Starling-May18 Projects/Katarina Stuart/KStuart.Starling-Aug18/Nc3_HihiSV/Data/2023-01-23.SVcalling

PDF Version generated by

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on

Aug 15, 2024 @03:06 PM NZST

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SV Var calling: 82 myna WGR

Choosing individuals that have high enough coverage:

13-28G dup mapped data for these samples

cd /nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling

ls -lh /nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/mapped_reads/*sorted.dup.bam | sed 's/G / /g' | sort -t' ' -k6,6n -k6,6V | cut -d' ' -f 10 | sed 's/.sorted.dup.bam//g' | tail -n 31 | sort | sed 's/V/t/g' | cut -f 9 > sample_file_ids_highcov.txt

Smoove:

For when you want to use lumpy on a large cohort use smoove: https://github.com/brentp/smoove

Step1: SV calling done separately for each sample

```
#!/bin/bash -e
#SBATCH --job-name=2024_01_26.SVCalling_hihiwgs_smoove_step1.sl
#SBATCH --account=uoa00338
#SBATCH --time=00-12:00:00
#SBATCH --mem=5GB
#SBATCH --output=%x %j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=2
#SBATCH --profile task
#SBATCH --array=1-31
#SBATCH --partition=milan
# load modules
module purge
module load smoove/0.2.8-Miniconda3
SAMPLE=$(sed
"${SLURM ARRAY TASK ID}q;d" /nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling/sample file ids highcov.txt)
echo "working with sample:" ${SAMPLE}
# set paths
DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/mapped_reads/
BAM=${DIR}/${SAMPLE}.sorted.dup.bam
GENOME=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/resources/Ncf_H98617_scaffolded_genome.fa
OUT_DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_smoove
```

```
#Run smoove step 1 smoove call --outdir ${OUT_DIR} --name ${SAMPLE} --fasta ${GENOME} -p 1 --genotype ${BAM}
```

Step2: Get the union of sites across all samples

```
#!/bin/bash -e
#SBATCH --job-name=2024 01 26.SVCalling hihiwgs smoove step2.sl
#SBATCH --account=uoa00338
#SBATCH --time=00-48:00:00
#SBATCH --mem=5GB
#SBATCH --output=%x %j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=2
#SBATCH --profile task
#SBATCH --partition=milan
# load modules
module purge
module load smoove/0.2.8-Miniconda3
# set paths
GENOME=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/resources/Ncf_H98617_scaffolded_genome.fa
OUT_DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_smoove
#Run smoove step 2
smoove merge --name merged -f ${GENOME} --outdir ${OUT_DIR} ${OUT_DIR}/*.genotyped.vcf.gz
# this will create ./merged.sites.vcf.gz
```

Step3: genotype each sample at those sites

```
#!/bin/bash -e
#SBATCH --job-name=2024 01 26.SVCalling hihiwgs smoove step3.sl
#SBATCH --account=uoa00338
#SBATCH --time=00-12:00:00
#SBATCH --mem=5GB
#SBATCH --output=%x %j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=4
#SBATCH --profile task
#SBATCH --array=1-31
#SBATCH --partition=milan
# load modules
module purge
module load smoove/0.2.8-Miniconda3
module load duphold/0.2.3
SAMPLE=$(sed
```

```
"${SLURM_ARRAY_TASK_ID}q;d" /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/sample_file_ids_highcov.txt)
echo "working with sample:" ${SAMPLE}

# set paths
DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/mapped_reads/
BAM=${DIR}/${SAMPLE}.sorted.dup.bam
GENOME=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/resources/Ncf_H98617_scaffolded_genome.fa
OUT_DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_smoove

#Run smoove step 3
smoove genotype -d -x -p 4 --name ${SAMPLE}_joint --outdir ${OUT_DIR} --fasta ${GENOME} --vcf ${OUT_DIR}/merged.sites.vcf.gz ${BAM}}
```

