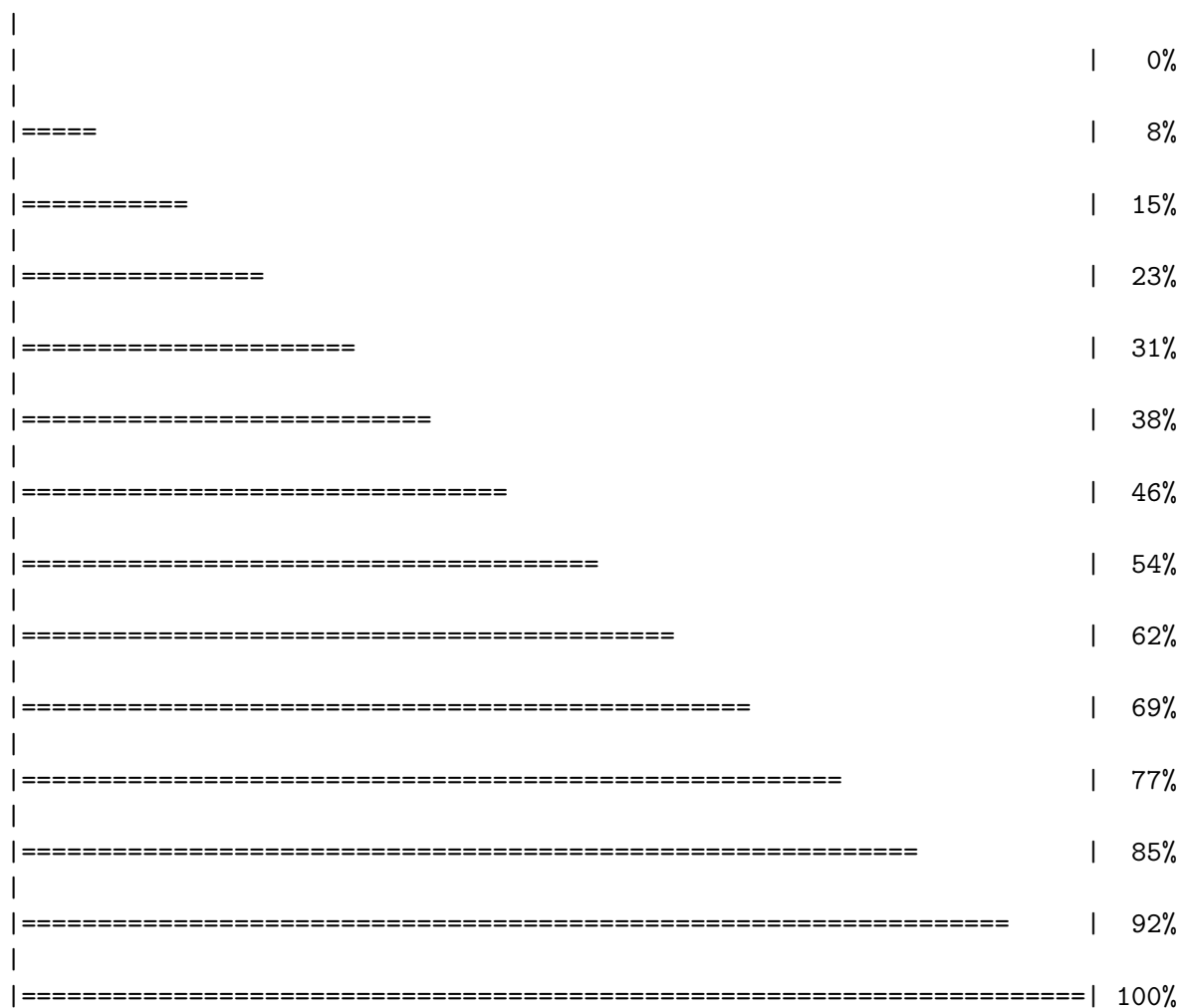
```
Warning in get.pdb(hits$ pdb.id, path = "pdbc", split = TRUE, gzip = TRUE): pdbc/
6HAP.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$ pdb.id, path = "pdbc", split = TRUE, gzip = TRUE): pdbc/
6HAM.pdb.gz exists. Skipping download
```

