

Class 10: Structural Bioinformatics 1

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PDB statistics

The Protein Data Bank (PDB) is a main repository of biomolecular structures. Let's see what it contains:

```
stats <- read.csv("~/Downloads/Data Export Summary.csv")
head(stats)
```

	Molecular.Type	X.ray	EM	NMR	Integrative	Multiple.methods
1	Protein (only)	178,795	21,825	12,773	343	226
2	Protein/Oligosaccharide	10,363	3,564	34	8	11
3	Protein/NA	9,106	6,335	287	24	7
4	Nucleic acid (only)	3,132	221	1,566	3	15
5	Other	175	25	33	4	0
6	Oligosaccharide (only)	11	0	6	0	1
	Neutron	Other	Total			
1	84	32	214,078			
2	1	0	13,981			
3	0	0	15,759			
4	3	1	4,941			
5	0	0	237			
6	0	4	22			

```
stats$X.ray
```

```
[1] "178,795" "10,363" "9,106" "3,132" "175" "11"
```

```
sum(stats$Neutron)
```

```
[1] 88
```

The comma in these numbers leads to the numbers here being read as characters.

```
c(100,10, "barry")
```

```
[1] "100"    "10"     "barry"
```

```
library(readr)
stats <- read_csv("~/Downloads/Data Export Summary.csv")
```

```
Rows: 6 Columns: 9
-- Column specification -----
Delimiter: ","
chr (1): Molecular Type
dbl (4): Integrative, Multiple methods, Neutron, Other
num (4): X-ray, EM, NMR, Total

i Use `spec()` to retrieve the full column specification for this data.
i Specify the column types or set `show_col_types = FALSE` to quiet this message.
```

```
head(stats)
```

```
# A tibble: 6 x 9
`Molecular Type` `X-ray`   EM   NMR Integrative `Multiple methods` Neutron
<chr>           <dbl> <dbl> <dbl>      <dbl>           <dbl> <dbl>
1 Protein (only) 178795 21825 12773      343          226    84
2 Protein/Oligosacch~ 10363  3564   34        8            11    1
3 Protein/NA       9106   6335   287       24            7    0
4 Nucleic acid (only) 3132   221   1566      3            15    3
5 Other             175    25    33        4            0    0
6 Oligosaccharide (o~ 11     0     6         0            1    0
# i 2 more variables: Other <dbl>, Total <dbl>
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
n.xray <- sum(stats$`X-ray`)
#n.em <-
n.total <- sum(stats$Total)

n.xray/n.total
```

```
[1] 0.8095077
```

```
n.em <- sum(stats$EM)
(n.xray/n.total)+(n.em/n.total)
```

```
[1] 0.937892
```

93.7892% percent of structures are solved by x-ray and EM.

Q2: What proportion of structures in the PDB are protein?

```
n.protein <- c(214078)
n.protein/n.total
```

```
[1] 0.8596889
```

85.96889%

Q3: SKIP...

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

One molecule is show to make it easier to see and it shows the interactions between water and the protein.

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have residue number is 301

Visualizing the HIV-1 protease structure

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.

