

Combining Rapid Antigen Testing and Syndromic Data Improves Sensitivity and Specificity in Real-World COVID-19 Detection

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1. Abstract

Background

The majority of the world's population live in low- and middle-income countries (LMICs) where access to gold-standard diagnostics like RT-PCR is often limited. Rapid antigen testing (RAT) and syndromic diagnosis are two alternative, inexpensive and easy-to-deploy surveillance methods but there are concerns that they lack the sensitivity and specificity to effectively guide practice.

Methods

Bangladesh's Institute of Epidemiology Disease Control And Research (IEDCR) identified potential COVID-19 patients in Dhaka using syndromic surveillance.

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A sample ($n = 1172$) of these patients was tested using RAT and syndromic data were collected. Models were fit to predict RT-PCR status using the RAT data, the syndromic data, and the two combined. Model performance was measured using predictive power and classification performance under three epidemiological scenarios: “Agnostic,” “Rising Cases” and “Low-Level Cases.”

Findings

Combined data models yielded equal or improved performance over syndromic- and RAT-only models across all three epidemiological scenarios and when compared as more generic prediction and classification engines. In the “Rising Cases” scenario, which most closely represents the current situation in many LMICs, the combined data model false negative rate is 26 percentage points lower than that of the RAT only model. Although the syndromic only model matches the combined models false negative rate, its false positive rate is 31 percentage points higher.

Interpretation

A few accurate tests may be less useful at the population level than many more imperfect ones. Small, scalable improvements in the accuracy of mass-deployed but imperfect tests can then make a very big difference for pandemic control.

We demonstrate that such improvements can be achieved by statistically utilising complementary strengths and weaknesses across two imperfect diagnostics, we can greatly improve the detection of COVID-19.

Funding

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2. Introduction

Identification and isolation of COVID-19 cases remains key to the pandemic response across the globe. The faster and more accurately we can identify cases, the more effectively we can provide clinical care, reduce transmission of infection and develop population-level interventions. RT-PCR testing has rapidly become the default, gold-standard test for COVID-19 in applied settings due to its high sensitivity and specificity for COVID-19 [2]. Most of the world’s population, however, live in low- and middle-income countries (LMICs) where the laboratory facilities needed to carry out RT-PCR tests are often scarce and hard to reach [4]. COVID-19 diagnosis worldwide, therefore, must be made accessible using inexpensive methods that can be carried out locally [6].

An increasingly popular alternative to RT-PCR is rapid antigen testing (RAT) [7]. Like RT-PCR, these tests have high specificity for COVID-19 while being less expensive, easier to implement, and faster to produce results [8]. RATs also require less commitment and discomfort for patients. For RT-PCR testing, patients must travel to a designated site (such as a hospital or testing booth) or have highly visible PPE-clad officials visit their home. Then, invasive nasopharyngeal swabs must be taken and there is a delay in receiving the result (between one day and a week in Bangladesh). In contrast, RAT can be conducted on nasal or saliva samples, completed in the home with minimal PPE and results

71 are available in 30 minutes. RATs can be taken by persons with limited training,
 72 thus decreasing the time and expense associated with identifying cases. Together,
 73 these traits make RATs an appealing alternative to RT-PCR. However, several
 74 concerns have been raised about the sensitivity of RAT [9] leading to more false
 75 negative diagnoses.

76 Another alternative to RT-PCR, one that has been used since the start of
 77 the pandemic, is identifying cases through symptom-thresholding [10]. In this
 78 approach, a patient presenting with a fever and one or more viral pneumonia
 79 symptoms is treated as a COVID-19 positive patient. The main advantage
 80 of this approach is the ease of implementation. As with RAT the process is
 81 faster, cheaper and less invasive than RT-PCR, but unlike RAT the process
 82 relies on minimal equipment and thus can be scaled quickly and easily. For
 83 example, in Bangladesh, an LMIC, much of the initial support and reporting of
 84 infections locally is provided by community support teams (CSTs) composed of
 85 local volunteers with basic training. The CSTs can easily collect symptomatic
 86 data in the community and provide care where the thresholds are met. However,
 87 these thresholds were developed early in the outbreak, and thus were necessarily
 88 drawn from clinical intuition, rather than data, and for different variants and
 89 populations than they are now applied to. Consequently, the relationship between
 90 the thresholds and the true COVID-19 status is often weak, with low specificity
 91 leading to a very large number of false positive diagnoses.

