

# Class 19: Investigating Pertussis Resurgence

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#1. Investigating pertussis cases by year

The CDC tracks cases of Pertussis in the US. We can get their data via web-scraping

Q1. With the help of the R “addin” package datapasta assign the CDC pertussis case number data to a data frame called cdc and use ggplot to make a plot of cases numbers over time.

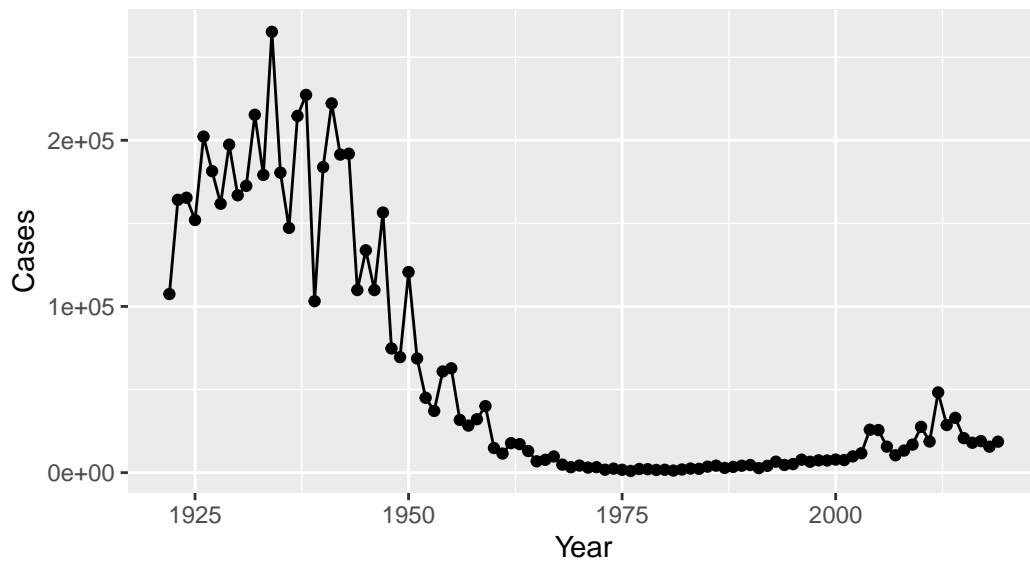
```
library (ggplot2)

baseplot <- ggplot(cdc) +
  aes (Year, Cases) +
  geom_point() +
  geom_line() +
  labs(title = "Cases of Pertussis in US from 1920 to 1999",
        subtitle = "Data from the CDC")

baseplot
```

## Cases of Pertussis in US from 1920 to 1999

Data from the CDC

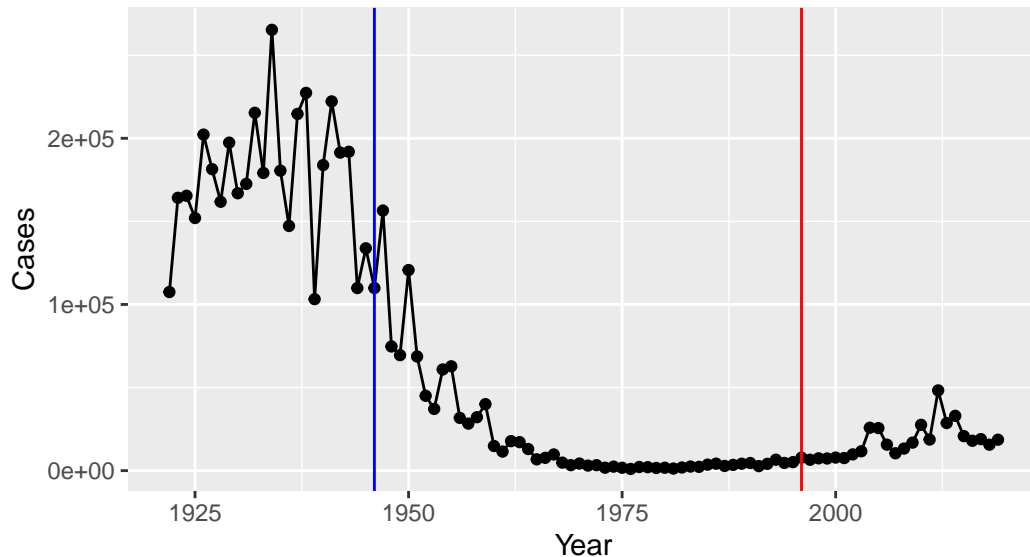


Q2. Using the ggplot `geom_vline()` function add lines to your previous plot for the 1946 introduction of the wP vaccine and the 1996 switch to aP vaccine (see example in the hint below). What do you notice?

```
baseplot +  
  geom_vline(xintercept = 1946, col = "blue") +  
  geom_vline(xintercept = 1996, col = "red")
```

## Cases of Pertussis in US from 1920 to 1999

Data from the CDC



Q3. Describe what happened after the introduction of the aP vaccine? Do you have a possible explanation for the observed trend?

