

Smooth modeling of covariate effects in bisulfite sequencing-derived measures of DNA methylation

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Overview



Please feel free to interrupt and ask questions at any time during the talk!

- ▶ Background and motivation
- ▶ †New method 1: SOMNiBUS (SmOoth ModeliNg of BisUlfite Sequencing)
- ▶ ‡New method 2: dSOMNiBUS (dispersion-adjusted SmOoth ModeliNg of BisUlfite Sequencing)
- ▶ *New method 3: sparseSOMNiBUS (SOMNiBUS with variable selection)

† Zhao, et.al (2020). A novel statistical method for modeling covariate effects in bisulfite sequencing derived measures of DNA methylation. *Biometrics*. Early-View

‡ Zhao, et.al (2020+). Detecting differentially methylated regions in bisulfite sequencing data using quasi-binomial mixed models with smooth covariate effect estimates.

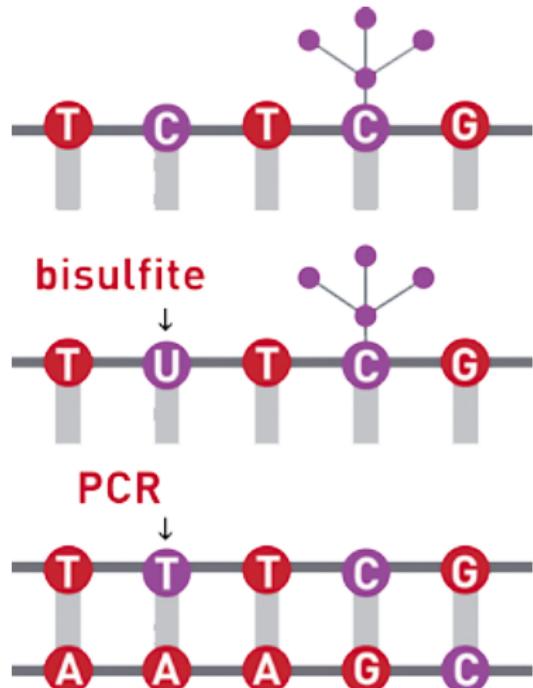
* Zhao, et.al (2020+). In preparation

Epigenetics and DNA Methylation



- ▶ change gene expression without changing DNA sequence
- ▶ can be altered by age, diet, stress and environmental exposures
- ▶ Localized abnormal methylation is a characteristic feature of many diseases

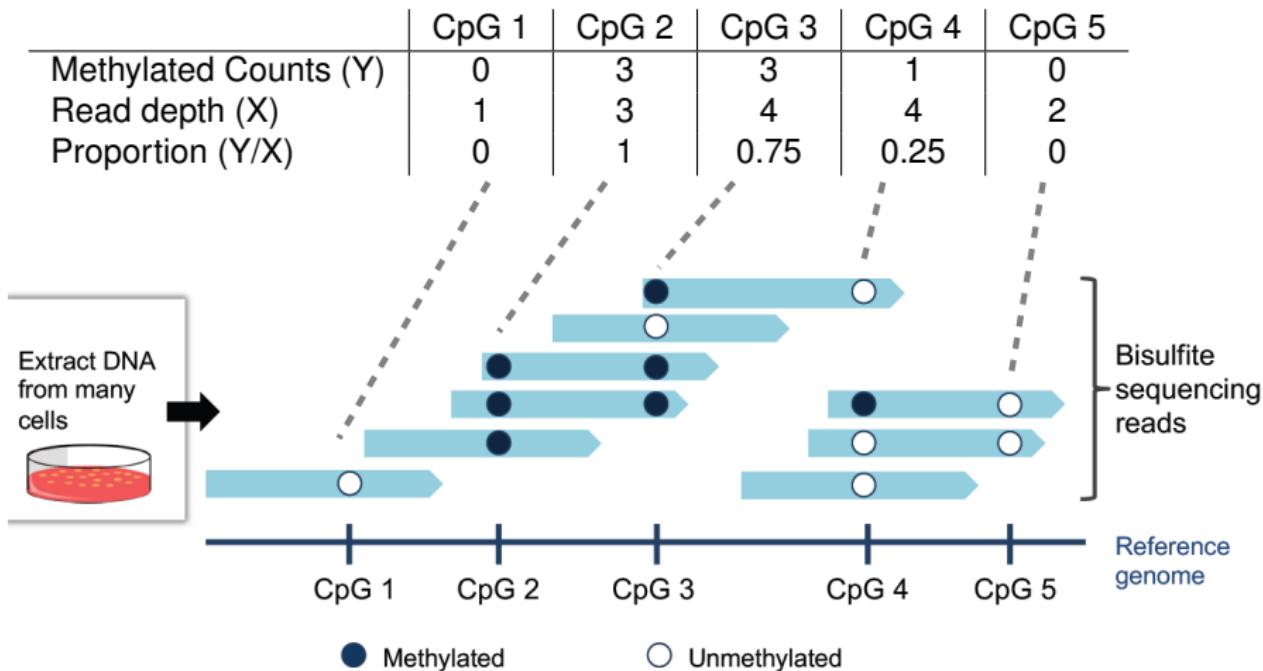
Bisulfite Sequencing & Methylation



Methylated cytosines are not converted by bisulfite treatment

<https://www.diagenode.com/en/applications/dna-bisulfite-conversion>

Sequencing-derived DNA methylation data



http://kkorthauer.org/talks/korthauer_aisc_2018_static.pdf

Motivating datasets

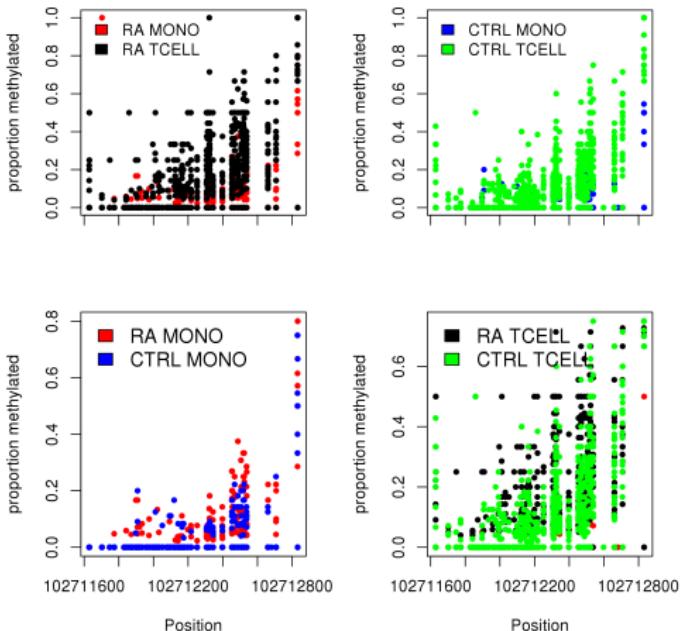
Methylation profiles of Rheumatoid Arthritis (RA) patients and controls
(from our collaborator Dr. Marie Hudson)



- ▶ Targeted Custom Capture Bisulfite Sequencing
 - predefined genomic regions
 - 5 million CpGs
- ▶ Cell-separated blood samples

	Monocytes	T cells
RA	10	12
Controls	8	13

- ▶ Small region on chromosome 4 near *BANK1*
- ▶ 123 CpGs



Goal



Find associations between

- ▶ methylation patterns in each targeted region, and
- ▶ phenotypes or covariates

Challenges / Opportunities



- Read depth at CpGs varies substantially
 - ▶ Need a model that can use all available data
- Cell-type mixture affects observed methylation levels
 - ▶ Adjust for this in model
- Sequencing errors, e.g. bisulfite conversion error
 - ▶ Build a model allowing for error
- Local correlations in methylation levels
 - ▶ Opportunity for imputing missing data or poorly measured signals
 - ▶ Opportunity for modelling smooth effects along the genome

Existing methods appropriate for regions



Method	regional	one-stage	count-based	read-depth variability	adjust for confounding	experimental errors
SOMNiBUS	✓	✓	✓	✓	✓	✓
BSmooth	✓			✗		
SMSC	✓			✗		✓
dmrseq	✓			✓	✓	
Biseq	✓			✗	✓	
GlobalTest	✓	✓			✓	

