

# Class 08: Machine Learning Mini Project

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## Exploratory data analysis

### Save your input data file into your Project directory

I downloaded this file and saved it into my project directory.

```
fna.data <- "WisconsinCancer.csv"
```

### Complete the following code to input the data and store as wisc.df

```
wisc.df <- read.csv(fna.data, row.names=1)
```

### We can use -1 here to remove the first column

```
wisc.data <- wisc.df[, -1]
```

### Create diagnosis vector for later

```
diagnosis <- as.factor(wisc.df[, 1])
```

Q1. How many observations are in this dataset?

There are 569 observations in this dataset.

```
nrow(wisc.data)
```

```
[1] 569
```

Q2. How many of the observations have a malignant diagnosis?

There are 212 observations that have a malignant diagnosis.

```
table(diagnosis)
```

```
diagnosis
```

```
  B    M  
357 212
```

Q3. How many variables/features in the data are suffixed with `_mean`?

There are 10 variables/features in the data that are suffixed with `_mean`.

```
colnames(wisc.data)
```

```
[1] "radius_mean"      "texture_mean"  
[3] "perimeter_mean"   "area_mean"  
[5] "smoothness_mean"  "compactness_mean"  
[7] "concavity_mean"   "concave.points_mean"  
[9] "symmetry_mean"    "fractal_dimension_mean"  
[11] "radius_se"        "texture_se"  
[13] "perimeter_se"     "area_se"  
[15] "smoothness_se"    "compactness_se"  
[17] "concavity_se"     "concave.points_se"  
[19] "symmetry_se"      "fractal_dimension_se"  
[21] "radius_worst"     "texture_worst"  
[23] "perimeter_worst"  "area_worst"  
[25] "smoothness_worst" "compactness_worst"  
[27] "concavity_worst"  "concave.points_worst"  
[29] "symmetry_worst"   "fractal_dimension_worst"
```

The function `grep()` could be useful here.

```
grep("_mean", colnames(wisc.data) )
```

```
[1] 1 2 3 4 5 6 7 8 9 10
```

```
length(grep("_mean", colnames(wisc.data)))
```

```
[1] 10
```

## Principal Component Analysis (PCA)

First we need to consider whether the data needs “scaling” to make our compassionate fair.

## Check column means and standard deviations

```
colMeans(wisc.data)
```

radius_mean	texture_mean	perimeter_mean
1.412729e+01	1.928965e+01	9.196903e+01
area_mean	smoothness_mean	compactness_mean
6.548891e+02	9.636028e-02	1.043410e-01
concavity_mean	concave.points_mean	symmetry_mean
8.879932e-02	4.891915e-02	1.811619e-01
fractal_dimension_mean	radius_se	texture_se
6.279761e-02	4.051721e-01	1.216853e+00
perimeter_se	area_se	smoothness_se
2.866059e+00	4.033708e+01	7.040979e-03
compactness_se	concavity_se	concave.points_se
2.547814e-02	3.189372e-02	1.179614e-02
symmetry_se	fractal_dimension_se	radius_worst
2.054230e-02	3.794904e-03	1.626919e+01
texture_worst	perimeter_worst	area_worst
2.567722e+01	1.072612e+02	8.805831e+02
smoothness_worst	compactness_worst	concavity_worst
1.323686e-01	2.542650e-01	2.721885e-01
concave.points_worst	symmetry_worst	fractal_dimension_worst
1.146062e-01	2.900756e-01	8.394582e-02

```
apply(wisc.data,2,sd)
```

radius_mean	texture_mean	perimeter_mean
3.524049e+00	4.301036e+00	2.429898e+01
area_mean	smoothness_mean	compactness_mean
3.519141e+02	1.406413e-02	5.281276e-02
concavity_mean	concave.points_mean	symmetry_mean
7.971981e-02	3.880284e-02	2.741428e-02
fractal_dimension_mean	radius_se	texture_se
7.060363e-03	2.773127e-01	5.516484e-01
perimeter_se	area_se	smoothness_se
2.021855e+00	4.549101e+01	3.002518e-03
compactness_se	concavity_se	concave.points_se
1.790818e-02	3.018606e-02	6.170285e-03
symmetry_se	fractal_dimension_se	radius_worst
8.266372e-03	2.646071e-03	4.833242e+00
texture_worst	perimeter_worst	area_worst
6.146258e+00	3.360254e+01	5.693570e+02
smoothness_worst	compactness_worst	concavity_worst
2.283243e-02	1.573365e-01	2.086243e-01
concave.points_worst	symmetry_worst	fractal_dimension_worst
6.573234e-02	6.186747e-02	1.806127e-02

