

Class 10: Structural Bioinformatics 1

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PDB statistics

The Protein Data Bank (PDB) is a main repository of biomolecular structures. Let's see what it contains:

```
stats <- read.csv("~/Downloads/Data Export Summary.csv")
head(stats)
```

	Molecular.Type	X.ray	EM	NMR	Integrative	Multiple.methods
1	Protein (only)	178,795	21,825	12,773	343	226
2	Protein/Oligosaccharide	10,363	3,564	34	8	11
3	Protein/NA	9,106	6,335	287	24	7
4	Nucleic acid (only)	3,132	221	1,566	3	15
5	Other	175	25	33	4	0
6	Oligosaccharide (only)	11	0	6	0	1

	Neutron	Other	Total
1	84	32	214,078
2	1	0	13,981
3	0	0	15,759
4	3	1	4,941
5	0	0	237
6	0	4	22

```
stats$X.ray
```

```
[1] "178,795" "10,363" "9,106" "3,132" "175" "11"
```

```
sum(stats$Neutron)
```

```
[1] 88
```

The comma in these numbers leads to the numbers here being read as characters.

```
c(100,10, "barry")
```

```
[1] "100"  "10"   "barry"
```

```
library(readr)
stats <- read_csv("~/Downloads/Data Export Summary.csv")
```

Rows: 6 Columns: 9

-- Column specification -----

Delimiter: ","

chr (1): Molecular Type

dbl (4): Integrative, Multiple methods, Neutron, Other

num (4): X-ray, EM, NMR, Total

i Use `spec()` to retrieve the full column specification for this data.

i Specify the column types or set `show_col_types = FALSE` to quiet this message.

```
head(stats)
```

A tibble: 6 x 9

	`Molecular Type` <chr>	`X-ray` <dbl>	EM <dbl>	NMR <dbl>	Integrative <dbl>	`Multiple methods` <dbl>	Neutron <dbl>
1	Protein (only)	178795	21825	12773	343	226	84
2	Protein/Oligosacch~	10363	3564	34	8	11	1
3	Protein/NA	9106	6335	287	24	7	0
4	Nucleic acid (only)	3132	221	1566	3	15	3
5	Other	175	25	33	4	0	0
6	Oligosaccharide (o~	11	0	6	0	1	0

i 2 more variables: Other <dbl>, Total <dbl>

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
n.xray <- sum(stats$`X-ray`)
#n.em <-
n.total <- sum(stats$Total)

n.xray/n.total
```

```
[1] 0.8095077
```

```
n.em <- sum(stats$EM)
(n.xray/n.total)+(n.em/n.total)
```

```
[1] 0.937892
```

93.7892% percent of structures are solved by x-ray and EM.

Q2: What proportion of structures in the PDB are protein?

