

# Class 10: Structural Bioinformatics 1

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## PDB Statistics

The Protein Data Bank (PDB) is the main repository of bio-molecular structures. Let's see what it contains:

```
stats <- read.csv("Data Export Summary.csv")
stats
```

	Molecular.Type	X.ray	EM	NMR	Integrative	Multiple.methods
1	Protein (only)	178,795	21,825	12,773	343	226
2	Protein/Oligosaccharide	10,363	3,564	34	8	11
3	Protein/NA	9,106	6,335	287	24	7
4	Nucleic acid (only)	3,132	221	1,566	3	15
5	Other	175	25	33	4	0
6	Oligosaccharide (only)	11	0	6	0	1
	Neutron	Other	Total			
1	84	32	214,078			
2	1	0	13,981			
3	0	0	15,759			
4	3	1	4,941			
5	0	0	237			
6	0	4	22			

```
sum(stats$Neutron)
```

```
[1] 88
```

The comma in these numbers leads to the numbers here being read as characters

```
c(100, "Barry")
```

```
[1] "100"   "Barry"
```

```
library(readr)
stats <- read_csv("Data Export Summary.csv")
```

```
Rows: 6 Columns: 9
-- Column specification -----
Delimiter: ","
chr (1): Molecular Type
dbl (4): Integrative, Multiple methods, Neutron, Other
num (4): X-ray, EM, NMR, Total

i Use `spec()` to retrieve the full column specification for this data.
i Specify the column types or set `show_col_types = FALSE` to quiet this message.
```

```
stats
```

```
# A tibble: 6 x 9
`Molecular Type`    `X-ray`     EM     NMR Integrative `Multiple methods` Neutron
<chr>                <dbl>    <dbl>   <dbl>      <dbl>                <dbl>    <dbl>
1 Protein (only)    178795  21825  12773      343        226     84
2 Protein/Oligosacch~ 10363   3564    34         8        11      1
3 Protein/NA          9106   6335   287       24         7      0
4 Nucleic acid (only) 3132    221   1566       3        15      3
5 Other                 175    25    33         4         0      0
6 Oligosaccharide (o~   11     0     6          0         1      0
# i 2 more variables: Other <dbl>, Total <dbl>
```

```
n.xray <- sum(stats$`X-ray`)
#n.em <-
n.total <- sum(stats$Total)

n.xray/n.total
```

```
[1] 0.8095077
```

Q1. What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
n.xray <- sum(stats$`X-ray`)
n.em <- sum(stats$EM)
n.total <- sum(stats$Total)

(n.xray/n.total) * 100
```

```
[1] 80.95077
```

```
(n.em/n.total) * 100
```

```
[1] 12.83843
```

Q2. What proportion of structures in the PDB are protein?

```
n.protein <- sum(stats$Total[grep("Protein", stats$`Molecular Type`)])
(n.protein / n.total) * 100
```

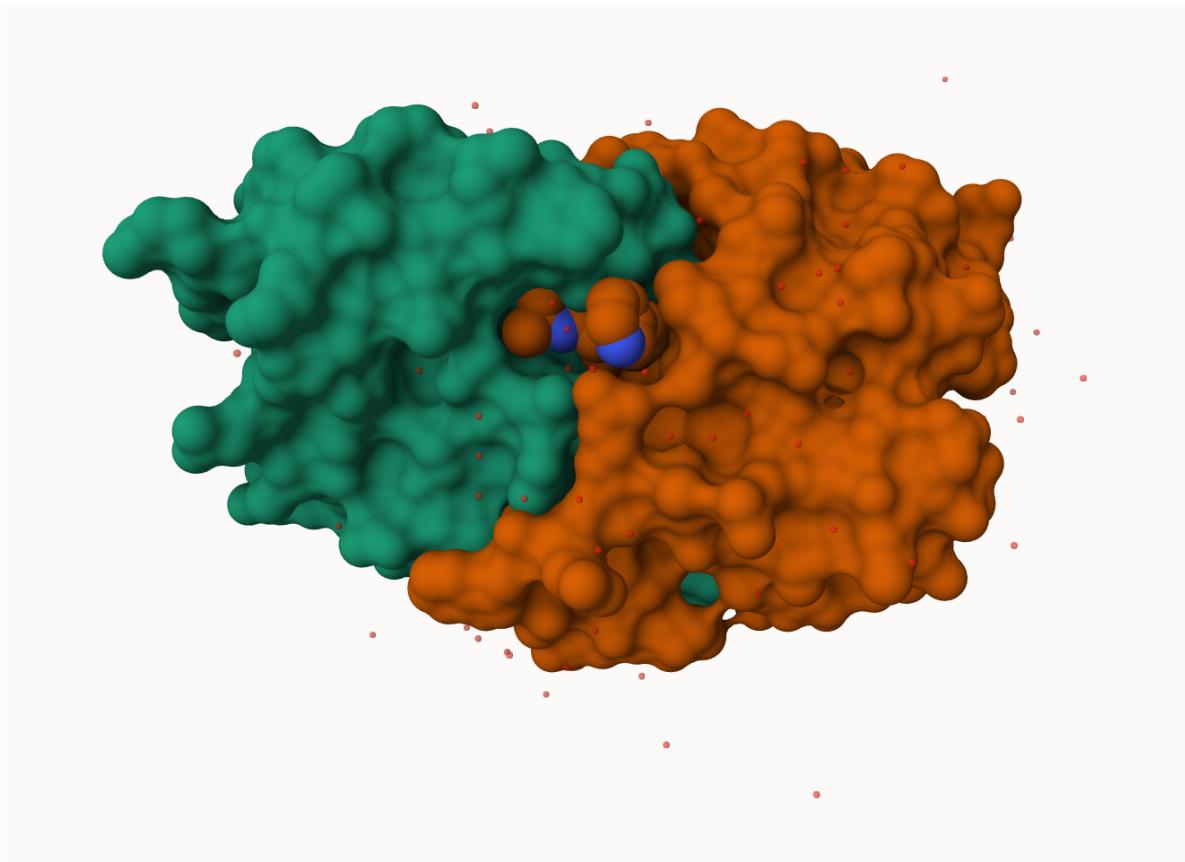
```
[1] 97.9118
```

Q3. Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

Skip...

### **Visualizing the HIV-1 protease structure**

We can use Molstar viewer online: <https://molstar.org/viewer/>



A clean image showing the catalytic ASP25 amino acids in both chains of the HIV-PR dimer, along with the inhibitor and all important active site water.



## Bio3D package for structural bioinformatics