We see an increase of cases after the introduction of aP vaccine. It remained for a bit, but began to rise higher to levels that have not been seen since 19. Potentially, it might be due to the potency of the vaccine or hesitancy to get vaccines. The vaccine switch was to minimize the symptoms caused by the wP (swelling, redness in baby).

#The CMI-PB project

The CMI-PB project is collecting data on aP and wP individuals and their immune response to infection and or booster shot.

CMI-PB returns data from it's API in JSON format \*like most APIs). We will use the jsonlite package to get data from this API.

```
library (jsonlite)

subject <- read_json("https://www.cmi-pb.org/api/subject",
                     simplifyVector = TRUE)

head(subject)
```

```
subject_id  infancy_vac  biological_sex      ethnicity  race
```

1	1	wP	Female Not Hispanic or Latino White
2	2	wP	Female Not Hispanic or Latino White
3	3	wP	Female Unknown White
4	4	wP	Male Not Hispanic or Latino Asian
5	5	wP	Male Not Hispanic or Latino Asian
6	6	wP	Female Not Hispanic or Latino White

	year_of_birth	date_of_boost	dataset
1	1986-01-01	2016-09-12	2020_dataset
2	1968-01-01	2019-01-28	2020_dataset
3	1983-01-01	2016-10-10	2020_dataset
4	1988-01-01	2016-08-29	2020_dataset
5	1991-01-01	2016-08-29	2020_dataset
6	1988-01-01	2016-10-10	2020_dataset

Q4. How many aP and wP infancy vaccinated subjects are in the dataset? Ans: aP 47, wP 49

```
table (subject$infancy_vac)
```

```
aP wP
47 49
```

Q5. How many Male and Female subjects/patients are in the dataset? ANS: Female 66, Male 30

```
table (subject$biological_sex)
```

```
Female  Male
66      30
```

Q6. What is the breakdown of race and biological sex (e.g. number of Asian females, White males etc...)? ANS:

```
table(subject$race, subject$biological_sex)
```

	Female	Male
American Indian/Alaska Native	0	1

Asian	18	9
Black or African American	2	0
More Than One Race	8	2
Native Hawaiian or Other Pacific Islander	1	1
Unknown or Not Reported	10	4
White	27	13

Q7. Using this approach determine (i) the average age of wP individuals, (ii) the average age of aP individuals; and (iii) are they significantly different?

```
library (lubridate)
```

Attaching package: 'lubridate'

The following objects are masked from 'package:base':

date, intersect, setdiff, union

```
today()
```

```
[1] "2023-03-14"
```

```
age_days <- today() - ymd(subject$year_of_birth)
age_years <- time_length (age_days, "years")
subject$age <- age_years
```

Filter the data for aP individuals in order to calculate days. Now find the average age of all individuals:

```
mean(subject$age)
```

```
[1] 31.05079
```

```
library (dplyr)
```

Attaching package: 'dplyr'

The following objects are masked from 'package:stats':

```
filter, lag
```

The following objects are masked from 'package:base':

```
intersect, setdiff, setequal, union
```

```
mean ( filter(subject, infancy_vac == "aP")$age)
```

```
[1] 25.5156
```

```
mean ( filter(subject, infancy_vac == "wP")$age)
```

```
[1] 36.36006
```

T-test

```
ap.age <- filter(subject, infancy_vac == "aP")$age  
wp.age <- filter(subject, infancy_vac == "wP")$age  
  
mean( ap.age )
```

```
[1] 25.5156
```

```
mean( wp.age )
```

```
[1] 36.36006
```

```
#T.test  
  
t.test(ap.age, wp.age)
```