92 A natural extension to these symptom-threshold approaches is syndromic
 93 modelling. Here, a patient presenting with a fever and one or more viral
 94 pneumonia symptoms is treated as a potential COVID-19 patient. However,
 95 rather than using a set of pre-determined criteria, a range of symptomatic and
 96 risk factor data are collected and then a sub-sample of patients is tested using
 97 RT-PCR for COVID-19 [11]. These data are used to fit a model that allows
 98 more accurate prediction of how likely a patient is to have COVID-19 through
 99 the identification of COVID-19 syndromes [13]. It is worth highlighting at this
 100 point that in resource-limited settings, there is very limited provision for testing
 101 of asymptomatic cases, despite their important role in disease transmission [14].
 102 Even while focusing solely on symptomatic patients, syndromic modelling is a
 103 complex and nuanced task. The strength of relationships between symptoms
 104 and diseases is not stable through time or across sampling strategies since the
 105 relative importance of each symptom for disease diagnosis, in part, depends on
 106 the prevalence of other diseases causing similar symptoms in the community [15].
 107 For example, if another disease for which loss of smell is a symptom becomes
 108 common, that symptom becomes a worse predictor for COVID-19. Similarly,
 109 if everyone who presents has a cough and thus is included in the sample, then
 110 coughing will likely have a very low correlation with COVID-19 (even if the
 111 two are strongly related in the general population). Symptoms are also inter-
 112 related, meaning that they cannot be interpreted independently. The majority of
 113 methods used currently do not account for these changes through time, symptom-
 114 to-symptom correlations or the relationship between the population sampled
 115 and the target population. Even then, the many types of common respiratory
 116 disease generally means that even then these models tend to have relatively high

JMC: Not
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JMC: Need
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117 false positive rates (low specificity) for COVID-19 [15], although much lower
118 than the symptom-threshold approach.

119 Poor sensitivity and specificity are problematic in diagnostics but higher
120 error rates than gold-standard methods may be tolerable depending on their
121 scale and impact given the local situation. Low specificity means a large number
122 of false positive classifications, where the patient is told they have COVID-19
123 but they actually do not. This might lead to patients unnecessarily self-isolating
124 and receiving support which can be expensive to the individuals and local public
125 health bodies, as well as reducing available resources for those who need them
126 [16]. Similarly, low sensitivity means more false negative classifications, where
127 the patient is told they do not have COVID-19 but they actually do, which can
128 lead to a health-risk for the individual and to the disease spreading further [17].
129 Although the default approach is generally to minimise both misclassification
130 rates (our “Agnostic” scenario below), the true costs of these misclassifications
131 will depend on local context. When the prevalence of the disease is low, false
132 positives may create local scepticism about the value of testing, or when there
133 are strong population-level mitigations already in place (such as a nationwide
134 lockdown), then false positives might be more costly than false negatives [16],
135 corresponding to our “Low-Level Cases” scenario. If the disease is abundant
136 or increasing rapidly then false negatives are likely to be more costly, as in our
137 “Rising Cases” scenario. Often the situation will be even more nuanced and a
138 different balance will need to be struck [4].

139 The “best” diagnostic, therefore, is not a single universal test. The two
140 dominant testing methods available in LMICs when not optimised for the
141 local situation are highly flawed. Relying solely on symptomatic diagnosis
142 will likely overestimate the number of individuals with COVID-19 due to its
143 lack of specificity. Conversely, RATs will give a false impression of control
144 due to the number of positive cases that will be missed. In this paper, we
145 demonstrate that by combining these two testing methods we can utilise their
146 complementary strengths, ameliorate their respective weaknesses, and optimise
147 them for different epidemiological scenarios. We aim to compare the performance
148 of these two testing methods and the combined approach both in terms of general
149 prediction and as diagnostics under three epidemiological scenarios with different
150 misclassification requirements. We show that the optimised combined data
151 models achieve equal-to-much-lower error rates than the next best method in all
152 metrics. We then discuss the role of statistically integrating data from multiple
153 imperfect testing methods in resource limited settings to improve the diagnosis
154 of diseases, particularly COVID-19.

155 3. Methods

156 Participants included in this study were identified for COVID-19 testing by
157 community support teams (CSTs). Recruitment took place across Dhaka (the
158 capital city of Bangladesh) between 19th May 2021 and 11th July 2021.

159 Patients were selected for further testing conditional on the presence of a
160 fever ($>38^{\circ}\text{C}$) at the point of testing and one or more of 14 additional symptoms