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
4K46.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
3GMT.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
4PZL.pdb.gz exists. Skipping download



Align all these structures

```
# Align related PDBs
pdbs <- pdbaln(files, fit = TRUE, exefile="msa")
```

Reading PDB files:

```
pdbs/split_chain/1AKE_A.pdb
pdbs/split_chain/6S36_A.pdb
pdbs/split_chain/6RZE_A.pdb
pdbs/split_chain/3HPR_A.pdb
pdbs/split_chain/1E4V_A.pdb
pdbs/split_chain/5EJE_A.pdb
pdbs/split_chain/1E4Y_A.pdb
pdbs/split_chain/3X2S_A.pdb
pdbs/split_chain/6HAP_A.pdb
pdbs/split_chain/6HAM_A.pdb
pdbs/split_chain/4K46_A.pdb
pdbs/split_chain/3GMT_A.pdb
pdbs/split_chain/4PZL_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
.... PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
...
```

Extracting sequences

```
pdb/seq: 1  name: pdbs/split_chain/1AKE_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 2  name: pdbs/split_chain/6S36_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 3  name: pdbs/split_chain/6RZE_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 4  name: pdbs/split_chain/3HPR_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 5  name: pdbs/split_chain/1E4V_A.pdb
pdb/seq: 6  name: pdbs/split_chain/5EJE_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 7  name: pdbs/split_chain/1E4Y_A.pdb
pdb/seq: 8  name: pdbs/split_chain/3X2S_A.pdb
pdb/seq: 9  name: pdbs/split_chain/6HAP_A.pdb
```



```

pdb/seq: 10  name: pdbs/split_chain/6HAM_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 11  name: pdbs/split_chain/4K46_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 12  name: pdbs/split_chain/3GMT_A.pdb
pdb/seq: 13  name: pdbs/split_chain/4PZL_A.pdb

```

pdbs

```

[Truncated_Name:1] 1AKE_A.pdb      1      .      .      .      40
-----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:2] 6S36_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:3] 6RZE_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:4] 3HPR_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:5] 1E4V_A.pdb      -----MRIILLGAPVAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:6] 5EJE_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:7] 1E4Y_A.pdb      -----MRIILLGALVAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:8] 3X2S_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:9] 6HAP_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:10] 6HAM_A.pdb     -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:11] 4K46_A.pdb     -----MRIILLGAPGAGKGTQAQFIMAKFGIPQIS
[Truncated_Name:12] 3GMT_A.pdb     -----MRLILLGAPGAGKGTQANFIKEKFGIPQIS
[Truncated_Name:13] 4PZL_A.pdb     TENLYFQSNMRIILLGAPGAGKGTQAKIIEQKYNIAHIS
                                   **~*****  *****  *  *~ *  **
1      .      .      .      40

41      .      .      .      80
TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
TGDMLRAAVKSGSELGKQAKDIMDACKLVDELVIALVKE
TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVKE
TGDMLRAAVKSGSELGKQAKDIMDCGKLVDELVIALVKE
TGDMLRAAVKSGSELGKQAKDIMDAGKLVDELVIALVRE
TGDMLRAAIKSGSELGKQAKDIMDAGKLVDEIIIALVKE
TGDMLRAAIKAGTELGKQAKSVIDAGQLVSDDIILGLVKE
TGDMLRAAVKAGTPLGVEAKTYMDEGKLPVDSLIIGLVKE
TGDMIRETIKSGSALGQELKKVLDAGELVSDEFIIVKVD
****~*  ~* *~ **  *  ~*  ** *  ^^ ~* ^^

