Genome annotation

Final Project

For the project I received a file with a fragment (30000 nt long) of genomic DNA of *Tepidiforma bonchosmolovskayae* gen. nov., sp. nov.. It is a moderately thermophilic bacterium isolated from a Chukotka hot spring. However, the genome of it was not well characterized because of its recent discovery. In addition, *T. bonchosmolovskayae* is an organism which belongs to a new class *Tepidiforma* within *Chloroflexi*, and no other organisms from this class have been sequenced so far. As a result, it is relevant to annotate genes of this bacterium and represent enough biological information about it using all the tools which were studied during the course.

The project is divided into the several steps:

- 1. Annotation of all coding and non-coding genes
- 2. Identification of functions for the found hypothetical proteins
- 3. Description of the operon structure and for the long operons (longer than 4 genes) identification of the known regulatory regions if it is possible
 - 4. Finding of genes obtained by the bacteria through horizontal gene transfer (HGT)
 - 5. Finding of genes of the bacteria associated with secondary metabolites

1. Annotation of all coding and non-coding genes

For annotation I took the file with my variant (27.fasta) from the server and then used Prokka which is appropriate for the annotation of prokaryotic genomes.

1) I identified the number of coding sequences (CDS) for coding genes with the command

```
prokka --outdir prokka_result --locustag
prokka --kingdom Bacteria 27.fasta
```

and found the result

organism: Genus species strain

contigs: 1 bases: 30000 CDS: 32

2) I identified the number of coding sequences (CDS) for non-coding genes with the command

prokka --outdir prokka_non-coding --rfam -locustag prokkar --kingdom Bacteria 27.fasta

and found the result

organism: Genus species strain

contigs: 1 bases: 30000 CDS: 32

The results are the same, it means that there are <u>no any non-coding genes</u> in the fragment.

With Prokka I identified that some of the proteins are hypothetical ones and one of the aims is to recognise which functions hypothetical proteins have.

2. Identification of functions for the found hypothetical proteins

Prokka found 18 hypothetical proteins and their functions were found with the tools (1 – BLAST, 2 – Pfam, 3 – TMHMM).

For each protein the steps of identification are described:

1) With BLAST https://blast.ncbi.nlm.nih.gov/Blast.cgi for hypothetical protein #1 the result is

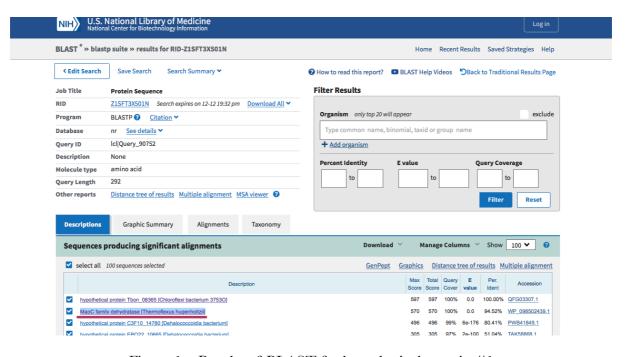


Figure 1 – Results of BLAST for hypothetical protein #1

The result is with good identity (94.52%), E-value (0.0), query cover (100%) and it can be suggested that our hypothetical protein has the same functions as <u>dehydratase</u> [Thermoflexus hugenholtzii].

2) The result for hypothetical protein #2

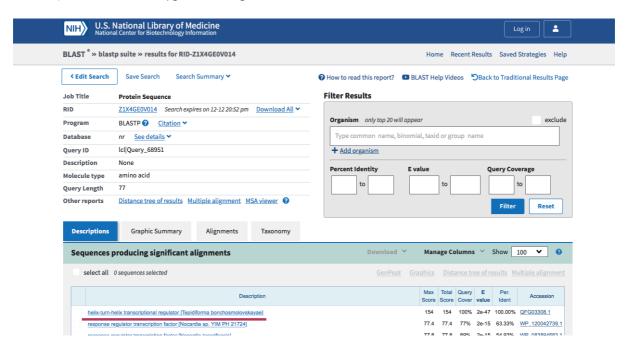


Figure 2 – Results of BLAST for hypothetical protein #2

The result is with good identity (100%), E-value (2e-47), query cover (100%) and it can be suggested that our hypothetical protein has the same functions as <u>helix-turn-helix</u> <u>transcriptional regulator [Tepidiforma bonchosmolovskayae].</u>

