# Package 'COMBIA'

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Description A comprehensive synergy/antagonism analyses of drug combinations with quality graphics and data. The analyses can be performed by Bliss independence and Loewe additivity models. 'COMBIA' provides improved statistical analysis and makes only very weak assumption of data variability while calculating bootstrap intervals (BIs). It is based on heteroscedasticity controlled resampling (bootstrapping) and includes a global (omnibus) test. Finally, package shows analyzed data, 2D and 3D plots ready to use in research publications. 'COMBIA' does not require manual data entry. Data can be directly input from wet lab experimental platforms for example fluostar, automated robots etc. One only needs to call a single function, analyzeCOMBO(), to perform all analysis (examples are provided with sample data).
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This function calculates significant synergy/antagonism according to Bliss or Loewe model and creates scientific publication ready graphs.

# Description

This function calculates significant synergy/antagonism according to Bliss or Loewe model and creates scientific publication ready graphs.

# Usage

```
analyzeCOMBO(
  filename,
  sheet = 1,
  model,
  inputFormates,
  platetype = "384",
  keyposition = 2,
  selectionkey = "65000",
  platekey = 7051,
  minThersholdForCVCal = 0.15,
  minThersholdForCV = 0.3,
  wells,
  yConcentration,
  xConcentration,
  xDrug,
  yDrug,
  cellLine,
  survivalFunc = function(x, y, z) { (x - z)/(y - z) },
  nBoot = 5000
)
```

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#### **Arguments**

filename Name of file containing experimental data. For MS Excel files, working version

of Perl must be present in the executable search path.

sheet Optional, sheet number if excel file is used for input.

model bliss or loewe.

inputFormates Any of these three formates "fmca", "macsynergy" and "others" are supported.

Example is provided with macsynergy format and test data for this example can

be found in installation directory ("extdata") of COMBIA. See files FluoOptima\_384\_2014-

03-28test\_M and testDataM in directory "extdata" for format details of "fmca"

and "macsynergy". "others" can be any other format.

platetype Optional default is 384. Only 384 and 96 well plates are supported.

keyposition Optional default is 2. Usefull for automated barcoded data.

selectionkey Optional default is 65000.

platekey Optional barcode.

minThersholdForCVCal

Optional default is 0.15.

minThersholdForCV

Optional default is 0.3.

wells wells argument should be in triplet form that is 1-Untreated control wells range,

2-empty wells range and 3-case wells range. Thus in example below (see well argument) experiment has four replicates. "13:110","m3:m10","b3:k10" is first replicate. Where "13:110" is the location of untreated control values in the test-Data.csv, "m3:m10" is the background/ empty well well values and "b3:k10" are

values after treatment.

yConcentration Y drug Concentrations.

xConcentration X drug Concentrations.

xDrug X drug name. yDrug Y drug name.

cellLine Cell/Experiment name.

survivalFunc Optional default is function (x,y,z) (x-z)/(y-z) i.e (treated - background)/ (un-

treated - background).

nBoot Optional Number of time to bootstrap default is 5000

## Value

Stores and show graph/data of synergy/antagonism analyses

#### Author(s)

Muhammad kashif

4 applyBliss

applyBliss

Function calculates Bliss Synergy, associated BIs and global BIs

## **Description**

Function calculates Bliss Synergy, associated BIs and global BIs

## Usage

```
applyBliss(noOfRows, noOfCols, rawDataPreProcessed, nBoot)
```

#### **Arguments**

noOfRows Number of rows in the experiment

noOfCols Number of columns in the experiment

rawDataPreProcessed

Data matrix

nBoot Number of bootstrap

## Value

Three lists, first list consists of Bliss Synergy/Antagonism, lower bound of BI and upper bound of BI. 2nd list consists of global BI of Maximun synergistic combiantion and 3rd list consists of global BI of maximum antagonistic combination.