```

	41	.	.	.	80
	81	.	.	.	120
[Truncated_Name:1] 1AKE_A.pdb		RIAQEDCRNGFLLDGFPR	TIPQADAMKEAGINVDY	VLEFD	
[Truncated_Name:2] 6S36_A.pdb		RIAQEDCRNGFLLDGFPR	TIPQADAMKEAGINVDY	VLEFD	
[Truncated_Name:3] 6RZE_A.pdb		RIAQEDCRNGFLLDGFPR	TIPQADAMKEAGINVDY	VLEFD	
[Truncated_Name:4] 3HPR_A.pdb		RIAQEDCRNGFLLDGFPR	TIPQADAMKEAGINVDY	VLEFD	
[Truncated_Name:5] 1E4V_A.pdb		RIAQEDCRNGFLLDGFPR	TIPQADAMKEAGINVDY	VLEFD	
[Truncated_Name:6] 5EJE_A.pdb		RIAQEDCRNGFLLDGFPR	TIPQADAMKEAGINVDY	VLEFD	
[Truncated_Name:7] 1E4Y_A.pdb		RIAQEDCRNGFLLDGFPR	TIPQADAMKEAGINVDY	VLEFD	
[Truncated_Name:8] 3X2S_A.pdb		RIAQEDSRNGFLLDGFPR	TIPQADAMKEAGINVDY	VLEFD	
[Truncated_Name:9] 6HAP_A.pdb		RICQEDSRNGFLLDGFPR	TIPQADAMKEAGINVDY	VLEFD	
[Truncated_Name:10] 6HAM_A.pdb		RICQEDSRNGFLLDGFPR	TIPQADAMKEAGINVDY	VLEFD	
[Truncated_Name:11] 4K46_A.pdb		RIAQDDCAKGFLLDGFP	R TIPQADGLKEVGVVVDY	VIEFD	
[Truncated_Name:12] 3GMT_A.pdb		RLKEADCANGYLFDFGP	RTIAQADAMKEAGVAIDY	VLEID	
[Truncated_Name:13] 4PZL_A.pdb		RISKNCNNGFLLDGVPR	TIPQAQELDKLGVNIDY	IIVEVD	
		*^	* *^* ** ***** **	^ ^*~^* *	
	81	.	.	.	120
	121	.	.	.	160
[Truncated_Name:1] 1AKE_A.pdb		VPDELIVDRIVGRRVHAP	SGRVYHVKNPPKVEGKDD	VDTG	
[Truncated_Name:2] 6S36_A.pdb		VPDELIVDKIVGRRVHAP	SGRVYHVKNPPKVEGKDD	VDTG	
[Truncated_Name:3] 6RZE_A.pdb		VPDELIVDAIVGRRVHAP	SGRVYHVKNPPKVEGKDD	VDTG	
[Truncated_Name:4] 3HPR_A.pdb		VPDELIVDRIVGRRVHAP	SGRVYHVKNPPKVEGKDD	GTG	
[Truncated_Name:5] 1E4V_A.pdb		VPDELIVDRIVGRRVHAP	SGRVYHVKNPPKVEGKDD	VDTG	
[Truncated_Name:6] 5EJE_A.pdb		VPDELIVDRIVGRRVHAP	SGRVYHVKNPPKVEGKDD	VDTG	
[Truncated_Name:7] 1E4Y_A.pdb		VPDELIVDRIVGRRVHAP	SGRVYHVKNPPKVEGKDD	VDTG	
[Truncated_Name:8] 3X2S_A.pdb		VPDELIVDRIVGRRVHAP	SGRVYHVKNPPKVEGKDD	VDTG	
[Truncated_Name:9] 6HAP_A.pdb		VPDELIVDRIVGRRVHAP	SGRVYHVKNPPKVEGKDD	VDTG	
[Truncated_Name:10] 6HAM_A.pdb		VPDELIVDRIVGRRVHAP	SGRVYHVKNPPKVEGKDD	VDTG	
[Truncated_Name:11] 4K46_A.pdb		VADSVIVERMAGRRAHL	ASGRTYHNVPKVEGKDD	VDTG	
[Truncated_Name:12] 3GMT_A.pdb		VPFSEIIERMSGRRTHP	ASGRTYHVKNPPKVEGKDD	VDTG	
[Truncated_Name:13] 4PZL_A.pdb		VADNLLIERITGRRIHP	ASGRTYHTKFNPPKVADKDD	VDTG	
		*	^^^ ^ *** * *** ** ^***** *** **		
	121	.	.	.	160
	161	.	.	.	200
[Truncated_Name:1] 1AKE_A.pdb		EELTTRKDDQEETVRKRL	VEYHQM TAPLIGYYSKEAE	AGN	
[Truncated_Name:2] 6S36_A.pdb		EELTTRKDDQEETVRKRL	VEYHQM TAPLIGYYSKEAE	AGN	
[Truncated_Name:3] 6RZE_A.pdb		EELTTRKDDQEETVRKRL	VEYHQM TAPLIGYYSKEAE	AGN	
[Truncated_Name:4] 3HPR_A.pdb		EELTTRKDDQEETVRKRL	VEYHQM TAPLIGYYSKEAE	AGN	
[Truncated_Name:5] 1E4V_A.pdb		EELTTRKDDQEETVRKRL	VEYHQM TAPLIGYYSKEAE	AGN	
[Truncated_Name:6] 5EJE_A.pdb		EELTTRKDDQEECVRKRL	VEYHQM TAPLIGYYSKEAE	AGN	

```

[Truncated_Name:7]1E4Y_A.pdb      EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
[Truncated_Name:8]3X2S_A.pdb      EELTTRKDDQEETVRKRLCEYHQM TAPLIGYYSKEAEAGN
[Truncated_Name:9]6HAP_A.pdb      EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
[Truncated_Name:10]6HAM_A.pdb     EELTTRKDDQEETVRKRLVEYHQM TAPLIGYYSKEAEAGN
[Truncated_Name:11]4K46_A.pdb     EDLVIREDDKEETVLARLGVYHNQ TAPLIAYYGKEAEAGN
[Truncated_Name:12]3GMT_A.pdb     EPLVQRDDDKKEETVKKRLDVYEA QTKPLITYYGDWARRGA
[Truncated_Name:13]4PZL_A.pdb     EPLITRTDDNEDTVKQRLSVYHAQ TAKLIDFYRNFSSSTNT
                                   * * * * * ^ * * * * * ^ *
                                   161 . . . 200

                                   201 . . 227
[Truncated_Name:1]1AKE_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:2]6S36_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:3]6RZE_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:4]3HPR_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:5]1E4V_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:6]5EJE_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:7]1E4Y_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:8]3X2S_A.pdb      T--KYAKVDGTKPVAEVRADLEKILG-
[Truncated_Name:9]6HAP_A.pdb      T--KYAKVDGTKPVCEVRADLEKILG-
[Truncated_Name:10]6HAM_A.pdb     T--KYAKVDGTKPVCEVRADLEKILG-
[Truncated_Name:11]4K46_A.pdb     T--QYLKFDGTKAVAESAELEKALA-
[Truncated_Name:12]3GMT_A.pdb     E-----NGLKAPA-----YRKISG-
[Truncated_Name:13]4PZL_A.pdb     KIPKYIKINGDQAVEKVSQDIFDQLNK
                                   *
                                   201 . . 227