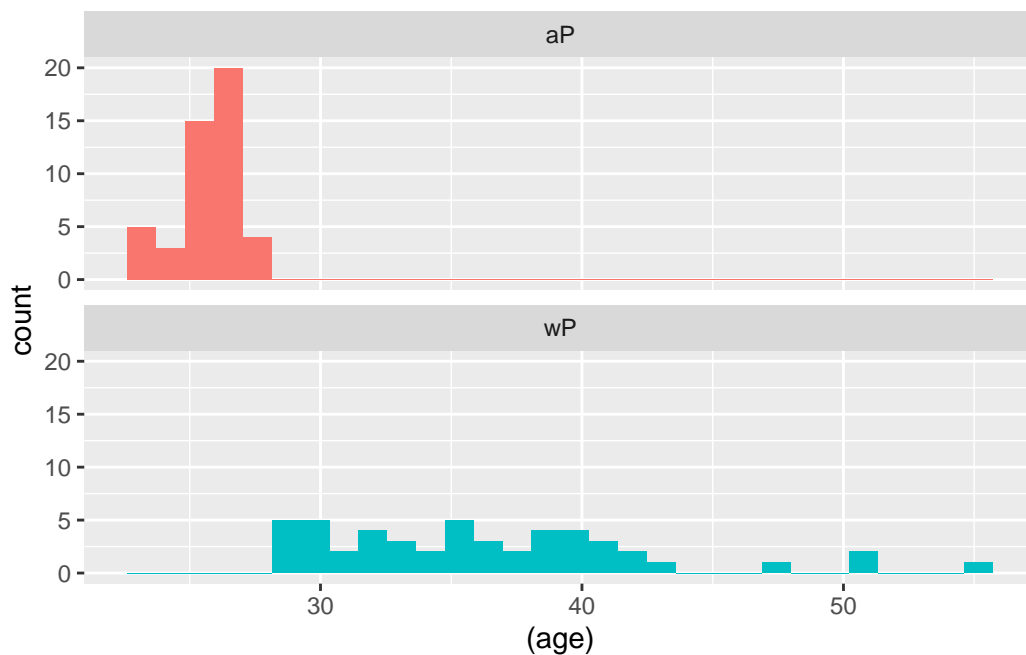
### Welch Two Sample t-test

```
data: ap.age and wp.age
t = -12.092, df = 51.082, p-value < 2.2e-16
alternative hypothesis: true difference in means is not equal to 0
95 percent confidence interval:
 -12.644857  -9.044045
sample estimates:
mean of x mean of y
 25.51560  36.36006
```

Q9. With the help of a faceted histogram (see below), do you think these two groups are significantly different?

```
ggplot(subject) +
  aes((age),
      fill=as.factor(infancy_vac)) +
  geom_histogram(show.legend=FALSE) +
  facet_wrap(vars(infancy_vac), nrow=2)
```

`stat\_bin()` using `bins = 30`. Pick better value with `binwidth`.



#Joining multiple tables

Read the specimen and ab\_titer tables into R and store the data as a specimen and titer named data frames.

```
specimen <- read_json("http://cmi-pb.org/api/specimen",
                      simplifyVector = TRUE)
titer <- read_json("http://www.cmi-pb.org/api/ab_titer",
                   simplifyVector = TRUE)
```

```
head(specimen)
```

	specimen_id	subject_id	actual_day_relative_to_boost
1	1	1	-3
2	2	1	736
3	3	1	1
4	4	1	3
5	5	1	7
6	6	1	11

	planned_day_relative_to_boost	specimen_type	visit
1	0	Blood	1
2	736	Blood	10
3	1	Blood	2
4	3	Blood	3
5	7	Blood	4
6	14	Blood	5

```
head (titer)
```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgE	FALSE	Total	1110.21154	2.493425
2	1	IgE	FALSE	Total	2708.91616	2.493425
3	1	IgG	TRUE	PT	68.56614	3.736992
4	1	IgG	TRUE	PRN	332.12718	2.602350
5	1	IgG	TRUE	FHA	1887.12263	34.050956
6	1	IgE	TRUE	ACT	0.10000	1.000000

	unit	lower_limit_of_detection
1	UG/ML	2.096133
2	IU/ML	29.170000
3	IU/ML	0.530000
4	IU/ML	6.205949



```
5 IU/ML          4.679535
6 IU/ML          2.816431
```

Q9. Complete the code to join specimen and subject tables to make a new merged data frame containing all specimen records along with their associated subject details:

```
meta <- inner_join(specimen, subject)
```

Joining with `by = join\_by(subject\_id)`

```
dim(meta)
```

```
[1] 729 14
```

```
head(meta)
```

```
specimen_id subject_id actual_day_relative_to_boost
1           1           1                        -3
2           2           1                       736
3           3           1                        1
4           4           1                        3
5           5           1                        7
6           6           1                       11
planned_day_relative_to_boost specimen_type visit infancy_vac biological_sex
1                           0         Blood    1          wP        Female
2                          736         Blood   10          wP        Female
3                           1         Blood    2          wP        Female
4                           3         Blood    3          wP        Female
5                           7         Blood    4          wP        Female
6                          14         Blood    5          wP        Female
ethnicity race year_of_birth date_of_boost dataset
1 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
2 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
3 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
4 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
5 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
6 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
age
```

```

1 37.19644
2 37.19644
3 37.19644
4 37.19644
5 37.19644
6 37.19644