We can use the Molecular viewer online: <https://molstar.org/viewer/>

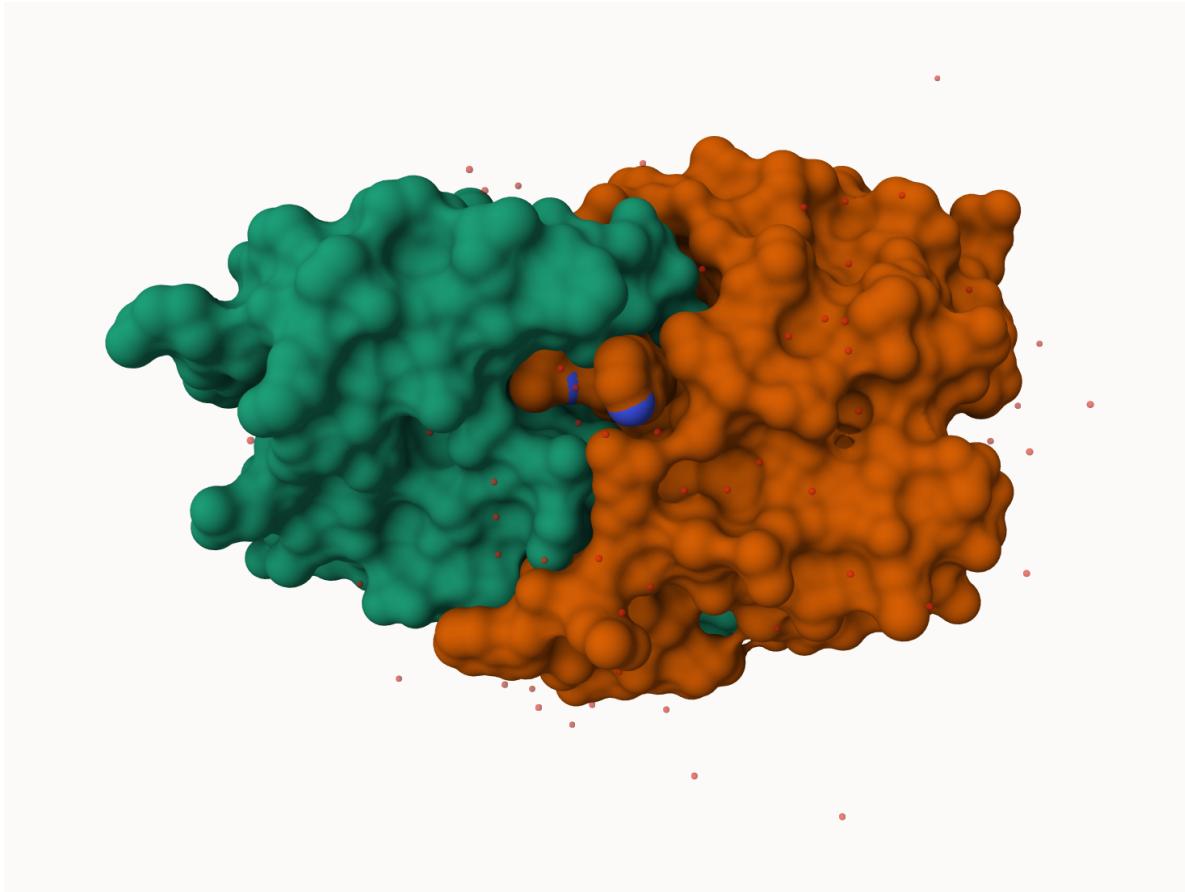
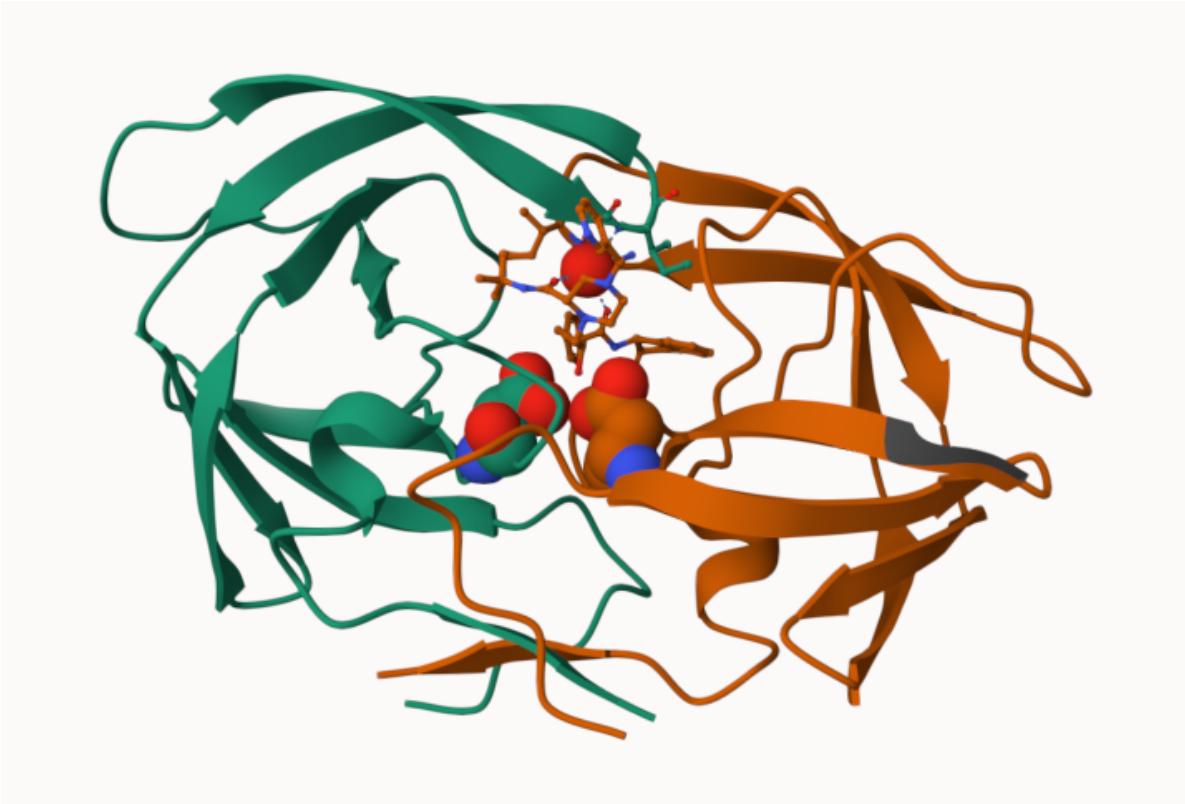


Figure 1: My First image of HIV-Pr with surface display shoing ligand binding

A new clean image showing the catalytic ASP25 amino acids in both chains of the HIV-PR dimer along with the inhibitor and the all important active site water.