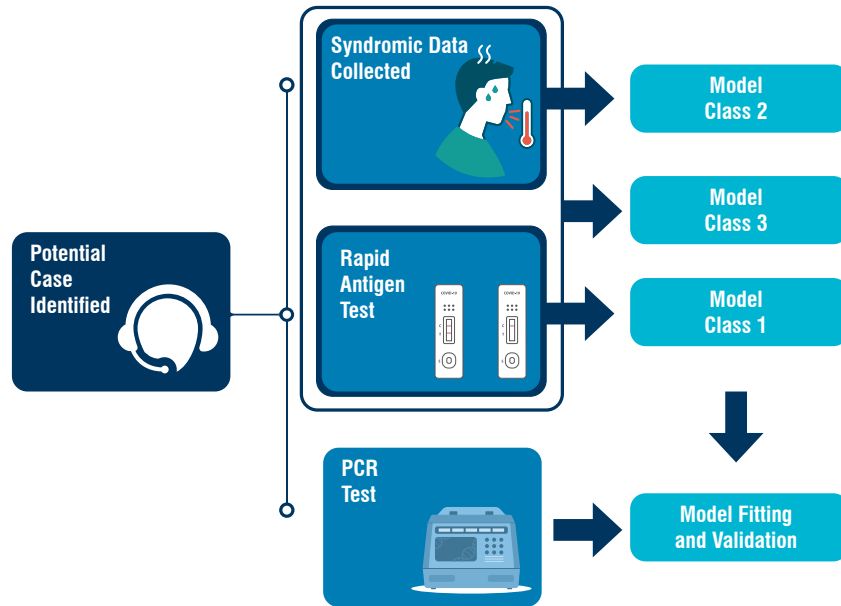


Figure 1: Schematic description of identification of likely COVID-19 patients by community support teams (CSTs), swab collection and model definitions. The teams collected syndromic data (age, gender and presence/absence of 14 predetermined symptoms), and two sets of naso-pharyngeal swabs (one each for Rapid Antigen Testing and RT-PCR). We then used rapid antigen testing (RAT) and syndromic data, two imperfect but inexpensive diagnostics, to generate three model classes: RAT result only in Model Class 1, syndromic data only in Model Class 2, and both RAT result and syndromic data in Model Class 3. The RT-PCR test result is used to train and test each model using temporal cross-validation.

161 associated with COVID-19 (breathing problems, coughing, diarrhoea, fever
 162 (ongoing), a headache, loss of taste, loss of smell, muscle pain, red eyes, a runny
 163 nose, a sore throat, tiredness, vomiting or a wet cough). The patient's age
 164 and gender were also recorded, but these data were not included in the patient
 165 selection criteria.

166 Nasal swabs and syndromic data were collected from the patient by medical
 167 technologists. One swab each was used for rapid antigen testing (RAT) and
 168 RT-PCR (gold-standard for COVID-19 status). The full questionnaire and
 169 testing protocols are provided in Appendix XX. Participants provided written
 170 informed consent to sample collection and for their test results to be analyzed in
 171 the study.

172 We examined the ability of the two imperfect identification methods, the
 173 syndromic profile and RAT result, to predict the patient's COVID-19 status
 174 when used separately and together. The different data combinations define three
 175 model classes (Figure 1).

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176 Model Class 1 uses only the RAT result and is the simplest of the three.
 177 It equates a positive RAT result with the patient being PCR positive, and a
 178 negative RAT result with PCR negativity. Model Class 2 uses only the syndromic
 179 data and Model Class 3 combines the RAT result with the syndromic data.

180 For Model Class 2, we used a Bayesian multivariate probit model [18]. The
 181 multivariate probit structure allows the model to account for the correlations
 182 between, and binary nature of, the symptoms (e.g. loss of taste is often correlated
 183 with loss of smell). By using a Bayesian formulation, we are able to quantify
 184 and propagate the uncertainty in the parameter estimates. Structurally, the
 185 multivariate probit model allows the symptoms and COVID-19 status to be
 186 treated as correlated binary outcomes with an intrinsic rate (the intercept for each
 187 variable) and the patient’s age and gender, while propagating and quantifying
 188 uncertainty.

189 In Model Class 3, we utilise the specificity of RAT by treating RAT positive
 190 patients as PCR positive patients. The RAT negative patients are then modelled
 191 using the sensitive syndromic approach using Model Class 2 to capture additional
 192 PCR-positive patients that are missed by the RAT. This approach leverages
 193 the fact that RAT-negative-PCR-positive patients may have different syndromic
 194 profiles than RAT-positive-PCR-positive patients and allows the model to adapt
 195 more specifically to that group. The models were fitted to the data using Bayesian
 196 inference techniques based on Hamiltonian Monte Carlo in the Stan programming
 197 language [19].

198 We conducted backwards model selection (starting with the most complex
 199 model feasible, with all 14 symptoms and both covariates) to identify a subset of
 200 models with the highest predictive power under temporal cross-validation (Figure
 201 2). Reducing the number of possible models to a small number of the most
 202 predictive models was necessary to reduce computational demand and reduce the
 203 risk of overfitting models to the test scenarios. The large number of symptoms
 204 means that there is a high number of potential model configurations ($>131\,000$
 205 for 14 symptoms and two covariates) which might, by chance, perform well on
 206 the test sets (even under the challenging conditions of temporal cross-validation)
 207 but lack transferability. By first using general predictive power to narrow down
 208 the number of candidate models and then testing those models under more
 209 specific scenarios, we are more likely to choose models which generalise well
 210 to new data. The number of candidate models used was not pre-determined.
 211 In fitting the models it became clear that there were “jumps” in performance
 212 (as defined below) between models containing five and four symptoms, so the
 213 models with zero to four symptoms were used as the candidate models.