Step4: paste all the single sample VCFs with the same number of variants to get a single, squared, joint-called file

```
#!/bin/bash -e
#SBATCH --job-name=2024 01 26.SVCalling hihiwgs smoove step4.sl
#SBATCH --account=uoa00338
#SBATCH --time=00-12:00:00
#SBATCH --mem=5GB
#SBATCH --output=%x %j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=4
#SBATCH --profile task
#SBATCH --partition=milan
# load modules
module purge
module load smoove/0.2.8-Miniconda3
# set paths
OUT_DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_smoove
cd ${OUT_DIR}
#Run smoove step 4
smoove paste --name smoove_hihi ${OUT_DIR}/*_joint-smoove.genotyped.vcf.gz
```

Step5: annotate the variants with exons, UTRs that overlap from a GFF and annotate high-quality heterozygotes:

```
#!/bin/bash -e

#SBATCH --job-name=2024_01_31.SVCalling_hihiwgs_smoove_step5.sl

#SBATCH --account=uoa02613

#SBATCH --time=00-12:00:00

#SBATCH --mem=5GB

#SBATCH --output=%x_%j.errout

#SBATCH --mail-user=katarina.stuart@auckland.ac.nz

#SBATCH --mail-type=ALL

#SBATCH --nodes=1

#SBATCH --ntasks=1

#SBATCH --cpus-per-task=4
```

```
#SBATCH --profile task
#SBATCH --partition=milan
# load modules
module purge
module load smoove/0.2.8-Miniconda3
# set paths
OUT\_DIR = /nesi/nobackup/uoa00338/kstuart\_projects/Nc3\_HihiSV/data/variant\_calling/SV\_smoove
cd ${OUT DIR}
module load AGAT/1.0.0-gimkl-2022a-Perl-5.34.1-R-4.2.1
GFF=/nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/annotation/Ncf H98617 scaffolded liftoff.gff
agat_convert_sp_gxf2gxf.pl --gff $GFF -o tsebra_fix.gff
#my annotation file didn't have a 'Name=" column and thus kept giving me an error message
#ERROR: no records found with 'gene' type in gff
#https://github.com/brentp/smoove/issues/184
#Manually add this field to the end of each gene line
awk -v search="gene" 'BEGIN { OFS="\t" } {
  if ($3 == search) {
     $9 = $9";Name=dummy"
  print
}' tsebra_fix.gff > tsebra_fix2.gff
#Run smoove step 5
smoove annotate --gff tsebra_fix2.gff smoove_hihi.smoove.square.vcf.gz | bgzip -c > smoove_hihi.smoove.square.anno.vcf.gz
```

Step6: Filtering

```
#!/bin/bash -e
#SBATCH --job-name=2024 01 31.SVCalling starlingwgs smoove step6.sl
#SBATCH --account=uoa02613
#SBATCH --time=00-12:00:00
#SBATCH --mem=1GB
#SBATCH --output=%x_%j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=4
#SBATCH --profile task
#SBATCH --partition=milan
# load modules
module purge
module load BCFtools/1.13-GCC-9.2.0
module load VCFtools/0.1.15-GCC-9.2.0-Perl-5.30.1
# set paths
OUT DIR=/nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling/SV smoove
GVCF=/nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling/SV smoove/smoove hihi.smoove.square.anno.vcf.gz
cd ${OUT_DIR}
```

Delly:

Step1: SV calling done separately for each sample

```
#!/bin/bash -e
#SBATCH --job-name=2024_01_26.SVCalling_hihiwgs_delly_step1.sl
#SBATCH --account=00338
#SBATCH --time=00-24:00:00
#SBATCH --mem=10GB
#SBATCH --output=%x_%j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=2
#SBATCH --profile task
#SBATCH --array=1-31
# load modules
module purge
module load Delly/1.1.3
SAMPLE=$(sed "${SLURM ARRAY TASK ID}q;d"
/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/sample_file_ids_highcov.txt)
echo "working with sample:" ${SAMPLE}
# set paths
```

```
DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/mapped_reads/
BAM=${DIR}/${SAMPLE}.sorted.dup.bam
GENOME=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/resources/Ncf_H98617_scaffolded_genome.fa
OUT_DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_delly

#Run delly step 1
delly call -0 ${OUT_DIR}/${SAMPLE}.bcf -g ${GENOME} ${BAM}
```