```
n.protein <- c(214078)
n.protein/n.total
```

```
[1] 0.8596889
```

85.96889%

Q3: SKIP...

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

One molecule is shown to make it easier to see and it shows the interactions between water and the protein.

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

residue number is 301

Visualizing the HIV-1 protease structure

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.

We can use the Molecular viewer online: <https://molstar.org/viewer/>

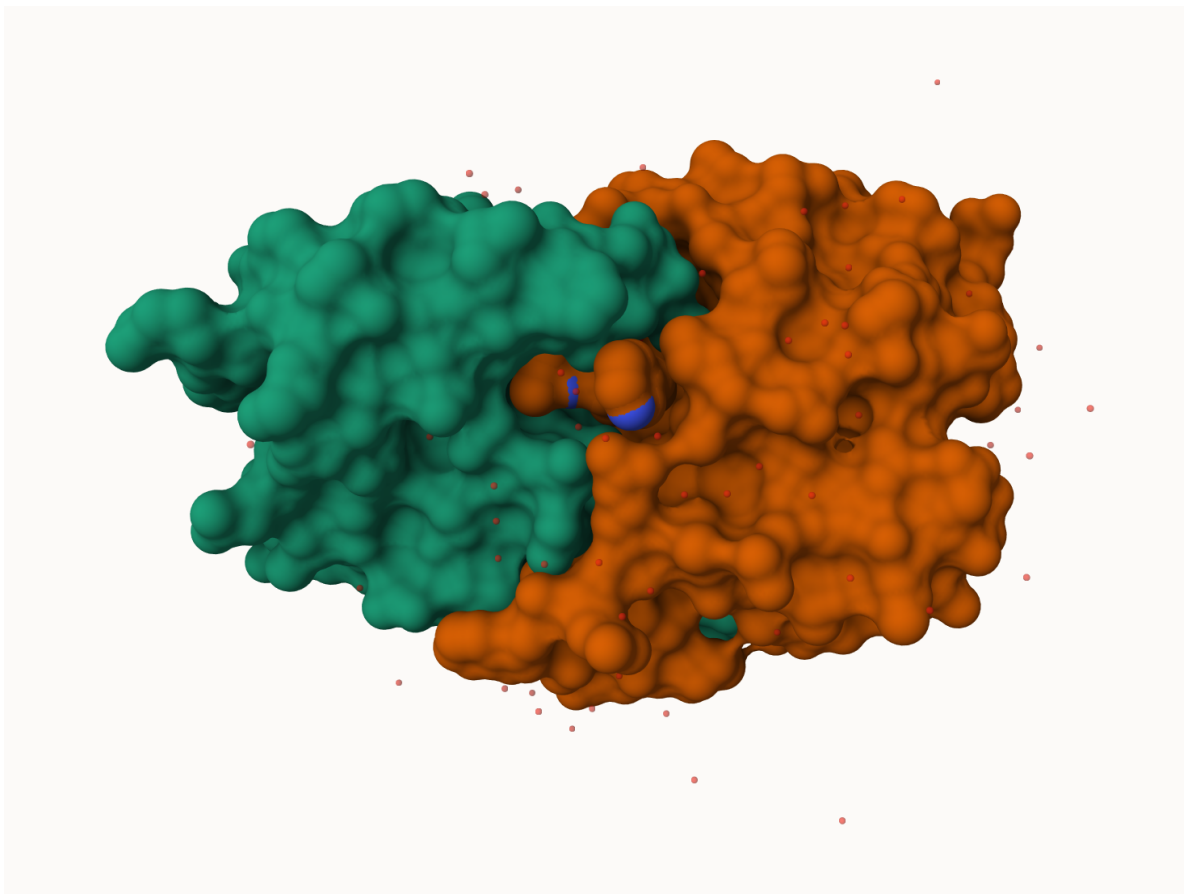
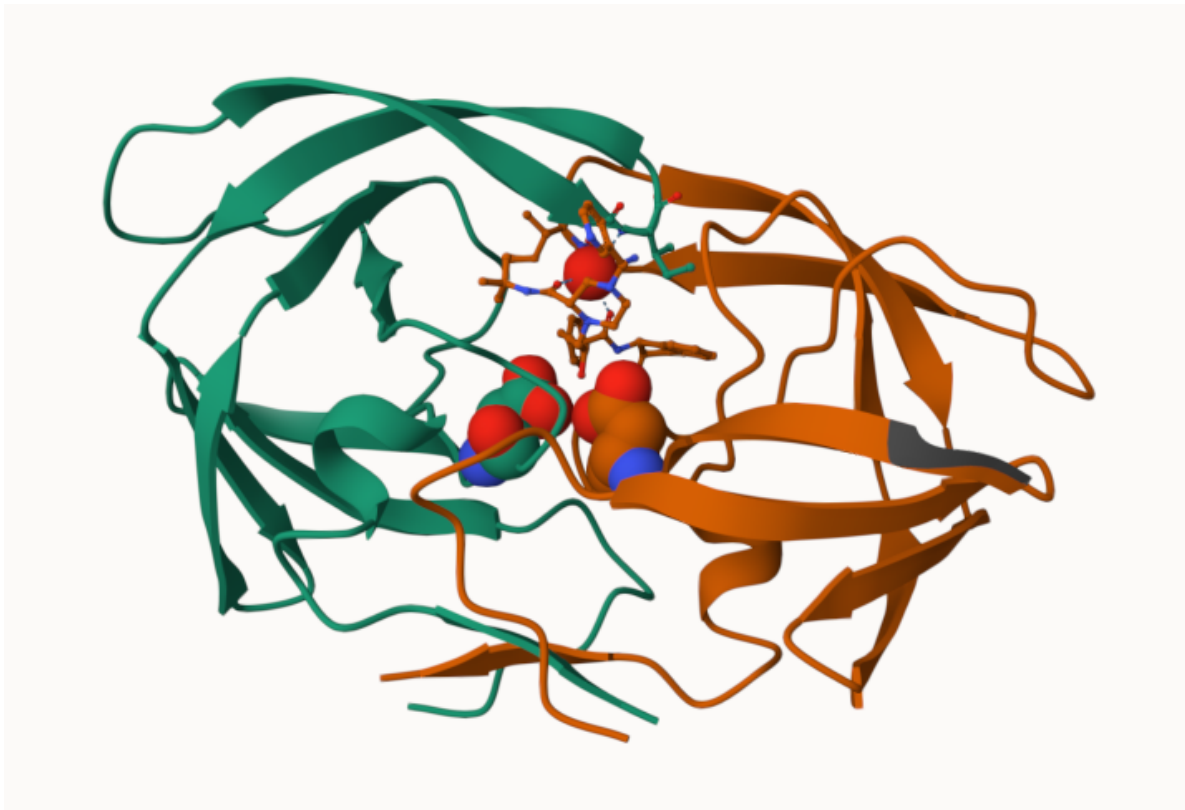


Figure 1: My First image of HIV-Pr with surface display shoing ligand binding

A new clean image showing the catalytic ASP25 amino acids in both chains of the HIV-PR dimer along with the inhibitor and the all important active site water.



Bio3D package for structural bioinformatics

```
library(bio3d)  
  
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
```

```
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

```

Protein sequence:

```

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

```

```

+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call

```

Q7: How many amino acid residues are there in this pdb object?

214 > Q8: Name one of the two non-protein residues?

MK1 > Q9: How many protein chains are in this structure?

2

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elasy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

Predicting fuctional motions of a single structure

Read an ADK structure from the PDB database

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

```
Call: read.pdb(file = "6s36")
```

```
Total Models#: 1
```

```
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)
```

```
Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 244 (residues: 244)
```

