

1      **Erosion of somatic tissue identity with loss of the X-linked**  
2                    **intellectual disability factor KDM5C**

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5      **Abstract**

6      Mutations in numerous chromatin-modifying enzymes cause neurodevelopmental disorders (NDDs) with  
7      unknown mechanisms. Loss of repressive chromatin regulators can lead to the aberrant transcription of  
8      tissue-specific genes outside of their intended context, however the mechanisms and consequences of  
9      their dysregulation are largely unknown. Here, we examine the role of lysine demethylase 5c (KDM5C),  
10     an eraser of histone 3 lysine 4 di and tri-methylation (H3K4me2/3) mutated in Claes-Jensen X-linked  
11     intellectual disability, in tissue identity. We found male *Kdm5c* knockout (-KO) mice, which recapitulate  
12     key human neurological phenotypes, aberrantly expresses many liver, muscle, ovary, and testis genes  
13     within the amygdala and hippocampus. Gonad-enriched genes misexpressed in the *Kdm5c*-KO brain are  
14     unique to germ cells, indicating an erosion of the soma-germline boundary. Germline genes are typically  
15     decommissioned in somatic lineages in the post-implantation epiblast, yet *Kdm5c*-KO epiblast-like cells  
16     (EpiLCs) aberrantly expressed key regulators of germline identity and meiosis, including *Dazl* and *Stra8*.  
17     Germline gene suppression is sexually dimorphic, as female EpiLCs required a higher dose of KDM5C  
18     to maintain germline gene suppression. Using a comprehensive list of mouse germline-enriched genes,  
19     we found KDM5C is selectively recruited to a subset of germline gene promoters that contain CpG islands  
20     (CGIs) to facilitate DNA CpG methylation (CpGme) during ESC to EpiLC differentiation. However, late stage  
21     spermatogenesis genes devoid of promoter CGIs can also become activated in *Kdm5c*-KO cells via ectopic  
22     activation by RFX transcription factors. Thus, distinct mechanisms govern the misexpression of germline  
23     gene classes, including activation by ectopic germline programs that mirror germ cell development within  
24     somatic tissues.

25      **Introduction**

26      A single genome holds the instructions to generate the myriad of cell types found within an organism.  
27      This is, in part, accomplished by chromatin regulators that can either promote or impede lineage-specific

28 gene expression through DNA and histone modifications<sup>1–5</sup>. Human genetic studies revealed mutations in  
29 chromatin regulators are a major cause of neurodevelopmental disorders (NDDs)<sup>6</sup> and many studies have  
30 identified their importance for regulating brain-specific transcriptional programs. Loss of some chromatin  
31 regulators can also result in the ectopic expression of tissue-specific genes outside of their target environment,  
32 such as the misexpression of liver-specific genes within adult neurons<sup>7</sup>. However, the mechanisms underlying  
33 ectopic gene expression and its impact upon neurodevelopment are poorly understood.

34 To elucidate the role of tissue identity in chromatin-linked NDDs, it is essential to first characterize the  
35 nature of the dysregulated genes and the molecular mechanisms governing their de-repression. Here we  
36 focus on lysine demethylase 5C (KDM5C, also known as SMCX or JARID1C), which erases histone 3 lysine  
37 4 di- and trimethylation (H3K4me2/3), a permissive chromatin modification enriched at gene promoters<sup>8</sup>.  
38 Pathogenic mutations in *KDM5C* cause Intellectual Developmental Disorder, X-linked, Syndromic, Claes-  
39 Jensen Type (MRXSCJ, OMIM: 300534). MRXSCJ is more common and severe in males and its neurological  
40 phenotypes include intellectual disability, seizures, aberrant aggression, and autistic behaviors<sup>9–11</sup>. Male  
41 *Kdm5c* knockout (-KO) mice recapitulate key MRXSCJ phenotypes, including hyperaggression, increased  
42 seizure propensity, and learning impairments<sup>12,13</sup>. RNA sequencing (RNA-seq) of the *Kdm5c*-KO hippocam-  
43 pus revealed ectopic expression of some germline genes within the brain<sup>13</sup>. However, it is unclear if other  
44 tissue-specific genes are aberrantly transcribed with KDM5C loss, at what point in development germline  
45 gene misexpression begins, and what mechanisms underlie their dysregulation.

46 Distinguishing between germ cells and somatic cells is a key feature of multicellularity<sup>14</sup> that occurs  
47 during early embryogenesis in many metazoans<sup>15</sup>. In mammals, chromatin regulators are crucial for  
48 decommissioning germline genes during the transition from naïve to primed pluripotency. Initially, germline  
49 gene promoters gain repressive histone H2A lysine 119 monoubiquitination (H2AK119ub1)<sup>16</sup> and histone 3  
50 lysine 9 trimethylation (H3K9me3)<sup>16,17</sup> in embryonic stem cells (ESCs) and are then decorated with DNA  
51 CpG methylation (CpGme) in the post-implantation embryo<sup>17–20</sup>. The contribution of KDM5C to this process  
52 remains unclear. Furthermore, studies on germline gene repression have primarily been conducted in  
53 males and focused on marker genes important for germ cell development rather than germline genes as a  
54 whole, given the lack of a curated germline-enriched gene list. Therefore, it is unknown if the mechanism  
55 of repression differs between sexes or for certain classes of germline genes, e.g. meiotic genes versus  
56 spermatid differentiation genes.

57 To illuminate KDM5C's role in tissue identity, here we characterized the aberrant expression of tissue-  
58 enriched genes within the *Kdm5c*-KO brain and epiblast-like stem cells (EpiLCs), an *in vitro* model of the  
59 post-implantation embryo. We curated a list of mouse germline-enriched genes, which enabled genome-wide  
60 analysis of germline gene silencing mechanisms for the first time. Based on the data presented below, we  
61 propose KDM5C plays a fundamental, sexually dimorphic role in the development of tissue identity during  
62 early embryogenesis, including the establishment of the soma-germline boundary.