```

Q10. Now using the same procedure join meta with titer data so we can further analyze this data in terms of time of visit aP/wP, male/female etc.

```
abdata <- inner_join(titer, meta)
```

Joining with `by = join\_by(specimen\_id)`

```
dim(abdata)
```

```
[1] 32675    21
```

```
head(abdata)
```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgE	FALSE	Total	1110.21154	2.493425
2	1	IgE	FALSE	Total	2708.91616	2.493425
3	1	IgG	TRUE	PT	68.56614	3.736992
4	1	IgG	TRUE	PRN	332.12718	2.602350
5	1	IgG	TRUE	FHA	1887.12263	34.050956
6	1	IgE	TRUE	ACT	0.10000	1.000000

	unit	lower_limit_of_detection	subject_id	actual_day_relative_to_boost
1	UG/ML	2.096133	1	-3
2	IU/ML	29.170000	1	-3
3	IU/ML	0.530000	1	-3
4	IU/ML	6.205949	1	-3
5	IU/ML	4.679535	1	-3
6	IU/ML	2.816431	1	-3

	planned_day_relative_to_boost	specimen_type	visit	infancy_vac	biological_sex
1	0	Blood	1	wP	Female
2	0	Blood	1	wP	Female
3	0	Blood	1	wP	Female
4	0	Blood	1	wP	Female

```

5           0      Blood      1      wP      Female
6           0      Blood      1      wP      Female
      ethnicity  race year_of_birth date_of_boost      dataset
1 Not Hispanic or Latino White  1986-01-01  2016-09-12 2020_dataset
2 Not Hispanic or Latino White  1986-01-01  2016-09-12 2020_dataset
3 Not Hispanic or Latino White  1986-01-01  2016-09-12 2020_dataset
4 Not Hispanic or Latino White  1986-01-01  2016-09-12 2020_dataset
5 Not Hispanic or Latino White  1986-01-01  2016-09-12 2020_dataset
6 Not Hispanic or Latino White  1986-01-01  2016-09-12 2020_dataset
      age
1 37.19644
2 37.19644
3 37.19644
4 37.19644
5 37.19644
6 37.19644

```

Q11. How many specimens (i.e. entries in abdata) do we have for each isotype?

```
table(abdata$isotype)
```

```

IgE  IgG  IgG1  IgG2  IgG3  IgG4
6698 1413 6141 6141 6141 6141

```

Q12. What do you notice about the number of visit 8 specimens compared to other visits?

```
table(abdata$visit)
```

```

  1    2    3    4    5    6    7    8
5795 4640 4640 4640 4640 4320 3920   80

```

Its drop to 90 on the 8th specimen. Decreasing values

Q13. Complete the following code to make a summary boxplot of Ab titer levels for all antigens:

```

ig1 <- abdata %>% filter(isotype == "IgG1", visit!=8)
head(ig1)

```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgG1	TRUE	ACT	274.355068	0.6928058
2	1	IgG1	TRUE	LOS	10.974026	2.1645083
3	1	IgG1	TRUE	FELD1	1.448796	0.8080941
4	1	IgG1	TRUE	BETV1	0.100000	1.0000000
5	1	IgG1	TRUE	LOLP1	0.100000	1.0000000
6	1	IgG1	TRUE	Measles	36.277417	1.6638332

	unit	lower_limit_of_detection	subject_id	actual_day_relative_to_boost
1	IU/ML	3.848750	1	-3
2	IU/ML	4.357917	1	-3
3	IU/ML	2.699944	1	-3
4	IU/ML	1.734784	1	-3
5	IU/ML	2.550606	1	-3
6	IU/ML	4.438966	1	-3

	planned_day_relative_to_boost	specimen_type	visit	infancy_vac	biological_sex
1	0	Blood	1	wP	Female
2	0	Blood	1	wP	Female
3	0	Blood	1	wP	Female
4	0	Blood	1	wP	Female
5	0	Blood	1	wP	Female
6	0	Blood	1	wP	Female