```
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
```

```
Protein sequence:
```

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV  
DELVIALVKERIAQEDCRNGFLDGFPRTPQADAMKEAGINVDYVLEFDVPDELIVDKI  
VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

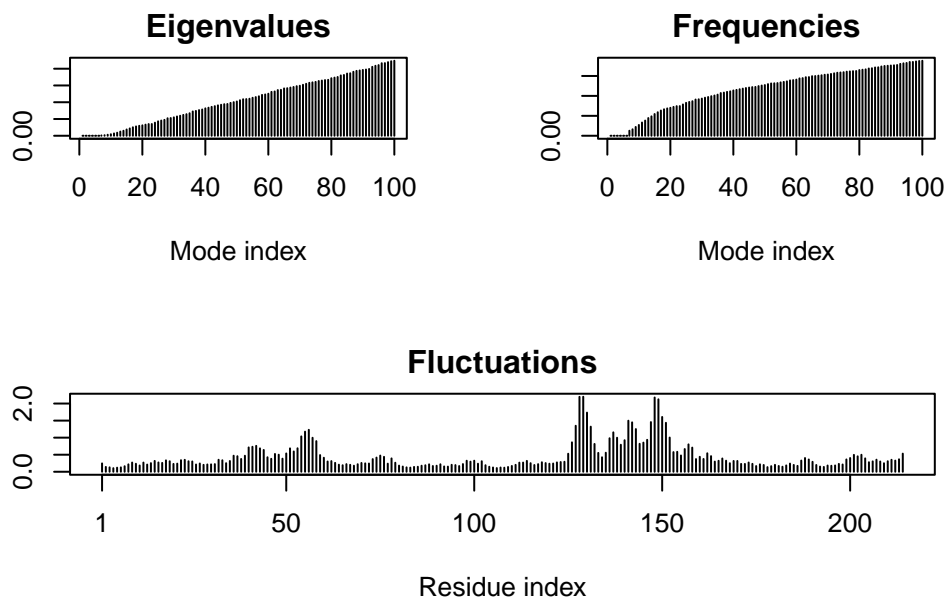
```
+ attr: atom, xyz, seqres, helix, sheet,  
      calpha, remark, call
```

```
# Perform protein flexibility prediction  
m <- nma(adk)
```

```
Building Hessian... Done in 0.034 seconds.
```

```
Diagonalizing Hessian... Done in 0.394 seconds.
```

```
plot(m)
```



write out our results as a wee trajectory/movie of predicted motions:

```
mktrj(m, file="adk_m7.pdb")
```

Comparative analysis with PCA

First step find an ADK sequence:

```
library(bio3d)
id <- "lake_A" ## Change this to run a different analysis
aa <- get.seq( id)
```

Warning in get.seq(id): Removing existing file: seqs.fasta

Fetching... Please wait. Done.

```
aa
```

```

pdb|1AKE|A      1      .      .      .      .      .      .      60
                MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMDLRAAVKSGSELGKQAKDIMDAGKLV
                1      .      .      .      .      .      .      60
```



```

        61      .      .      .      .      .      .      120
pdb|1AKE|A  DELVIALVKERIAQEDCRNGFLLDGFRTIPQADAMKEAGINVDYVLEFDVPDELIVDRI
        61      .      .      .      .      .      .      120

        121     .      .      .      .      .      .      180
pdb|1AKE|A  VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTRKDDQEETVRKRLVEYHQMTAPLIG
        121     .      .      .      .      .      .      180

        181     .      .      .      214
pdb|1AKE|A  YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
        181     .      .      .      214

```

Call:

```
read.fasta(file = outfile)
```

Class:

```
fasta
```

Alignment dimensions:

```
1 sequence rows; 214 position columns (214 non-gap, 0 gap)
```

```
+ attr: id, ali, call
```

DB database for all related entries:

```
blast <- blast.pdb(aa)
```

Searching ... please wait (updates every 5 seconds) RID = SPYKXYWE014

.....

Reporting 96 hits