```

Call:

```
pdbaln(files = files, fit = TRUE, exeFile = "msa")
```

Class:

```
pdbs, fasta
```

Alignment dimensions:

```
13 sequence rows; 227 position columns (204 non-gap, 23 gap)
```

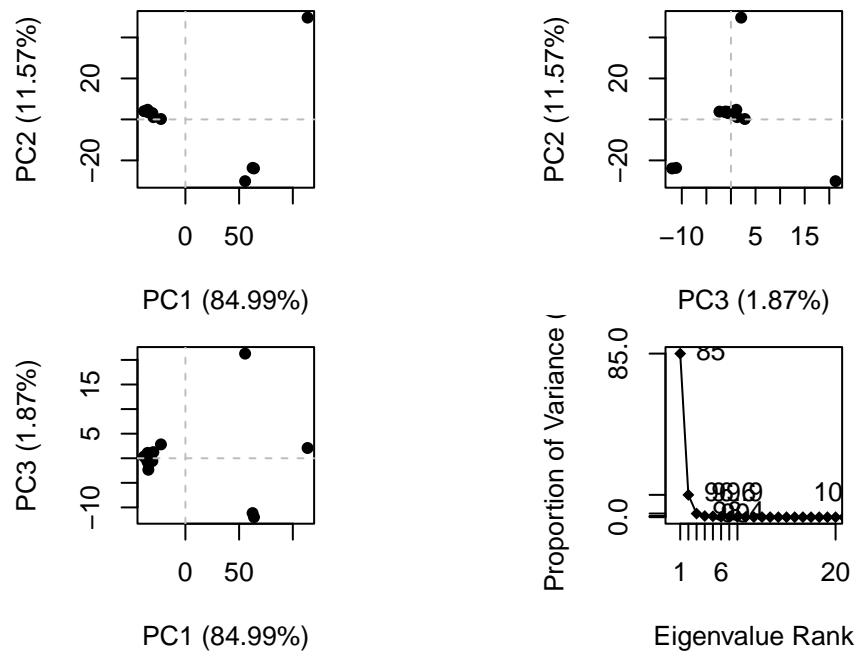
```
+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