BSmooth: Hansen, 2012

SMSC: Lakhal-Chaieb, 2017

dmrseq: Korthauer, 2018

BiSeq: Hebestreit, 2013

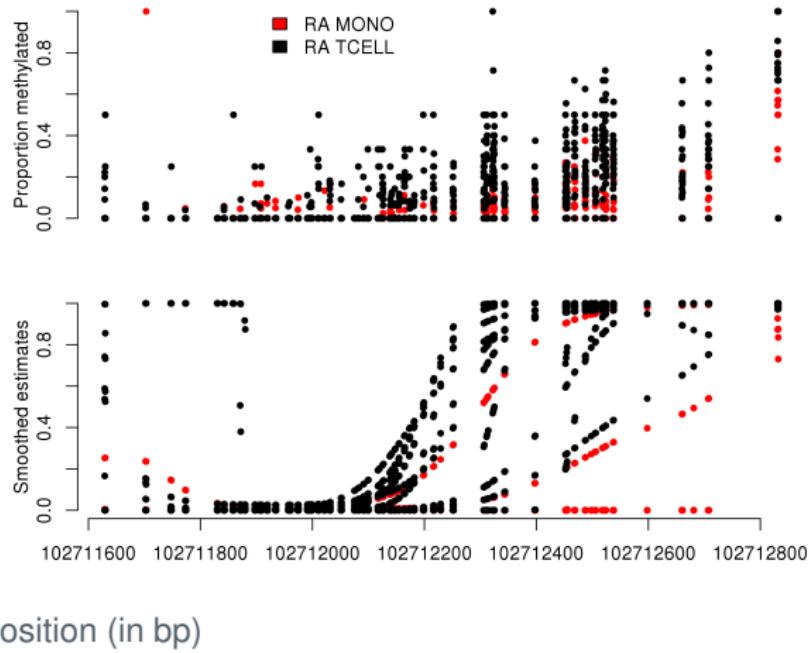
GlobalTest: Goeman, 2006

An example of two-stage method

Raw data & per-sample smoothed estimates



Results from SMSC (Lakhal-Chaieb, 2017)





Method	regional	one-stage	count-based	read-depth variability	adjust for confounding	experimental errors
SOMNiBUS	✓	✓	✓	✓	✓	✓
BSmooth	✓			✗		
SMSC	✓			✗		✓
dmrseq	✓			✓	✓	
Biseq	✓			✗	✓	
GlobalTest	✓	✓			✓	

Motivation: a novel **one-stage** method that

- ▶ collapses smoothing and testing steps into a single step
- ▶ allows for experimental errors, variable read depths and test samples with a mixture of cell types
- ▶ provides **rigorous uncertainty assessment** for differentially methylated regions

Overview



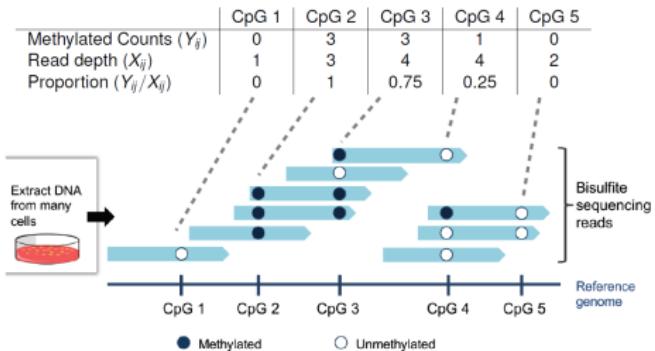
- ▶ Background and motivation
- ▶ [†]**New method 1: SOMNiBUS (SmOoth ModeliNg of BisUlfite Sequencing)**
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[‡] Zhao, et.al (2020+). Detecting differentially methylated regions in bisulfite sequencing data using quasi-binomial mixed models with smooth covariate effect estimates.

Notations

- ▶ X_{ij} : total number of reads aligned to CpG j from sample i
- ▶ Y_{ij} : **observed** methylated counts at CpG j for sample i . $Y_{ij} = \sum_{k=1}^{X_{ij}} Y_{ijk}$
- ▶ S_{ij} : **true** methylated counts at CpG j for sample i . $S_{ij} = \sum_{k=1}^{X_{ij}} S_{ijk}$



- ▶ t_{ij} : the genome position (in bp) for sample i at CpG j
- ▶ $Z_{1i}, Z_{2i}, \dots, Z_{Pi}$ are the P covariates.
- ▶ π_{ij} : the methylation proportion parameter for sample i , CpG j

SOMNiBUS[†]: Model

- ▶ Assume **known error parameters** p_0 and p_1 ,

$$\begin{aligned} p_0 &= \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 0) \\ p_1 &= \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 1). \end{aligned}$$

- ▶ Specify the model

$$\begin{aligned} S_{ij} \mid \mathbf{Z}_I, X_{ij} &\sim \text{Binomial}(X_{ij}, \pi_{ij}) \\ \log \left\{ \frac{\pi_{ij}}{1 - \pi_{ij}} \right\} &= \beta_0(t_{ij}) + \beta_1(t_{ij})Z_{1i} + \beta_2(t_{ij})Z_{2i} + \dots + \beta_P(t_{ij})Z_{Pi}, \end{aligned}$$

- ▶ Smooth curves along the genome for
 - ▶ Overall methylation
 - ▶ Covariate effects

[†]R package: <https://github.com/kaiqiong/SOMNiBUS>.

Technical details 1: splines

- ▶ Use splines for smoothing

$$\beta_p(t_{ij}) = \sum_{l=1}^{L_p} \alpha_{pl} B_l^{(p)}(t_{ij}) \text{ for } p = 0, 1, \dots, P.$$

- ▶ Penalize roughness of effect curves $\beta_p(t_{ij})$.

$$\mathcal{L}^{\text{Penalization}} = \sum_{p=0}^P \lambda_p \int (\beta_p''(t))^2 dt = \sum_{p=0}^P \lambda_p \alpha_p^T \mathbf{A}_p \alpha_p = \alpha^T \mathbf{A}_\lambda \alpha,$$

$\{\lambda_0, \lambda_1, \dots, \lambda_P\}$ are the smoothing parameters.

- ▶ Penalties go onto second derivatives
- ▶ $P + 1$ penalization parameters for P covariates

Technical details 2: E-M algorithm

E step: Calculate $\eta_{ij}^* = \mathbb{E}(S_{ij} \mid Y_{ijk}; \alpha^*)$

M step: \ddagger Maximize $Q(\alpha, \lambda \mid \alpha^*) = l(\eta^*; \alpha) - \frac{1}{2}\alpha^T A_\lambda \alpha + \frac{1}{2} \log \{|A_\lambda|_+\}$

- ▶ Estimate α given the value of λ : P-IRLS

$$\hat{\alpha}_\lambda = \operatorname{argmax}_\alpha \left\{ l(\eta^*; \alpha) - \frac{1}{2}\alpha^T A_\lambda \alpha \right\}$$

- ▶ Estimate λ : maximize the Laplace-approximated restrictive (or marginal) likelihood

$$L^M(\lambda) = \int \exp \{ Q(\alpha, \lambda \mid \alpha^*) \} d\alpha \approx \text{Laplace}(\lambda; \hat{\alpha}_\lambda).$$

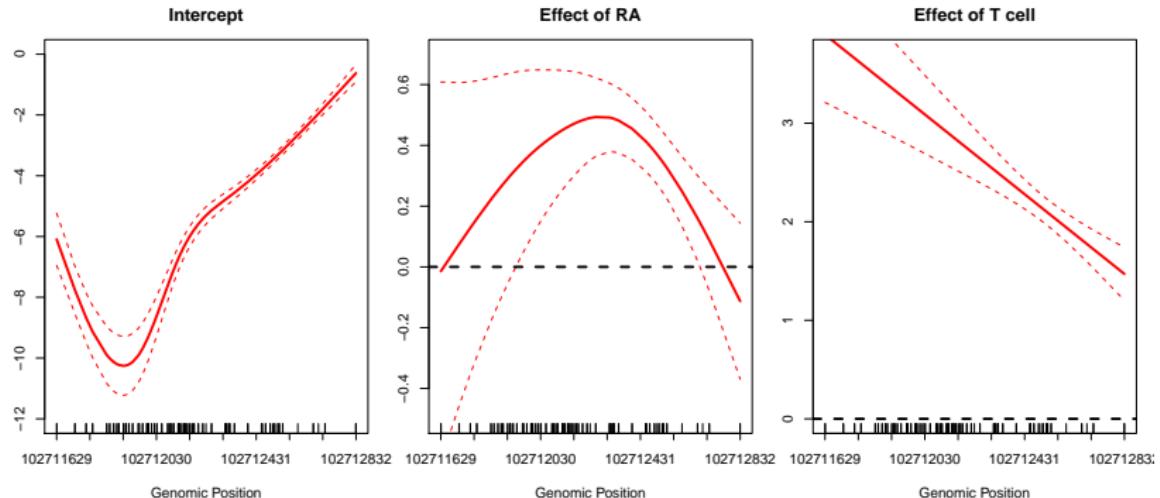
\ddagger Wood (2011), JRSSB; R package mgcv