```
hits <- plot(blast)
```

```

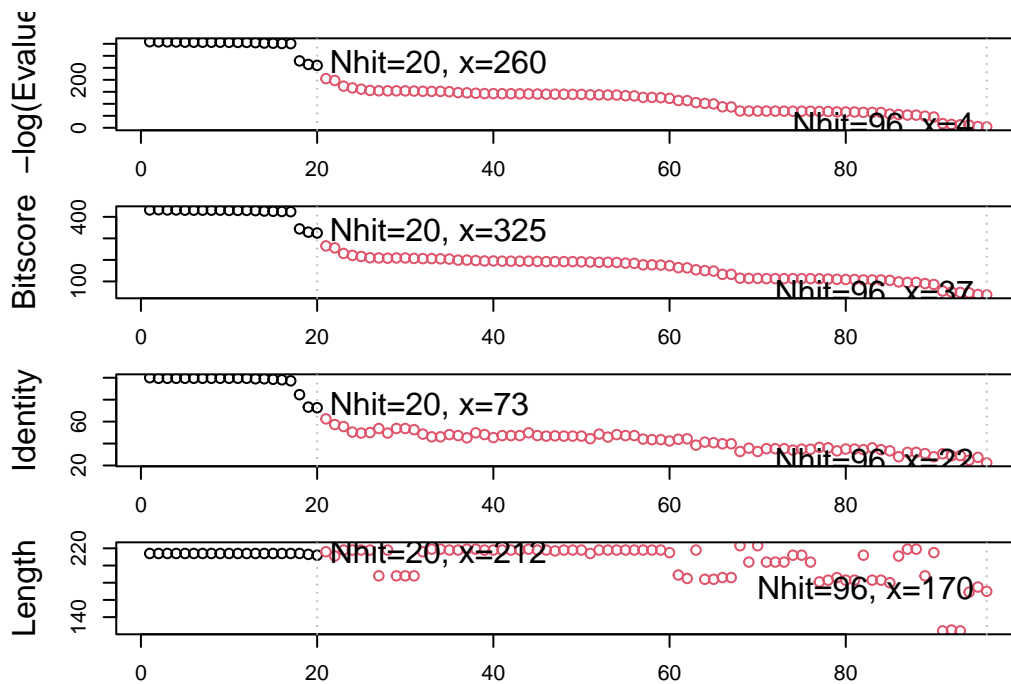
* Possible cutoff values:    260 3
    Yielding Nhits:         20 96

```

```

* Chosen cutoff value of:    260
    Yielding Nhits:         20

```



```
head(blast$hit.tbl)
```

	queryid	subjectids	identity	alignmentlength	mismatches	gapopens	q.start		
1	Query_6403663	1AKE_A	100.000	214	0	0	1		
2	Query_6403663	8BQF_A	99.533	214	1	0	1		
3	Query_6403663	4X8M_A	99.533	214	1	0	1		
4	Query_6403663	6S36_A	99.533	214	1	0	1		
5	Query_6403663	9R6U_A	99.533	214	1	0	1		
6	Query_6403663	9R71_A	99.533	214	1	0	1		
	q.end	s.start	s.end	evalue	bitscore	positives	mlog.evalue	pdb.id	acc
1	214	1	214	1.78e-156	432	100.00	358.6267	1AKE_A	1AKE_A
2	214	21	234	2.91e-156	433	100.00	358.1351	8BQF_A	8BQF_A
3	214	1	214	3.18e-156	432	100.00	358.0464	4X8M_A	4X8M_A
4	214	1	214	4.67e-156	432	100.00	357.6621	6S36_A	6S36_A
5	214	1	214	1.04e-155	431	99.53	356.8615	9R6U_A	9R6U_A
6	214	1	214	1.23e-155	431	99.53	356.6937	9R71_A	9R71_A

The “top hits” are in the `hits` object. Now we can download these to our computer. Put these in a sub-folder (directory) called `pdbs`