214 We scored the models’ predictive power using cross-entropy. Cross-entropy
 215 measures the accuracy of models that generate probabilities of binary outcomes,
 216 rather than make binary classifications, similar in concept to a mean square error
 217 for normally-distributed data, but adapted for binary data [20]. A cross-entropy
 218 value close to zero corresponds to high levels of accuracy, with larger values
 219 indicating lower accuracy. As the score only uses the predicted probability and
 220 true values, it is possible to directly compare the predictions of any model for
 221 the same test set. More details on the model structure and selection process,

222 including code, are available in Appendix XX.

223 We then compared models as classifiers using their false positive and false
224 negative rates in three epidemiological scenarios. In applied settings, models
225 must often be evaluated on their performance as classifiers rather than just as
226 prediction engines (i.e. their ability to say a patient is COVID-19 positive or
227 negative, not simply the probability the patient might be COVID-19 positive or
228 negative). To generate a classification, a probability threshold must be chosen
229 over which patients are classified as COVID-19 positive.

230 Classifier performance was compared using receiver operating characteristic
231 (ROC) curves and error rates under three epidemiological scenarios. ROC
232 curves show the true and false positive rates that each model can achieve. To
233 extract the error rate under the epidemiological scenarios (described below
234 and in Table 1), we use the ROC calculations, to identify the probability
235 threshold which most closely meets the scenario requirement (see Table 1 for
236 requirements and Appendix XX for calculation details). Comparing specific
237 scenarios allows classifier performance to be demonstrated in relevant scenarios.
238 Whether measuring classifier performance in specific scenarios or more generally,
239 decisions need to be made about the relative cost and acceptable levels of the two
240 types of misclassification (false positives and negatives). We strongly emphasise
241 that local context should be the guide in applying these methods.

242 In Scenario 1, we do not consider epidemiological context but simply costing
243 false negative and false positive rates equally. We do this by maximising the two
244 correct classification rates both individually and in total, as measured by the
245 harmonic mean (as opposed to the arithmetic mean which would only maximise
246 the rates in total). Scenario 2 corresponds to the current situation in Bangladesh
247 at time of writing (July 2021), with COVID-19 cases beginning to rapidly increase
248 again. Under these circumstances, false negatives are extremely costly relative
249 to false positives due to the exponential growth of the disease. In Scenario 3,
250 the pandemic is not declining but maintaining a steady rate of cases. In this
251 situation, policy-makers may be keen to keep false positive diagnoses low to
252 prevent lockdown fatigue and to keep the workforce active.

Add ROC
calculations
to appendix

253 4. Results

254 Of 1241 subjects surveyed, a total of 1172 subjects had complete data available
255 for the current analyses with the remained removed due to missed symptoms or
256 ambiguous coding of dates or symptoms. The mean age of women participants
257 (47% of the sample) was 37 (SD = 14), and for men (53% of the sample) was 36
258 (SD = 14). Participants were identified by the community support teams (CSTs)
259 and drawn from across Dhaka.

260 Model selection for Model Class 2 (syndromic data only) and 3 (syndromic
261 and RAT data), each retained age as an explanatory variable and showed a
262 marked decline in predictive power at more than 4 symptoms. The final four
263 symptoms in order of importance (i.e. the most important symptom was retained
264 in all of the final 4 models, the least important symptom was only retained in
265 the 4 symptom model) were wet cough, runny nose, loss of smell and breathing