```
# Vector containing PDB codes for figure axis
ids <- basename.pdb(pdb$id)
```

```
# Draw schematic alignment
# plot(pdb, labels=ids)
```

Jump to PCA

```
# Perform PCA
pc.xray <- pca(pdb)
plot(pc.xray)
```



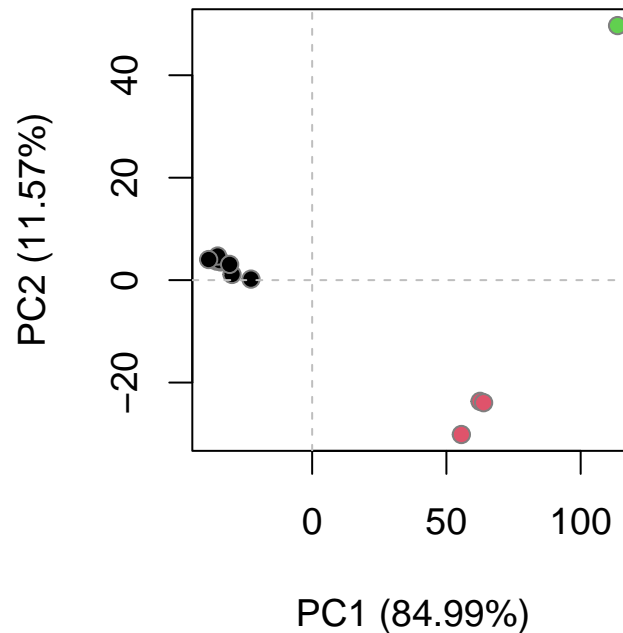
Calculate a prettier plot of the analysis:

```
# Calculate RMSD
rd <- rmsd(pdb)
```

Warning in rmsd(pdb): No indices provided, using the 204 non NA positions

```
# Structure-based clustering
hc.rd <- hclust(dist(rd))
grps.rd <- cutree(hc.rd, k=3)

plot(pc.xray, 1:2, col="grey50", bg=grps.rd, pch=21, cex=1)
```



Optional further visualization

To visualize the major structural variations in the

```
# Visualize first principal component
pc1 <- mkrtrj(pc.xray, pc=1, file="pc_1.pdb")
```

We can also plot our main PCA results using ggplot:

```
#Plotting results with ggplot2
library(ggplot2)
library(ggrepel)