3) For hypothetical protein #3 there are no any relevant results with BLAST (only hypothetical proteins were found) and Pfam (no any Pfam-A matches), but with TMHMM the result is

TMHMM result

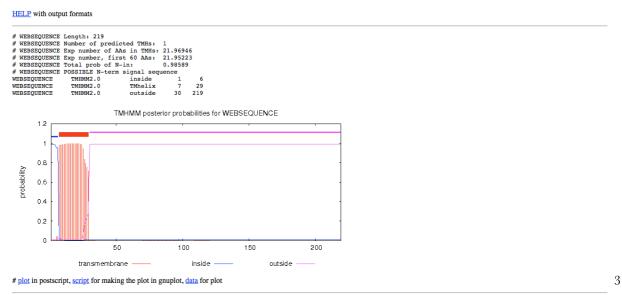


Figure 3 – Results of TMHMM for hypothetical protein #3

With TMHMM a transmembrane segment was found, it means that hypothetical protein #3 has the functions of transmembrane proteins.

4) For hypothetical protein #4 the relevant result was found by TMHMM



Figure 4 – Results of TMHMM for hypothetical protein #4

With TMHMM a transmembrane segment was found, it means that hypothetical protein #4 has the functions of transmembrane proteins.

5) For hypothetical protein #5 the relevant result was found by Pfam

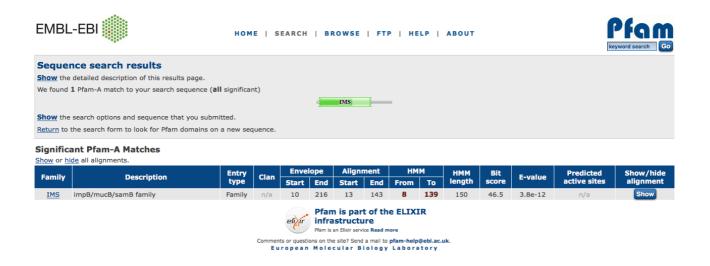


Figure 5 – Results of Pfam for hypothetical protein #5

The protein belongs to impB/mucB/samB family (the proteins of it are involved in UV protection).

- 6) For hypothetical protein #6 no any appropriate functions with all of the described tools were found.
 - 7) For hypothetical protein #7 the relevant result was found by Pfam



Figure 6 – Results of Pfam for hypothetical protein #7

The protein belongs to RNase_H superfamily and has the functions like the other proteins of this family – numerous enzymes which are involved in nucleic acid metabolism and implicated in many biological processes, including replication, homologous recombination, DNA repair, transposition and RNA interference.

8) For hypothetical protein #8 the result is

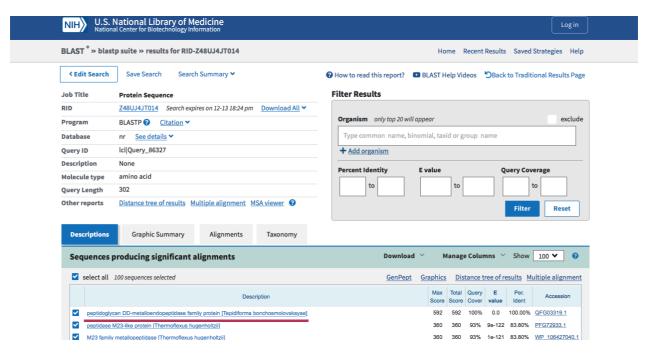


Figure 7 – Results of BLAST for hypothetical protein #8

The result is with good identity (100%), E-value (0.0), query cover (100%) and it can be suggested that our hypothetical protein has the same functions as <u>peptidoglycan DD-metalloendopeptidase family protein [Tepidiforma bonchosmolovskayae].</u>