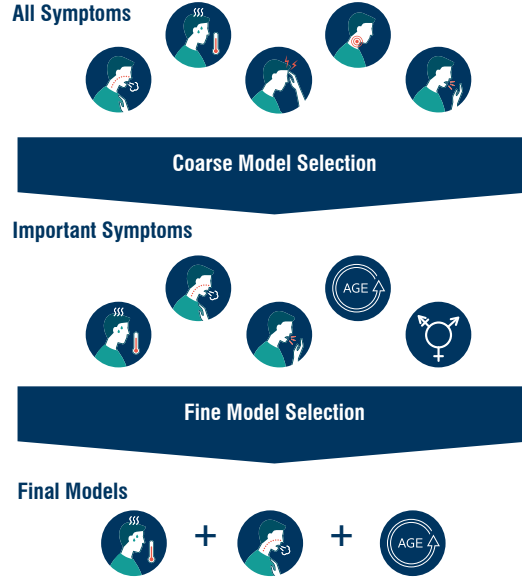


Figure 2: Schematic for rounds of model selection in the multivariate probit component of Model Classes 2 and 3. With 14 symptoms (only 5 shown here for demonstration purposes) and two covariates there are over 131000 possible model combinations. To make exploring these possible models computationally feasible and to reduce the risk of overfitting, we carried out two rounds of model selection. First, the data are divided into temporal cross-validation sets. The multivariate probit connects symptoms to the RT-PCR result through a correlation matrix. In the coarse model selection, the most complex feasible model (all symptoms and covariates) is fit to the training data. The estimated correlations between each symptom and the RT-PCR result are compared for each cross-validation set. The symptoms that have non-zero correlations in a systematic direction (i.e. all positively or all negatively correlated with RT-PCR result) are retained. The process is then repeated on each retained set of symptoms until the four symptoms in each model class with the strongest correlation to RT-PCR result. We then conduct a more exhaustive model selection on all the possible permutations of the four symptoms and two covariates. In this round, each model is fit to training data and used to predict for the test set, and the quality of those predictions is measured using cross-entropy scoring. The cross-entropy score is then used to select the best predictive model for each level of model complexity. Only these final models are then used for classification. This reduces the set of models tested as classifiers from $>131\,000$ to just four per model class.

Table 1: For each epidemiological scenario there is a requirement and a performance criterion. The requirement refers to a base level of performance the model must achieve; in general this will be a maximum acceptable error rate of some kind. These requirements were determined in discussion with members of the Institute of Epidemiology, Disease Control and Research, Ministry of Health, Bangladesh (IEDCR). The requirement determines a probability threshold for each model which most closely meets that requirement. The performance criterion is then used to determine which model performs the 'best' given that the requirement has been met.

Scenario Name	Requirement	Performance Criterion (Error)
1 Agnostic	Maximise correct classification rates	Sum of error rates
2 Rising Cases	Max. 20% false negative rate	False negative rate
3 Low-Level Cases	Max. 20% false positive rate	False positive rate

problems for Model Class 2, and fever, wet cough, tiredness and diarrhoea for Model Class 3. For both Model Class 2 and Model Class 3 model selection retained age but not gender as a covariate.

In the comparison of model predictive performance, Model Class 1 (RAT only) performed worst with an out-of-sample cross-entropy of 3.24 (cross-entropy values further from zero correspond to worse predictive performance). The median cross-entropy values were between 2.53 and 2.59 for models in Class 2 (syndromic data only). Models in Class 3 (combined data model) performed best with cross-entropy values between 1.44 and 1.47 (see Figure 3).

General model classification performance is shown by the full ROC curves for each model (Figure 4).

Scenario specific classification performance is shown in Figure 5. Across all three scenarios (defined in Table 1), the best models in Class 3 performed equally well or better than the other two model classes. In Scenario 1 ("Agnostic"), models in Classes 1 and 3 performed equally well and distinctly better than models in Class 2. In Scenario 2 ("Rising Cases"), Model Class 1 failed to meet the requirement and so was excluded (effectively infinite error), and Model Class 3 once again outperformed Class 2. In Scenario 3 ("Low-Level Cases"), all three model classes met the requirement. Once again, Model Class 2 performed worst and Model Class 3 achieved the lowest error, with Model Class 1 falling in between the two (closer to Class 3 than 2). Across all the scenarios and both Classes 2 and 3, the number of symptoms made relatively little difference within the final four candidate models in terms of median performance, although the more complex models have higher precision. It should be noted that the candidate models are chosen as a result of a selection process and performed much better than more complex models (i.e. those with 5 or more symptoms) or simpler models (with no symptoms but an intercept and covariates) in terms of cross-entropy and ROC, indicating they would likely also perform worse in these scenarios. These models were not tested on the scenarios to minimise the