# Author(s)

Muhammad kashif

```
dataFile <- system.file( "extdata", "rawDataPreProcessed.csv", package="COMBIA" )
dataSample <- read.csv(dataFile, header=FALSE )
nR <- 8
nC <- 10
rslt <- applyBliss(nR, nC, as.matrix(dataSample ), 500)</pre>
```

applyLoewe 5

BIs	applyLoewe	This function calculates Loewe synergy/antagonism and associated BIs
-----	------------	--

# Description

This function calculates Loewe synergy/antagonism and associated BIs

# Usage

```
applyLoewe(rawDataPreProcessed, xConcentration, yConcentration, nBoot)
```

## **Arguments**

```
rawDataPreProcessed
Raw preprocessed experimental data

xConcentration X drug concentrations

yConcentration Y drug concentrations

nBoot Number of times to bootstrap
```

#### Value

Three lists, first list consisting of Loewe Synergy/Antagonism, lower bound of BI and upper bound of BI. 2nd list consists of global BI for maximum synergy and 3rd list consists of global BI of maximum antagonistic combination.

#### Author(s)

Muhammad kashif

```
## Not run:
dataFile <- system.file("extdata", "rawDataPreProcessed.csv", package="COMBIA")
dataSample <- read.csv(dataFile, header=FALSE)
xConc <- c(0.00, 0.20, 0.39, 0.78, 1.56,3.12, 6.25, 12.50, 25.00, 50)
yConc <- c(128, 64, 32, 16, 8, 4, 2, 0)
noOFBoot <- 500 # a large number is recomended
rslt <- applyLoewe(as.matrix(dataSample), xConc, yConc, noOFBoot)
## End(Not run)</pre>
```

6 calculateSi

calculateSi

Calculates survival indices (S.Is) for a range of wells (casewells). S.Is for a range of wells are calculated, that range is specified at the third place of wells argument list. This function call the rangemean function to calculate the mean of the range of the specified range. S.I is calculated by (Case well-meanofemptyrange/mean of controlwell-meanofemptyrange). In the wells argument one should provide arguments in the triplet form that is first one is control data range, second one is the empty data range while third one is the control range.

#### **Description**

Calculates survival indices (S.Is) for a range of wells (casewells). S.Is for a range of wells are calculated, that range is specified at the third place of wells argument list. This function call the rangemean function to calculate the mean of the range of the specified range. S.I is calculated by (Case well- meanofemptyrange/mean of controlwell- meanofemptyrange). In the wells argument one should provide arguments in the triplet form that is first one is control data range, second one is the empty data range while third one is the control range.

#### Usage

calculateSi(hashedplates, platekey, platetype, rowsperexperiment, wells)

## **Arguments**

hashedplates A hash table of picked plates. It is the output of function "selectPlate".

platekey It is the key of the plate whose S.I is needed to be calculated.

platetype It is the type of plate (386 and 96).

rowsperexperiment

It is the argument that specifies if the same experiment is reptead and how many times in a plate. If an experiment is repeated twice in adjacent rows then average

of its values will be used in the SI calculation.

wells This argument can take a list of arguments in the triplet form. Where first ar-

gument of triplet is the range of control wells, second argument is the range of empty wells while third one is the range of case wells. It is made so that in labs plates layouts can differ greatly. By using this triplet scheme one can handel a

number of palte layouts.

# Value

A matrix with S.I showing values where they are actually exist on the plate.