```
library(bio3d)  
  
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
 pdb
```

```
Call: read.pdb(file = "1hsg")  
  
Total Models#: 1  
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)  
  
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)  
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

```
Protein sequence:
```

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

```
attributes(pdb)
```

```
$names
```

```
[1] "atom"    "xyz"     "seqres"  "helix"   "sheet"   "calpha"  "remark" "call"
```

```
$class
```

```
[1] "pdb"    "sse"
```

```
head(pdb$atom)
```

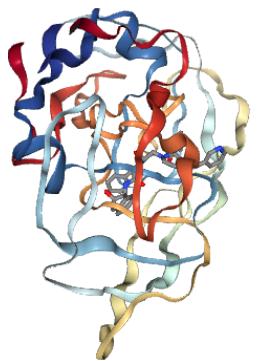
	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

```
library(bio3dview)
library(knitr)
library(webshot2)
```

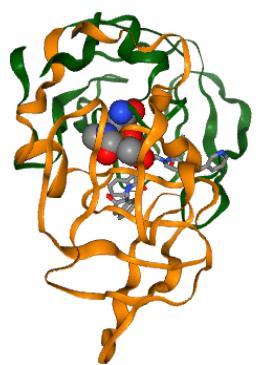
```
view.pdb(pdb)
```



```
library(knitr)
library(webshot2)

# Select the important ASP 25 residue
sele <- atom.select(pdb, resno=25)

# Highlight them in spacefill representation
view.pdb(pdb, cols=c("darkgreen","darkorange"),
          highlight = sele,
          highlight.style = "spacefill")
```



## Predicting functional motions of a single structure

Read an ADK structure from the PDB database:

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file  
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

```
Call: read.pdb(file = "6s36")

Total Models#: 1
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]

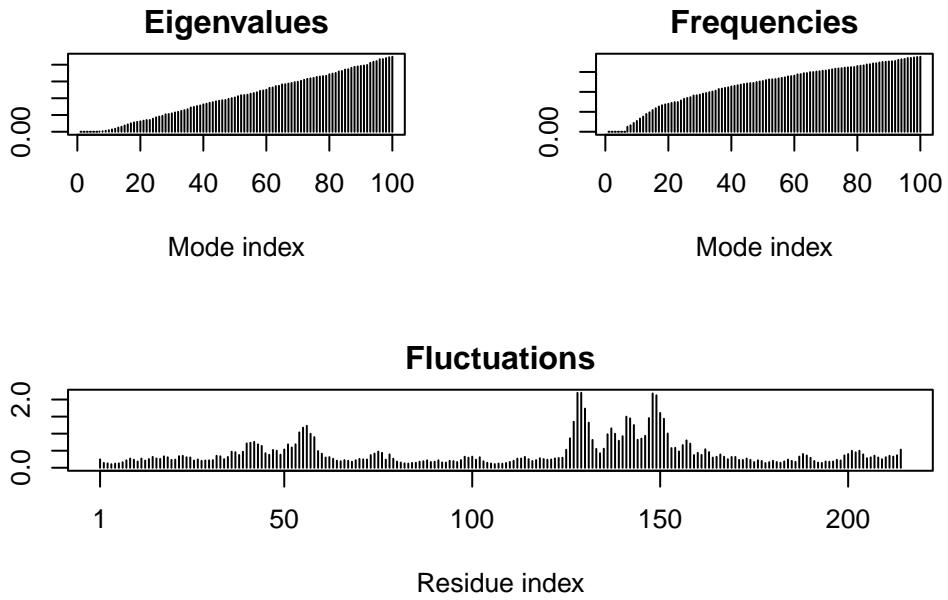
Protein sequence:
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIDMAGKLVT
DELVIALVKERIAQEDCRNGFLLDGFPRTRIPQADAMKEAGINVDTVLEFDVPDELVDKI
VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTRKDDQEETVRKRLVEYHQMTAPLIG
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG

+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call
```

```
m <- nma(adk)
```

```
Building Hessian...      Done in 0.045 seconds.
Diagonalizing Hessian... Done in 0.672 seconds.
```

```
plot(m)
```



Write out our results as a trajectory/movie of predicted motions:

```
mktrj(m, file="adk_m7.pdb")
```