Bio3D package for structural bioinformatics

```
library(bio3d)  
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
 pdb
```

```
Call: read.pdb(file = "1hsg")  
  
Total Models#: 1  
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

Protein sequence:

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPCKMIGGIGGFVKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWPCKMIGGIGGFVKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

Q7: How many amino acid residues are there in this pdb object?

214 > Q8: Name one of the two non-protein residues?

MK1 > Q9: How many protein chains are in this structure?

2

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

Predicting functional motions of a single structure

Read an ADK structure from the PDB database

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

Call: read.pdb(file = "6s36")

Total Models#: 1

Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)

Non-protein/nucleic resid values: [CL (3), HOH (238), MG (2), NA (1)]

Protein sequence:

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMLRAAVKSGSELGKQAKDIDMAGKLVT  
DELVIALVKERIAQEDCRNGFLLDGFPRТИPQADAMKEAGINVDYVLEFDVPDELIVDKI  
VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

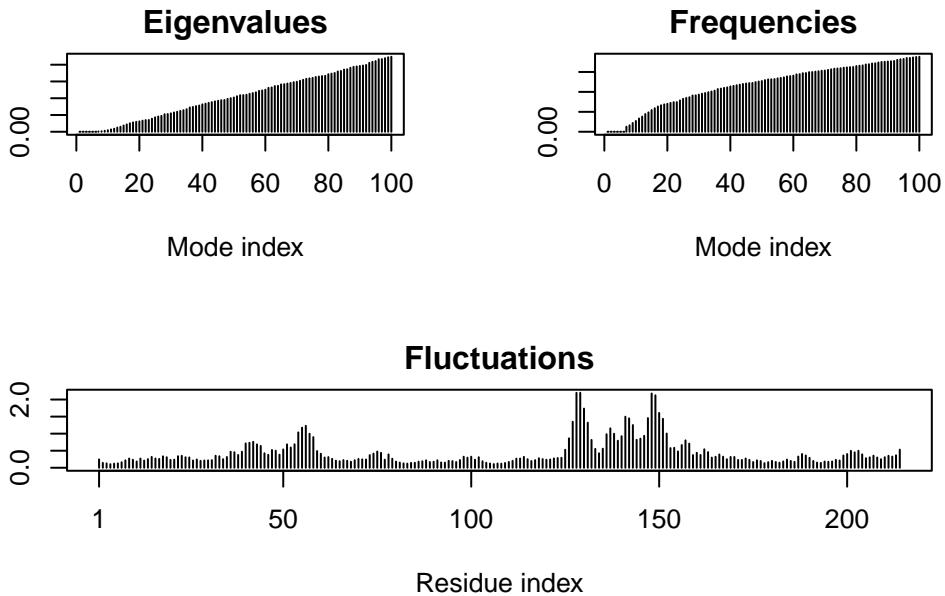
+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call

```
# Perform protein flexibility prediction  
m <- nma(adk)
```

Building Hessian... Done in 0.034 seconds.

Diagonalizing Hessian... Done in 0.394 seconds.

```
plot(m)
```



write out our results as a wee trajectory/movie of predicted motions:

```
mktrj(m, file="adk_m7.pdb")
```

Comparative analysis with PCA

First step find an ADK sequence:

```
library(bio3d)
id <- "1AKE_A" ## Change this to run a different analysis
aa <- get.seq(id)
```

Warning in get.seq(id): Removing existing file: seqs.fasta

Fetching... Please wait. Done.

```
aa
```

1 pdb 1AKE A 1	.	60 MRIILLGAPGAGKGTQAAQFIMEKYGIPQISTGMLRAAVKSGSELGKQAKDIMDAGKLVT 1
----------------------	---	---

	61	120
pdb 1AKE A	DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVVDYVLEFDVPDELIVDRI							
	61	120
	121	180
pdb 1AKE A	VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG							
	121	180
	181	.	.	.	214			
pdb 1AKE A	YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG							
	181	.	.	.	214			

Call:

```
read.fasta(file = outfile)
```

Class:

fasta

Alignment dimensions:

1 sequence rows; 214 position columns (214 non-gap, 0 gap)

+ attr: id, ali, call

DB database for all related entries:

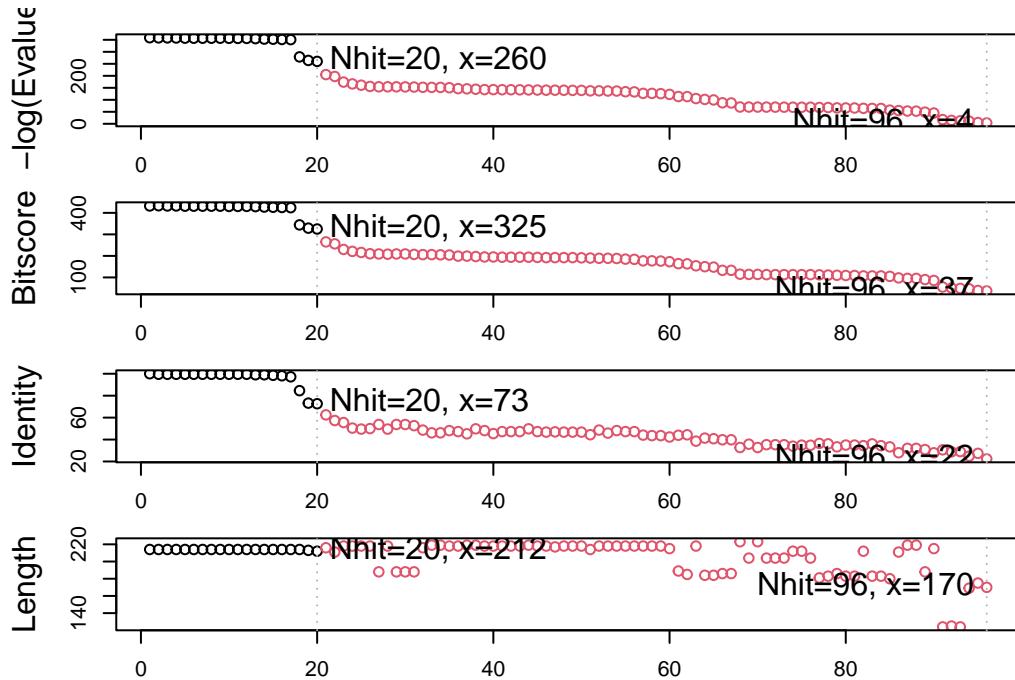
```
blast <- blast.pdb(aa)
```

Searching ... please wait (updates every 5 seconds) RID = SPYKXYWE014
.....
Reporting 96 hits