```
## Download related PDB files
```

```
files <- get.pdb(hits$pdb.id, path="pdb", split=TRUE, gzip=TRUE)
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/1AKE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/8BQF.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/4X8M.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/6S36.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/9R6U.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/9R71.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/8Q2B.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/8RJ9.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/6RZE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/4X8H.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/3HPR.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdb", split = TRUE, gzip = TRUE):  
pdb/1E4V.pdb.gz exists. Skipping download
```

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/5EJE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4Y.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3X2S.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAP.pdb.gz exists. Skipping download

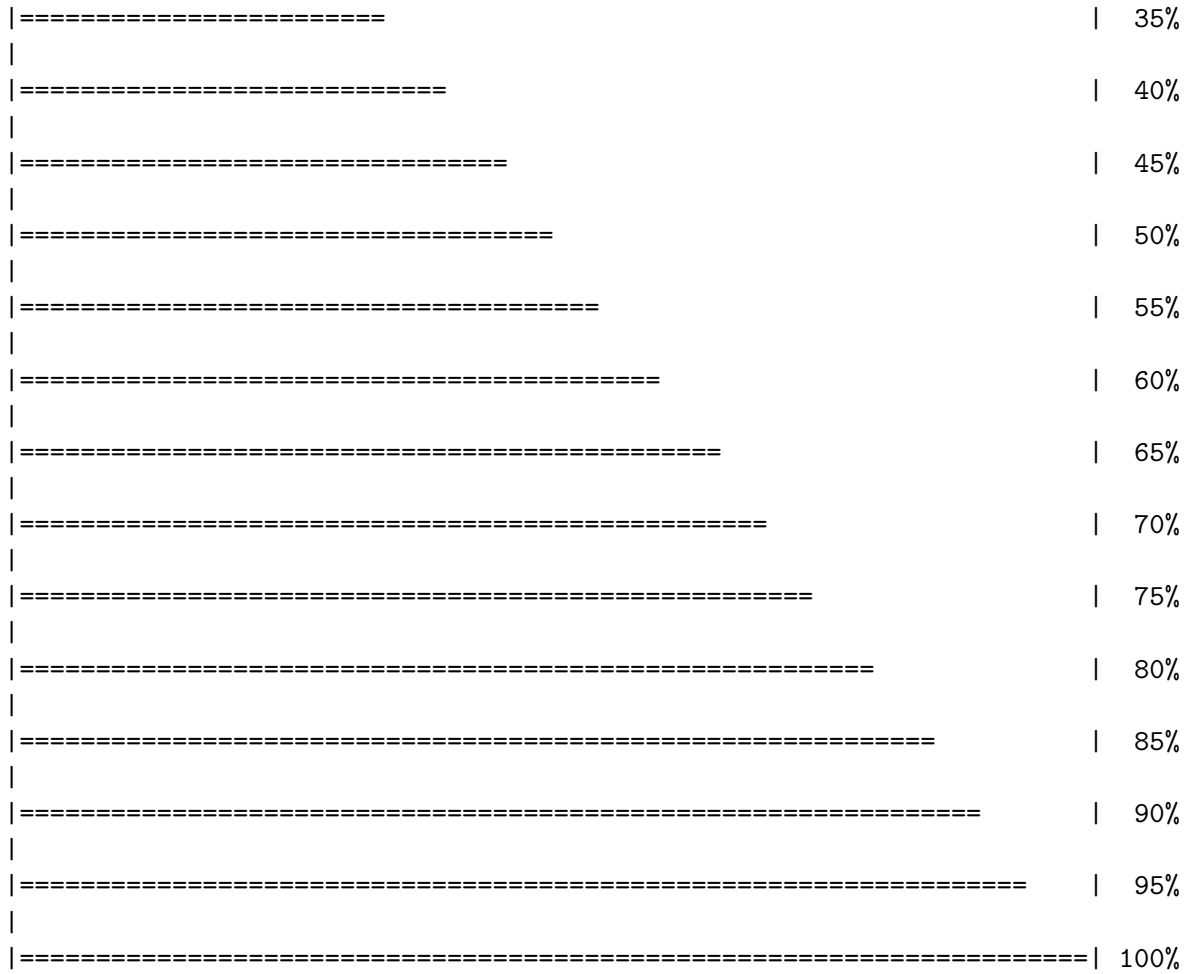
Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAM.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/8PVW.pdb.gz exists. Skipping download

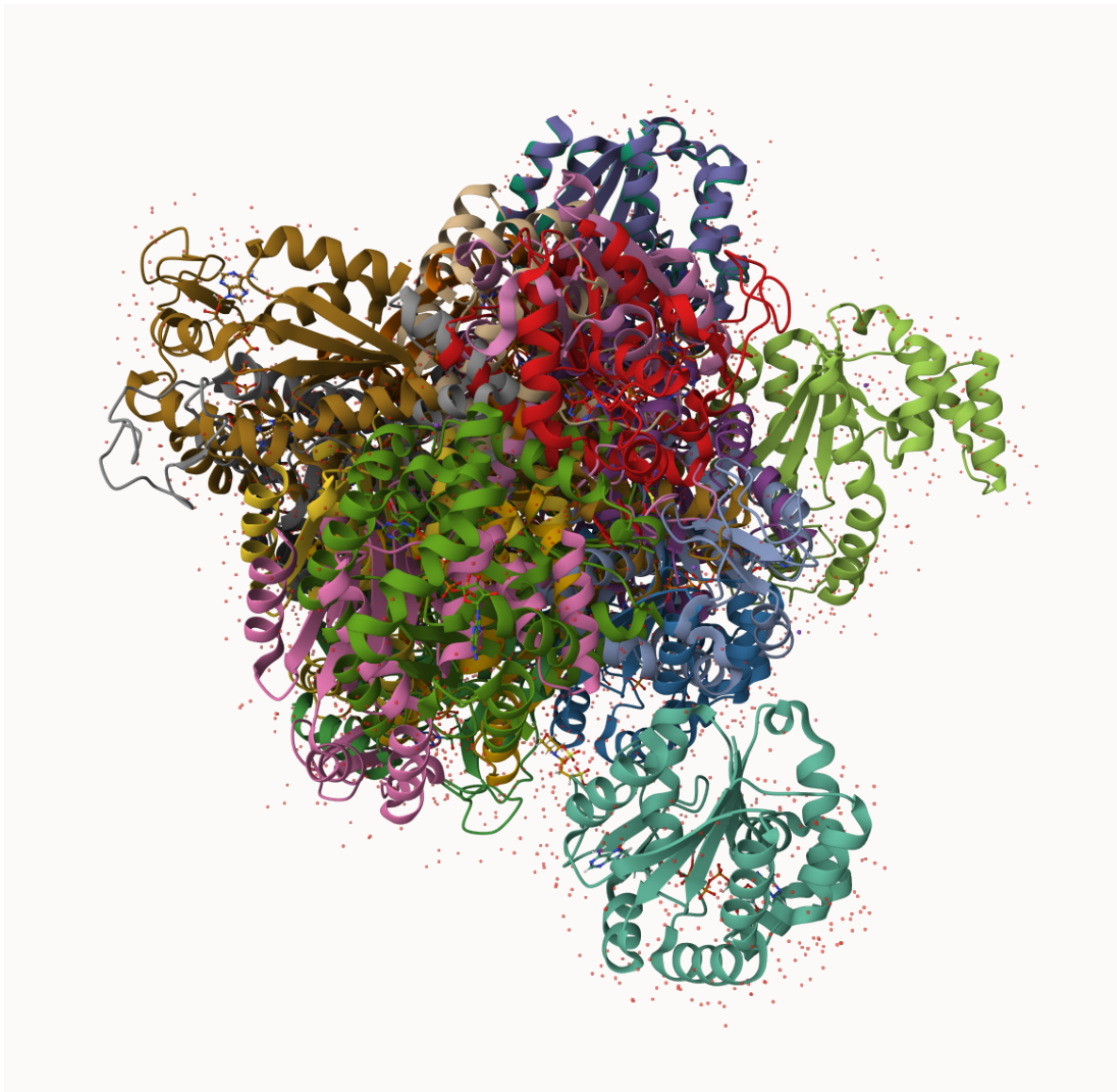
Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4K46.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4NP6.pdb.gz exists. Skipping download

	0%
====	5%
=====	10%
=====	15%
=====	20%
=====	25%
=====	30%



These look like a hot mess



Next we will use the `pdbsaln()` function to align and also superpose the PDB structure.

This requires a BioConductor package called “msa” that we need to install. First we install BiocManager. Then we use `BiocManager::install("msa")`