## Perform PCA on wisc.data by completing the following code

```
wisc.pr <- prcomp( wisc.data, scale = TRUE )

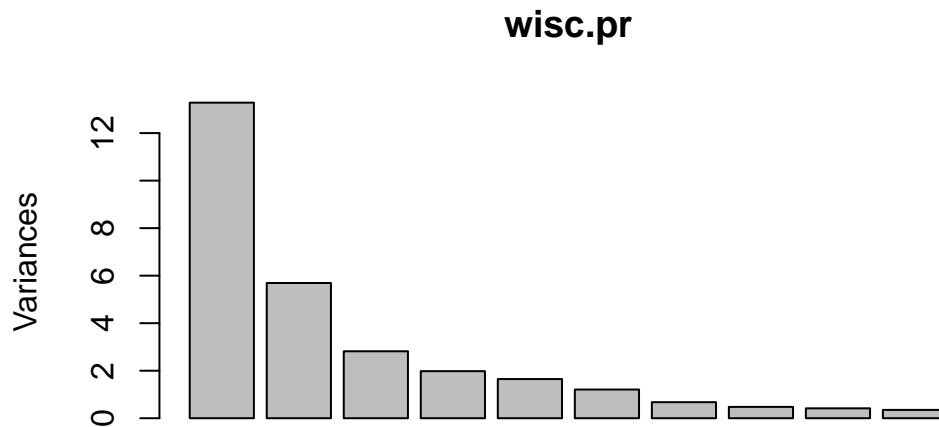
summary(wisc.pr)
```

Importance of components:

	PC1	PC2	PC3	PC4	PC5	PC6	PC7
Standard deviation	3.6444	2.3857	1.67867	1.40735	1.28403	1.09880	0.82172
Proportion of Variance	0.4427	0.1897	0.09393	0.06602	0.05496	0.04025	0.02251
Cumulative Proportion	0.4427	0.6324	0.72636	0.79239	0.84734	0.88759	0.91010
	PC8	PC9	PC10	PC11	PC12	PC13	PC14
Standard deviation	0.69037	0.6457	0.59219	0.5421	0.51104	0.49128	0.39624
Proportion of Variance	0.01589	0.0139	0.01169	0.0098	0.00871	0.00805	0.00523
Cumulative Proportion	0.92598	0.9399	0.95157	0.9614	0.97007	0.97812	0.98335
	PC15	PC16	PC17	PC18	PC19	PC20	PC21
Standard deviation	0.30681	0.28260	0.24372	0.22939	0.22244	0.17652	0.1731
Proportion of Variance	0.00314	0.00266	0.00198	0.00175	0.00165	0.00104	0.0010

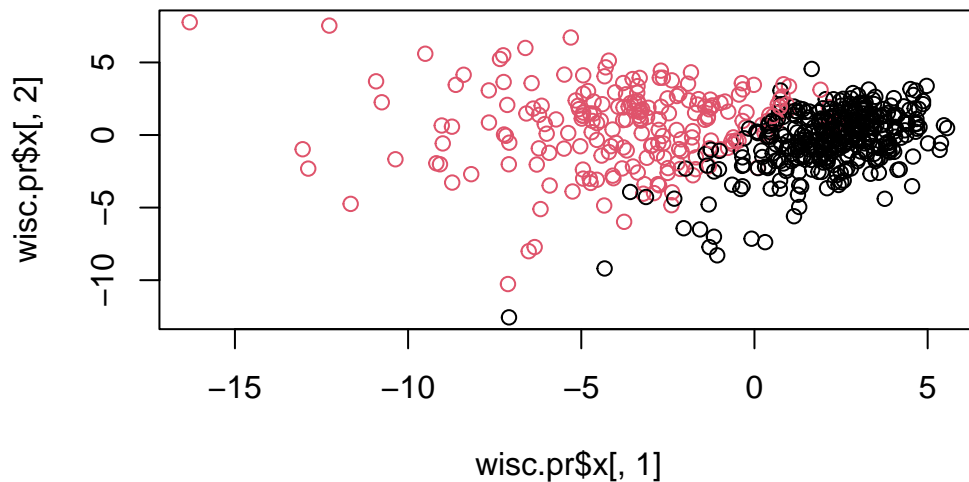
Cumulative Proportion	0.98649	0.98915	0.99113	0.99288	0.99453	0.99557	0.9966
	PC22	PC23	PC24	PC25	PC26	PC27	PC28
Standard deviation	0.16565	0.15602	0.1344	0.12442	0.09043	0.08307	0.03987
Proportion of Variance	0.00091	0.00081	0.0006	0.00052	0.00027	0.00023	0.00005
Cumulative Proportion	0.99749	0.99830	0.9989	0.99942	0.99969	0.99992	0.99997
	PC29	PC30					
Standard deviation	0.02736	0.01153					
Proportion of Variance	0.00002	0.00000					
Cumulative Proportion	1.00000	1.00000					

```
plot(wisc.pr)
```



Let's make a PC plot (a.k.a. "score plot" or "PC1 vs PC2" etc. plot)

```
plot(wisc.pr$x[,1], wisc.pr$x[,2], col = diagnosis)
```



Q4. From your results, what proportion of the original variance is captured by the first principal components (PC1)?