Step2: Merge SV sites into a unified site list

```
#!/bin/bash -e
#SBATCH --job-name=2024_01_26.SVCalling_hihiwgs_delly_step2.sl
#SBATCH --account=uoa00338
#SBATCH --time=00-24:00:00
#SBATCH --mem=10GB
#SBATCH --output=%x %j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=2
#SBATCH --profile task
# load modules
module purge
module load Delly/1.1.3
#create sample BCF file input list
BCF_LIST=""
for SAMPLE in $(cat /nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling/sample file ids highcov.txt)
BCF_LIST="${BCF_LIST} /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_delly/${SAMPLE}.bcf"
done
echo $BCF_LIST
# set paths
OUT_DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_delly
#Run delly step2
delly merge -o ${OUT_DIR}/merged_sites.bcf ${BCF_LIST}
```

Step3: Genotype this merged SV site list across all samples

```
#!/bin/bash -e

#SBATCH --job-name=2024_01_26.SVCalling_hihiwgs_delly_step3.sl

#SBATCH --account=uoa00338

#SBATCH --time=00-24:00:00

#SBATCH --mem=20GB

#SBATCH --output=%x_%j.errout

#SBATCH --mail-user=katarina.stuart@auckland.ac.nz

#SBATCH --mail-type=ALL

#SBATCH --nodes=1

#SBATCH --ntasks=1
```

```
#SBATCH --cpus-per-task=2
#SBATCH --profile task
#SBATCH --array=1-31
# load modules
module purge
module load Delly/1.1.3
SAMPLE=$(sed
"${SLURM_ARRAY_TASK_ID}q;d" /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/sample_file_ids_highcov.txt)
echo "working with sample:" ${SAMPLE}
# set paths
GENOME=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/resources/Ncf_H98617_scaffolded_genome.fa
OUT_DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_delly
DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/mapped_reads/
BAM=${DIR}/${SAMPLE}.sorted.dup.bam
#Run delly step3
delly call -g ${GENOME} -v ${OUT_DIR}/merged_sites.bcf -o ${OUT_DIR}/${SAMPLE}.rep.geno.bcf ${BAM}
```

Step4: Merge all genotyped samples to get a single VCF/BCF using BCFtools merge

```
#!/bin/bash -e
#SBATCH --job-name=2024_01_26.SVCalling_hihiwgs_delly_step4.sl
#SBATCH --account=uoa00338
#SBATCH --time=00-04:00:00
#SBATCH --mem=10GB
#SBATCH --output=%x %j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=2
#SBATCH --profile task
# load modules
module purge
module load BCFtools/1.13-GCC-9.2.0
#create sample BCF file input list
BCF_LIST=""
for SAMPLE in $(cat /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/sample_file_ids_highcov.txt)
BCF_LIST="${BCF_LIST} /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_delly/${SAMPLE}.rep.geno.bcf"
done
echo $BCF_LIST
OUT DIR=/nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling/SV delly
#Run delly step4
bcftools merge -m id -O b -o ${OUT DIR}/merged rep geno.bcf ${BCF LIST}
```

Step5: Convert BCF to VCF

```
#!/bin/bash -e
#SBATCH --job-name=2024_01_26.SVCalling_hihiwgs_delly_step5.sl
#SBATCH --account=uoa00338
#SBATCH --time=00-04:00:00
#SBATCH --mem=10GB
#SBATCH --output=%x_%j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=2
#SBATCH --profile task
# load modules
module purge
module load BCFtools/1.13-GCC-9.2.0
# set paths
OUT_DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_delly
#Run delly step5
bcftools view ${OUT_DIR}/merged_rep_geno.bcf -o ${OUT_DIR}/merged_rep_geno.vcf
```

