

GERMLINE EXOME ANALYSIS REPORT

PATIENT

Patient ID: DŽ1601

Diagnosis: Testicular myeloid sarcoma **Specimen type:** Peripheral blood **Date of collection:** 6.4.2023

METHODOLOGY*

Method used: Whole-exome sequencing **Target enrichment:** KAPA HyperExome

Sequencing device: NextSeq 500 **Date of sequencing:** 26.4.2023

Report issued: 2.5.2023

RESULTS

Gene	Variant (cDNA/protein)	Zygosity	MAF (%)	Associated phenotype	Variant classification
ERCC2	c.1847G>C/p.R616P	Het.	0,026	Xeroderma pigmentosum, group D (AR)	Pathogenic (class 5)
				Trichothiodystrophy type I, photosensitive (AR)	
				Cerebrooculofacioskeletal syndrome type 2 (AR)	

MAF – minor allele frequency – Non-Finnish European population (gnomAD database)

AD - autosomal dominant inheritance

AR - autosomal recessive inheritance

XLR - X-linked recessive

Variant classification:

Pathogenic (class 5) Likely pathogenic (class 4) Variant of uncertain significance (VUS, class 3) Likely benign (class 2) Benign (class 1)

DFP – disease-associated and/or functional polymorphism (according to the HGMD database)

A classification created according to Richards et al.: Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015 May; 17(5):405-24.

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^{*}Details regarding the methodology are listed on page 3.

INTERPRETATION

The result interpretation part is divided into the following sections:

I) Variant found

- i. Gene/protein function
- ii. Diseases and/or cancer types associated with alterations in the respective gene/protein
- iii. Variant description with supporting database and/or literature information regarding its effect on protein function (if available). Prediction algorithms results (in novel variants or variants of uncertain significance)
- iv. Additional comments, alternative nomenclature (if relevant)

1) c.1847G>C/p.R616P variant was found in the ERCC2 gene.

- i. The ERCC2 gene encodes for a DNA helicase that is a member of the RAD3/XPD family of protein helicases. ERCC2 is a regulator of the nucleotide excision repair (NER) pathway, which is used by cells to repair bulky DNA lesions that are caused by environmental mutagens, UV irradiation, and certain chemotherapeutic agents, such as cisplatin (reviewed in PMID: 24086042). ERCC2 functions as an ATP-dependent 5' to 3' helicase that unwinds damaged DNA, enabling other NER factors to access the DNA for subsequent repair (PMID: 9351836) (provided by the **OncoKB database**).
- ii. Germline variants in *ERCC2* lead to trichothiodystrophy, xeroderma pigmentosum, and Cockayne syndrome (reviewed in PMID: 19809470) (provided by the **OncoKB database**).
- iii. c.1847G>C/p.R616P in ERCC2 (NM_000400.4, exon 20) has been listed in the HGMD database as causal for trichothiodystrophy based on a primary literature report published by Broughton et al. (1994, PMID: 7920640). Subsequently, the variant has been reported in one patient with xeroderma pigmentosum by Schäfer et al. (2013, PMID: 23800062). The variant was also listed in several reports focused on mapping germline alterations in different cohorts of cancer patients. In the ClinVar database, the variant has been classified as "Pathogenic" by multiple submitters with no conflict in association with xeroderma pigmentosum group D, cerebrooculofacioskeletal syndrome type 2, and trichothiodystrophy type I.

Given the lower coverage of the respective genomic region, it is recommended to verify the variant's presence by an independent method.

METHODOLOGY

Library preparation and sequencing

DNA for whole-exome sequencing was extracted from peripheral blood using the QIAmp DNA Micro Kit (Qiagen, Germany). Sequencing libraries were prepared using KAPA HyperExome Kit (Roche, Switzerland). Sequencing was carried out on the NextSeq 500 device using NextSeq 500/550 Mid Output Kit v2,5 (150 cycles) (Illumina, CA, USA).

Patient-specific information regarding the analysis

The sequencing was performed on the 26th of April, 2023. 91% of the target regions were covered at least 30 times.

Bioinformatic workflow

Data processing: BWA (alignment to GRCh38), Picard (marking duplicates), Samtools (sorting/indexing)

Quality Control: Qualimap, PicardTools CollectHsMetrics, FastQC (aggregated in MultiQC)

Variant Calling: GATK HaplotypeCaller, VarDict, Strelka (union approach)

Annotation: Ensembl Variant Effect Predictor + in-house annotation scripts for Gene, Transcript

assignment, Variant Consequence, Population databases (1000 Genomes, GnomAD, ExAC), Clinical databases (dbSNP, CancerGeneCensus, COSMIC, HGMD, NHLBI ESP, TruSight, ClinVar, MD Anderson, Foundation One CDx), Protein structure predictors (SIFT, PolyPhen2)

Limitations

Mutations outside of the coding regions, in other genes, copy number changes, and changes that cannot be detected at present technical possibilities and current level of knowledge cannot be excluded.

Not all detected variants are reported. Reported variants were pre-selected based on their known or potential significance in the disease. All cancer-associated variants found are listed in the protocol supplement.

VARIANT CLASSIFICATION SYSTEM

The classification system used in this report is based on recommendations of the American College of Medical Genetics and Genomics (ACMG), initially published in 2015 by Richards *et al.* (PMID: 25741868). Gene selection is focused solely on cancer-associated genes if not specified otherwise. Additionally, 72 genes defined by ACMG that should be evaluated in individuals undergoing clinical exome/genome sequencing based on the medical actionability of the associated condition were evaluated as well (for more details, see Miller *et al.*, 2022; PMID: 35802134). Any potential incidental/secondary findings in such genes are reported and consulted directly with a clinical geneticist.

Pathogenic – the variant has sufficient evidence to classify as pathogenic (capable of causing disease). Targeted testing of at-risk family members and appropriate changes in medical management (i.e., high-risk surveillance) for pathogenic mutation carriers are recommended. Pathogenic variants are always included in results reports.

Likely pathogenic – the variant has strong evidence in favor of pathogenicity. Targeted testing of at-risk family members and appropriate changes in medical management (i.e., high-risk surveillance) for carriers are recommended. Likely pathogenic variants are always included in results reports.

Variant of uncertain significance (VUS) – the variant has limited and/or conflicting evidence regarding pathogenicity. VUS in cancer-associated genes is typically included in results reports, however, can be omitted for heterozygous variants in genes associated with autosomal recessive disorders that predict rather benign by available prediction algorithms.

Likely benign – the variant has strong evidence against pathogenicity. Likely benign variants are not included in the results reports.

Benign – the variant has very strong evidence against pathogenicity. Likely benign variants are not included in the results reports.

Disease-associated and/or functional polymorphism – this category was defined by the HGMD database and describes variants that are either (i) disease-associated and of likely functional significance, or (ii) of clear functional significance even though no associated clinical phenotype may have been identified to date.