The proportion of the original variance that is captured by the first principal components (PC1) is 44.27%.

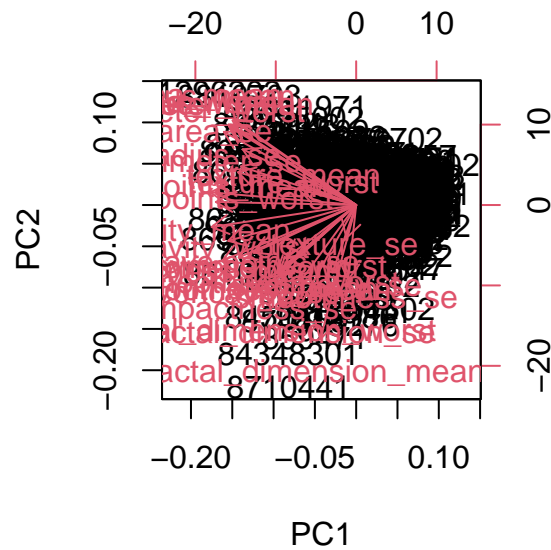
Q5. How many principal components (PCs) are required to describe at least 70% of the original variance in the data?

Three principal components (PCs) are required to describe at least 70% of the original variance in the data. This information can be found by looking at the cumulative proportion of PC3 (72.636%).

Q6. How many principal components (PCs) are required to describe at least 90% of the original variance in the data?

There are seven principal components that are required to describe at least 90% of the original variance in the data. This information can be found by looking at the cumulative proportion of PC7 (91.010%).

```
biplot(wisc.pr)
```

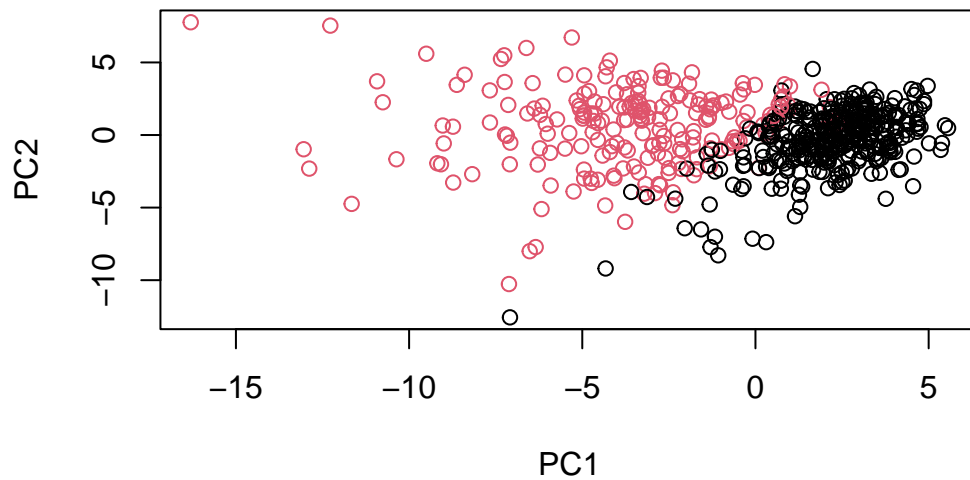


Q7. What stands out to you about this plot? Is it easy or difficult to understand? Why?

What stands out to me about this plot is that all of the data is very compact and difficult to analyze. I can tell that there are data labels and numerical values within the plot, but they are very hard to decipher, and therefore hard to understand.

## Scatter plot observations by components 1 and 2

```
plot( wisc.pr$x[,1], wisc.pr$x[,2] , col = diagnosis ,
      xlab = "PC1", ylab = "PC2")
```