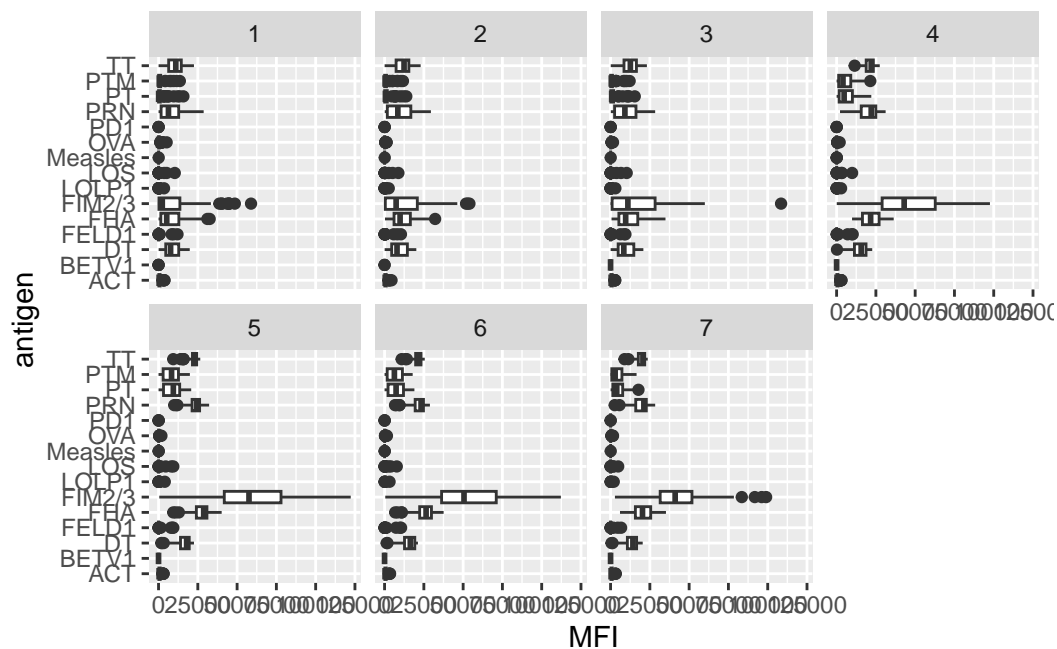
  

	ethnicity	race	year_of_birth	date_of_boost	dataset
1	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
2	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
3	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
4	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
5	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
6	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset

	age
1	37.19644
2	37.19644
3	37.19644
4	37.19644
5	37.19644
6	37.19644

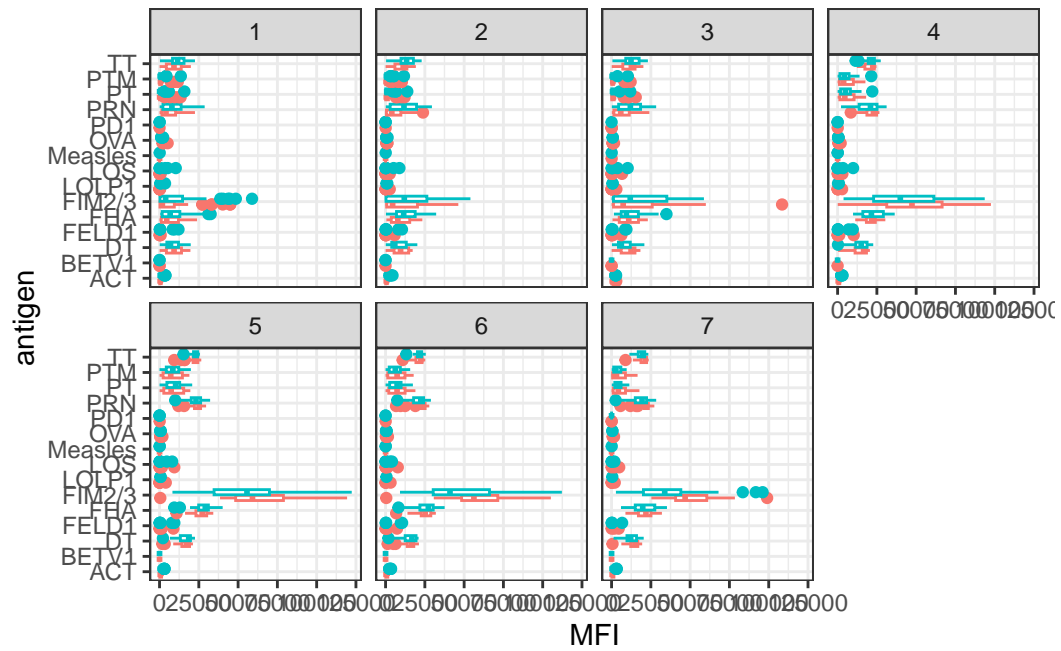
```
ggplot(ig1) +
  aes(MFI, antigen) +
  geom_boxplot() +
  facet_wrap(vars(visit), nrow=2)
```