DHC: out-of sample vs test set cross-entropy

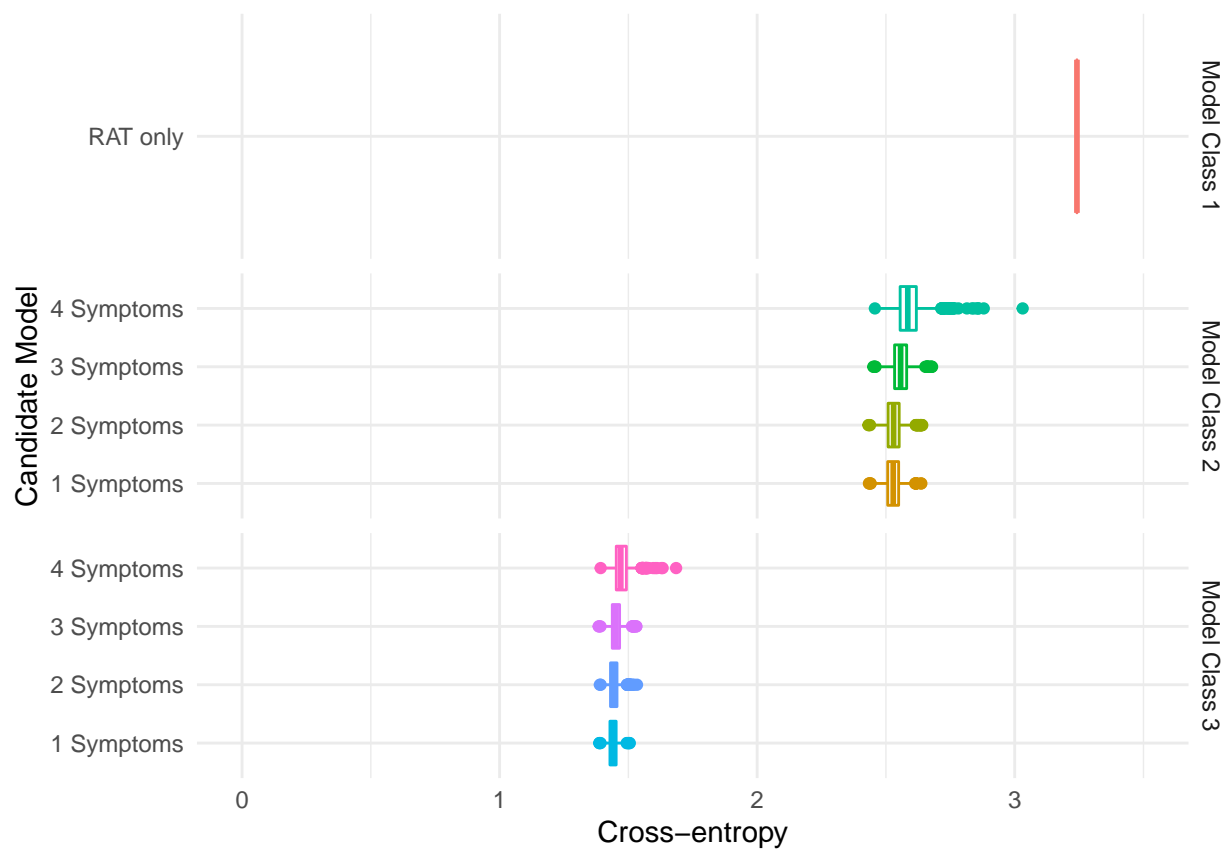


Figure 3: Predictive performance of candidate models. Interquartile ranges for the posterior cross-entropy of the best candidate models at each level of model complexity tested under temporal cross-validation. cross-entropy is a measure of distance from the truth, so values closer to zero indicate better models. The intermediate complexity models perform best at prediction, although performance is similar across all the models within each model class (1: rapid antigen testing (RAT) only; 2: syndromic data only; and 3: combined RAT and syndromic data).

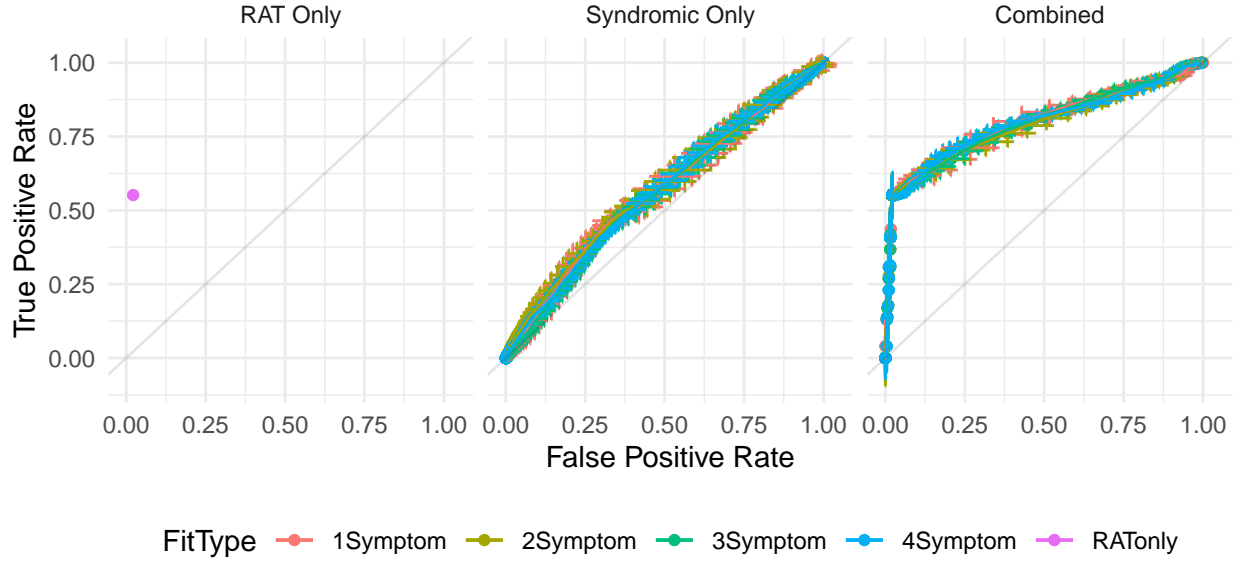


Figure 4: Receiver operating characteristics for rapid antigen testing (RAT) only approach (Model Class 1) and posterior mean (\pm posterior standard deviation) receiver operating characteristics for Class 2 (syndromic data only) and 3 (syndromic and RAT data) models. These curves demonstrate the performance of the model for any hypothetical scenario as defined by the axes (as opposed to Figure 5 which demonstrates model performance in specific epidemiological scenarios which are realisations of a single point in this space).