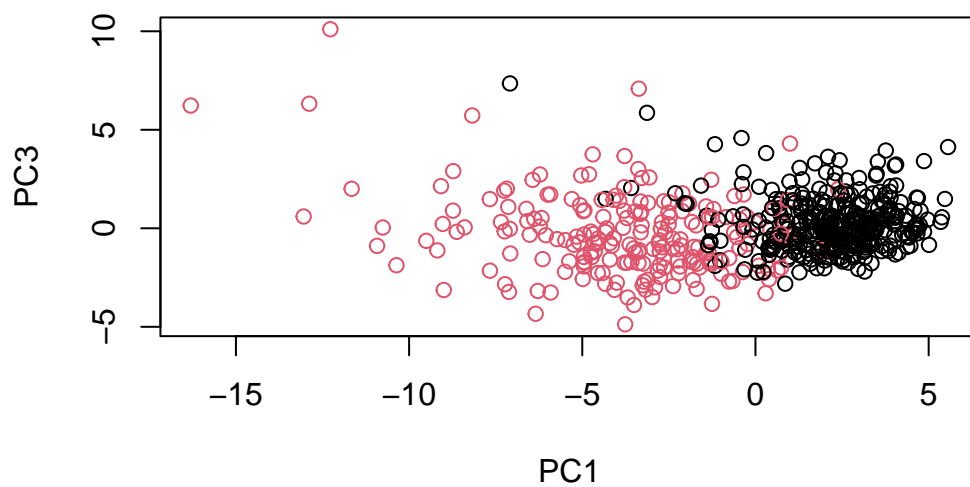
## Repeat for components 1 and 3

Q8. Generate a similar plot for principal components 1 and 3. What do you notice about these plots?

The scatterplots for PC1 vs PC2 and PC1 vs PC3 are much easier to analyze and comprehend compared to the biplot made previously. The graph effectively shows the differences in clustering between the two diagnosis groups.

```
plot( wisc.pr$x[,1], wisc.pr$x[,3] , col = diagnosis ,  
      xlab = "PC1", ylab = "PC3")
```





## Create a data.frame for ggplot

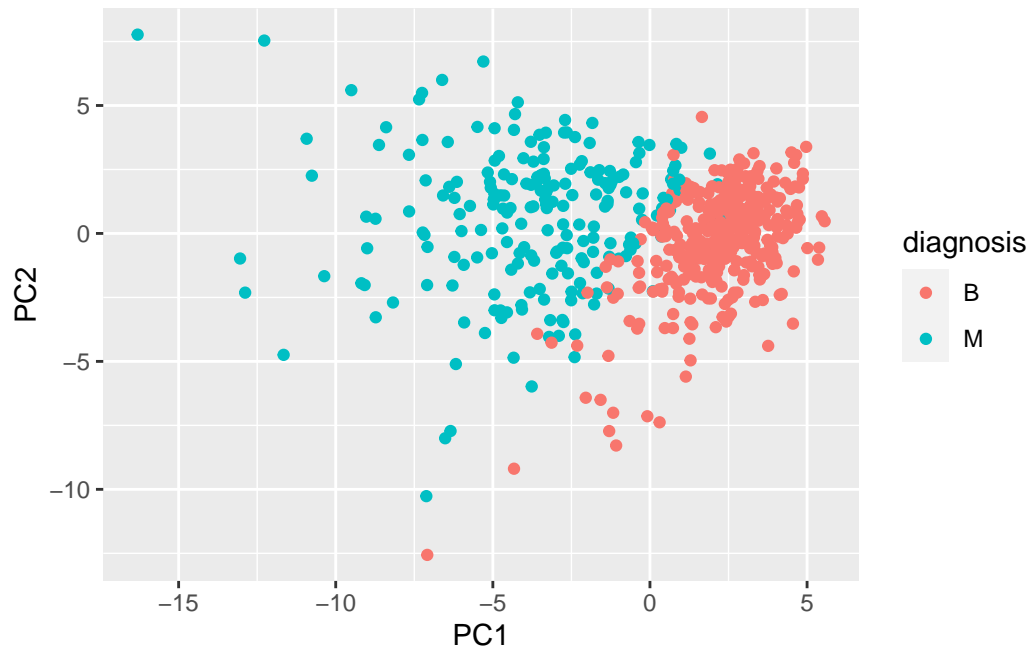
```
df <- as.data.frame(wisc.pr$x)
df$diagnosis <- diagnosis
```