Technical details 3: inference



- ▶ Pointwise confidence intervals
- ▶ Regional tests for non-zero covariate effects
 - ▶ for each covariate, or
 - ▶ for the combined effects of multiple covariates
- ▶ Penalization affects effective degree of freedom

Results in *BANK1* region



$$p = 1.11e - 16$$

$$p = 6.37e - 218$$

- Error parameters $p_0 = 0.003$ and $1 - p_1 = 0.1^{\ddagger}$

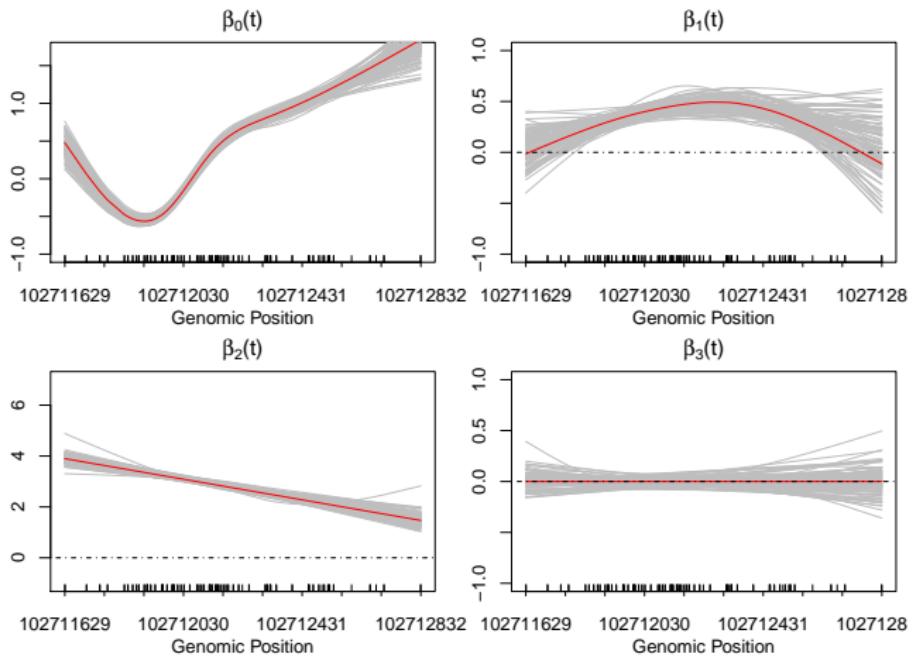
[†] Prochenka et al. (2015) *Bioinformatics*.

Simulation study

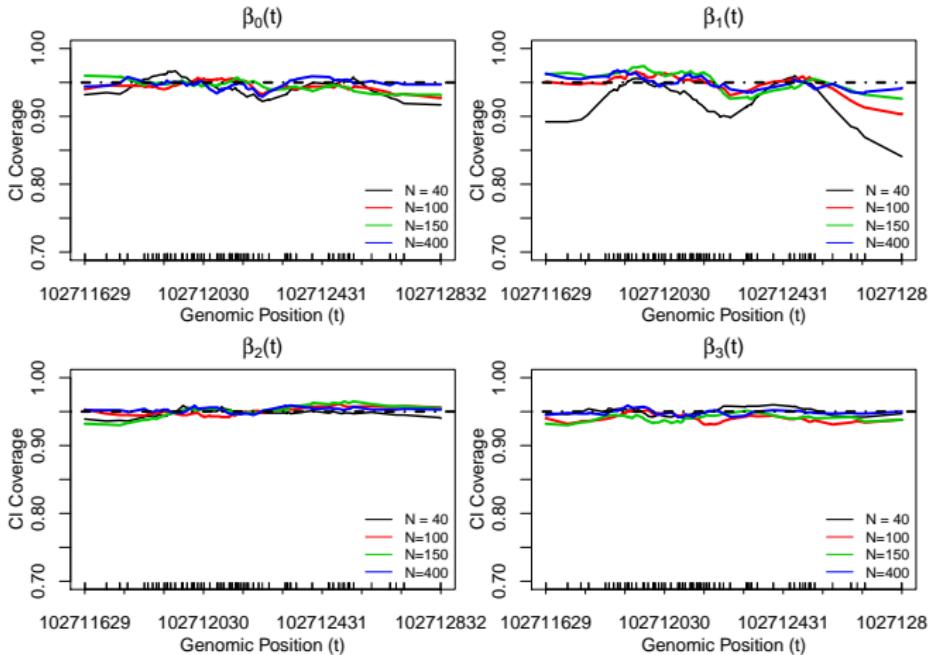
- ▶ Simulated dataset similar to the *BANK1* example
- ▶ One “null” covariate with no effect
- ▶ Two covariates with effects like those seen near *BANK1*
- ▶ Simulate the observed methylated counts Y_{ij} from

$$Y_{ij} \mid S_{ij} \sim \text{Binomial}(S_{ij}, p_1) + \text{Binomial}(X_{ij} - S_{ij}, p_0).$$