- 9) For hypothetical protein #9 no any appropriate functions with all of the described tools were found.
- 10) For hypothetical protein #10 no any appropriate functions with all of the described tools were found.
 - 11) For hypothetical protein #11 the result is

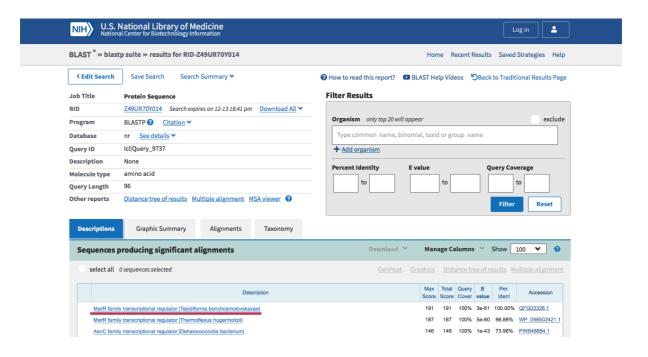


Figure 8 – Results of BLAST for hypothetical protein #11

The result is with good identity (100%), E-value (3e-61), query cover (100%) and it can be suggested that our hypothetical protein has the same functions as <u>MarR family transcriptional regulator [Tepidiforma bonchosmolovskayae].</u>

12) For hypothetical protein #12 the result is

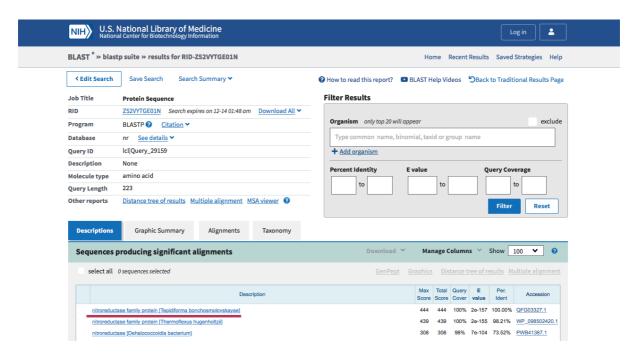


Figure 9 – Results of BLAST for hypothetical protein #12

The result is with good identity (100%), E-value (2e-157), query cover (100%) and it can be suggested that our hypothetical protein has the same functions as <u>nitroreductase family protein [Tepidiforma bonchosmolovskayae].</u>

13) For hypothetical protein #13 the result is

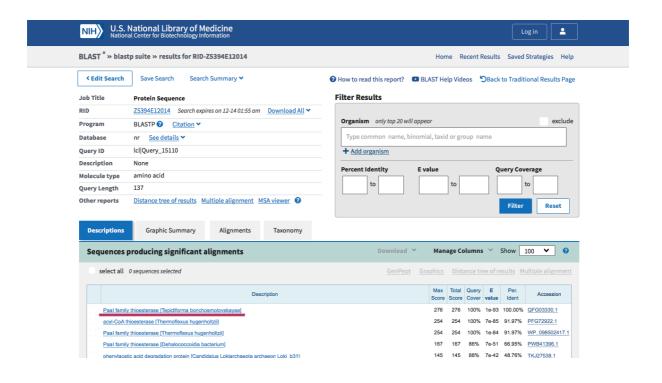


Figure 10 – Results of BLAST for hypothetical protein #13

The result is with good identity (100%), E-value (1e-93), query cover (100%) and it can be suggested that our hypothetical protein has the same functions as <u>PaaI family thioesterase</u> [Tepidiforma bonchosmolovskayae].

- 14) For hypothetical protein #14 no any appropriate functions with all of the described tools were found.
 - 15) For hypothetical protein #15 the relevant result was found by Pfam



Figure 11 – Results of Pfam for hypothetical protein #15

The protein belongs to DinB superfamily and has the functions like the other proteins of this family – metalloenzymes to perform functions such as redox reactions.