## Load the ggplot2 package

```
library(ggplot2)
```

## Make a scatter plot colored by diagnosis

```
ggplot(df) +
  aes(PC1, PC2, col=diagnosis) +
  geom_point()
```



### Variance explained

### Calculate variance of each component

```
pr.var <- wisc.pr$sdev^2  
head(pr.var)
```

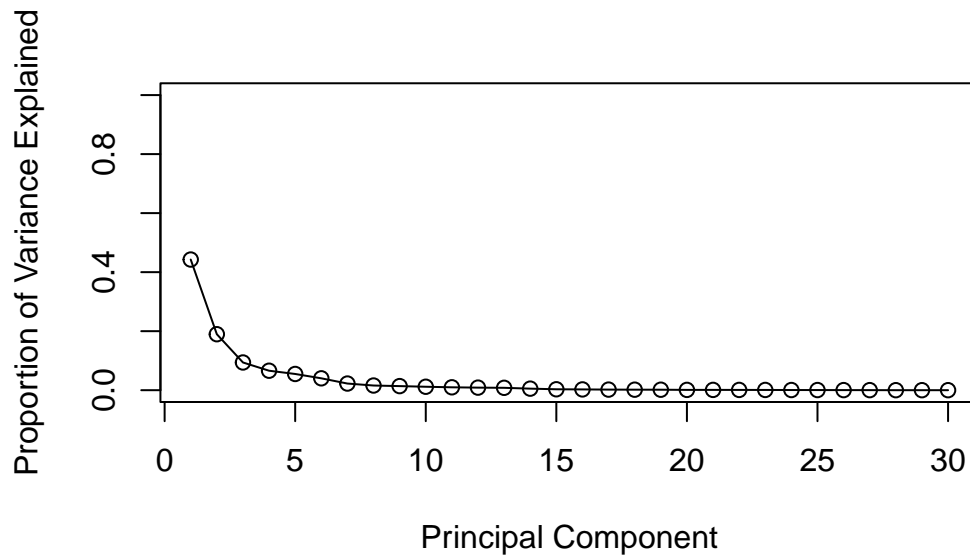
```
[1] 13.281608  5.691355  2.817949  1.980640  1.648731  1.207357
```

### Variance explained by each principal component: pve

```
pve <- pr.var / sum(pr.var)
```

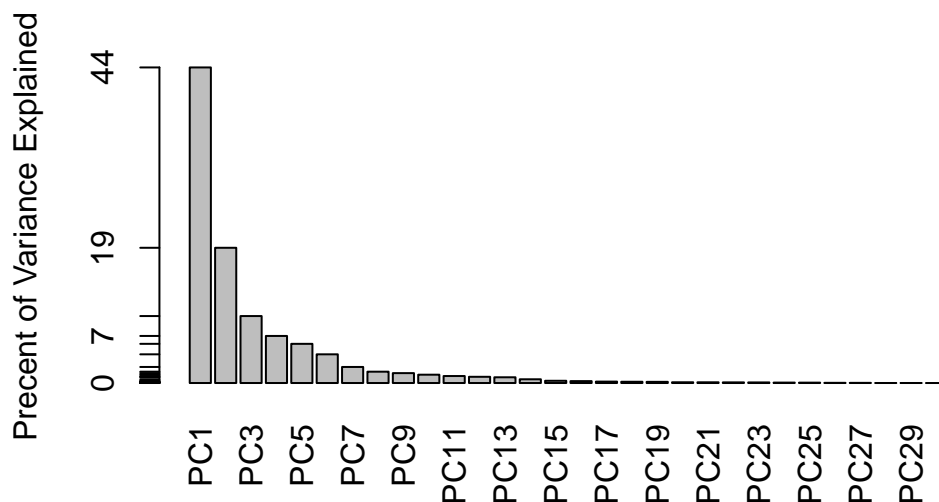
## Plot variance explained for each principal component

```
plot(pve, xlab = "Principal Component",  
     ylab = "Proportion of Variance Explained",  
     ylim = c(0, 1), type = "o")
```



## Alternative scree plot of the same data, note data driven y-axis

```
barplot(pve, ylab = "Precent of Variance Explained",  
        names.arg=paste0("PC",1:length(pve)), las=2, axes = FALSE)  
axis(2, at=pve, labels=round(pve,2)*100 )
```



## Communicating PCA results

Q9. For the first principal component, what is the component of the loading vector (i.e. `wisc.pr$rotation[,1]`) for the feature `concave.points_mean`?

For the first principal component, the component of the loading vector for the feature ‘`concave.points_mean`’ is -0.26085376.

```
wisc.pr$rotation[,1]
```

radius_mean	texture_mean	perimeter_mean
-0.21890244	-0.10372458	-0.22753729
area_mean	smoothness_mean	compactness_mean
-0.22099499	-0.14258969	-0.23928535
concavity_mean	concave.points_mean	symmetry_mean
-0.25840048	-0.26085376	-0.13816696
fractal_dimension_mean	radius_se	texture_se
-0.06436335	-0.20597878	-0.01742803
perimeter_se	area_se	smoothness_se
-0.21132592	-0.20286964	-0.01453145
compactness_se	concavity_se	concave.points_se

-0.17039345	-0.15358979	-0.18341740
symmetry_se	fractal_dimension_se	radius_worst
-0.04249842	-0.10256832	-0.22799663
texture_worst	perimeter_worst	area_worst
-0.10446933	-0.23663968	-0.22487053
smoothness_worst	compactness_worst	concavity_worst
-0.12795256	-0.21009588	-0.22876753
concave.points_worst	symmetry_worst	fractal_dimension_worst
-0.25088597	-0.12290456	-0.13178394

Q10. What is the minimum number of principal components required to explain 80% of the variance of the data?