```
hits <- plot(blast)
```

* Possible cutoff values: 260 3
 Yielding Nhits: 20 96

* Chosen cutoff value of: 260
Yielding Nhits: 20



```
head(blast$hit.tbl)
```

	queryid	subjectids	identity	alignmentlength	mismatches	gapopens	q.start		
	q.end	s.start	s.end	evalvalue	bitscore	positives	mlog.evalvalue	pdb.id	acc
1	Query_6403663	1AKE_A	100.000		214	0	0	1AKE_A	1
2	Query_6403663	8BQF_A	99.533		214	1	0	8BQF_A	1
3	Query_6403663	4X8M_A	99.533		214	1	0	4X8M_A	1
4	Query_6403663	6S36_A	99.533		214	1	0	6S36_A	1
5	Query_6403663	9R6U_A	99.533		214	1	0	9R6U_A	1
6	Query_6403663	9R71_A	99.533		214	1	0	9R71_A	1

The “top hits” are in the `hits` object. Now we can download these to our computer. Put these in a sub-folder (directory) called pdbs

```
## Download related PDB files
files <- get.pdb(hits$pdb.id, path="pdbs", split=TRUE, gzip=TRUE)
```

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1AKE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/8BQF.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4X8M.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6S36.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/9R6U.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/9R71.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/8Q2B.