```
# Align related PDBs
pdbs <- pdbsaln(files, fit = TRUE, exefile="msa")
```

```
Reading PDB files:
pdbs/split_chain/1AKE_A.pdb
```

```

pdbs/split_chain/8BQF_A.pdb
pdbs/split_chain/4X8M_A.pdb
pdbs/split_chain/6S36_A.pdb
pdbs/split_chain/9R6U_A.pdb
pdbs/split_chain/9R71_A.pdb
pdbs/split_chain/8Q2B_A.pdb
pdbs/split_chain/8RJ9_A.pdb
pdbs/split_chain/6RZE_A.pdb
pdbs/split_chain/4X8H_A.pdb
pdbs/split_chain/3HPR_A.pdb
pdbs/split_chain/1E4V_A.pdb
pdbs/split_chain/5EJE_A.pdb
pdbs/split_chain/1E4Y_A.pdb
pdbs/split_chain/3X2S_A.pdb
pdbs/split_chain/6HAP_A.pdb
pdbs/split_chain/6HAM_A.pdb
pdbs/split_chain/8PVW_A.pdb
pdbs/split_chain/4K46_A.pdb
pdbs/split_chain/4NP6_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
..   PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
..   PDB has ALT records, taking A only, rm.alt=TRUE
..   PDB has ALT records, taking A only, rm.alt=TRUE
....  PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
..

```

Extracting sequences

```

pdb/seq: 1    name: pdbs/split_chain/1AKE_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 2    name: pdbs/split_chain/8BQF_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 3    name: pdbs/split_chain/4X8M_A.pdb
pdb/seq: 4    name: pdbs/split_chain/6S36_A.pdb
    PDB has ALT records, taking A only, rm.alt=TRUE

```