The minimum number of principal components required to explain 80% of the variance of the data is 5. This information can be found by looking at the cumulative proportion for PC5 (84.734%).

```
summary(wisc.pr)
```

Importance of components:

	PC1	PC2	PC3	PC4	PC5	PC6	PC7
Standard deviation	3.6444	2.3857	1.67867	1.40735	1.28403	1.09880	0.82172
Proportion of Variance	0.4427	0.1897	0.09393	0.06602	0.05496	0.04025	0.02251
Cumulative Proportion	0.4427	0.6324	0.72636	0.79239	0.84734	0.88759	0.91010
	PC8	PC9	PC10	PC11	PC12	PC13	PC14
Standard deviation	0.69037	0.6457	0.59219	0.5421	0.51104	0.49128	0.39624
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Cumulative Proportion	0.92598	0.9399	0.95157	0.9614	0.97007	0.97812	0.98335
	PC15	PC16	PC17	PC18	PC19	PC20	PC21
Standard deviation	0.30681	0.28260	0.24372	0.22939	0.22244	0.17652	0.1731
Proportion of Variance	0.00314	0.00266	0.00198	0.00175	0.00165	0.00104	0.0010
Cumulative Proportion	0.98649	0.98915	0.99113	0.99288	0.99453	0.99557	0.9966
	PC22	PC23	PC24	PC25	PC26	PC27	PC28
Standard deviation	0.16565	0.15602	0.1344	0.12442	0.09043	0.08307	0.03987
Proportion of Variance	0.00091	0.00081	0.0006	0.00052	0.00027	0.00023	0.00005
Cumulative Proportion	0.99749	0.99830	0.9989	0.99942	0.99969	0.99992	0.99997
	PC29	PC30					
Standard deviation	0.02736	0.01153					
Proportion of Variance	0.00002	0.00000					
Cumulative Proportion	1.00000	1.00000					

## Hierarchical Clustering

### Scale the wisc.data data using the “scale()” function

```
data.scaled <- scale(wisc.data)
```

```
data.dist <- dist(data.scaled)
```

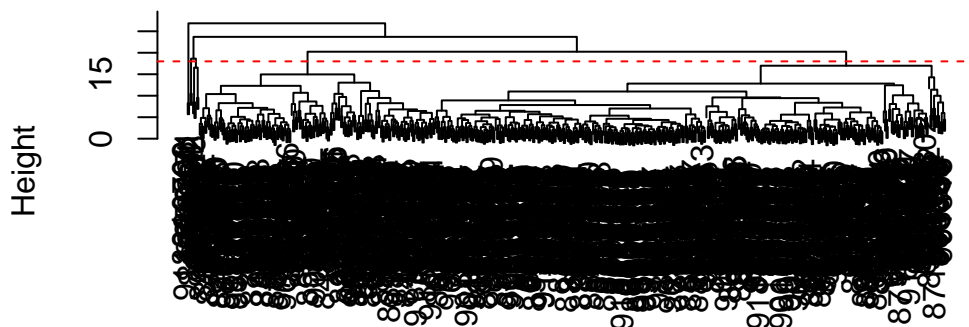
```
wisc.hclust <- hclust(data.dist, method = "complete")
```

Q11. Using the plot() and abline() functions, what is the height at which the clustering model has 4 clusters?

The height at which the clustering model has four clusters is 18.

```
plot(wisc.hclust)  
abline(h=18, col="red", lty=2)
```

### Cluster Dendrogram



data.dist  
hclust (\*, "complete")

```
wisc.hclust.clusters <- cutree(wisc.hclust, k = 4)
```

```
table(wisc.hclust.clusters, diagnosis)
```

	diagnosis	
wisc.hclust.clusters	B	M
1	12	165
2	2	5
3	343	40
4	0	2

Q12. Can you find a better cluster vs diagnoses match by cutting into a different number of clusters between 2 and 10?

There is not a better cluster than four because the two diagnoses (benign and malignant) there is a very clear distinction between these values.