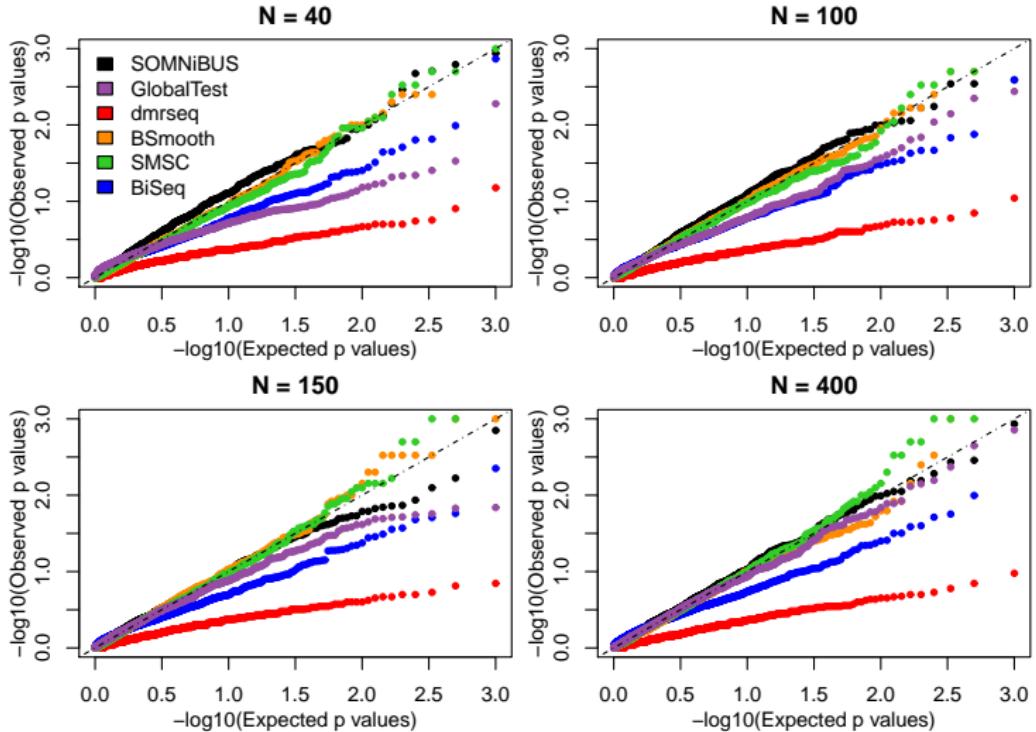
Little bias in the curve estimates



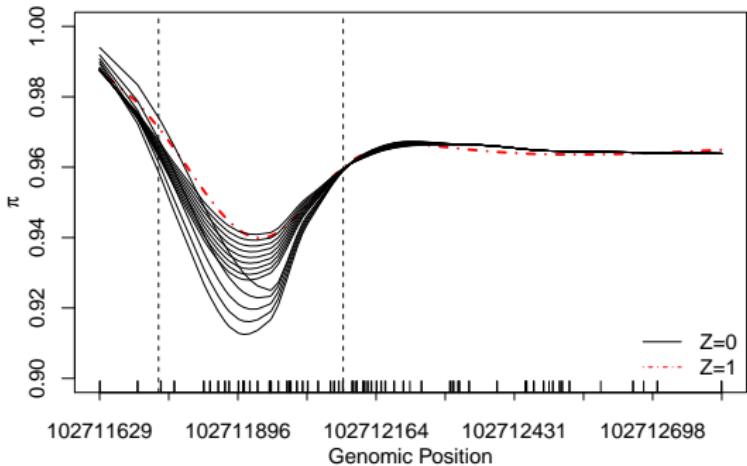
Empirical confidence interval coverages



Accurate type I error rates



Simulation to evaluate power

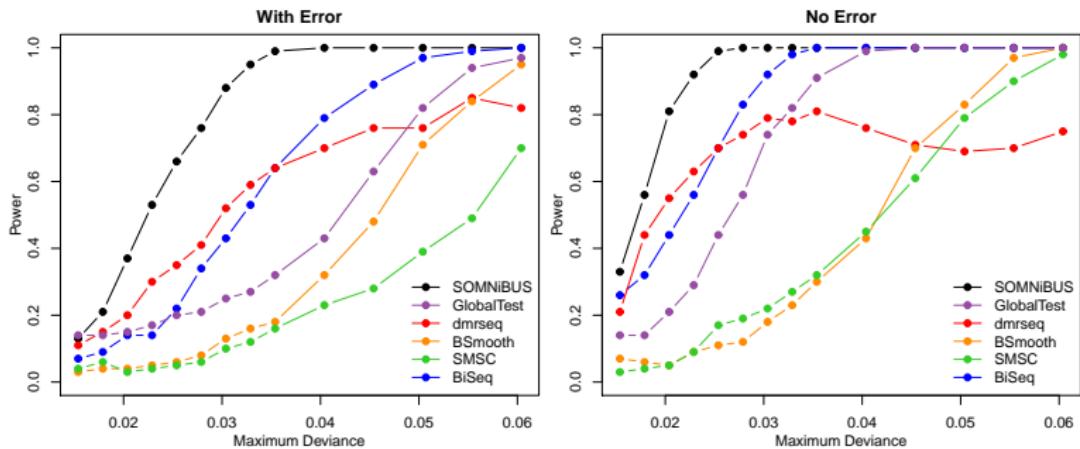


$Z = 1$ curve in red (fixed)

$Z = 0$ curve varied to give various sizes of differences

Increased power to detect DMRs

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Maximum difference between curves

- With Error: $p_0 = 0.003, p_1 = 0.9$
- No Error: $p_0 = 0, p_1 = 1$



Advantages

- ▶ Able to use data from many more CpGs where univariate analysis fails / power gain
- ▶ One-stage nature
- ▶ Explicitly allows for experimental errors
- ▶ Inference!



Advantages

- ▶ Able to use data from many more CpGs where univariate analysis fails / power gain
- ▶ One-stage nature
- ▶ Explicitly allows for experimental errors
- ▶ Inference!

Room for improvements

- ▶ Its underlying binomial assumption may be overly restrictive
- ▶ It is only applicable for data with negligible (within-group) variability (such as data from inbred animal or cell line experiments)

Overview



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Motivating datasets

(from our collaborator Dr. Sasha Bernatsky)



- ▶ CARTaGENE is an ongoing population-based cohort, including ~43,000 participants aged 40 to 69 years in Quebec
- ▶ The level of anti-citrullinated protein antibodies (ACPA) is a marker of rheumatoid arthritis (RA) risk that often presents prior to any clinical manifestations
- ▶ **Aim:** detect differentially methylated regions (DMRs) associated with ACPA

Motivating datasets

(from our collaborator Dr. Sasha Bernatsky)

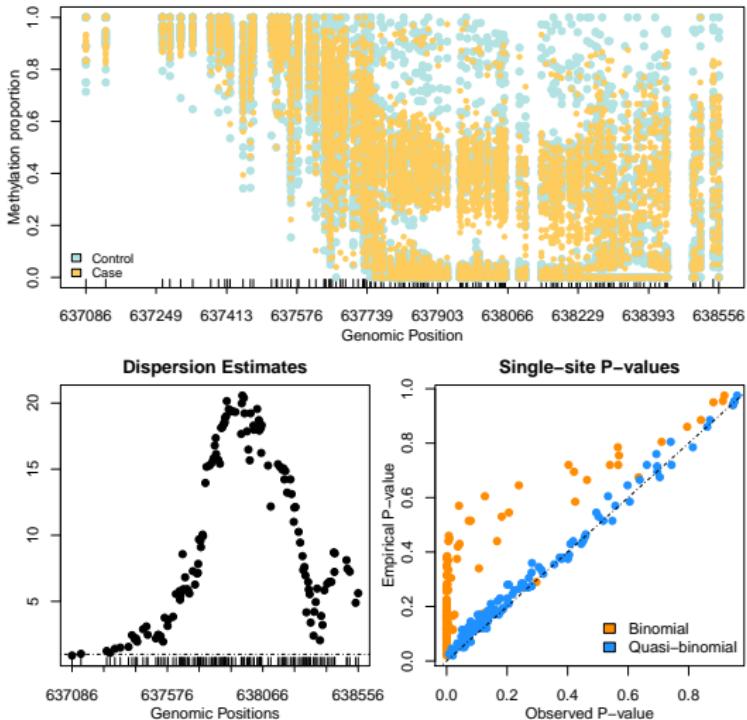


- ▶ blood samples of ACPA positive and ACPA negative subjects
 - **covariate of primary interest:** ACPA status
 - **adjusting variables:** age, sex, smoking status and cell type composition(captured by the top 4 PCs)
- ▶ two batches of data, referred to as data 1 and data 2, were collected in 2017 and 2019, respectively.

	data 1 (N = 116)	data 2 (N = 102)
ACPA Positives	55	48
ACPA Negatives	61	54
Number of targeted regions (with at least 50 CpGs)	10,759	12,985

Observed dispersion in a targeted region

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New method 2: dSOMNiBUS

(dispersion-adjusted SmOoth ModeliNg of BisUlfite Sequencing)

- ▶ The same error model

$$\begin{aligned} p_0 &= \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 0) \\ p_1 &= \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 1). \end{aligned}$$

- ▶ A quasi-binomial mixed model with the **combination** of
 - a *multiplicative* dispersion, ϕ
 - an *additive* dispersion, \mathbf{u} , (i.e. a subject-specific RE)

$$\log \frac{\pi_{ij}}{1 - \pi_{ij}} = \beta_0(t_{ij}) + \beta_1(t_{ij})Z_{1i} + \beta_2(t_{ij})Z_{2i} + \dots + \beta_P(t_{ij})Z_{Pi} + u_i,$$
$$u_i \stackrel{iid}{\sim} N(0, \sigma_0^2)$$

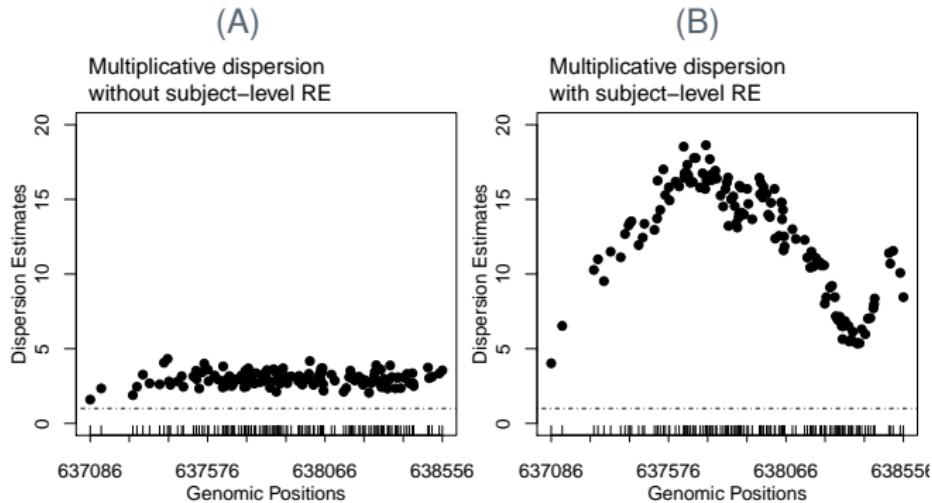
$$\text{Var}(S_{ij} \mid u_i) = \phi X_{ij} \pi_{ij} (1 - \pi_{ij})$$

- ▶ Smoothness parameters to penalize the roughness of effect curves.