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/8RJ9.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6RZE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4X8H.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3HPR.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4V.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/5EJE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4Y.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3X2S.pdb.gz exists. Skipping download

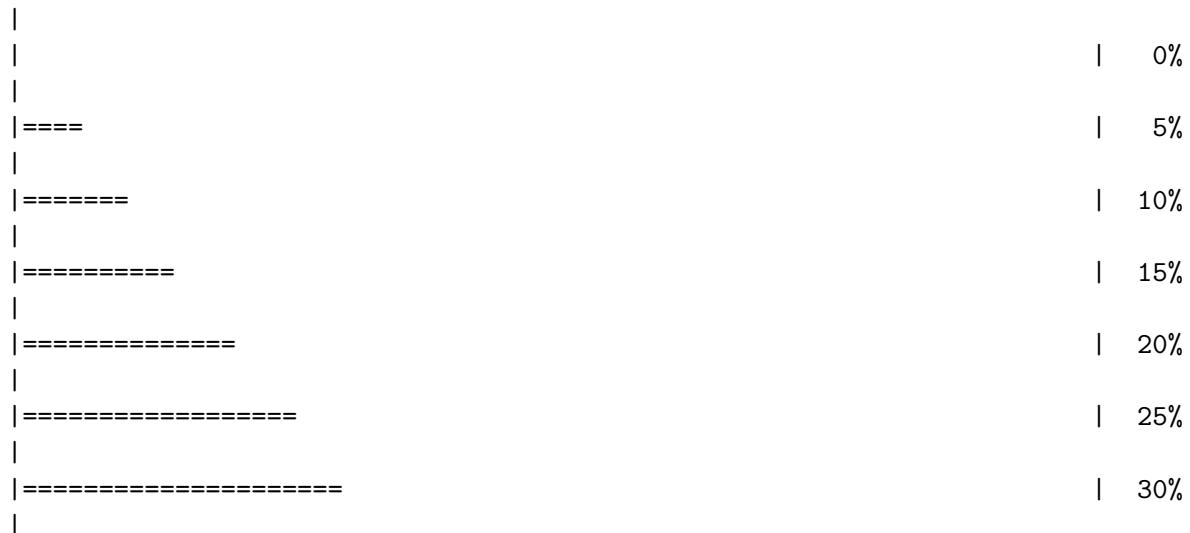
Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAP.pdb.gz exists. Skipping download

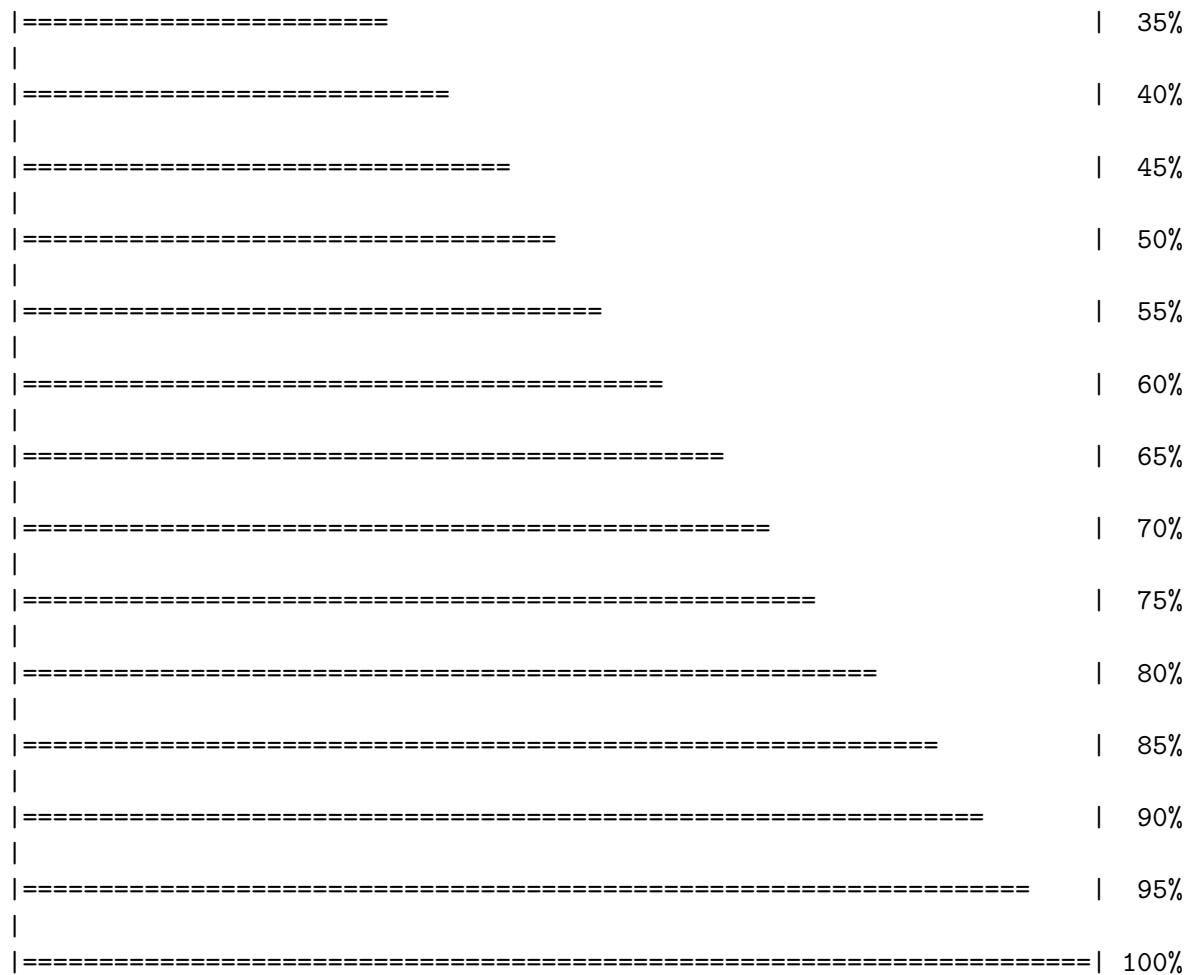
Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAM.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/8PVW.pdb.gz exists. Skipping download

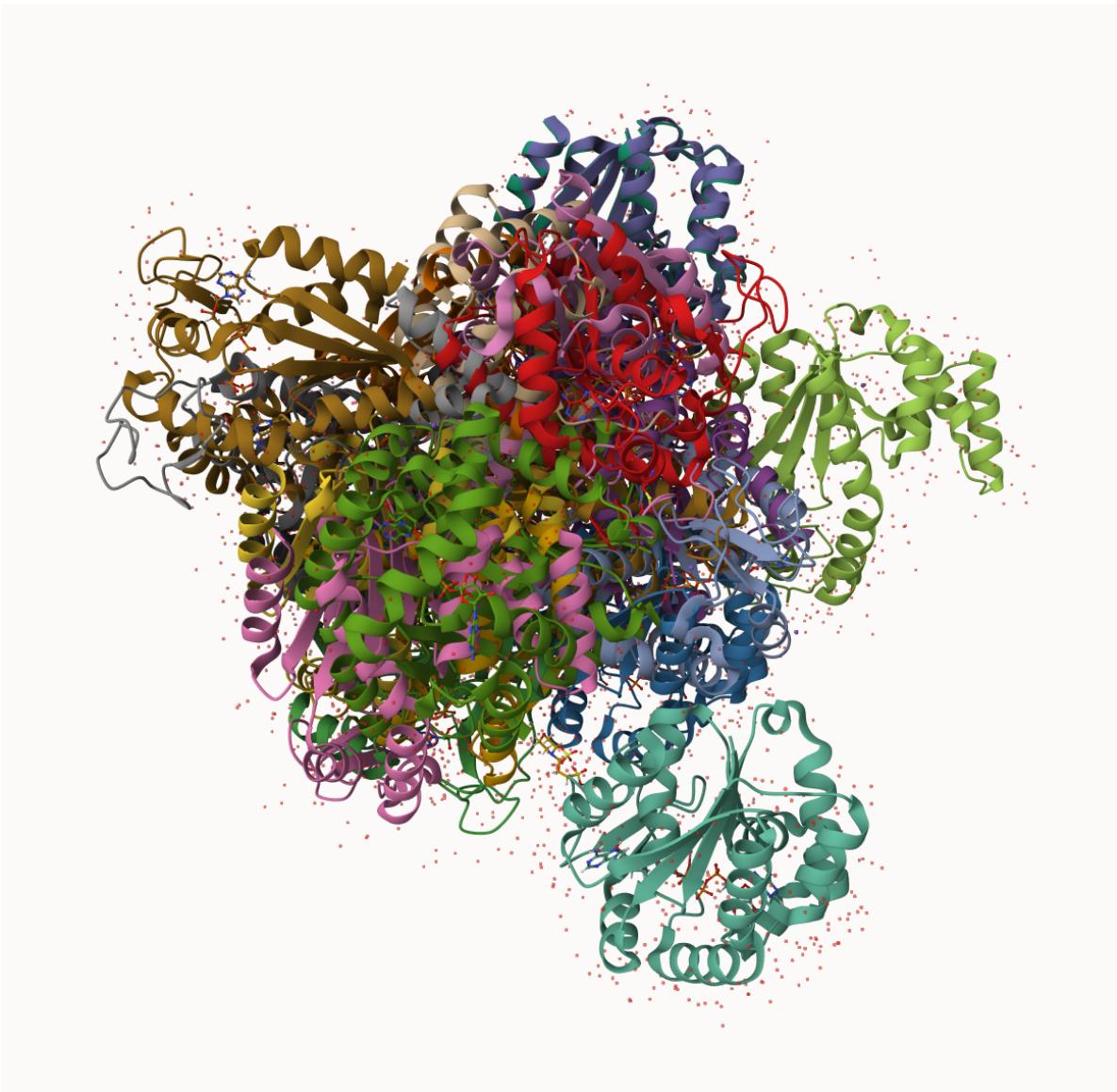
Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4K46.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4NP6.pdb.gz exists. Skipping download





These look like a hot mess



Next we will use the `pdbaln()` function to align and also superpose the PDB structure.

This requires a BioConductor package called “msa” that we need to install. First we install BiocManager. Then we use `BiocManager::install("msa")`