```

pdb/seq: 5    name: pdbc/split_chain/9R6U_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 6    name: pdbc/split_chain/9R71_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 7    name: pdbc/split_chain/8Q2B_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 8    name: pdbc/split_chain/8RJ9_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 9    name: pdbc/split_chain/6RZE_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 10   name: pdbc/split_chain/4X8H_A.pdb
pdb/seq: 11   name: pdbc/split_chain/3HPR_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 12   name: pdbc/split_chain/1E4V_A.pdb
pdb/seq: 13   name: pdbc/split_chain/5EJE_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 14   name: pdbc/split_chain/1E4Y_A.pdb
pdb/seq: 15   name: pdbc/split_chain/3X2S_A.pdb
pdb/seq: 16   name: pdbc/split_chain/6HAP_A.pdb
pdb/seq: 17   name: pdbc/split_chain/6HAM_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 18   name: pdbc/split_chain/8PVW_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 19   name: pdbc/split_chain/4K46_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 20   name: pdbc/split_chain/4NP6_A.pdb

```

Have a look at this new “alignment object” pdbc

pdbc

	1	.	.	.	40
[Truncated_Name:1] 1AKE_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAA				
[Truncated_Name:2] 8BQF_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAA				
[Truncated_Name:3] 4X8M_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAA				
[Truncated_Name:4] 6S36_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAA				
[Truncated_Name:5] 9R6U_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAA				
[Truncated_Name:6] 9R71_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAA				
[Truncated_Name:7] 8Q2B_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAA				
[Truncated_Name:8] 8RJ9_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAA				
[Truncated_Name:9] 6RZE_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAA				
[Truncated_Name:10] 4X8H_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAA				

[Truncated_Name:1]3HPR_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMDLRAA	1	.	.	.	40
[Truncated_Name:12]1E4V_A.pdb	--MRIILLGAPVAGKGTQAQFIMEKYGIPQISTGMDLRAA					
[Truncated_Name:13]5EJE_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMDLRAA					
[Truncated_Name:14]1E4Y_A.pdb	--MRIILLGALVAGKGTQAQFIMEKYGIPQISTGMDLRAA					
[Truncated_Name:15]3X2S_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMDLRAA					
[Truncated_Name:16]6HAP_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMDLRAA					
[Truncated_Name:17]6HAM_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMDLRAA					
[Truncated_Name:18]8PVW_A.pdb	--MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMDLRAA					
[Truncated_Name:19]4K46_A.pdb	--MRIILLGAPGAGKGTQAQFIMAKFGIPQISTGMDLRAA					
[Truncated_Name:20]4NP6_A.pdb	NAMRIILLGAPGAGKGTQAQFIMEKFGIPQISTGMDLRAA					
	***** ***** *~*****					
		41	.	.	.	80
[Truncated_Name:1]1AKE_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:2]8BQF_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQE---					
[Truncated_Name:3]4X8M_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:4]6S36_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:5]9R6U_A.pdb	VKSGSELGAQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:6]9R71_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:7]8Q2B_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:8]8RJ9_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:9]6RZE_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:10]4X8H_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:11]3HPR_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:12]1E4V_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:13]5EJE_A.pdb	VKSGSELGKQAKDIMDACKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:14]1E4Y_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:15]3X2S_A.pdb	VKSGSELGKQAKDIMDCGKLVTDDELVIALVKERIAQEDSR					
[Truncated_Name:16]6HAP_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVRERICQEDSR					
[Truncated_Name:17]6HAM_A.pdb	IKSGSELGKQAKDIMDAGKLVTDDEIIIALVKERICQEDSR					
[Truncated_Name:18]8PVW_A.pdb	VKSGSELGKQAKDIMDAGKLVTDDELVIALVKERIAQEDCR					
[Truncated_Name:19]4K46_A.pdb	IKAGTELGKQAKSVIDAGQLVSDDIILGLVKERIAQDDCA					
[Truncated_Name:20]4NP6_A.pdb	IKAGTELGKQAKAVIDAGQLVSDDIILGLIKERIAQADCE					
	~* *~*** *** ~* **~*~^~^~*~^*** *					
		41	.	.	.	80
		81	.	.	.	12
[Truncated_Name:1]1AKE_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVD					
[Truncated_Name:2]8BQF_A.pdb	-GFLLDGFPRTIPQADAMKEAGINVDYVIEFDVPDELIVD					
[Truncated_Name:3]4X8M_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVD					
[Truncated_Name:4]6S36_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVD					
[Truncated_Name:5]9R6U_A.pdb	NGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVD					

[Truncated_Name:6] 9R71_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDALIVD		
[Truncated_Name:7] 8Q2B_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:8] 8RJ9_A.pdb	NGFLLAGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:9] 6RZE_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:10] 4X8H_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:11] 3HPR_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:12] 1E4V_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:13] 5EJE_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:14] 1E4Y_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:15] 3X2S_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:16] 6HAP_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:17] 6HAM_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:18] 8PVW_A.pdb	NGFLLDGFPR TIPQADAMKEAGINVDYVLEFDVPDELIVD		
[Truncated_Name:19] 4K46_A.pdb	KGFLLDGFPR TIPQADGLKEVGVVVDYVIEFDVADSVIVE		
[Truncated_Name:20] 4NP6_A.pdb	KGFLLDGFPR TIPQADGLKEMGINVDYVIEFDVADDVIVE		
	**** *****^~** *^ ***** * ^**^		
	81 . . . 120		
	121 . . . 160		
[Truncated_Name:1] 1AKE_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:2] 8BQF_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:3] 4X8M_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:4] 6S36_A.pdb	KIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:5] 9R6U_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:6] 9R71_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:7] 8Q2B_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKA		
[Truncated_Name:8] 8RJ9_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:9] 6RZE_A.pdb	AIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:10] 4X8H_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:11] 3HPR_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDGTGEELTTRKD		
[Truncated_Name:12] 1E4V_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:13] 5EJE_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:14] 1E4Y_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:15] 3X2S_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:16] 6HAP_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:17] 6HAM_A.pdb	RIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTGEELTTRKD		
[Truncated_Name:18] 8PVW_A.pdb	RILKR--GETSGRV-----D		
[Truncated_Name:19] 4K46_A.pdb	RMAGRRRAHLASGR TYHNVNPPKVEGKDDVTGEDLVIRE		
[Truncated_Name:20] 4NP6_A.pdb	RMAGRRRAHLPSGR TYHVVNPPKVEGKDDVTGEDLVIRE		
	^ * ***		
	121 . . . 160		
	161 . . . 200		

[Truncated_Name:1] 1AKE_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:2] 8BQF_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:3] 4X8M_A.pdb	DQEETVRKRLVEWHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:4] 6S36_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:5] 9R6U_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:6] 9R71_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:7] 8Q2B_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:8] 8RJ9_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:9] 6RZE_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:10] 4X8H_A.pdb	DQEETVRKRLVEYHQM TAA LIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:11] 3HPR_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:12] 1E4V_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:13] 5EJE_A.pdb	DQEECVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:14] 1E4Y_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:15] 3X2S_A.pdb	DQEETVRKRLCEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:16] 6HAP_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:17] 6HAM_A.pdb	DQEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:18] 8PVW_A.pdb	DNEETVRKRLVEYHQM TAPLIGYYSKEAEAGNTKYAKVDG
[Truncated_Name:19] 4K46_A.pdb	DKEETVLARLGVYHNQ TAPLIAYYGKEAEAGNTQYLKFDG
[Truncated_Name:20] 4NP6_A.pdb	DKEETVRARLNVYHTQ TAPLIEYYGKEAAAGKTQYLKFDG
	* * * * * ^ * * * * * * * * * *
161	. . . 200

	201	216
[Truncated_Name:1] 1AKE_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:2] 8BQF_A.pdb	TKPVAEVRADLEKIL-	
[Truncated_Name:3] 4X8M_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:4] 6S36_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:5] 9R6U_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:6] 9R71_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:7] 8Q2B_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:8] 8RJ9_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:9] 6RZE_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:10] 4X8H_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:11] 3HPR_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:12] 1E4V_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:13] 5EJE_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:14] 1E4Y_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:15] 3X2S_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:16] 6HAP_A.pdb	TKPVCEVRADLEKILG	
[Truncated_Name:17] 6HAM_A.pdb	TKPVCEVRADLEKILG	
[Truncated_Name:18] 8PVW_A.pdb	TKPVAEVRADLEKILG	
[Truncated_Name:19] 4K46_A.pdb	TKA VA EVS AELEKALA	

