Starling-May18 Projects/Katarina Stuart/KStuart.Starling-Aug18/At1_Genome/Analysis/2023-01-18.SizeGenome

PDF Version generated by

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on

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2023-01-18.SizeGenome



Genome Size

Jellyfish

https://bioinformatics.uconn.edu/genome-size-estimation-tutorial/

```
#!/bin/bash -e
#SBATCH --job-name=2023_02_14.genomesize_jellyfish.sl
#SBATCH --account=uoa02613
#SBATCH --time=00-04:00:00
#SBATCH --mem=50GB
#SBATCH --output=%x_%j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=4
#SBATCH --profile task
#SBATCH --array=1-4
module load Jellyfish/2.3.0-gimkl-2020a
KMER=$(sed "${SLURM_ARRAY_TASK_ID}q;d" /nesi/nobackup/uoa02613/kstuart_projects/At1_MynaGenome/analysis/genomesize/kmer_list.txt)
echo "working with kemr:" $KMER
cd /nesi/nobackup/uoa02613/kstuart_projects/At1_MynaGenome/analysis/genomesize
R1=/nesi/nobackup/uoa02613/Myna\_ONT\_2022/Myna\_ONT\_RAW\_dec/Basecalling\_guppy 6.2.1/pilon\_polishing/Myna\_10x\_processed\_R1\_val\_1.fq.gz
R2 = /nesi/nobackup/uoa02613/Myna\_ONT\_2022/Myna\_ONT\_RAW\_dec/Basecalling\_guppy 6.2.1/pilon\_polishing/Myna\_10x\_processed\_R2\_val\_2.fq.gz
jellyfish count -t 16 -C -m ${KMER} -s 10G -o ${KMER}mer_out --min-qual-char=? <(zcat $R1) <(zcat $R2)
jellyfish histo -o ${KMER}mer out.histo ${KMER}mer out
```

genome size calculations from histogram

```
module load R/4.2.1-gimkl-2022a
R

setwd("/nesi/nobackup/uoa02613/kstuart_projects/At1_MynaGenome/analysis/genomesize")

dataframe20 <- read.table("20mer_out.histo")

pdf("histo2.pdf")
plot(dataframe20[3:200,], type="l", xlab="K-mer Length", ylab="K-mer Count")
abline(v = 31, col="red", lwd=3)
abline(v = 7, col="blue", lwd=3)
dev.off()
```

31: 36167636

sum(as.numeric(dataframe20[7:9999,1]*dataframe20[7:9999,2]))

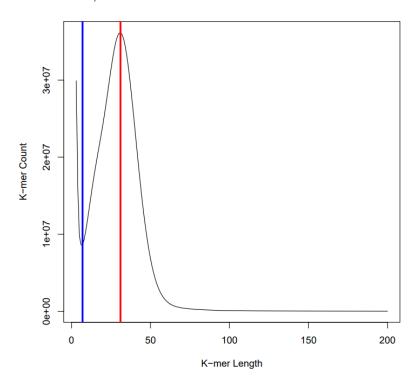
36,028,219,169

sum(as.numeric(dataframe20[7:9999,1]*dataframe20[7:9999,2]))/31

Est. Size: 1,162,200,618

1040603622 /1162200618

= 0.8953735% complete



Some additional numbers from the other K-mer values:

module load R/4.2.1-gimkl-2022a R

setwd("/nesi/nobackup/uoa02613/kstuart_projects/At1_MynaGenome/analysis/genomesize")

dataframe18 <- read.table("18mer_out.histo")
sum(as.numeric(dataframe18[7:9999,1]*dataframe18[7:9999,2]))/33 #1142465245

dataframe19 <- read.table("19mer_out.histo")
sum(as.numeric(dataframe19[7:9999,1]*dataframe19[7:9999,2]))/32 #1150752569

dataframe21 <- read.table("21mer_out.histo")
sum(as.numeric(dataframe21[6:9999,1]*dataframe21[6:9999,2]))/30 #1177818515

dataframe22 <- read.table("22mer_out.histo")
sum(as.numeric(dataframe18[7:9999,1]*dataframe18[7:9999,2]))/29 #1300046659

Merqury QV assessment

https://au-mynotebook.labarchives.com/share/BABS%2520Genome/NDk1LjN8MjkyMjMvMzgxL1RyZWVOb2RlLzExODM5MTI5MTB8MTI1Ny4zMtps://au-mynotebook.labarchives.com/share/BABS%2520Genome/NDk1LjN8MjkyMjMvMzgxL1RyZWVOb2RlLzExODM5MTI5MTB8MTI1Ny4zMtps://au-mynotebook.labarchives.com/share/BABS%2520Genome/NDk1LjN8MjkyMjMvMzgxL1RyZWVOb2RlLzExODM5MTI5MTB8MTI1Ny4zMtps://au-mynotebook.labarchives.com/share/BABS%2520Genome/NDk1LjN8MjkyMjMvMzgxL1RyZWVOb2RlLzExODM5MTI5MTB8MTI1Ny4zMtps://au-mynotebook.labarchives.com/share/BABS%2520Genome/NDk1LjN8MjkyMjMvMzgxL1RyZWVOb2RlLzExODM5MTI5MTB8MTI1Ny4zMtps://au-mynotebook.labarchives.com/share/BABS%2520Genome/NDk1LjN8MjkyMjMvMzgxL1RyZWVOb2RlLzExODM5MTI5MTB8MTI1Ny4zMtps://au-mynotebook.labarchives.com/share/BABS%2520Genome/NDk1LjN8MjkyMjMvMzgxL1RyZWVOb2RlLzExODM5MTI5MTB8MTI1Ny4zMtps://au-mynotebook.labarchives.com/share/BABS%2520Genome/NDk1LjN8MjkyMjMvMzgxL1RyZWVOb2RlLzExODM5MTI5MTB8MTI3MTB8MTI