```
# Align related PDBs
pdbs <- pdbaln(files, fit = TRUE, exefile="msa")
```

```
Reading PDB files:
pdbs/split_chain/1AKE_A.pdb
```

```
pdb/split_chain/8BQF_A.pdb
pdb/split_chain/4X8M_A.pdb
pdb/split_chain/6S36_A.pdb
pdb/split_chain/9R6U_A.pdb
pdb/split_chain/9R71_A.pdb
pdb/split_chain/8Q2B_A.pdb
pdb/split_chain/8RJ9_A.pdb
pdb/split_chain/6RZE_A.pdb
pdb/split_chain/4X8H_A.pdb
pdb/split_chain/3HPR_A.pdb
pdb/split_chain/1E4V_A.pdb
pdb/split_chain/5EJE_A.pdb
pdb/split_chain/1E4Y_A.pdb
pdb/split_chain/3X2S_A.pdb
pdb/split_chain/6HAP_A.pdb
pdb/split_chain/6HAM_A.pdb
pdb/split_chain/8PVW_A.pdb
pdb/split_chain/4K46_A.pdb
pdb/split_chain/4NP6_A.pdb

    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
..   PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
....  PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
..
.
```

Extracting sequences

```
pdb/seq: 1  name: pdb/split_chain/1AKE_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 2  name: pdb/split_chain/8BQF_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 3  name: pdb/split_chain/4X8M_A.pdb
pdb/seq: 4  name: pdb/split_chain/6S36_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
```