#### Author(s)

Muhammad Kashif

#### **Examples**

 ${\tt combineDataFromMultipleFiles}$ 

Combine data from multiple files

#### **Description**

Combine data from multiple files

# Usage

```
combineDataFromMultipleFiles(
  yConcentration,
  xConcentration,
  replNo,
  file,
  totalNumberofReplicates,
  siReplicates
)
```

## Arguments

```
\begin{array}{lll} \mbox{yConcentration} & \mbox{Y drug concentrations} \\ \mbox{xConcentration} & \mbox{X drug concentrations} \\ \mbox{replNo} & \mbox{Number of Replicates in all files} \\ \mbox{file} & \mbox{File name} \\ \mbox{totalNumberofReplicates} \\ \mbox{Total number of replicates per files} \\ \mbox{siReplicates} & \mbox{data} \\ \end{array}
```

#### Value

Combined data of replicate survival indices from multiple experiments

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#### Author(s)

Muhammad kashif

## **Examples**

```
xConc <- c(0.00, 0.20, 0.39, 0.78, 1.56, 3.12, 6.25, 12.50, 25.00, 50)
yConc <- c(128, 64, 32, 16, 8, 4, 2, 0)
rN <- 4
fN <- 1
trN <- 4
dataFile <- system.file("extdata", "rawDataPreProcessed.csv", package="COMBIA")
dataSample <- read.csv(dataFile, header=FALSE)
replList <- list(vector, 4)
for( i in 1:4)
{ replList[[i]] <- dataSample[i,] }
rslt <- combineDataFromMultipleFiles(list(yConc),
list(xConc), rN,fN,trN, replList )</pre>
```

createUniquePertbs

Function to make unique perturbations of the replicates these will be used incase if CV is greater than threshold.

# Description

Function to make unique perturbations of the replicates these will be used incase if CV is greater than threshold.

## Usage

```
createUniquePertbs(totalNumberofReplicates)
```

## **Arguments**

```
totalNumberofReplicates
Total replicate number
```

#### Value

unique possible perturbations

# Author(s)

Muhammad kashif

```
rslt <- createUniquePertbs(5)</pre>
```

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cVCal

This function calculates CV

## **Description**

This function calculates CV

# Usage

```
cVCal(vals)
```

# Arguments

vals

Values

#### Value

cv of input values

## Author(s)

Muhammad kashif

# **Examples**

```
mData <- matrix(1:10, 2,5)
rslt <- cVCal(mData)</pre>
```

extractKey

Extracts the keyvalues (Barcode) from a dataset, every plate needs barcode. Keyvalues are extracted from the header of the plates at the position specified by keyposition argument.

# Description

Extracts the keyvalues (Barcode) from a dataset, every plate needs barcode. Keyvalues are extracted from the header of the plates at the position specified by keyposition argument.

# Usage

```
extractKey(keyposition, rawdata, numberofrowsperplate, doubleplateexperiment)
```

#### **Arguments**

keyposition Position of keyvalue in the header of plate.

rawdata An object(dataframe) of rawdata.

numberofrowsperplate

This argument is not needed when you call function "readFluostarPlates". The number of rows depend upon the geometry of the plates. These are 16 in case of 384well paltes.

doubleplateexperiment

This parameter can have TRUE & FALSE values only. It is set to TRUE when an experiment is performed twice and we only want to choose only one of them.

## Value

A complete set of keyvalues.

#### Author(s)

Muhammad Kashif

## **Examples**

extractReplicateValues

This function will takes a list of ranges removes case wells and extract replicate values separately

#### **Description**

This function will takes a list of ranges removes case wells and extract replicate values separately

# Usage

```
extractReplicateValues(
  rawDataUnProcessed,
  wellRanges,
  wellplace = 3,
  simple = FALSE,
  excelFormate = FALSE
)
```

#### **Arguments**

rawDataUnProcessed

A data matrix

wellRanges Ranges of wells

wellplace Place of treated (case) well range

simple TRUE if survival values are already calculated otherwise it is FALSE

excelFormate True if ranges are in excel formate

#### Value

Replicate values

#### Author(s)

Muhammad kashif

#### **Examples**

extractValuesFromRange

This function extract numerical indices of a given range e-g B2

## **Description**

This function extract numerical indices of a given range e-g B2

#### Usage

```
extractValuesFromRange(range, excelFormate)
```

## **Arguments**

range Range e-g B2

excelFormate TRUE if range is in spreadsheet formate

## Value

Number of starting row, ending row, starting column and ending column

#### Author(s)

Muhammad kashif

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#### **Examples**

```
rng <- c("B2")
exclF <- TRUE
rslt <- extractValuesFromRange(rng, exclF)</pre>
```

loeweModel

This function applies Loewe Model

# Description

This function applies Loewe Model

## Usage

loeweModel(xConcentration, yConcentration, drugYObs\_Mean, drugXObs\_Mean)