Merqury uses Meryl to make kmer databases and then compares kmer profiles from genome assemblies with those from raw short read data.

install

conda create -n merqury -c conda-forge -c bioconda merqury openjdk=11

environment variable 'MERQURY' set to /nesi/nobackup/uoa02613/kstuart_projects/programs/miniconda/envs/mamba/envs/merqury/share/merqury

- # To activate this environment, use
- # \$ conda activate merqury
- # To deactivate an active environment, use
- # \$ conda deactivate

module load Miniconda3

CONDA_BASE=\$(conda info --base)

source \${CONDA_BASE}/etc/profile.d/conda.sh

c/nesi/nobackup/uoa02613/kstuart_projects/programs/miniconda/envs/mcclintock

conda create -n merqury -c conda-forge -c bioconda merqury openjdk=11

conda create --prefix /nesi/nobackup/uoa02613/kstuart_projects/programs/miniconda/envs/mamba/envs/merqury/share/merqury

conda activate /nesi/nobackup/uoa02613/kstuart_projects/programs/miniconda/envs/mamba/envs/merqury/share/merqury

environment variable 'MERQURY' set to /nesi/nobackup/uoa02613/kstuart_projects/programs/miniconda/envs/mamba/envs/merqury/share/merqury

- # To activate this environment, use
- # \$ conda activate merqury
- # To deactivate an active environment, use
- # \$ conda deactivate

setting up meryl database

First, select the best k based on the genome size (21 is good for most).

cd /nesi/nobackup/uoa02613/kstuart_projects/At1_MynaGenome/analysis/genomesize/merqury

MERQURY=/nesi/nobackup/uoa02613/kstuart_projects/programs/miniconda/envs/mamba/envs/merqury/share/merqury

GSIZE=1.040e9

\${MERQURY}/best_k.sh \$GSIZE

genome: 1.040e9

tolerable collision rate: 0.001

19.9591

k=20

Next, set up a variable pointing to the kmer read files and generate individual meryl databases, e.g.: need 24 hrs on the below settings(but actually ran in 15 mins???)

#!/bin/bash -e

#SBATCH --job-name=2023_02_08.genomesize_meryl.sl

```
#SBATCH --account=uoa02613
#SBATCH --time=00-12:00:00
#SBATCH --mem=5GB
#SBATCH --output=%x_%j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=2
#SBATCH --profile task
cd /nesi/nobackup/uoa02613/kstuart_projects/At1_MynaGenome/analysis/genomesize/merqury
module load Miniconda3
CONDA_BASE=$(conda info --base)
source ${CONDA_BASE}/etc/profile.d/conda.sh
conda activate /nesi/nobackup/uoa02613/kstuart_projects/programs/miniconda/envs/mamba/envs/merqury
KMERREADS = "/nesi/nobackup/uoa02613/Myna\_ONT\_2022/Myna\_ONT\_RAW\_dec/Basecalling\_guppy 6.2.1/pilon\_polishing/Myna\_10x\_processed\_R*\_val\_*.fq.gz" and the contraction of the contraction 
# Build meryl dbs
for FASTQ in $KMERREADS; do
meryl k=$k count output $(basename ${FASTQ} .fq.gz).meryl $FASTQ
MERYLPARTS="*.meryl"
MERYLDB=myna.10x.meryl
meryl union-sum output $MERYLDB $MERYLPARTS
```

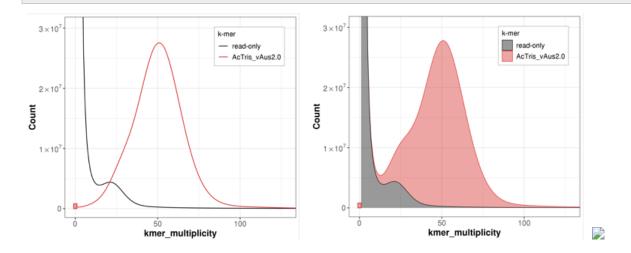
Running Merqury

NOTE: Merqury is very fussy about file paths and wants everything local. It seems to work best with things in the current directory for a single run:

Run Merqury on each assembly:

```
#!/bin/bash -e
#SBATCH --job-name=2023_02_10.genomesize_merqury.sl
#SBATCH --account=uoa02613
#SBATCH --time=00-12:00:00
#SBATCH --mem=15GB
#SBATCH --output=%x_%j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=16
#SBATCH --profile task
cd /nesi/nobackup/uoa02613/kstuart projects/At1 MynaGenome/analysis/genomesize/merqury
module load Miniconda3
CONDA_BASE=$(conda info --base)
source ${CONDA_BASE}/etc/profile.d/conda.sh
conda activate /nesi/nobackup/uoa02613/kstuart_projects/programs/miniconda/envs/mamba/envs/merqury
module load R/4.2.1-gimkl-2022a #also made sure to install.packages("argparse")
GENOME=/nesi/nobackup/uoa02613/kstuart_projects/At1_MynaGenome/analysis/curation/step4_scaffolding/ragtag_atris_synteny/renamed/AcTris_vAus2.0.fasta
MERQURY=/nesi/nobackup/uoa02613/kstuart_projects/programs/miniconda/envs/mamba/envs/merqury/share/merqury
MERYLDB=myna.10x.meryl
$MERQURY/merqury.sh $MERYLDB $GENOME AcTris_vAus
```

#\$MERQURY/eval/spectra-cn.sh \$MERYLDB \$GENOME \$(basename \$GENOME .fasta)



AcTris_vAus2.0.completeness.stats:
AcTris_vAus2.0 all 972627157 1066771063 91.1749