295 risk of over-fitting to the scenarios, in the case of the more complex models, and
 296 because they are not policy relevant, in the case of the simpler models.

297 5. Discussion

298 We have demonstrated that combining rapid antigen tests (RATs) with
 299 syndromic modelling yields better prediction of COVID-19 status and greater
 300 flexibility than each diagnostic individually. These improvements are non-trivial
 301 in real-world settings. In Bangladesh, there are currently 15 000 new cases being
 302 identified every day, using only the limited supply of RT-PCR, the pandemic
 303 growth is accelerating and every missed case has a compounding effect. Scenario
 304 2 (“Rising Cases”) was developed with the need to keep false negative rates
 305 low and maps well onto the situation in Bangladesh (see Table 1). In this
 306 scenario, the combined data model (Model Class 3) false negative rate is 26
 307 percentage points lower than that of the RAT only model (Model Class 1). Although
 308 the syndromic only model (Model Class 2) matches the combined models false
 309 negative rate, its false positive rate is 31 percentage points higher. These are
 310 large performance gains for any diagnostic but when deployed at the scale of
 311 Bangladesh and similar countries, these improvements represent catching tens
 312 of thousands of cases that would otherwise be missed. Furthermore, this boost

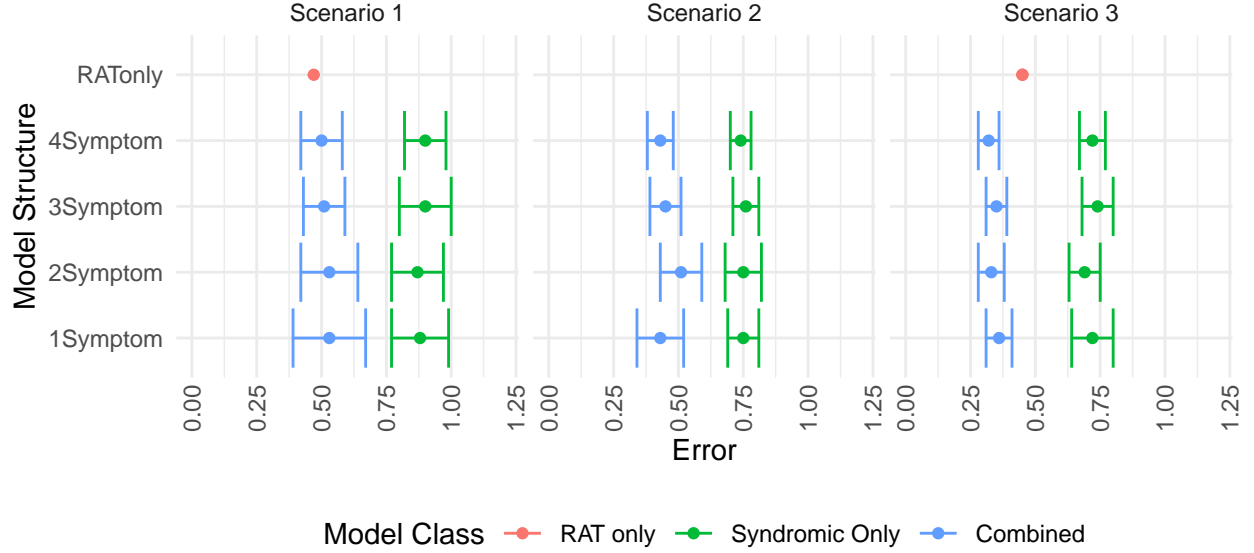


Figure 5: Performance of models under each scenario measured by errors defined in Table 2. Low errors correspond to better model performance. There is no error rate defined for the Model Class 1 (RAT only model) in Scenario 2 as the model failed to meet the requirement for that scenario (making the error functionally infinite).

is achieved with data that are already being collected in Bangladesh and other low- and middle- income countries (LMICs). Outwith developing and rerunning the models presented in this paper, these improvements are essentially cost-free and eminently scalable.

The pattern is similar in epidemiological Scenarios 1 (“Agnostic”) and 3 (“Low-Level Cases”), with the combined model class performing performing equally well or better than the other two classes (Figure 5). These three scenarios only offer snapshots of performance, however, and we strongly advocate defining model performance in terms of false negative and false positive rates with reference to local conditions. An indication of how these models will perform under any condition can be obtained by comparing the more generic model performance metrics for prediction and classification (Figures 3 and 4, respectively). These figures demonstrate both the added flexibility of the more complex model classes that allow them to be tailored to specific needs and the need to combine the high-quality but inflexible RAT results with the more flexible but lower quality syndromic data.