Step 6: filtering

```
#!/bin/bash -e
#SBATCH --job-name=2024_01_31.SVCalling_hihiwgs_delly_step6.sl
#SBATCH --account=uoa02613
#SBATCH --time=00-12:00:00
#SBATCH --mem=1GB
#SBATCH --output=%x_%j.errout
#SBATCH --mail-user=katarina.stuart@auckland.ac.nz
#SBATCH --mail-type=ALL
#SBATCH --nodes=1
#SBATCH --ntasks=1
#SBATCH --cpus-per-task=4
#SBATCH --profile task
# load modules
module purge
module load BCFtools/1.13-GCC-9.2.0
module load PLINK/1.09b6.16
module load VCFtools/0.1.15-GCC-9.2.0-Perl-5.30.1
cd /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_delly
VCF=/nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling/SV delly/merged rep geno.vcf
bcftools view -f PASS ${VCF} > merged_delly_pass.vcf
#generate file with info (like length and type)
grep -v "^#" merged_delly_pass.vcf | cut -f1-8 | sed 's/;\t/g' | cut -f1,2,3,5,8,11 | sed 's/END=//g' | awk '{print $0,"\t",$6-$2}'
> merged delly pass info.txt
```

```
#count number of (genotype) PASS per SNP
grep -v "^#" merged_delly_pass.vcf | grep -o -n 'PASS' | cut -d : -f 1 | uniq -c > merged_delly_pass_infopass.txt
#final info file has the final column with a count of how many snps had a pass
paste merged_delly_pass_info.txt <(sed 's/[[:space:]]\+/t/g' merged_delly_pass_infopass.txt | cut -f 2 ) > merged_delly_pass_infoall.txt
#generate list of SNPs to keep (75% + PASS for samples)
awk '$8>24' merged_delly_pass_infoall.txt | cut -f3 > merged_delly_pass_VARs.snpID
vcftools --vcf merged delly pass.vcf --snps merged delly pass VARs.snpID --keep-INFO-all --recode --out merged delly SNPfilt
#filter out z and w chrom contigs
SEX=/nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/annotation/Ncf sex linked contigs.txt
LIST=
for CHROM in $(cut -f 1 $SEX)
LIST="${LIST} "--not-chr" $CHROM"
echo $LIST
vcftools --vcf merged delly SNPfilt.recode.vcf $LIST --keep-INFO-all --recode --out merged delly autosomes
#Filter to just the superscaffolds
cat <(zgrep "/#" merged delly autosomes.recode.vcf) <(zgrep -v "/#" merged delly autosomes.recode.vcf | grep -v "Ncf contig" ) >
merged delly chroms.vcf
```

Manta:

Manta SV calling requires just a single line (and a bit of patience)

```
#!/bin/bash -e

#SBATCH --job-name=2024_01_26.SVCalling_hihi_manta.sl

#SBATCH --account=uoa02613

#SBATCH --time=00-150:00:00

#SBATCH --mem=150GB

#SBATCH --output=%x_%j.errout

#SBATCH --mail-user=katarina.stuart@auckland.ac.nz

#SBATCH --mail-type=ALL

#SBATCH --ndes=1

#SBATCH --ntasks=1

#SBATCH --rcpus-per-task=16

#SBATCH --profile task

# load modules

module load manta/1.6.0-gimkl-2020a-Python-2.7.18
```

```
# set paths
GENOME=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/resources/Ncf_H98617_scaffolded_genome.fa
OUT_DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_manta
# create list of input BAM files for Manta
FILE LIST=""
for SAMPLE_NUMBER in {1..31}
do
SAMPLE=$(sed "${SAMPLE NUMBER}q;d"
/nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling/sample file ids highcov.txt)
DIR=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/mapped_reads
BAM=${DIR}/${SAMPLE}.sorted.dup.bam
FILE_LIST="${FILE_LIST} "--bam" ${BAM} "
done
echo ${FILE_LIST}
# This created a runWorkflow.py file for the job
configManta.py ${FILE_LIST} --referenceFasta ${GENOME} --runDir ${OUT_DIR}
# run manta
/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_manta/runWorkflow.py
```

Fixing inversions

```
cd /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_manta/results/variants

GENOME=/nesi/nobackup/uoa00338/kstuart_projects/Nc2_HihiWGS/data/resources/Ncf_H98617_scaffolded_genome.fa

SAMTOOLS=/opt/nesi/CS400_centos7_bdw/SAMtools/0.1.19-gimkl-2017a/bin/samtools

cp diploidSV.vcf.gz diploidSV2.vcf.gz
gunzip diploidSV2.vcf.gz

module purge
module load Python/2.7.18-gimkl-2020a

python convertInversion.py ${SAMTOOLS} ${GENOME} diploidSV.vcf > diploidSV_inversions.vcf
```