R package: <https://github.com/kaiqiong/SOMNiBUS>

RE term enables flexible dispersion patterns in a region

A byproduct of introducing a subject-level RE to a model with smooth covariate effects is a regional dispersion pattern of varying degree.



$$\text{Var}(S_{ij}) \approx X_{ij}\pi_{ij}^*(1 - \pi_{ij}^*) \left\{ \phi + \sigma_0^2 (X_{ij} - \phi) \pi_{ij}^*(1 - \pi_{ij}^*) \right\}$$

Technical details 1: difficulties



- ▶ Three sets of unknown parameters
 - conditional mean parameters (REs): $\mathcal{B} = (\boldsymbol{\alpha}, \mathbf{u}) \in \mathbb{R}^{N+\sum_0^P L_p}$
 - variance component parameters: $\Theta = (\lambda, \sigma_0^2) \in \mathbb{R}^{P+2}$
 - **multiplicative dispersion parameter:** ϕ
- ▶ The conditional ‘distribution’ of $\mathbf{S} | \mathcal{B}$ is not available
- ▶ Joint estimation of ϕ and Θ is required, as $\text{Laplace}(\phi, \Theta; \widehat{\mathcal{B}}) \neq f(\phi)g(\Theta)$
- ▶ In the presence of data errors, one cannot easily estimate ϕ using the EM algorithm
 - The estimating equation for ϕ is not linear in the unknown methylated counts \mathbf{S}

Technical details 2: Estimation & Inference

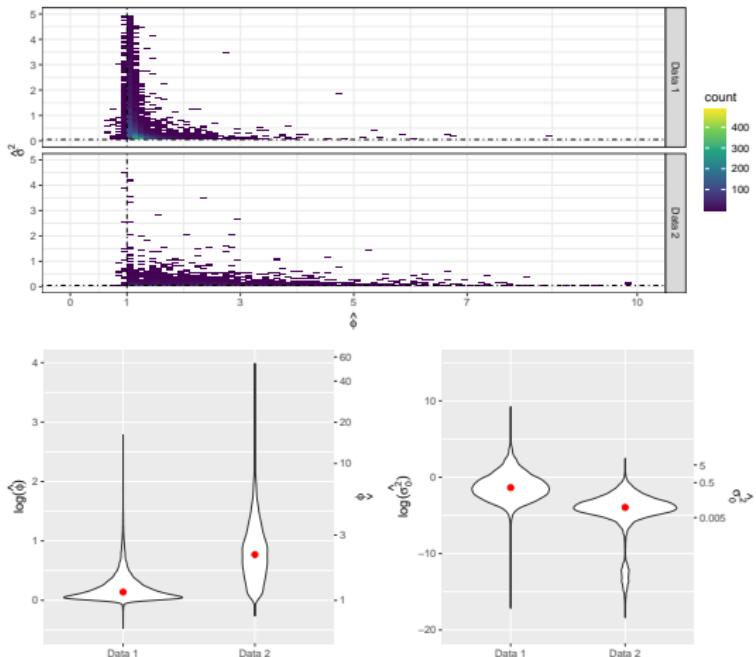


- ▶ Use the notation of **extended quasi-likelihood** to write the conditional quasi-likelihood function
- ▶ Calculate **Laplace-approximated marginal quais-likelihood** function and its derivatives
- ▶ **A hybrid ES algorithm**
 - A plug-in estimator for ϕ by exploiting its relationship with the dispersion for the contaminated outcome \mathbf{Y}
 - Estimate \mathcal{B} and Θ using ES iterations[†] assuming ϕ is fixed and known
- ▶ Inference using the observed **quasi-Fisher information**

[†] Efron (1986), Jorgensen (1987), McCullagh and Nelder (1989)

[‡] Elashoff and Ryan (2004)

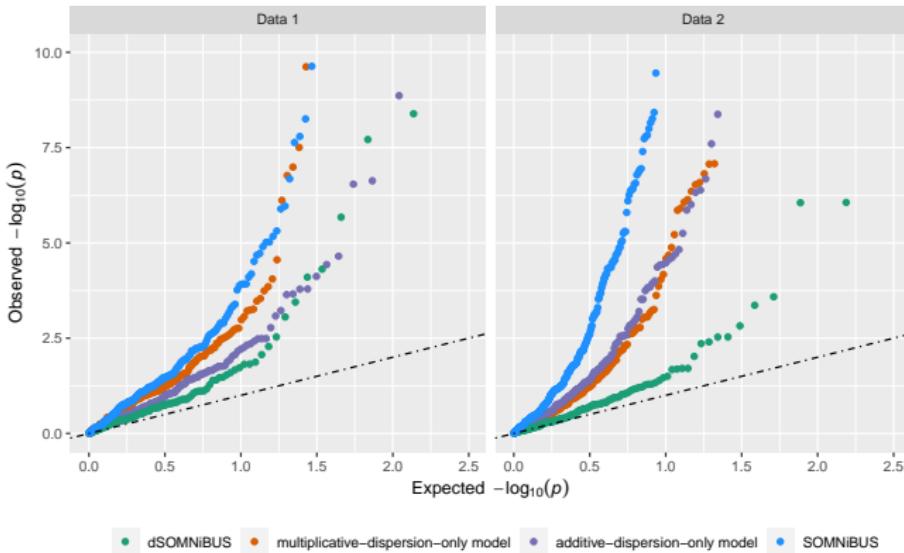
Both additive and multiplicative dispersion is present in the data



The distribution of estimated $\hat{\phi}$ and $\hat{\sigma}_0^2$ for the 10,759 and 12,985 regions in dataset 1 and 2, respectively.

Ignoring either type of dispersion leads to inflated type I errors

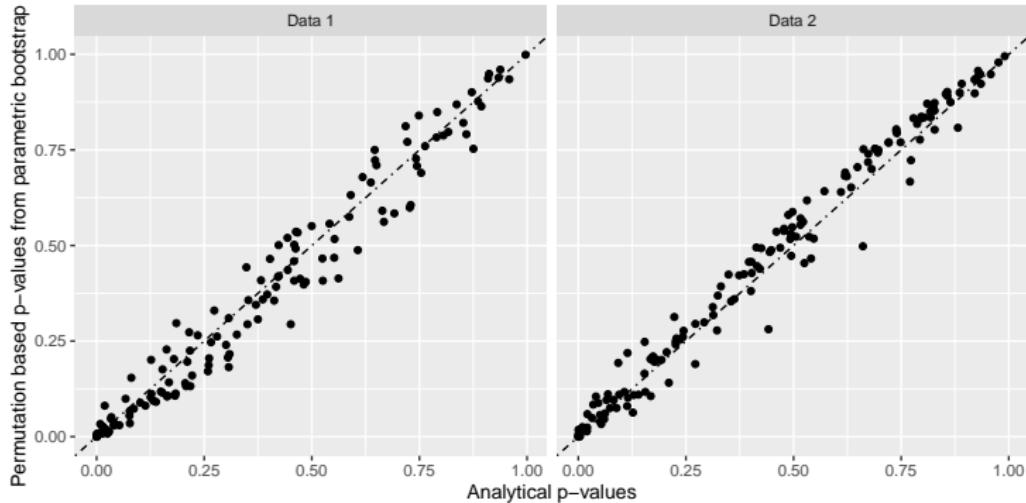
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dSOMNiBUS: $\phi > 0, \sigma_0^2 > 0$; multiplicative-dispersion-only model: $\phi > 0, \sigma_0^2 = 0$

SOMNiBUS: $\phi = 1, \sigma_0^2 = 0$; additive-dispersion-only model: $\phi = 1, \sigma_0^2 > 0$

Analytical v.s. bootstrap based p-values



Our inference procedure provides well-calibrated regional p-values.