# Arguments

```
xConcentration X drug concentrations
yConcentration Y drug concentrations
drugYObs_Mean Mean of y drug observations
drugXObs_Mean Mean of x drug observations
```

## Value

Loewe Model values

## Author(s)

Muhammad kashif

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readFile	Reads experimental data from a file. This function reads the data from specified (excel,log, txt etc) file and store it in a data frame.
	specifiea (excei,iog, ixi eic) fue ana store ii in a aaia frame.

#### **Description**

Reads experimental data from a file. This function reads the data from specified (excel,log, txt etc) file and store it in a data frame.

#### Usage

```
readFile(
   filename,
   separator,
   sheet,
   noofrows_skip,
   readplates,
   numberofrowsperplate,
   platetype
)
```

# **Arguments**

filename Filename.ext. separator Any character(,; 'etc) that is used as a separator in specified file. Need to use only when reading excel files. It is the number of the excel sheet to sheet be read in a worksheet. Number of the rows in the file that should be skipped before starting the data noofrows\_skip reading. readplates Number of the plates that you want to read from a set of plates in a file. This parameter can only be used with excel files. Otherwise it will be ignored. numberofrowsperplate It is calculated on the basis of type of plates i-e number of rows per plates are 17 for 384 well plates(16 lines from plates + 1 header lines) and 9 for 96 well plates (8 lines from plates + 1 header lines). type of plate used i-e 384 or 96 well plate. platetype

#### Value

Data frame of file data.

#### Author(s)

Muhammad Kashif

```
f <- system.file("extdata", "optima.log", package="COMBIA")
fileDF <- readFile(filename = f, separator ="\t", sheet=1, noofrows_skip=0,
readplates=1, numberofrowsperplate=17, platetype="384")</pre>
```

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readFluostarPlates

Read a file and process it to calculate the Survival indeces(S.I). This function calls other functions to complete its task. It reads a file to separate and regenerate the missing platekeys. Checks are performed to keep regenerated missing keyvalues in sync with data. It calculates survival indeces of the provided control wells, where wells should always be in triplet form that is control well range, empty well range and case well range. It can also handle the double plate experiments in which one plate is read twice and only one of them is selected in S.I calculations. Secondly it can also read the data from the file where a plate is read only one time, still it cope with variations if an experiment is repeated twice or many time in adjacent rows in the file.

#### **Description**

Read a file and process it to calculate the Survival indeces(S.I). This function calls other functions to complete its task. It reads a file to separate and regenerate the missing platekeys. Checks are performed to keep regenerated missing keyvalues in sync with data. It calculates survival indeces of the provided control wells, where wells should always be in triplet form that is control well range, empty well range and case well range. It can also handle the double plate experiments in which one plate is read twice and only one of them is selected in S.I calculations. Secondly it can also read the data from the file where a plate is read only one time, still it cope with variations if an experiment is repeated twice or many time in adjacent rows in the file.

## Usage

```
readFluostarPlates(
   filename,
   separator = ",",
   noofrows_skip = 0,
   sheet = "1",
   readplates = 1,
   platetype,
   doubleplateexperiment = TRUE,
   keyposition,
   selectionkey,
   platekey,
   rowsperexperiment = 1,
   wells
)
```

## **Arguments**

filename value of this argument should be path and filename.ext e=g "e:/optima.txt".

separator is the sepration character within the file assigned to filename.

noofrows\_skip Number of the rows in the file that should be skipped before starting the data

reading.

sheet Need to use only when reading excel files. It is the number of the excel sheet to

be read in a worksheet.

