

# 1 Error Model for Mass Spectrometry

## 1.1 Context

In order to understand the Nf- $\kappa$ B signaling pathway of the cell a good model of the process has to be obtained. The state of the art approach is used which is to define a structure of the model with certain parameters left to be fitted such that the model agrees with the measured data. This structure is generated by pure thought and prior knowledge of the systems. A vital part of this pursuit is to determine the functional dependencies of values in the model and their degree of freedom represented by the number of parameters that can adjust these dependencies. It is of high interest to limit this number of parameters to a minimum that just allows the model to fit all realistic systems of the kind it is supposed to model but not more, as it could get “overfitted” which results in unrealistic predictions.

Since measured data is not free of noise this noise has to be accounted for when trying to fit the model. Thus the model should not predict the measured data exactly but the measured data should be distributed around the prediction just as the measurement is distributed around the real values due to the noise of measurement. Since these distributions are not known they have to be fitted as part of the model. The right type of distribution has to be picked and the parameters fitted such that it represents the distribution of the measurements around the prediction and therefore hopefully the real value.

With the error model obtained we can calculate the probability of a certain measurement to be taken and hence the probability to measure a set of certain values like the measurements already taken. This likelihood to measure the existing data will be used as the goodness of fit. Thus fitting the selected model means trying to find a set of parameters for the model that maximizes this likelihood.

## 1.2 Goal

Here we want to analyze the mechanics of the procedure to measure the data in order to obtain an understanding of the noise which is produced relative to the real values. We hope to derive a general functional dependency of the measured values to their errors with only a few parameters to be fitted. Experience in this field of science has shown that such mechanistic approaches usually do not work perfectly due to the lack of accurate knowledge about the underlying process. Hence alternative models will be worked out and compared by means of AIC, AIC<sub>C</sub>, BIC, etc. If sufficient data is provided we

will also compute Shapiro-Wilk tests and the like to analyse the distribution of error around an established prediction, e.g. multiple measurements of the same dilution of a certain protein. If there is more than one of such data sets we could also analyze the errors dependency to properties of the protein as size or lipophilicity.

### 1.3 Mechanic Description

The cells which ought to be analyzed has a certain volume  $V_C$  and contains a discrete number  $N_C$  of the protein of interest. This number can also be represented by the concentration of the protein in the cell  $C_C$  by

$$N_C = V_C \cdot C_C$$

This cell lives in a culture of  $n$  mostly equal cells which is lysated to acquire the lysate with volume  $V_L$ , number of proteins  $N_L = N_C \cdot n$  and an concentration  $C_L = \frac{N_L}{V_L}$  which is the average of the protein concentrations of the individual cells of the culture. In the analysis of a constant state this averaging can be an advantage as it corrects for errors due to biological diversity and absorbs some deviations of outliers. But in the observation of a dynamic time dependent process it could be source of a major falsification of the data. The stimulus given to the culture does not reach each cell at an equal moment of time and the cells probably do not react equally resulting in a diverse set of reaction curves with peaks at different times and maybe even completely different shapes. An averaging at each individual point of time over all the cells would result in a reaction curve that is not representative for any typical cell reaction. Another uncertainty is whether intercellular communication can result in long forced delays of the measured process in some cells. However the resulting deviation of the measured curve to the true cell response is hard to compute and has to be ignored for now. As the process which is to be analyzed does only occur over several hours and is measured in time steps of 30 minutes to 2 hours we hope to be minimally affected by the diversity of responses in time.

However all the following steps to generate the resulting measurement can be modeled in a lot easier fashion and will be addressed now in a much more satisfying manner. As the protein travels from the lysate through the filters, ionisation, splicing and finally will be detected by a fraction of focus it has probability of  $1 - p$  to get lost and a probability  $p$  to be detected.