```
[Truncated_Name:20]4NP6_A.pdb    TKQVSEVSADIAKALA
                                ** * ** *~* *
                                201      .      216
```

Call:

```
pdbaln(files = files, fit = TRUE, exefile = "msa")
```

Class:

```
pdb, fasta
```

Alignment dimensions:

```
20 sequence rows; 216 position columns (182 non-gap, 34 gap)
```

```
+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

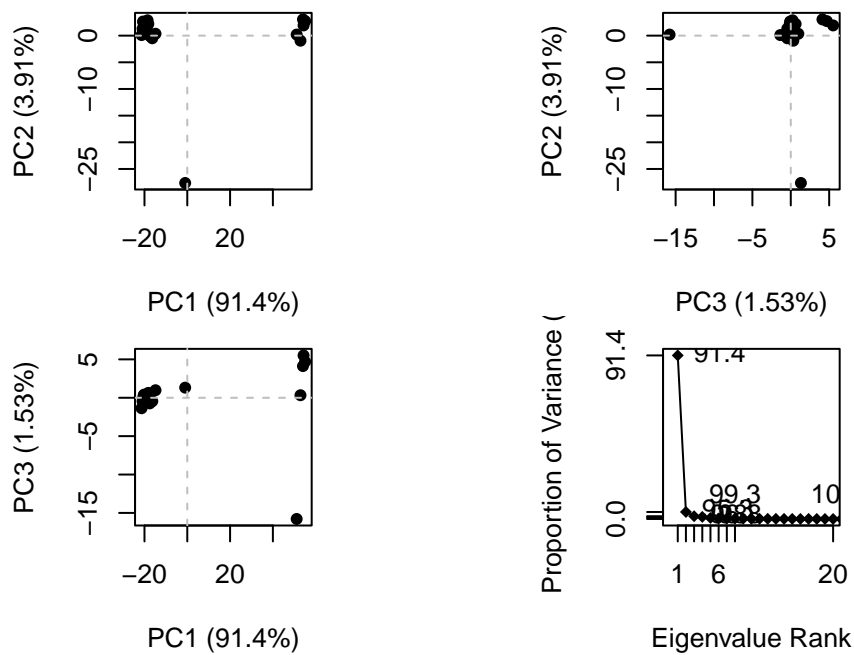
We could view these in R with **bio3dview** `view.pdb()` function.

```
# library(bio3dview)
# view.pdb(pdb, colorScheme = "residue")
```

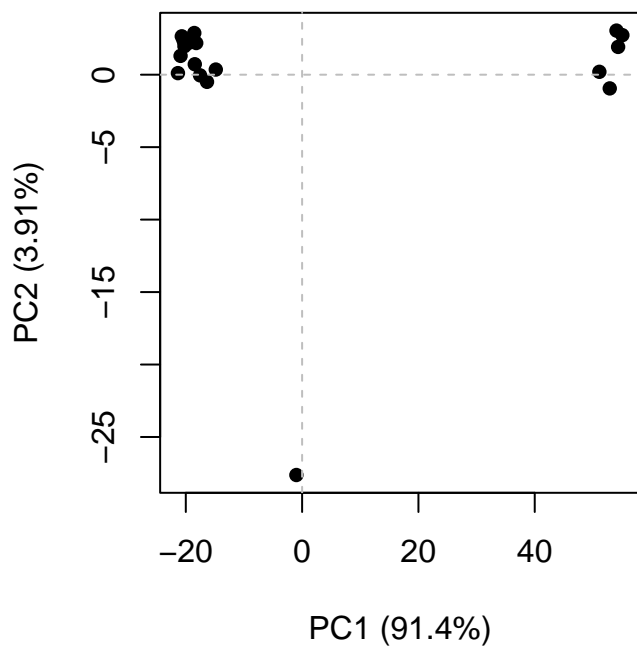
PCA

We can run PCA on our `pdb` object using the `pca()` function from **bio3d**

```
pc.xray <- pca(pdb)
plot(pc.xray)
```



```
plot(pc.xray,1:2)
```



We can make a visualization of the major conformational difference (i.e. large scale structure change) captured by our PCA analysis with `mktrj()` function.

```
pc1 <- mktbj(pc.xray, pc=1, file="pc_1.pdb")
```

Let's see in Molstar