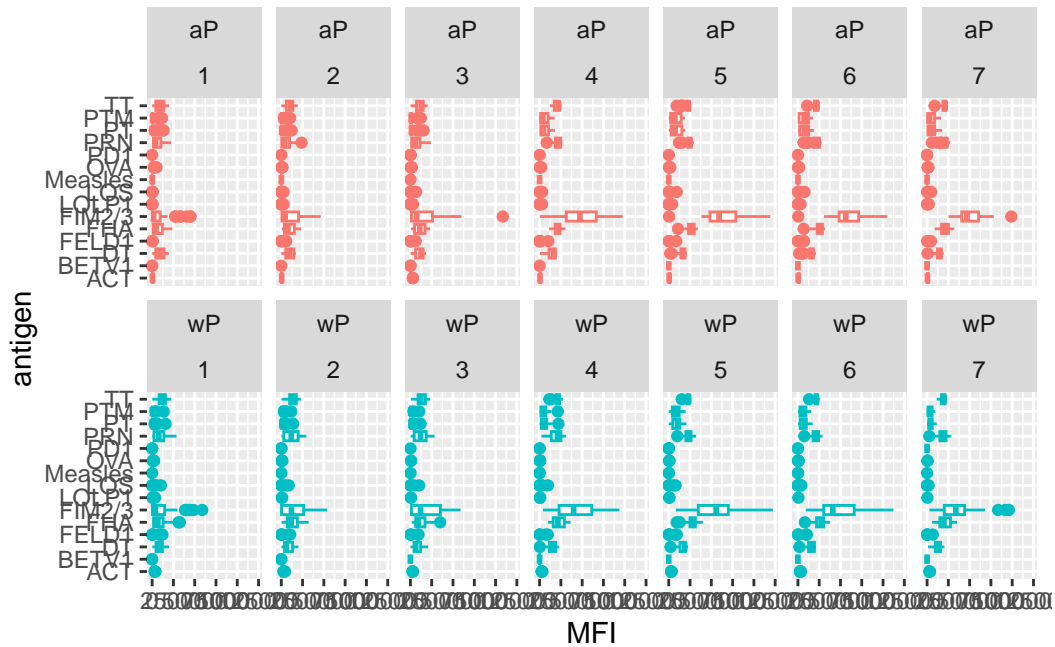
Q14. What antigens show differences in the level of IgG1 antibody titers recognizing them over time? Why these and not others?

FIM2/3, FHA, PT are all in the aP boost vaccine.

```
ggplot(ig1) +
  aes(MFI, antigen, col=infancy_vac ) +
  geom_boxplot(show.legend = FALSE) +
  facet_wrap(vars(visit), nrow=2) +
  theme_bw()
```

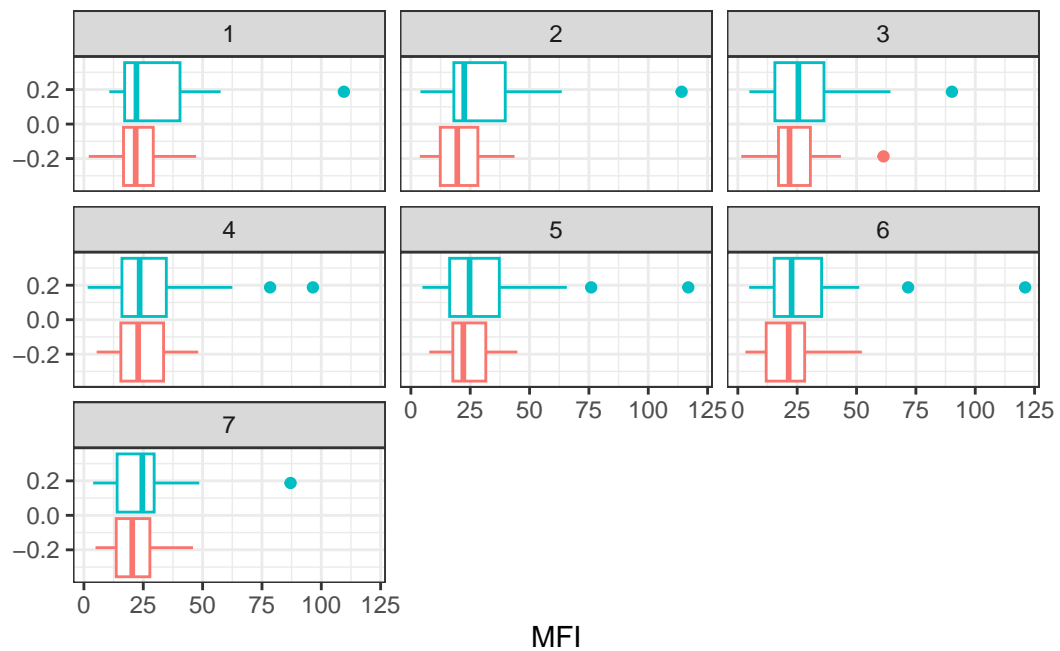


```
ggplot(ig1) +
  aes(MFI, antigen, col=infancy_vac ) +
  geom_boxplot(show.legend = FALSE) +
  facet_wrap(vars(infancy_vac, visit), nrow=2)
```