We have deliberately not emphasised the final symptoms chosen through model selection in this paper as we are focusing on prediction and classification for a unique sub-population: self-referring, symptomatic patients. We do, however, highlight that while fever and loss of smell were the two most important symptoms in the two classes of syndromic models, the other symptoms retained were different

334 (with cough and wet cough retained in the combined syndromic and RAT model,
 335 Class 3, and loss of taste and vomiting in the syndromic only model, Class 2).
 336 Further research is needed to understand the mechanisms by which symptoms
 337 predict COVID-19 and by which RAT misses COVID-19. Of particular interest
 338 is whether individuals that are missed by RAT are less infectious, which could
 339 be explored by using Threshold Cycle (Ct) values from the RT-PCR to compare
 340 viral load with respect to prediction by the different methods [21]. We note
 341 also that, as expected, age was retained in model selection. We were, however,
 342 surprised that gender was removed during model selection. Gender is thought to
 343 play a major role in infection risk [23]. As we are looking to predict symptomatic
 344 COVID-19 in symptomatic individuals, generalised risk of infection is perhaps
 345 less predictive than expected, potentially due to the balancing of risk and burden
 346 [24].

347 Using a large sample collected under field-realistic conditions, we have rigorously
 348 tested our approach. By taking a statistical modelling approach to case
 349 identification, we are able to update our diagnostic process in real time, allowing
 350 this method to readily adapt to new variants (or even new diseases) or new
 351 priorities for resource allocation. The modelling frameworks we have used are
 352 also sufficiently flexible to accommodate new data sources. Of particular interest
 353 are extensions to include the “pandemic context” in the model using space-time
 354 data. Furthermore, by using more sophisticated modelling structures that work
 355 at the scale of probabilities, rather than binary tests, it is possible to tune error
 356 rates to better reflect the local relative costs of false positives and false negatives.
 357 Naturally, these strengths have complementary limitations. Our models require
 358 updating in real-time and can only achieve good performance if the validation
 359 data are of high quality. Similarly, targeting error rates is only sensible if those
 360 rates properly reflect local conditions which is hard to do in practice. These
 361 limitations should be seriously considered but the alternatives for imperfect
 362 testing methods are diagnostics that cannot be tailored to local conditions at all
 363 (and, as such may perform worse than a method which is sub-optimally tailored
 364 to local conditions) or diagnostics which make these decisions implicitly and not
 365 explicitly. We choose to make these decisions explicitly to allow them to be more
 366 readily challenged, researched and improved upon. We also emphasise the need
 367 for rigorous experimental design to ensure findings from the sample population
 368 are applicable to the target population and the need for further research into
 369 understanding error rate trade-offs in applied settings.

370 We believe that the combined syndromic and rapid antigen testing approach
 371 represents the most promising approach to large-scale testing in LMICs for
 372 COVID-19 at present. By using the small amount of RT-PCR testing possible
 373 and formally integrating multiple imperfect, non-gold-standard methods, we can
 374 tune these diagnostics to our local conditions. Where data collection is being
 375 coordinated electronically (e.g. through mobile applications), the models can be
 376 used for diagnosis in the field and updated in real-time. We have demonstrated
 377 that these improvements can be impressive in real-world scenarios, and will have
 378 a large impact when scaled to the population sizes in LMICs. The methodology
 379 we have outlined here is applicable to a wide range of diseases and settings across

LMICs. One of the biggest challenges in diagnosing and tracking many diseases in resource-limited settings is the low availability of access to gold-standard testing (such as RT-PCR in the case of COVID-19) and high error rates of alternative testing methods. In this paper, we have outlined the process for coupling a small number of gold-standard tests with formal statistical integration of alternative testing methods, to generate high quality diagnostic models. This process readily maps onto many other case identification problems, including the diagnosis of several neglected tropical diseases. For example, malaria (gold standard (GS) is also RT-PCR, imperfect methods (IM) include antigen tests, syndromic diagnosis and blood smears), schistosomiasis (GS: RT-PCR or autopsy; IM: Kato Katz egg counts, antibody detection) and rabies (GS: fluorescent antibody test; IM: light microscopy, differential diagnosis).

The management of global pandemics can only be done with global testing. While the quest to achieve this using only gold-standard diagnostic methods is laudable, it is also often impractical. Imperfect diagnostics are frequently imperfect in different ways, and these differences are ripe for statistical treatment. What is more, these approaches are often more agile than gold-standard diagnostics in situations of flux, for example, in the early stages of new pandemics or disease strains, when fast responses are essential. By investing in understanding how to utilise the complementary strengths of imperfect testing and deploy the limited gold-standard testing available for validation, we can provide good quality testing at the scale needed to fight infectious diseases.

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