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readplates Number of the plates to read from a set of plates from an excel file, This feature

is only workable with xls files.

platetype Two types of plate formates are supported 384 and 96 wells.

doubleplateexperiment

This parameter can have TRUE & FALSE values only. It is set to TRUE when

an experiment is read twice.

keyposition 
It is the position of key in the header. Currently it is located at the second

position but it can be at any position in the header.

selectionkey value, that will be used during the selection of plate. Current value is 65000.

platekey barcode of the plate whose wells you want to measure for Survival index

rowsperexperiment

It is the argument that specifies if the same experiment is repeated and how many times in a plate. If an experiment is repeated twice in adjacent rows then average

of its values will be used in the SI calculation.

wells This argument can take a list of arguments in the triplet form. Where first ar-

gument of triplet is the range of control wells, second argument is the range of empty wells while third one is the range of case wells. It is made so that in labs plates layouts can differ greatly. By using this triplet scheme one can handel a number of palte layouts. Values should be given in the according to plate range e-g a4:d5 means start from the a(1) row and first column and continue to d(4)

row 5th column.

#### Value

Matrix of S.I.

#### Author(s)

Muhammad Kashif

#### **Examples**

readFMCAValues

Read data from raw FMCA format and clean for outliers

## Description

Read data from raw FMCA format and clean for outliers

16 readFMCAValues

#### Usage

```
readFMCAValues(
   file,
   platetype,
   keyposition,
   selectionkey,
   platekey,
   wells,
   minThersholdForCVCal,
   minThersholdForCV,
   yConcentration,
   xConcentration
)
```

## **Arguments**

file Name of file to be read platetype 384 etc keyposition Bar code position

selectionkey 65000 platekey Barcode

wells Wells ranges minThersholdForCVCal

militine should be eveal

Thresolld for data outliears in CV

minThersholdForCV

Thresold of values in CV not to remove

yConcentration Concentrations of y drug xConcentration Concentrations of x drug

# Value

Matrix of replicated survival values

## Author(s)

Muhammad kashif

readMacSynergyValues Read data from macsynergyII formate and clean for outliers

#### **Description**

Read data from macsynergyII formate and clean for outliers

#### Usage

```
readMacSynergyValues(
    file,
    sheet,
    nrow = 41,
    wellRangesExcel,
    minThersholdForCVCal,
    minThersholdForCV,
    survivalFunc
)
```

## **Arguments**

```
file Name of fiele to be read

sheet Sheet Number

nrow Number of rows in the sheet

wellRangesExcel

TRUE if wells in excel formate

minThersholdForCVCal

Thresolld for data outliears in CV

minThersholdForCV

Thresold of values in CV not to remove

survivalFunc <- function (x,y,z) (x-z)/(y-z) # It can be any function
```

#### Value

Matrix of replicated values

#### Author(s)

Muhammad kashif

18 readOtherValues

readOtherValues

Read data from raw format and clean for outliers

#### **Description**

Read data from raw format and clean for outliers

# Usage

```
readOtherValues(
   file,
   sheet,
   rskip = 0,
   cStart = 1,
   wellRangesExcel,
   platetype,
   minThersholdForCVCal,
   minThersholdForCV,
   survivalFunc,
   xConcentration,
   yConcentration
```

## **Arguments**

file Name of fiele to be read

sheet Sheet

rskip Number of rows to skip before reading data, default rskip=0 cStart Number of column to start reading data, default cStart=1

wellRangesExcel

well ranges in excel formate

platetype 384 or 96 minThersholdForCVCal

Thresolld for data outliears in CV

minThersholdForCV

Thresold of values in CV not to remove

survivalFunc A function to calculate survival values

xConcentration Concentrations of drug at x-axis yConcentration Concentrations of drugs at y-axis