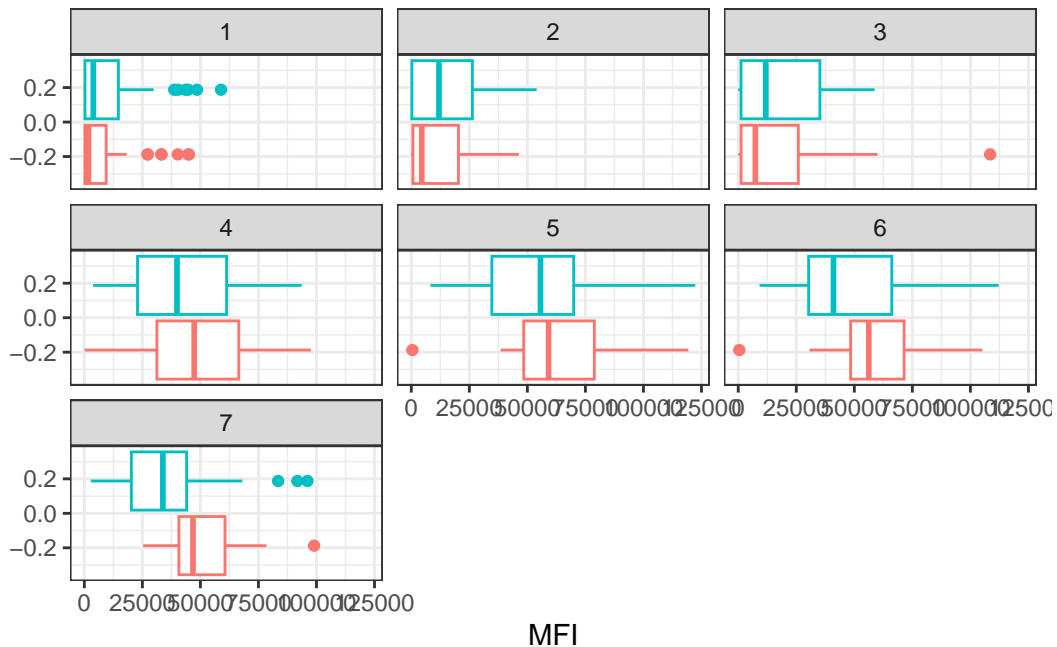
Q15. Filter to pull out only two specific antigens for analysis and create a boxplot for each. You can chose any you like. Below I picked a “control” antigen (“Measles”, that is not in our vaccines) and a clear antigen of interest (“FIM2/3”, extra-cellular fimbriae proteins from *B. pertussis* that participate in substrate attachment).

```
filter(ig1, antigen=="Measles") %>%
  ggplot() +
  aes(MFI, col=infancy_vac) +
  geom_boxplot(show.legend = FALSE) +
  facet_wrap(vars(visit)) +
  theme_bw()
```



```
filter(ig1, antigen== "FIM2/3") %>%
  ggplot() +
  aes(MFI, col=infancy_vac) +
  geom_boxplot(show.legend = FALSE) +
  facet_wrap(vars(visit)) +
  theme_bw()
```





Q16. What do you notice about these two antigens time course and the FIM2/3 data in particular?

Ans: We see that FIM2/3 is on the rise, increasing faster than measles

Q17. Do you see any clear difference in aP vs. wP responses?