- 16) For hypothetical protein #16 no any appropriate functions with all of the described tools were found.
 - 17) For hypothetical protein #17 the result is

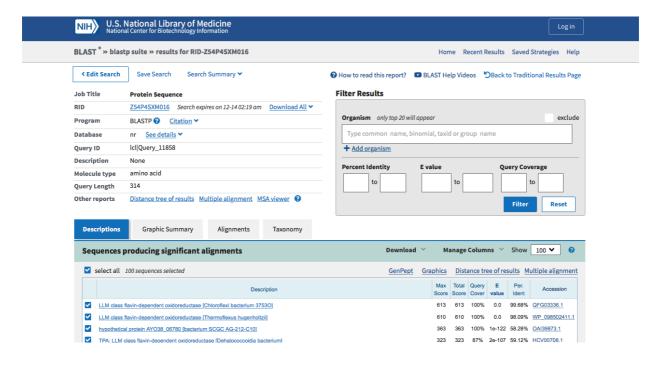


Figure 12 – Results of BLAST for hypothetical protein #17

The result is with good identity (99.68%), E-value (0.0), query cover (100%) and it can be suggested that our hypothetical protein has the same functions as <u>LLM class flavin-dependent oxidoreductase [Chloroflexi bacterium 3753O].</u>

18) For hypothetical protein #18 no any appropriate functions with all of the described tools were found.

3. Description of the operon structure

From Prokka results it is shown that the such operons exist

1-7 (long operon), 8, 9, 10, 11, 12-14, 15-16, 17, 18-23 (long operon), 24-25, 26, 27-29, 30-31, 32.

The distance between the genes in the same operon <150bp.

GNU nano 2.	2.6		File: P	ROKKA_12:	112019	.gff		
#gff-version	3							
#sequence-region Tepidiforma 1 30000								
repidiforma	Prodigal:2.6	CDS	423	1892		_	0	ID=result_non-coding_00001;eC_num
repidiforma Fepidiforma	Prodigal:2.6	CDS	1971	2849		_	0	ID=result_non-coding_00002;infere
repidiforma	Prodigal:2.6	CDS	2898	3131		_	0	ID=result_non-coding_00003;infere
repidiforma	Prodigal:2.6	CDS	3219	3878		_	ø	ID=result_non-coding_00004;infere
repidiforma	Prodigal:2.6	CDS	3940	4329		_	0	ID=result_non-coding_00005;infere
repidiforma	Prodigal:2.6	CDS	4442	7150		_	ø	ID=result_non-coding_00006;eC_num
repidiforma	Prodigal:2.6	CDS	7249	8142		_	ø	ID=result_non-coding_00007;infere
repidiforma	Prodigal:2.6	CDS	8262	10760		+	ø	ID=result non-coding 00008;eC num
[epidiforma	Prodigal:2.6	CDS	10777	10989			0	ID=result_non-coding_00009;infere
[epidiforma	Prodigal:2.6	CDS	11049	12125		_	0	ID=result non-coding 00010;eC num
[epidiforma	Prodigal:2.6	CDS	12839	13699		+	0	ID=result_non-coding_00011;eC_num
[epidiforma	Prodigal:2.6	CDS	13868	15103		_	0	ID=result_non-coding_00012;infere
[epidiforma	Prodigal:2.6	CDS	15141	16484		_	0	ID=result non-coding 00013;eC num
Tepidiforma	Prodigal:2.6	CDS	16588	17496		_	0	ID=result_non-coding_00014;infere
[epidiforma	Prodigal:2.6	CDS	17602	17814		+	0	ID=result_non-coding_00015;infere
[epidiforma	Prodigal:2.6	CDS	17804	18304		+	0	ID=result_non-coding_00016;eC_num
[epidiforma	Prodigal:2.6	CDS	18286	18900		-	0	ID=result_non-coding_00017;eC_num
[epidiforma	Prodigal:2.6	CDS	19007	20614		+	0	ID=result_non-coding_00018;Name=g
[epidiforma	Prodigal:2.6	CDS	20717	21157		+	0	ID=result_non-coding_00019;infere
[epidiforma	Prodigal:2.6	CDS	21177	21479		+	0	ID=result_non-coding_00020;eC_num
[epidiforma	Prodigal:2.6	CDS	21568	21858		+	0	ID=result_non-coding_00021;infere
[epidiforma	Prodigal:2.6	CDS	21903	22574		+	0	ID=result_non-coding_00022;infere
[epidiforma	Prodigal:2.6	CDS	22682	23476		+	0	ID=result_non-coding_00023;eC_num
[epidiforma	Prodigal:2.6	CDS	23473	23955		-	0	ID=result_non-coding_00024;Name=y
[epidiforma	Prodigal:2.6	CDS	23945	24358		-	0	ID=result_non-coding_00025;infere
[epidiforma	Prodigal:2.6	CDS	24418	24756		+	0	ID=result_non-coding_00026;infere
[epidiforma	Prodigal:2.6	CDS	24753	25244		-	0	ID=result_non-coding_00027;infere
[epidiforma	Prodigal:2.6	CDS	25292	25480		-	0	ID=result_non-coding_00028;infere
[epidiforma	Prodigal:2.6	CDS	25487	26923		-	0	ID=result_non-coding_00029;eC_num
[epidiforma	Prodigal:2.6	CDS	27101	28414		+	0	ID=result_non-coding_00030;Name=t
[epidiforma	Prodigal:2.6	CDS	28491	29435		+	0	ID=result_non-coding_00031;db_xre
[epidiforma	Prodigal:2.6	CDS	29461	29703		-	0	ID=result_non-coding_00032;infere
##FASTA								