```

pdb/seq: 5 name: pdbs/split_chain/9R6U_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 6 name: pdbs/split_chain/9R71_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 7 name: pdbs/split_chain/8Q2B_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 8 name: pdbs/split_chain/8RJ9_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 9 name: pdbs/split_chain/6RZE_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 10 name: pdbs/split_chain/4X8H_A.pdb
pdb/seq: 11 name: pdbs/split_chain/3HPR_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 12 name: pdbs/split_chain/1E4V_A.pdb
pdb/seq: 13 name: pdbs/split_chain/5EJE_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 14 name: pdbs/split_chain/1E4Y_A.pdb
pdb/seq: 15 name: pdbs/split_chain/3X2S_A.pdb
pdb/seq: 16 name: pdbs/split_chain/6HAP_A.pdb
pdb/seq: 17 name: pdbs/split_chain/6HAM_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 18 name: pdbs/split_chain/8PVW_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 19 name: pdbs/split_chain/4K46_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 20 name: pdbs/split_chain/4NP6_A.pdb

```

Have a look at this new “alignment object” pdbs

pdb

	1	40
[Truncated_Name:1] 1AKE_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA	
[Truncated_Name:2] 8BQF_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA	
[Truncated_Name:3] 4X8M_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA	
[Truncated_Name:4] 6S36_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA	
[Truncated_Name:5] 9R6U_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA	
[Truncated_Name:6] 9R71_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA	
[Truncated_Name:7] 8Q2B_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA	
[Truncated_Name:8] 8RJ9_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA	
[Truncated_Name:9] 6RZE_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA	
[Truncated_Name:10] 4X8H_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA	

[Truncated_Name:11] 3HPR_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA
[Truncated_Name:12] 1E4V_A.pdb	--MRIILLGAPVAGKGTQAQFIMEKYGIPQISTGDMRLAA
[Truncated_Name:13] 5EJE_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA
[Truncated_Name:14] 1E4Y_A.pdb	--MRIILLGALVAGKGTQAQFIMEKYGIPQISTGDMRLAA
[Truncated_Name:15] 3X2S_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA
[Truncated_Name:16] 6HAP_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA
[Truncated_Name:17] 6HAM_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA
[Truncated_Name:18] 8PVW_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAA
[Truncated_Name:19] 4K46_A.pdb	--MRIILLGAPGAGKGTQAQFIMAKFGIPQISTGDMRLAA
[Truncated_Name:20] 4NP6_A.pdb	NAMRIILLGAPGAGKGTQAQFIMEKFGIPQISTGDMRLAA ***** 40
1	
41 80
[Truncated_Name:1] 1AKE_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:2] 8BQF_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQE---
[Truncated_Name:3] 4X8M_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:4] 6S36_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:5] 9R6U_A.pdb	VKSGSELGAQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:6] 9R71_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:7] 8Q2B_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:8] 8RJ9_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:9] 6RZE_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:10] 4X8H_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:11] 3HPR_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:12] 1E4V_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:13] 5EJE_A.pdb	VKSGSELGKQAKDIMDACKLVTDELVIALVKERIAQEDCR
[Truncated_Name:14] 1E4Y_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:15] 3X2S_A.pdb	VKSGSELGKQAKDIMDCGKLVTDELVIALVKERIAQEDSR
[Truncated_Name:16] 6HAP_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVRERICQEDSR
[Truncated_Name:17] 6HAM_A.pdb	IKSGSELGKQAKDIMDAGLVTDEIIIALVKERICQEDSR
[Truncated_Name:18] 8PVW_A.pdb	VKSGSELGKQAKDIMDAGLVTDELVIALVKERIAQEDCR
[Truncated_Name:19] 4K46_A.pdb	IKAGTELGKQAKSVIDAGQLVSDDIILGLVKERIAQDDCA
[Truncated_Name:20] 4NP6_A.pdb	IKAGTELGKQAKAVIDAGQLVSDDIILGLIKERIAQADCE ^* ^*** *** ^* **^~^~^~^~^*** *
41 80
81 120
[Truncated_Name:1] 1AKE_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVDTVLEFDVPDELIVD
[Truncated_Name:2] 8BQF_A.pdb	-GFLLDGFPRTIPQADAMKEAGINVDTVIEFDVPDELIVD
[Truncated_Name:3] 4X8M_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVDTVLEFDVPDELIVD
[Truncated_Name:4] 6S36_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVDTVLEFDVPDELIVD
[Truncated_Name:5] 9R6U_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVDTVLEFDVPDELIVD

[Truncated_Name:6] 9R71_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDALIVD		
[Truncated_Name:7] 8Q2B_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:8] 8RJ9_A.pdb	NGFLLAGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:9] 6RZE_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:10] 4X8H_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:11] 3HPR_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:12] 1E4V_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:13] 5EJE_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:14] 1E4Y_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:15] 3X2S_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:16] 6HAP_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:17] 6HAM_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:18] 8PVW_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVD		
[Truncated_Name:19] 4K46_A.pdb	KGFLLDGFPTIPQADGLKEVGVVVDYVIEFDVADSVIVE		
[Truncated_Name:20] 4NP6_A.pdb	KGFLLDGFPTIPQADGLKEMGINVDYVIEFDVADDVIVE		
	***** *****^~** *^ ****^**** * ^**^		
81	.	.	120
121	.	.	160
	RIVGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKD		
	RIVGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKD		
	RIVGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKD		
	KIVGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKD		
	RIVGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKD		
	RILKR--GETSGRV-----D		
	RMAGRRAHLASGRTYHNVNPPKVEGKDDVTGEDLVIRED		
	RMAGRRAHLPGRRTYHVNVNPPKVEGKDDVTGEDLVIRED		
	^ * ***		
121	.	.	160
161	.	.	200

[Truncated_Name:1] 1AKE_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:2] 8BQF_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:3] 4X8M_A.pdb DQEETVRKRLVEWHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:4] 6S36_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:5] 9R6U_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:6] 9R71_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:7] 8Q2B_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:8] 8RJ9_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:9] 6RZE_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:10] 4X8H_A.pdb DQEETVRKRLVEYHQMTAALIGYYSKAEAGNTKYAKVDG
[Truncated_Name:11] 3HPR_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:12] 1E4V_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:13] 5EJE_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:14] 1E4Y_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:15] 3X2S_A.pdb DQEETVRKRLCEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:16] 6HAP_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:17] 6HAM_A.pdb DQEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:18] 8PVW_A.pdb DNEETVRKRLVEYHQMTAPLIGYYSKAEAGNTKYAKVDG
[Truncated_Name:19] 4K46_A.pdb DKEETVLARLCVYHNQTAPIIAYYGKEAEAGNTQYLKFDG
[Truncated_Name:20] 4NP6_A.pdb DKEETVRARLNVYHTQTAPIIEYYGKEAAAGKTQYLKFDG

161 200

		201	.	216
[Truncated_Name:1]	1AKE_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:2]	8BQF_A.pdb	TKPVAEVRADEKIL-		
[Truncated_Name:3]	4X8M_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:4]	6S36_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:5]	9R6U_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:6]	9R71_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:7]	8Q2B_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:8]	8RJ9_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:9]	6RZE_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:10]	4X8H_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:11]	3HPR_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:12]	1E4V_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:13]	5EJE_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:14]	1E4Y_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:15]	3X2S_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:16]	6HAP_A.pdb	TKPVCEVRADLEKILG		
[Truncated_Name:17]	6HAM_A.pdb	TKPVCEVRADLEKILG		
[Truncated_Name:18]	8PVW_A.pdb	TKPVAEVRADEKILG		
[Truncated_Name:19]	4K46_A.pdb	TKVAEVSAELEKALA		