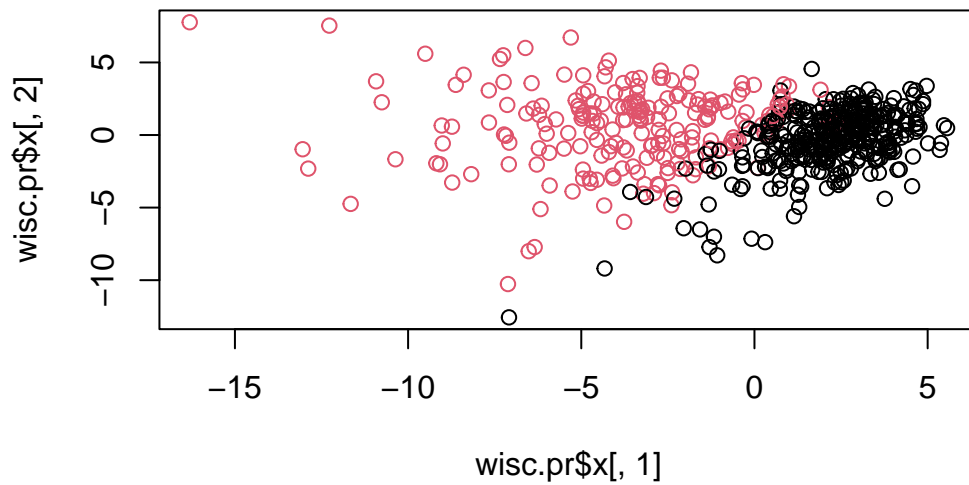
Q13. Which method gives your favorite results for the same data.dist dataset? Explain your reasoning.

“ward.d2” is my preferred method for giving results for the same data.dist dataset. This is because the “ward.d2” function allows for the clusters to be more organized in more sepecific ways. Within each cluster, there is less variance.

## Combining Methods

I want to cluster in “PC sapce.”

```
plot(wisc.pr$x[,1], wisc.pr$x[,2], col = diagnosis)
```



The `hclust()` function wants a distance matrix as input...

```
d <- dist(wisc.pr$x[, 1:7])
wisc.pr.hclust <- hclust(d, method="ward.D2")
```

```
grps <- cutree(wisc.pr.hclust, k=2)
table(grps)
```

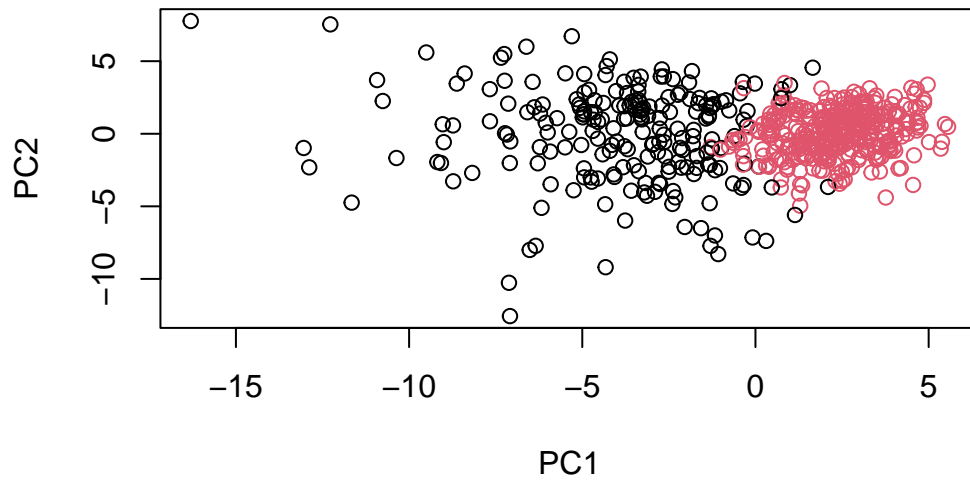
```
grps
  1  2
216 353
```

```
table(grps, diagnosis)
```

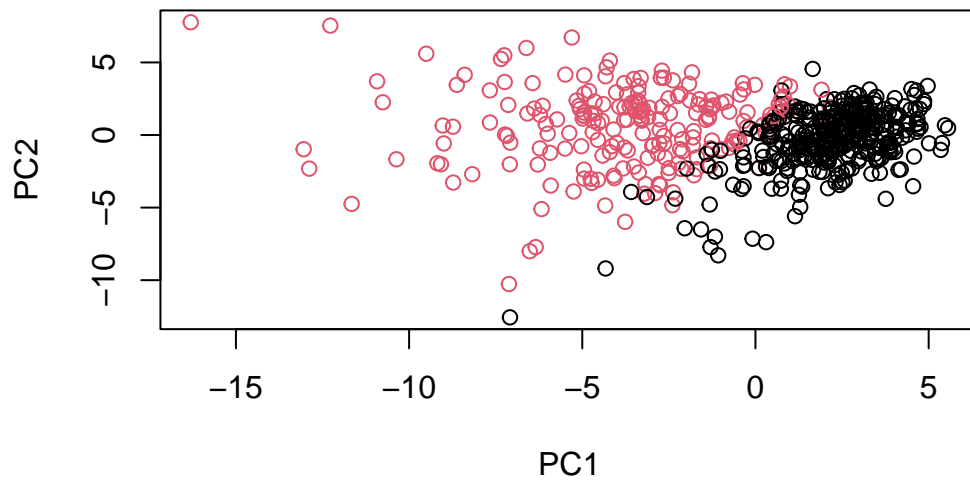
```
      diagnosis
grps   B    M
  1   28 188
  2  329  24
```



```
plot(wisc.pr$x[,1:2], col=grps)
```



```
plot(wisc.pr$x[,1:2], col=diagnosis)
```



```
g <- as.factor(grps)
levels(g)
```