Figure 13 – Results of Prokka for operon structure task

For long operons (1-7 and 18-23) the regulatory regions should be identified if possible. With software Softberry it can be done (http://www.softberry.com).

For 18-23 operon (+)

Figure 14 – Results of Softberry for operon 18-23

If we look at Figure 13 and coordinates of operons we see that right promoter has

pos: 18975

LDF - 1.43

However, no TF were found for this promoter

```
Oligonucleotides from known TF binding sites:
For promoter at 17602:
     rpoH3: CTCCCCCT at position 17574 Score -
    rpoS17: CCCCCTCC at position 17576 Score -
                                                   17
For promoter at 21533:
                                   21510 Score -
     metR: TTTTTCA at position
     poD15: TTTTCACG at position ompR: TCATATTT at position
    rpoD15:
                                   21512 Score -
                                    21520 Score -
     argR2: CATATTTT at position
                                    21521 Score -
For promoter at 26953:
       ihf: ATCATACA at position 26953 Score -
                                                   13
No such sites for promoter at
                              8261
No such sites for promoter at 18975
For promoter at 23658:
      cysB: TCTTGCAT at position
                                    23622 Score -
    rpoD16: TTCAATCT at position
                                    23650 Score -
No such sites for promoter at 12838
For promoter at 29474:
    rpoD16: CCTACAAT at position
                                    29460 Score -
                              4444
No such sites for promoter at
For promoter at
                   220:
    rpoS17: CCCCCTCC at position
                                      195 Score - 17
No such sites for promoter at 27604
No such sites for promoter at 15513
For promoter at
                  3905:
            TTCCTCCT at position
    rpoD17:
                                     3883 Score -
    rpoD19:
            TCCTGCTA at position
                                     3887 Score -
No such sites for promoter at 22932
No such sites for promoter at
                               7019
No such sites for promoter at 26402
```