```
[Truncated_Name:20]4NP6_A.pdb    TKQVSEVSADIAKALA
                           ** * ** *^* *
                           201      .      216
```

Call:

```
pdbaln(files = files, fit = TRUE, exefile = "msa")
```

Class:

```
pdb, fasta
```

Alignment dimensions:

```
20 sequence rows; 216 position columns (182 non-gap, 34 gap)
```

```
+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

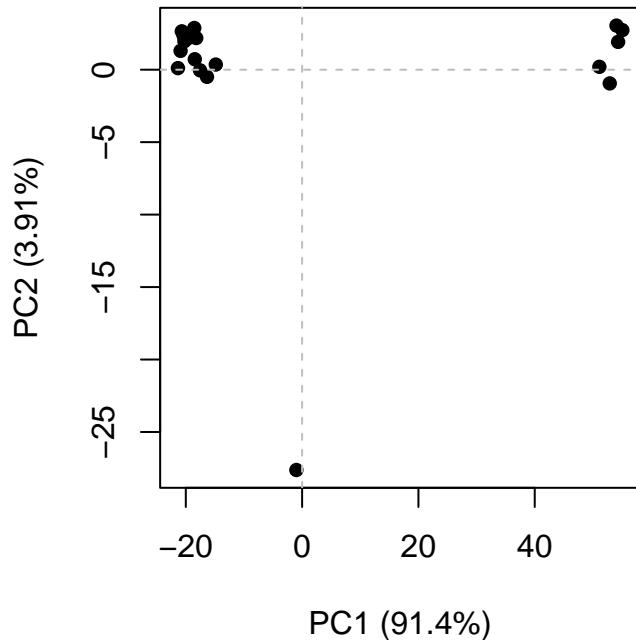
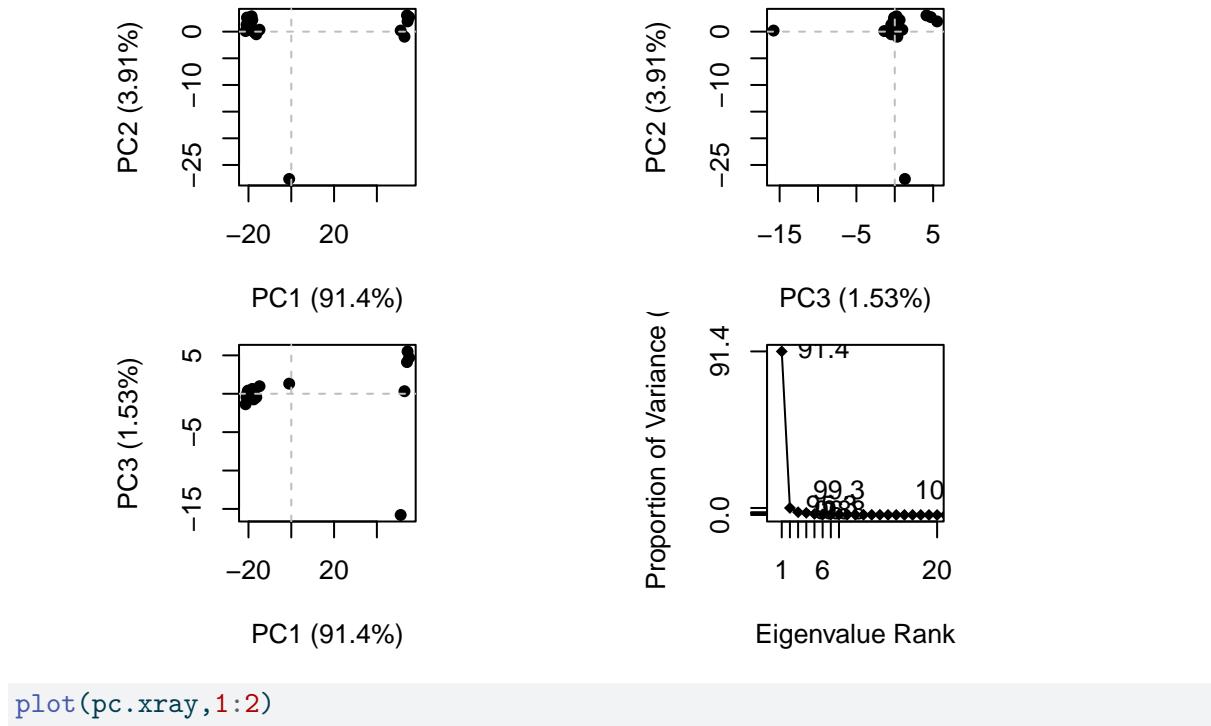
We could view these in R with **bio3dview view.pdb()** function.

```
# library(bio3dview)
# view.pdb(pdb, colorScheme = "residue")
```

PCA

We can run PCA on our **pdb** object using the **pca()** function from **bio3d**

```
pc.xray <- pca(pdb)
plot(pc.xray)
```



We can make a visualization of the major conformational difference (i.e. large scale structure change) captured by our PCA analysis with `mktrj()` function.

```
pc1 <- mktrj(pc.xray, pc=1, file="pc_1.pdb")
```

Let's see in Molstar