aP vaccines seem to have a higher antigen response in comparison to wp

```
url <- "https://www.cmi-pb.org/api/v2/rnaseq?versioned_ensembl_gene_id=eq.ENS00000211896."
rna <- read_json(url, simplifyVector = TRUE)

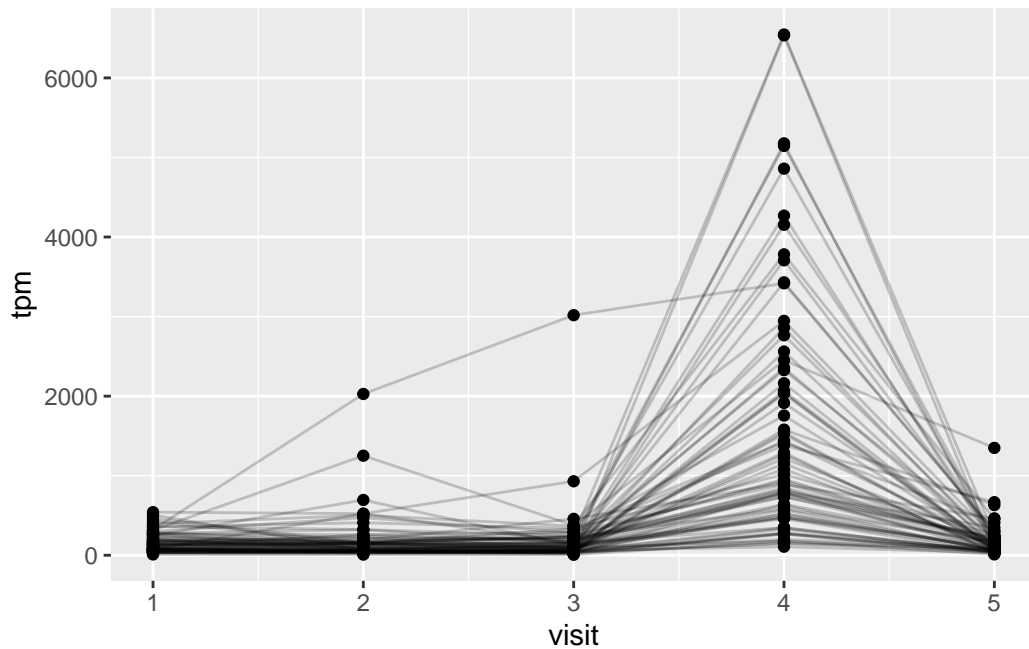
#meta <- inner_join(specimen, subject)
ssrna <- inner_join(rna, meta)
```

Joining with `by = join\_by(specimen\_id)`

Q18. Make a plot of the time course of gene expression for IGHG1 gene (i.e. a plot of visit vs. tpm).

```
ggplot(ssrna) +
  aes(visit, tpm, group=subject_id) +
```

```
geom_point() +  
geom_line(alpha=0.2)
```



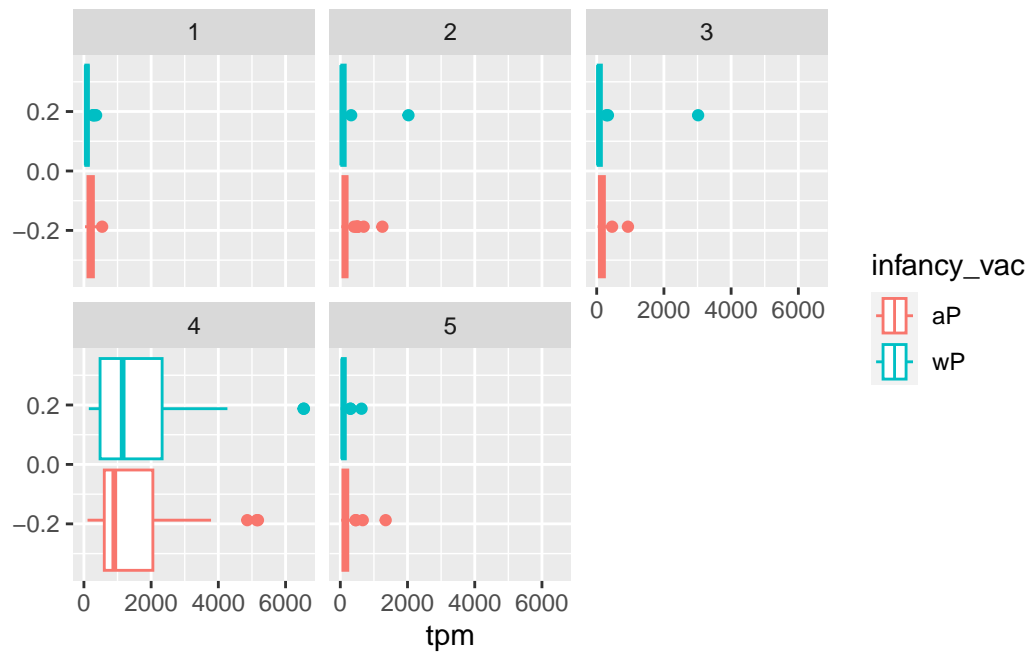
Q19. What do you notice about the expression of this gene (i.e. when is it at it's maximum level)?

Ans. Maximum level at the 4th visit. Expression spiked during the 4th visit.

Q20. Does this pattern in time match the trend of antibody titer data? If not, why not?

Nope it does not make sense since antibodies can last for a while. A sudden spike on the 4th doesn't make sense. It should be constant or spike then decline.

```
ggplot(ssrna) +  
  aes(tpm, col=infancy_vac) +  
  geom_boxplot() +  
  facet_wrap(vars(visit))
```



```
ssrna %>%
  filter(visit==4) %>%
  ggplot() +
    aes(tpm, col=infancy_vac) + geom_density() +
    geom_rug()
```