Figure 15 – Results of Softberry for operon 18-23

For 1-7 operon we should reverse our sequence (-) and then use Softberry

```
> test sequence
                          30000
Length of sequence-
 Threshold for promoters - 0.20
Number of predicted promoters -
                                         11
Promoter Pos: 21832 LDF- 2.90
-10 box at pos. 21817 ttctattct Score -35 box at pos. 21797 ttgcag Score
                                                 57
Promoter Pos: 6566 LDF- 2.37
-10 box at pos. 6551 gggtaaact Score
-35 box at pos. 6526 tagccg Score
                                                 74
-35 box at pos. 6526 tagccg
Promoter Pos: 3055 LDF- 2.03
-10 box at pos. 3040 gtgtatgat Score
-35 box at pos. 3023 ctcacg Score
                                                 68
Promoter Pos: 17225 LDF- 1.89
-10 box at pos. 17210 tgctacact Score
 -35 box at pos. 17192 atgtcg
Promoter Pos: 9471 LDF- 1.02
-10 box at pos. 9456 gaggaccat Score
-35 box at pos. 9436 gtgacg
Promoter Pos: 16315 LDF- 0.89
-10 box at pos. 16300 ggttattcg Score
                                                28
 -35 box at pos. 16282 ttgccg
Promoter Pos: 23476 LDF- 0.71
                                                 35
-10 box at pos. 23461 agctgtact Score
-35 box at pos. 23441 ttacct
                                                33
Promoter Pos: 8484 LDF- 0.66
 -10 box at pos. 8469 gtgaaaaat Score
                                                41
 -35 box at pos.
                    8451 gtgtca
Promoter Pos: 16890 LDF- 0.58
-10 box at pos. 16875 ggtgaccat Score
-35 box at pos. 16855 ttcagt Score
                                                 20
Promoter Pos: 13503 LDF- 0.37
-10 box at pos. 13488 cggcagaat Score
-35 box at pos. 13468 cggccg Score
                                                 47
                                                -17
Promoter Pos: 10287 LDF- 0.37
                                                 14
 -10 box at pos. 10272 ctggaggat Score
 -35 box at pos. 10252 atgacg
                                                 30
Oligonucleotides from known TF binding sites:
```

Figure 16 – Results of Softberry for operon 1-7

It is shown that no promoters for operon was found.

4. Finding of genes obtained by the bacteria through horizontal gene transfer (HGT)

All the found proteins with high identity are from very close related species. I suppose horizontal gene transfer (HGT) in my fragment sequence has not been concerned.

5. Finding of genes of the bacteria associated with secondary metabolites

KEGG tool was used for finding of metabolic pathways https://www.kegg.jp.

- 1. Trehalose-6-phosphate synthase is in metabolic pathway trehalose synthesis
- 2. Valine - tRNA ligase synthesis of tRNA
- 3. Prodigiosin synthesizing transferase PigC biosynthesis of the red antibiotic prodigiosin

. . .

Visualisation part

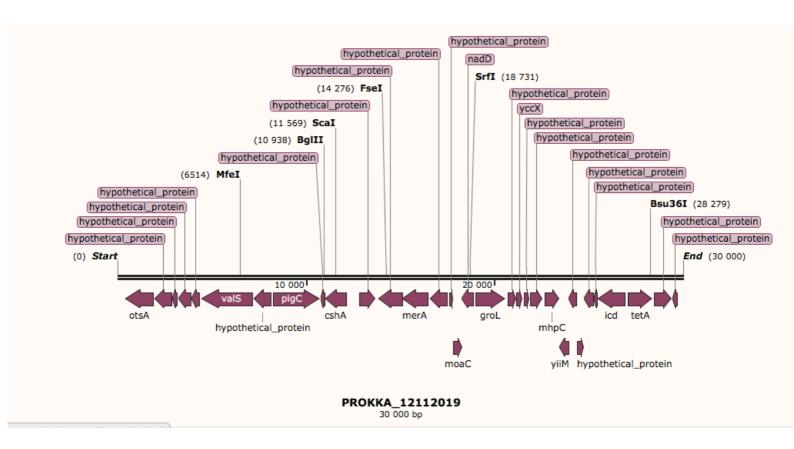


Figure 17 – SnapGene visualisation