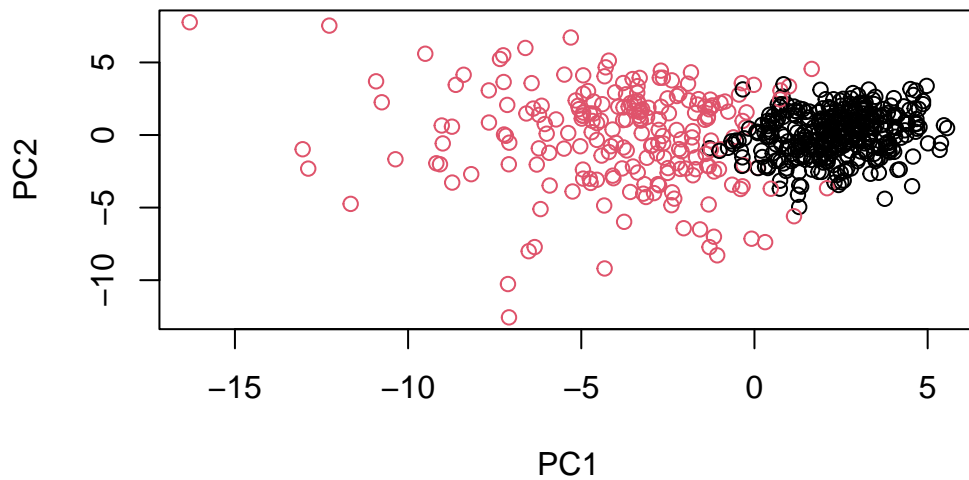
```
[1] "1" "2"
```

```
g <- relevel(g,2)
levels(g)
```

```
[1] "2" "1"
```

## Plot using our re-ordered factor

```
plot(wisc.pr$x[,1:2], col=g)
```



Use the distance along the first 7 PCs for clustering i.e. `wisc.pr$x[, 1:7]`

```
wisc.pr.hclust <- hclust(d, method="ward.D2")
```

```
wisc.pr.hclust.clusters <- cutree(wisc.pr.hclust, k=2)
```

Q15. How well does the newly created model with two clusters separate out the two diagnoses?

The newly created model with two clusters separates out the two diagnoses works much better than the model with four clusters. This is due to the fact that the two cluster model creates a much clearer distinction between the malignant and benign diagnoses.

## Compare to actual diagnoses

```
table(wisc.pr.hclust.clusters, diagnosis)
```

diagnosis

```
wisc.pr.hclust.clusters  B  M
                        1 28 188
                        2 329 24
```

Q16. How well do the k-means and hierarchical clustering models you created in previous sections (i.e. before PCA) do in terms of separating the diagnoses? Again, use the `table()` function to compare the output of each model (`wisc.km$cluster` and `wisc.hclust.clusters`) with the vector containing the actual diagnoses.

The `wisc.hclust.clusters` method seems to be better because it separates the benign and malignant diagnoses in a more distinct way.

```
table(wisc.pr.hclust.clusters, diagnosis)
```

```

              diagnosis
wisc.pr.hclust.clusters  B  M
                        1 28 188
                        2 329 24
```

```
table(wisc.hclust.clusters, diagnosis)
```

```

              diagnosis
wisc.hclust.clusters  B  M
                        1 12 165
                        2  2  5
                        3 343 40
                        4  0  2
```

Q17. Which of your analysis procedures resulted in a clustering model with the best specificity? How about sensitivity?

`wisc.hclust.clusters` has a the best sensitivity because it has a higher sensitivity than `wisc.pr.hclust.clusters`. `wisc.hclust.clusters` has a the best specificity because it has a higher specificity than `wisc.pr.hclust.clusters`. This result shows that the `wisc.hclust.clusters` method is more sensitive and specific.

FN TP TN FP

**Sensitivity  $TP/(TP+FN)$**

For `wisc.pr.hclust.clusters`

$188/(188+28)$

[1] 0.8703704

For `wisc.hclust.clusters`

$165/(165+12)$

[1] 0.9322034

## **Specificity $TN/(TN+FN)$**

For `wisc.pr.hclust.clusters`

$329/(329+28)$

[1] 0.9215686

For `wisc.hclust.clusters`

$343/(343+12)$

[1] 0.9661972