Filtering

```
# load modules
module purge
module load BCFtools/1.13-GCC-9.2.0
module load PLINK/1.09b6.16
module load VCFtools/0.1.15-GCC-9.2.0-Perl-5.30.1

cd /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_manta/results/variants

VCF=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_manta/results/variants/diploidSV_inversions.vcf

bcftools view -f PASS ${VCF} > diploidSV_pass.vcf

#filter out z and w chrom contigs

SEX=/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/annotation/Ncf_sex_linked_contigs.txt

LIST=

for CHROM in $(cut -f 1 $SEX)
```

```
do
LIST="${LIST} "--not-chr" $CHROM"
done

echo $LIST

vcftools --vcf diploidSV_pass.vcf $LIST --keep-INFO-all --recode --out manta_autosomes

#Filter to just the superscaffolds
cat <(zgrep "^#" manta_autosomes.recode.vcf) <(zgrep -v "^#" manta_autosomes.recode.vcf | grep -v "Ncf_contig" ) > manta_chroms.vcf
```

SURVIVOR

Installing

cd /nesi/nobackup/uoa02613/kstuart_projects/programs git clone https://github.com/fritzsedlazeck/SURVIVOR.git cd SURVIVOR/Debug make

cd /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_survivor
In -s /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_smoove/smoove_hihi_genofiltered.vcf
In -s /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_delly/merged_delly_chroms.vcf
In -s /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_manta/results/variants/manta_chroms.vcf

Splitting up the currnet SVCF files so we have 1 file per individual PER SVcaller, so I can work/merge with them individually.

```
module load VCFtools/0.1.15-GCC-9.2.0-Perl-5.30.1

mkdir split_vcfs

for SAMPLE in $(cat /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/sample_file_ids_highcov.txt);
do
echo ${SAMPLE}}

mkdir split_vcfs/${SAMPLE}}

vcftools --vcf smoove_hihi_genofiltered.vcf --indv $SAMPLE --recode --recode-INFO-all --out

split_vcfs/${SAMPLE}/smoove_autosomes.${SAMPLE}}

vcftools --vcf merged_delly_chroms.vcf --indv $SAMPLE --recode --recode-INFO-all --out

split_vcfs/${SAMPLE}/delly_autosomes.${SAMPLE}}

vcftools --vcf manta_chroms.vcf --indv $SAMPLE --recode --recode-INFO-all --out

split_vcfs/${SAMPLE}/delly_autosomes.${SAMPLE}}

vcftools --vcf manta_chroms.vcf --indv $SAMPLE --recode --recode-INFO-all --out

split_vcfs/${SAMPLE}/manta_autosomes.${SAMPLE}}

done
```