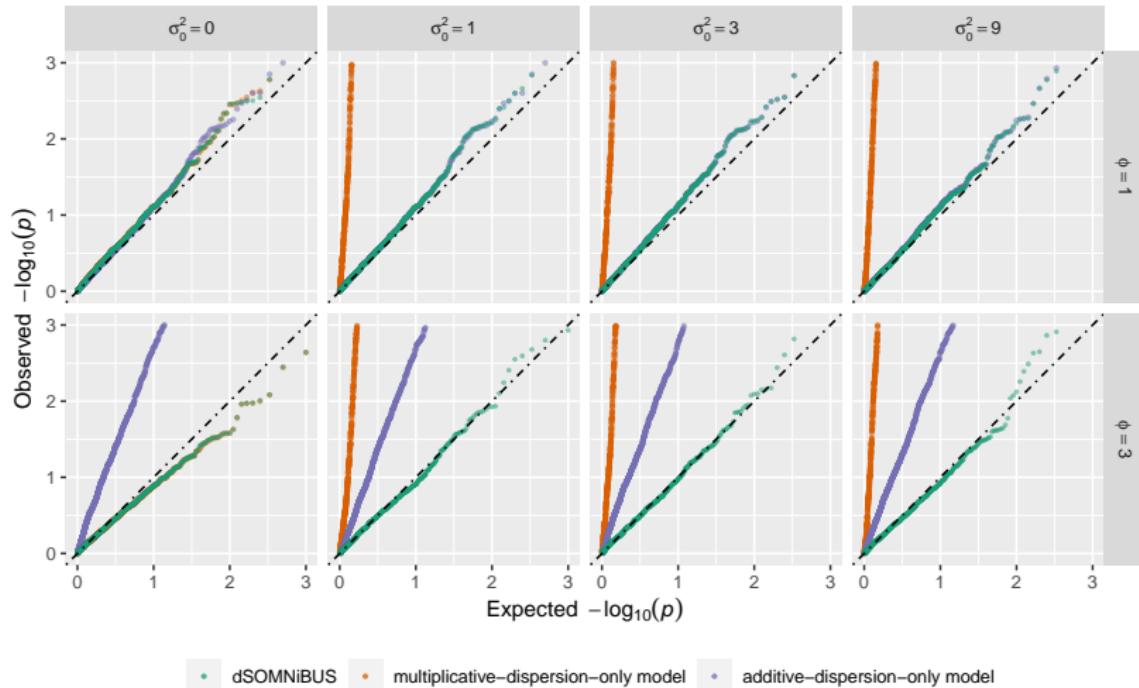
Simulation

- ▶ Specify the same $\beta_p(t)$ and Z_p as paper 1.
- ▶ $S_{ij} \sim \text{Beta-binomial} \left(\mu_{ij} = \pi_{ij}, \rho_{ij} = \frac{\phi - 1}{X_{ij} - 1}, \text{size} = X_{ij} \right)$
- ▶ In this way, we can always guarantee $\frac{\text{Var}(S_{ij})}{X_{ij}\pi_{ij}(1 - \pi_{ij})} \equiv \phi$.
- ▶ Recall: If $S \sim \text{Beta-binomial}(\mu, \rho, \text{size} = X)$,

$$\text{Var}(S) = \underbrace{[1 + (X - 1)\rho]}_{\text{dispersion}} \underbrace{X\mu(1 - \mu)}_{V(\mathbb{E}(Y))}.$$

The impact of dispersion

$$p_0 = 0.003, p_1 = 0.9$$

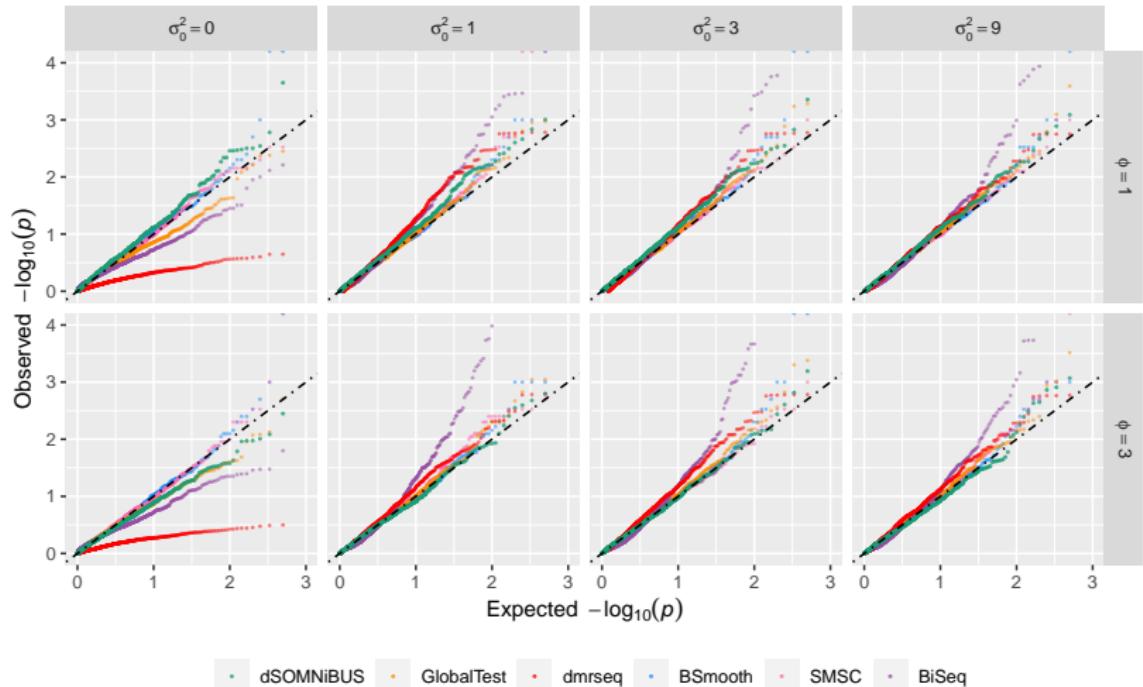


Type I Error

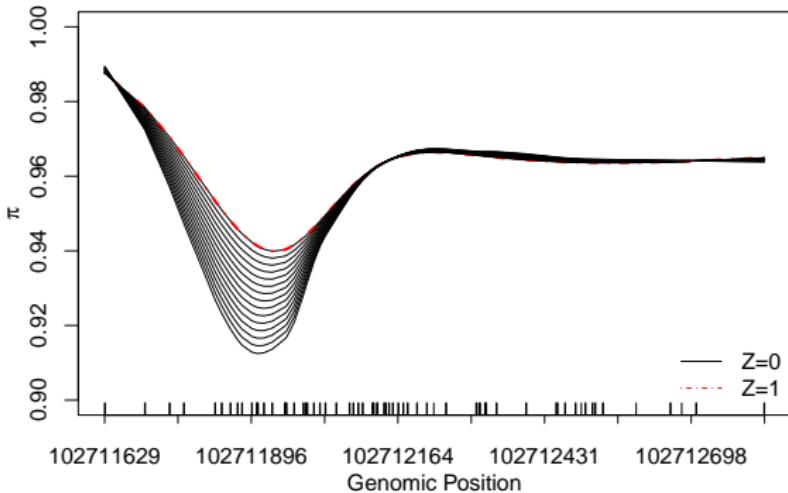
$p_0 = 0.003, p_1 = 0.9$



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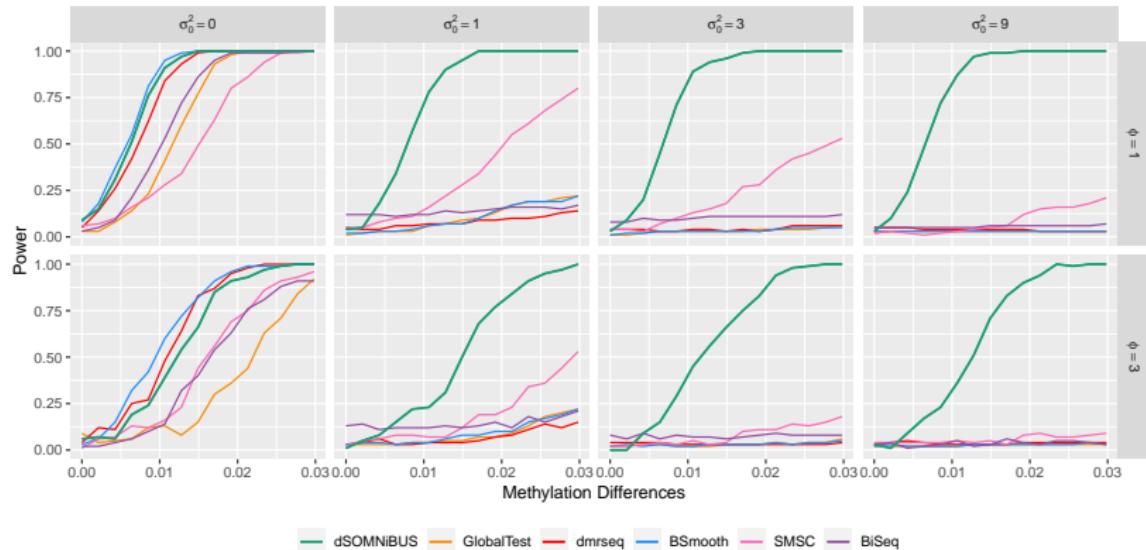
Simulation to evaluate power