## Value

Matrix of survival values of experimental replicates

## Author(s)

Muhammad kashif

removeOutliers 19

#### **Examples**

removeOutliers

This function Remove Outliers

## **Description**

This function Remove Outliers

#### Usage

removeOutliers(arrangeReplicates, minThersholdForCVCal, minThersholdForCV)

#### **Arguments**

```
arrangeReplicates
A data matrix
minThersholdForCVCal
Threshold for value removal in CV
minThersholdForCV
Values to be excluded
```

#### Value

Replicate values

#### Author(s)

Muhammad kashif

20 selectPlate

selectPlate

Select one of the two read plates and built a hashtable. One plate from each pair of the read plate is selected in case of double plate experinment on the basis of presence of minimum selection key and if none have maxed out values then one with highest mean value is picked.

#### **Description**

Select one of the two read plates and built a hashtable. One plate from each pair of the read plate is selected in case of double plate experiment on the basis of presence of minimum selection key and if none have maxed out values then one with highest mean value is picked.

#### Usage

```
selectPlate(
  rawdata,
  processedbarcode,
  numberofrowsperplate,
  selectionkey,
  doubleplateexperiment
)
```

#### **Arguments**

rawdata

An object(dataframe) of rawdata.

processedbarcode

A vector of regenerated missing keyvalues. In this case it is the output of function "extractKey".

number of row sperplate

This argument is not needed when you call function "readFluostarPlates". The number of rows depends upon the geometry of the plates. These are 16 in case of 384well paltes.

selectionkey

keyvalue on basis of which a plate is slected from a pair of plates read in double plate experiment.

doubleplateexperiment

This parameter can have TRUE & FALSE values only. It is set to TRUE when an experiment is read twice.

#### Value

A hashtable of picked plates.

# Author(s)

Muhammad Kashif

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#### **Examples**

synAntPlot

This function plots the synergy analysis 2D and 3D graphs

# Description

This function plots the synergy analysis 2D and 3D graphs

## Usage

```
synAntPlot(
  processedData,
  xConcentration,
  yConcentration,
  xDrug,
  yDrug,
  cellLine
)
```

# **Arguments**

```
processedData A matrix to plot
xConcentration X drug concentrations
yConcentration Y drug concentrations
xDrug X drug name
yDrug Y drug name
cellLine Cell line name
```

## Value

Plot the values

# Author(s)

Muhammad kashif

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#### **Examples**

```
dataFile <- system.file("extdata", "processedData.csv", package="COMBIA")
procData <- read.csv( dataFile, header=FALSE)
xConc <- c(0.00, 0.20, 0.39, 0.78, 1.56, 3.12, 6.25, 12.50, 25.00, 50)
yConc <- c(128, 64, 32, 16, 8, 4, 2, 0)
xD <- "X_Drug"
yD <- "Y_Drug"
clN <- "myCell"
rslt <- synAntPlot(as.matrix(procData),xConc,yConc, xD, yD, clN)</pre>
```

synergySignificant

Function calculates significant synergy/antagonism

## **Description**

Function calculates significant synergy/antagonism

# Usage

```
synergySignificant(
   synergyCalculationLists,
   noOfRows,
   noOfCols,
   xDrug,
   yDrug,
   cellLine
)
```

# Arguments

synergyCalculationLists

List of synergy antagonism calculations

noOfRows Number of rows
noOfCols Number of columns

xDrug Name of drug on x-axis
yDrug Name of drug on y-axis
cellLine Cell Line

#### Value

Processed data

## Author(s)

Muhammad kashif

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```
dataFile <- system.file("extdata", "rawDataPreProcessed.csv", package="COMBIA")
dataSample <- read.csv(dataFile, header=FALSE)
nR <- 8
nC <- 10
rslt <- applyBliss(nR, nC, as.matrix(dataSample), 100)
synergySignificant(rslt, nR, nC,"A", "B", "Cell")</pre>
```

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