Merging with genotype info

Splitting up each individual sample's 3 VCF files into het, homref, and homalt & merging across tools with SURVIVOR (but within samples)

```
module load BCFtools/1.16-GCC-11.3.0
cd /nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling/SV survivor
DIR=/nesi/nobackup/uoa02613/kstuart projects/programs/SURVIVOR/Debug/
for SAMPLE in $(cat /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/sample_file_ids_highcov.txt);
cd split vcfs/${SAMPLE}/
grep "^#\|0/0" delly autosomes.${SAMPLE}.recode.vcf > delly ${SAMPLE} homref.vcf
grep "^#\|0/1" delly_autosomes.${SAMPLE}.recode.vcf > delly_${SAMPLE}_het.vcf
grep "^#\|1/1" delly autosomes.${SAMPLE}.recode.vcf > delly ${SAMPLE} homalt.vcf
grep "^#\|0/0" manta autosomes.${SAMPLE}.recode.vcf > manta ${SAMPLE} homref.vcf
grep "/#\|0/1" manta autosomes.${SAMPLE}.recode.vcf > manta ${SAMPLE} het.vcf
grep "^#\\1/1" manta_autosomes.${SAMPLE}.recode.vcf > manta _${SAMPLE} homalt.vcf
grep "/#\|0/0" smoove autosomes.${SAMPLE}.recode.vcf > lumpy ${SAMPLE} homref.vcf
grep "^#\|0/1" smoove autosomes.${SAMPLE}.recode.vcf > lumpy ${SAMPLE} het.vcf
grep "^#\|1/1" smoove autosomes.${SAMPLE}.recode.vcf > lumpy ${SAMPLE} homalt.vcf
Is *${SAMPLE}*homref.vcf > homref ${SAMPLE}
Is *${SAMPLE}*het.vcf > het ${SAMPLE}
ls *${SAMPLE}*homalt.vcf > homalt ${SAMPLE}
#merging WITHIN genotype to make sure genotype is also in consensus (because SURVIVOR doesn't have a
genotype option)
${DIR}/SURVIVOR merge homref_${SAMPLE} 1000 2 1 1 0 30 ${SAMPLE}_survivor_homref.vcf
${DIR}/SURVIVOR merge het ${SAMPLE} 1000 2 1 1 0 30 ${SAMPLE} survivor het.vcf
${DIR}/SURVIVOR merge homalt ${SAMPLE} 1000 2 1 1 0 30 ${SAMPLE} survivor homalt.vcf
grep "^#" ${SAMPLE} survivor homref.vcf > ${SAMPLE} survivor header
grep -v "^#" ${SAMPLE} survivor homref.vcf > ${SAMPLE} survivor homref SNPs.vcf
grep -v "^#" ${SAMPLE} survivor het.vcf > ${SAMPLE} survivor het SNPs.vcf
grep -v "^#" ${SAMPLE}_survivor_homalt.vcf > ${SAMPLE}_survivor_homalt_SNPs.vcf
cat ${SAMPLE} survivor header ${SAMPLE} survivor homref SNPs.vcf ${SAMPLE} survivor het SNPs.vcf
${SAMPLE} survivor homalt SNPs.vcf > ${SAMPLE} survivor unsorted.vcf
bcftools sort ${SAMPLE}_survivor_unsorted.vcf > ${SAMPLE}_survivor_v2.vcf
```

```
echo ${SAMPLE}
grep -v "^#" ${SAMPLE}_survivor_v2.vcf | wc -l

#Make sure the final column contains GT info
awk -F'\t' 'BEGIN{OFS="\t"} { if ($12 == "./.:NaN:0:0,0:--:NaN:NaN:NAN:NAN:NAN") $12 = $11; print }' ${SAMPLE}_survivor_v2.vcf

> ${SAMPLE}_survivor_v3.vcf

cd ../../
done
```

The absent and present VCF files should have at least 2 genotypes (that are the same).

The final per-sample VCF should have only one genotype in it, because we merged across genotypes.

What to do about insertions?

At this stage the dataset may not have many insertions (INS). This is because LUMPY doesn't pick them up, and so it depends on the consensus between MANTA and DELLY as to how many you will have in your final data set. The way I see it, your obvious choices at this stage are:

- 1) Not care about it, and just proceed with the SVs you have (many papers do this). DONE
- 2) Choose to use a 4th program that picks up INS. You could even swap out LUMPY. Other programs I have looked into are GRIDDS, didn't work but I didn't troubleshoot it for too long.
- 3) Carry some of the DELLY and MANTA insertions back into the dataset using some other level of criteria