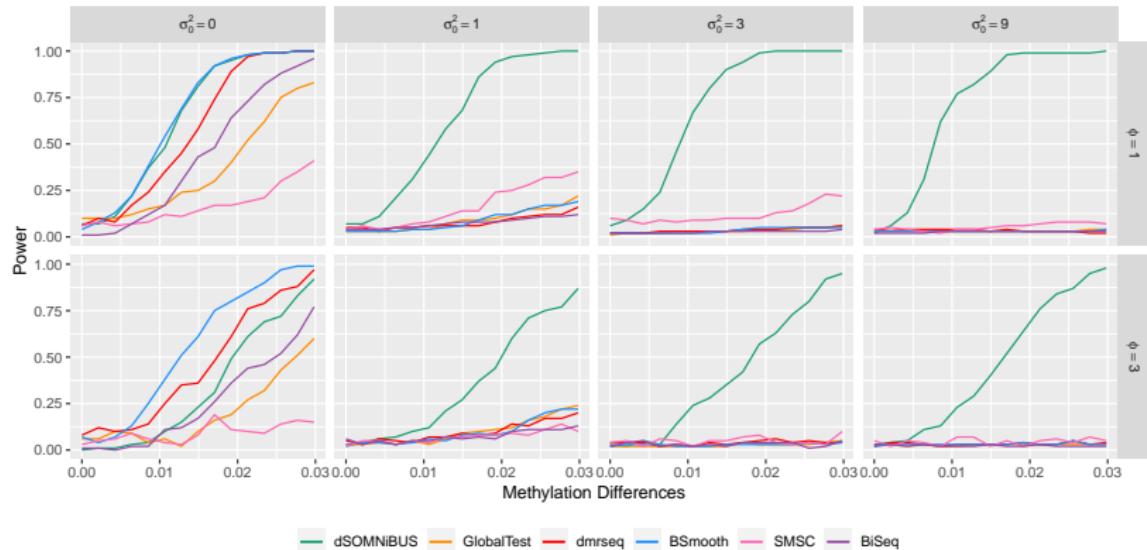
$Z = 1$ curve in red (fixed)

$Z = 0$ curve varied to give various sizes of differences

Power without errors: $p_0 = 0, p_1 = 1$



Power with errors: $p_0 = 0.003, p_1 = 0.9$



Summary



- ▶ an **adequate representation of realistic dispersion trends** in regional methylation data
- ▶ **well-founded theoretical properties** accounting for all (known) sources of data variability and possible experimental errors
- ▶ **increased power**; correct control of the type I error rate

- ▶ methodologies can be generally applied to other types of count data
 - allele-specific gene expression (ASE) measured from RNA-seq data
 - any type of count data for a more comprehensive representation of dispersion
 - varying-coefficients models in other context, e.g. temporal trend

Next step plans



- ▶ integrate SNP information (automatic variable selection)
- ▶ covariates (eg. disease status) may influence the variability/dispersion of DNA methylation (model $\phi(Z)$)
- ▶ correlated samples (additional set of random effects)



methylation QTL mapping

Given: a set of CpGs & a set of nearby SNPs ($P \gg N$)

Output: genetic variants associated with methylation levels in the test region

sparseSOMNiBUS

- ▶ a sparsity-smoothness penalty on each functional component $\beta_p(t)$

$$J(\beta_p) = \lambda \sqrt{(1 - \alpha)J_1(\beta_p) + \alpha J_2(\beta_p)}$$

where

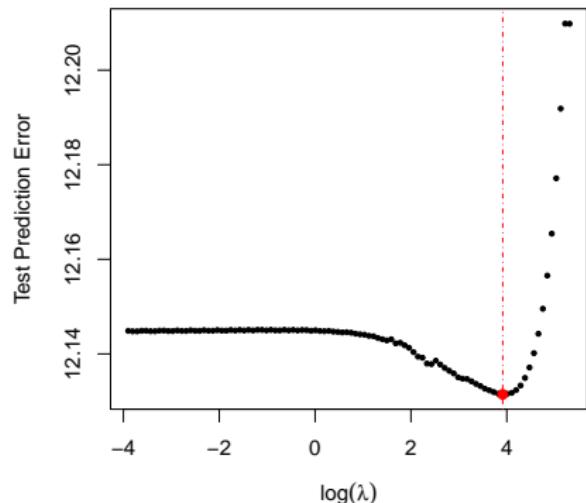
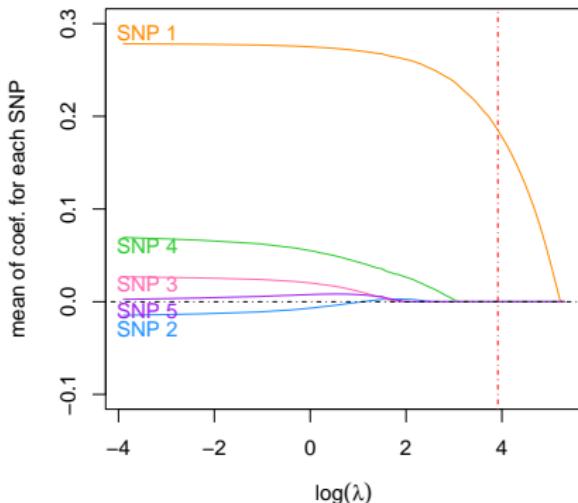
$$J_1(\beta_p) = \int (\beta_p(t))^2 dt$$

$$J_2(\beta_p) = \int (\beta_p''(t))^2 dt$$

- ▶ proximal gradient descent + backtracking line search (Rcpp)
- ▶ tuning parameters λ and α , selected by cross-validation

A simple illustration of sparseSOMNiBUS

- ▶ a methylation region with 123 CpG sites
- ▶ 5 SNPs: 1 mQTL and 4 negative controls



under the best chosen $\alpha = 0.55$

Acknowledgement

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- ▶ Dr. Celia Greenwood and Dr. Karim Oualkacha
- ▶ Dr. Lajmi Lakhal-Chaieb, Dr. Aurélie Labbe
- ▶ Dr. Yi Yang
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- ▶ Dr. Sasha Bernatsky, Dr. Marie Hudson, Dr. Inés Colmegna
- ▶ the CARTaGENE study investigators
- ▶ the participants in the CARTaGENE study

**Fonds de recherche
Santé**





Thanks

Questions & Comments

SOMNiBUS[†]: Model



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- ▶ Assume **known error parameters** p_0 and p_1 ,

$$\begin{aligned} p_0 &= \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 0) \\ p_1 &= \mathbb{P}(Y_{ijk} = 1 \mid S_{ijk} = 1). \end{aligned}$$

- ▶ Specify the model

$$\begin{aligned} S_{ij} \mid \mathbf{Z}_I, X_{ij} &\sim \text{Binomial}(X_{ij}, \pi_{ij}) \\ \log \left\{ \frac{\pi_{ij}}{1 - \pi_{ij}} \right\} &= \beta_0(t_{ij}) + \beta_1(t_{ij})Z_{1i} + \beta_2(t_{ij})Z_{2i} + \dots + \beta_P(t_{ij})Z_{Pi}, \end{aligned}$$

- ▶ Consider basis expansion: $\beta_p(t_{ij}) = \sum_{l=1}^{L_p} \alpha_{pl} B_l(t_{ij})$ for $p = 0, 1, \dots, P$.
- ▶ [‡]Smoothness parameters to penalize the roughness of effect curves

$$\mathcal{L}^{\text{Smooth}} = \sum_{p=0}^P \lambda_p \int (\beta_p''(t))^2 dt = \sum_{p=0}^P \lambda_p \boldsymbol{\alpha}_p^T \mathbf{A}_p \boldsymbol{\alpha}_p = \boldsymbol{\alpha}^T \mathbf{A}_{\lambda} \boldsymbol{\alpha},$$

[†]R package: <https://github.com/kaiqiong/SOMNiBUS>. [‡]Wahba (1980), Parker and Rice (1985)