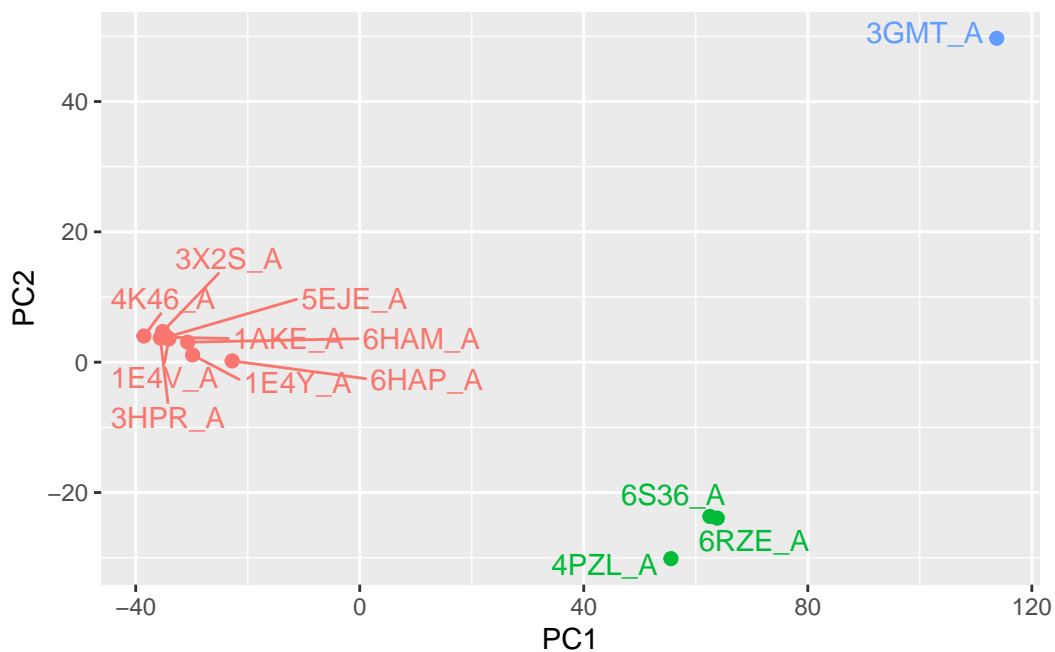
df <- data.frame(PC1=pc.xray$z[,1],
```

```

PC2=pc.xray$z[,2],
col=as.factor(grps.rd),
ids=ids)

p <- ggplot(df) +
  aes(PC1, PC2, col=col, label=ids) +
  geom_point(size=2) +
  geom_text_repel(max.overlaps = 20) +
  theme(legend.position = "none")
p

```



Normal Mode Analysis

Use the `nma()` function to provide a normal mode analysis on both of the single structures or a complete structure ensemble. This facilitates characterizing and comparing flexibility profiles of related protein structures.

```

# NMA of all structures
modes <- nma(pdb)

```

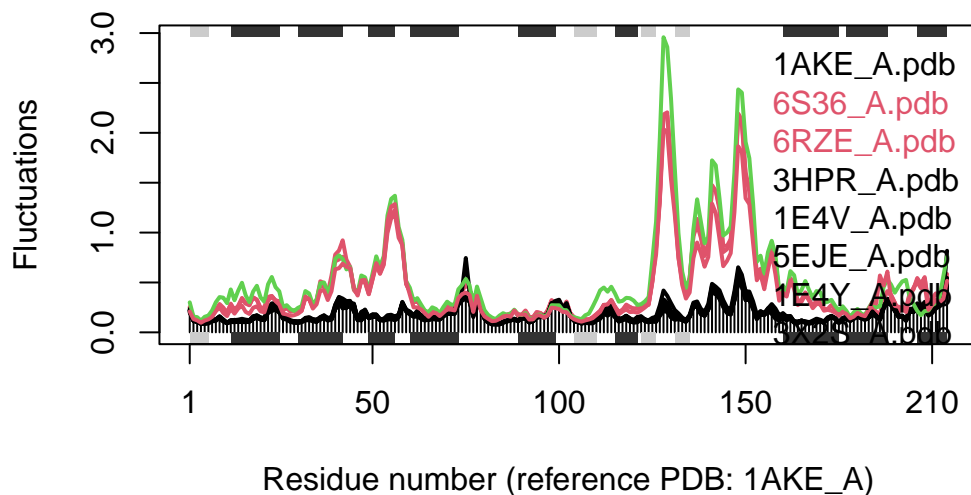
Details of Scheduled Calculation:

```
... 13 input structures
... storing 606 eigenvectors for each structure
... dimension of x$U.subspace: ( 612x606x13 )
... coordinate superposition prior to NM calculation
... aligned eigenvectors (gap containing positions removed)
... estimated memory usage of final 'eNMA' object: 36.9 Mb
```

	0%
=====	8%
=====	15%
=====	23%
=====	31%
=====	38%
=====	46%
=====	54%
=====	62%
=====	69%
=====	77%
=====	85%
=====	92%
=====	100%

```
plot(modes, pdbs, col=grps.rd)
```

Extracting SSE from pdbs\$sse attribute



Q14. What do you note about this plot? Are the black and colored lines similar or different? Where do you think they differ most and why?

I note that these residue numbers are plotted with how many fluctuations occur between specific clusters. The black and colored lines are different, however the colors themselves represent a certain cluster of residues that are grouped together, as we can see on the right of the graph with the different structures being colored accordingly to how similar they are. They differ the most around the 150 region because this seems to be where there is the biggest peak which means this is the region with the most fluctuations meaning differences.