Then merge across samples

```
cd /nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling/SV survivor
Is */*/* survivor v3.vcf > allsample files
DIR=/nesi/nobackup/uoa02613/kstuart_projects/programs/SURVIVOR/Debug/
${DIR}/SURVIVOR merge allsample_files 1000 1 1 1 0 30 merged_rep_new.vcf
#exclude long SVs and high missingness
grep -v "^#" merged_rep_new.vcf | sed 's/;/\t/g' | cut -f 1,2,3,10 | sed 's/SVLEN=\|-//g' | awk '$4 > 50000' | cut -f3 > snpid_longSvs.txt
module load VCFtools/0.1.15-GCC-9.2.0-Perl-5.30.1
vcftools --vcf merged rep new.vcf --exclude snpid longSvs.txt --max-missing 0.5 --mac 1 --recode --recode-INFO-all --out hihi size
#remove TRA
grep -v "SVTYPE=TRA" hihi_size.recode.vcf > hihi_size2.recode.vcf
#remove duplicates
grep -v "^#" hihi size2.recode.vcf | cut -f 3 | sort | uniq -d > snpid duplicatedSvs.txt
vcftools --vcf hihi_size2.recode.vcf --exclude snpid_duplicatedSvs.txt --recode --recode-INFO-all --out hihi_filtered
#THINKING: not included yet - probably will
#Remove variants without (https://onlinelibrary.wiley.com/doi/10.1111/eva.13652)
#count number of genotypes for each SNP
```

```
grep -v "^#" hihi _filtered.recode.vcf | cut -f3 > genofilter_SNPID.txt
grep -v "^#" hihi filtered.recode.vcf | awk '{print gsub(/0V0/, "&")}' > genofilter homr.txt
grep -v "^#" hihi_filtered.recode.vcf | awk '{print gsub(/0V1/, "&")}' > genofilter_het.txt
grep -v "^#" hihi_filtered.recode.vcf | awk '{print gsub(/1V1/, "&")}' > genofilter_hom.txt
#final info file has the final column with a count of how many snps had a pass
paste genofilter_SNPID.txt <(sed 's/[[:space:]]\+\/t/g' genofilter_homr.txt | cut -f 2 ) <(sed 's/[[:space:]]\+\/t/g' genofilter_het.txt | cut -f 2 | cu
s/[:space:]\+/t/g' genofilter_hom.txt | cut -f 2) > genofilter_both.txt
###old method:
#needs at least 2 hets and 2 refs or alts
awk '$3 > 1 && ($2 > 1 || $4 > 1)' genofilter_both.txt | cut -f1 > genofilter1_snplist.txt
vcftools --vcf hihi_filtered.recode.vcf --snps genofilter1_snplist.txt --recode --recode-INFO-all --out hihi_genofiltered
#decided to actually reinclude these, and exclude non-het SNPs let
#so for now I had to generate a new VCF files of the ones I gnored in the first round of manual curation, so I can make their plots using my
code on the curation page
vcftools --vcf hihi filtered.recode.vcf --exclude genofilter1 snplist.txt --recode --recode-INFO-all --out hihi genofilteredextra
###current method: make list of SNPs to exclude later
# and exclude ones that don't have any het (it's just 22), and 8 that remain in the final dataset.
awk '$3 < 1' genofilter_both.txt | cut -f1 > SVs_to_remove.txt
```

Plotting PCA and looking at het

```
module load PLINK/1.09b6.16
module load VCFtools/0.1.15-GCC-9.2.0-Perl-5.30.1
cd /nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/variant_calling/SV_survivor
vcftools --vcf hihi filtered.recode.vcf --het --out hihi filtered
vcftools --vcf hihi_filtered.recode.vcf --plink --out hihi_filtered.plink
plink --file hihi_filtered.plink --pca --out hihi_filtered --make-rel --allow-extra-chr --chr-set 27
module load R/4.1.0-gimkl-2020a
R
library(ggplot2)
setwd("/nesi/nobackup/uoa00338/kstuart projects/Nc3 HihiSV/data/variant calling/SV survivor")
# PASS
pca.eigenvec <- read.table("hihi filtered.eigenvec", sep=" ")</pre>
pca g1 <- data.frame(PC1 = pca.eigenvec$V3, # the first eigenvector
             PC2 = pca.eigenvec$V4, # the second eigenvector
             PC3 = pca.eigenvec$V5, # the second eigenvector
             stringsAsFactors = FALSE)
population <- read.table("/nesi/nobackup/uoa00338/kstuart_projects/Nc3_HihiSV/data/sample_metadata.txt", header = T, sep = "\t")
pca_plot <- cbind(population,pca_g1)</pre>
```

```
ggplot(pca_plot ,aes(x=PC1,y=PC2, col = BAM_SIZE_G))+
geom_point(size=5,alpha=1)+
theme_classic(base_size = 18)
summary(lm(PC1 ~ BAM_SIZE_G, data=pca_plot )) #nonsig
summary(lm(PC2 ~ BAM_SIZE_G, data=pca_plot )) #nonsig
#need to also test insert size: <a href="https://accio.github.io/bioinformatics/2020/03/10/filter-bam-by-insert-size.html">https://accio.github.io/bioinformatics/2020/03/10/filter-bam-by-insert-size.html</a>
```