Technical detail 1: E-M algorithm



Complete joint likelihood

- \dagger Random-effect view of the smoothness penalty: $\alpha \sim MVN(\mathbf{0}, \mathbf{A}_\lambda^{-1})$
- $I^{\text{complete}}(\mathbf{S}; \alpha, \lambda) = I(\mathbf{S}; \alpha) - \frac{1}{2} \alpha^T \mathbf{A}_\lambda \alpha + \frac{1}{2} \log \{|\mathbf{A}_\lambda|_+\}$

E step: Calculate $\eta_{ij}^* = \mathbb{E}(S_{ij} \mid Y_{ijk}; \alpha^*)$

M step: \ddagger Maximize $Q(\alpha, \lambda \mid \alpha^*) = I(\eta^*; \alpha) - \frac{1}{2} \alpha^T \mathbf{A}_\lambda \alpha + \frac{1}{2} \log \{|\mathbf{A}_\lambda|_+\}$

- Estimate α given the value of λ : P-IRLS

$$\hat{\alpha}_\lambda = \operatorname{argmax}_\alpha \left\{ I(\eta^*; \alpha) - \frac{1}{2} \alpha^T \mathbf{A}_\lambda \alpha \right\}$$

- Estimate λ : maximize the Laplace-approximated restrictive likelihood

$$L^M(\lambda) = \int \exp \{ Q(\alpha, \lambda \mid \alpha^*) \} d\alpha \approx \text{Laplace}(\lambda; \hat{\alpha}_\lambda).$$

\dagger Wahba (1983), JRSSB; Silverman (1985), JRSSB. \ddagger Wood (2011), JRSSB; R package `mgcv`

Technical detail 2: Inference



- ▶ Conditional on the values of smoothing parameter λ
- ▶ Estimate the variance of EM estimator $\hat{\alpha}$, V , using the observed Fisher information[†]
- ▶ Hypothesis testing for a regional zero effect $H_0 : \beta_p(t) = 0$.
 - Wald-type statistic

$$T_p = \widehat{\alpha}_p^T \{V_p\}^{-1} \widehat{\alpha}_p \sim \chi_{\tau_p}^2$$

- Penalization affects effective degree of freedom[‡]; $\tau_p < L_p = \dim(\alpha_p)$

$$\tau_p = \sum_{l=a_p}^{b_p} (2F - FF)_{(l,l)}, \text{ for } p = 0, 1, \dots, P,$$

- F is the ‘hat’ matrix and has the form $F = (\mathbb{X}^T \widehat{W} \mathbb{X} + A_{\hat{\lambda}})^{-1} \mathbb{X}^T \widehat{W} \mathbb{X}$

[†] Oakes, D. (1999) Direct calculation of the information matrix via the EM. JRSSB

[‡] Wood, S.N. (2013) On p-values for smooth components of an extended generalized additive model. Biometrika

- ▶ Random-effect view of the smoothness penalty: $\alpha \sim MVN(\mathbf{0}, \mathbf{A}_\lambda^-)$
- ▶ conditional mean parameters (REs): $\mathcal{B} = (\alpha, \mathbf{u}) \in \mathbb{R}^{N + \sum_0^P L_p}$
- ▶ variance component parameters: $\Theta = (\lambda, \sigma_0^2) \in \mathbb{R}^{P+2}$
- ▶ **multiplicative dispersion parameter:** ϕ

Complete joint log-quasi-likelihood function

$$\begin{aligned}
 q\ell^{(S, \mathcal{B})}(\mathcal{B}, \phi, \Theta) &= q\ell^{(S|\mathcal{B})}(\mathcal{B}, \phi) - \underbrace{\frac{1}{2} \alpha^T \mathbf{A}_\lambda \alpha - \frac{1}{2\sigma_0^2} \mathbf{u}^T \mathbf{u}}_{-\frac{1}{2\phi} \mathcal{B}^T \Sigma_\Theta \mathcal{B}} \\
 &\quad + \underbrace{\frac{1}{2} \log \{ |\mathbf{A}_\lambda|_+ \} + \frac{N}{2} \log \left(1/\sigma_0^2 \right)}_{1/2 \log \{ |\Sigma_\Theta / \phi|_+ \}}
 \end{aligned}$$



Conditional quasi-likelihood function

$$qL^{(S|\mathcal{B})}(\mathcal{B}, \phi) \propto \exp \left\{ -\frac{1}{2\phi} \sum_{i,j} d_{ij}(S_{ij}, \pi_{ij}) - \frac{M}{2} \log \phi \right\},$$

- ▶ $d_{ij}(S_{ij}, \pi_{ij}) = -2 \int_{S_{ij}/X_{ij}}^{\pi_{ij}} \frac{S_{ij} - X_{ij}\pi_{ij}}{\pi_{ij}(1-\pi_{ij})} d\pi_{ij}$ is the quasi-deviance function
- ▶ This is the extended quasi-likelihood for the joint parameter (\mathcal{B}, ϕ)
- ▶ It exhibits the properties of log-likelihood, with respect to both \mathcal{B} (exact) and ϕ (approximate)
- ▶ [†]The assumptions required are that ϕ be small and that $\kappa_r = O(\phi^{r-1})$

[†] Efron (1986), Jorgensen (1987), McCullagh and Nelder (1989)

dSOMNiBUS: Estimation



► Marginal quasi-likelihood function

$$qL^M(\phi, \Theta) = \int \exp \left\{ q\ell^{(S, \mathcal{B})}(\mathcal{B}, \phi, \Theta) \right\} d\mathcal{B} \approx \text{Laplace}(\phi, \Theta; \widehat{\mathcal{B}}) \neq f(\phi)g(\Theta).$$

► A similar E-M algorithm

Initialize $\Theta^{(0)}, \phi^{(0)}, \mathcal{B}^{(0)}$ (estimates ignoring errors); Choose $\varepsilon = 10^{-6}$; Set $\ell = 0$;

repeat

- E step: $\eta_{ij}^{(\ell)} = \mathbb{E}(S_{ij} \mid Y_{ij}; \mathcal{B}^{(\ell)})$;
- M step: $(\mathcal{B}^{(\ell)}, \phi^{(\ell)}, \Theta^{(\ell)}) = \underset{\mathcal{B}, \phi, \Theta}{\operatorname{argmax}} \ell^{\text{Joint}}(\mathcal{B}, \phi, \Theta; \eta_{ij}^{(\ell)})$. Specifically repeat

• Solve $\mathbf{U}(\mathcal{B}; \Theta^{(s)}) = \mathbf{0}$ to obtain $\mathcal{B}^{(s)}$ using data $\eta_{ij}^{(\ell)}$;

• Newton's update for the Laplace approximated marginal likelihood evaluated at data $\eta_{ij}^{(\ell)}$:

$$(\phi, \Theta)^{(s+1)} = (\phi, \Theta)^{(s)} - [\nabla^2 \text{Laplace}(\mathcal{B}^{(s)})]^{-1} \nabla \text{Laplace}(\mathcal{B}^{(s)});$$

$s \leftarrow s + 1$;

until $\|\mathcal{B}^{(s)} - \mathcal{B}^{(s-1)}\|_2 < \varepsilon$;

$\ell \leftarrow \ell + 1$;

until $\|\mathcal{B}^{(\ell)} - \mathcal{B}^{(\ell-1)}\|_2 < \varepsilon$;

Return $\Theta^{(\ell)}, \mathcal{B}^{(\ell)}, \phi^{(\ell)}$;

► Estimating ϕ

- Likelihood-based estimator
- Moment-based estimator (better)

Inference for smooth covariate effects

- ▶ Estimate the variance of EM estimator $\hat{\alpha}$, V , using the observed (quasi-)Fisher information[†]
- ▶ Hypothesis testing for a regional zero effect $H_0 : \beta_p(t) = 0$.
 - Regional statistic

$$T_p = \frac{\hat{\alpha}_p^T \{ \hat{V}_p \}^{-1} \hat{\alpha}_p}{\tau_p} \sim F_{\tau_p, M - \tau}$$

- τ_p : EDF for smooth term $\beta_p(t)$. τ : total EDF of the model
- This F null distribution relies on the assumption that $(M - \tau)\hat{\phi}/\phi \sim \chi^2_{M - \tau}$, which is approximately true for moment-based dispersion estimator

[†] Elashoff and Ryan (2004) An EM algorithm for estimating equations. Journal of Computational and Graphical Statistics