63 **Results**

64 **Tissue-enriched genes are aberrantly expressed in the *Kdm5c*-KO brain**

65 Previous RNA sequencing (RNA-seq) of the adult male *Kdm5c*-KO hippocampus revealed ectopic  
66 expression of some germline genes unique to the testis<sup>13</sup>. It is currently unknown if the testis is the only  
67 tissue type misexpressed in the *Kdm5c*-KO brain. We thus systematically tested whether other tissue-specific  
68 genes are misexpressed in the male brain with constitutive knockout of *Kdm5c* (*Kdm5c*<sup>-y</sup>, 5CKO)<sup>21</sup> by using  
69 a published list of mouse tissue-enriched genes<sup>22</sup>.

70 We found a large proportion of significantly upregulated genes (DESeq2<sup>23</sup>, log2 fold change > 0.5,  
71 q < 0.1) within the male *Kdm5c*-KO amygdala and hippocampus are typically enriched within non-brain  
72 tissues (Amygdala: 35%, Hippocampus: 24%) (Figure 1A-B). For both the amygdala and hippocampus,  
73 the majority of tissue-enriched differentially expressed genes (DEGs) were testis genes (Figure 1A-B).  
74 Even though the testis has the largest total number of tissue-biased genes (2,496 genes) compared to any  
75 other tissue, testis-biased DEGs were significantly enriched for both brain regions (Amygdala p = 1.83e-05,  
76 Odds Ratio = 5.13; Hippocampus p = 4.26e-11, Odds Ratio = 4.45, Fisher's Exact Test). An example of a  
77 testis-enriched gene misexpressed in the *Kdm5c*-KO brain is *FK506 binding protein 6* (*Fkbp6*), a known  
78 regulator of PIWI-interacting RNAs (piRNAs) and meiosis<sup>24,25</sup> (Figure 1C).

79 Interestingly, we also observed significant enrichment of ovary-biased DEGs in both the amygdala and  
80 hippocampus (Amygdala p = 0.00574, Odds Ratio = 18.7; Hippocampus p = 0.048, Odds Ratio = 5.88,  
81 Fisher's Exact) (Figure 1A-B). Ovary-enriched DEGs included *Zygotic arrest 1* (*Zar1*), which sequesters  
82 mRNAs in oocytes for meiotic maturation<sup>26</sup> (Figure 1D). Given that the *Kdm5c*-KO mice we analyzed are  
83 male, these data demonstrate that the ectopic expression of gonad-enriched genes is independent of  
84 organismal sex.

85 Although not consistent across brain regions, we also found significant enrichment of DEGs biased  
86 towards two non-gonadal tissues - the liver (Amygdala p = 0.04, Odds Ratio = 6.58, Fisher's Exact Test) and  
87 muscles (Hippocampus p = 0.01, Odds Ratio = 6.95, Fisher's Exact Test) (Figure 1A-B). *Apolipoprotein C-I*  
88 (*Apoc1*) a lipoprotein metabolism and transport gene, is among the liver-biased DEG derepressed in both  
89 the hippocampus and amygdala<sup>27</sup> and its brain overexpression has been implicated in Alzheimer's disease<sup>28</sup>  
90 (Figure 1E).

91 For all *Kdm5c*-KO tissue-enriched DEGs, aberrantly expressed mRNAs are polyadenylated and spliced  
92 into mature transcripts (Figure 1C-E). Of note, we observed little to no dysregulation of brain-enriched genes  
93 (Amygdala p = 1, Odds Ratio = 1.22; Hippocampus p = 0.74, Odds Ratio = 1.22, Fisher's Exact), despite the  
94 fact these are brain samples and the brain has the second highest total number of tissue-enriched genes  
95 (708 genes). Altogether, these results suggest the aberrant expression of tissue-enriched genes within the  
96 brain is a major effect of KDM5C loss.

97 **Germline genes are misexpressed in the *Kdm5c*-KO brain**

98     *Kdm5c*-KO brain expresses testicular germline genes<sup>13</sup>, however the testis also contains somatic cells that  
99 support hormone production and germline functions. To determine if *Kdm5c*-KO results in ectopic expression  
100 of somatic testicular genes, we first evaluated the known functions of testicular DEGs through gene ontology.  
101 We found *Kdm5c*-KO testis-enriched DEGs had high enrichment of germline-relevant ontologies, including  
102 spermatid development (GO: 0007286, p.adjust = 6.2e-12) and sperm axoneme assembly (GO: 0007288,  
103 p.adjust = 2.45e-14) (Figure 2A).

104     We then evaluated testicular DEG expression in wild-type testes versus testes with germ cell depletion<sup>29</sup>,  
105 which was accomplished by heterozygous *W* and *Wv* mutations in the enzymatic domain of *c-Kit* (*Kit*<sup>W/Wv</sup>)<sup>30</sup>.  
106 Almost all *Kdm5c*-KO testis-enriched DEGs lost expression with germ cell depletion (Figure 2B). We then  
107 assessed testis-enriched DEG expression in a published single cell RNA-seq dataset that identified cell  
108 type-specific markers within the testis<sup>31</sup>. Some *Kdm5c*-KO testis-enriched DEGs were classified as specific  
109 markers for different germ cell developmental stages (e.g. spermatogonia, spermatocytes, round spermatids,  
110 and elongating spermatids), yet none marked somatic cells (Figure 2C). Together, these data demonstrate  
111 that the *Kdm5c*-KO brain aberrantly expresses germline genes but not somatic testicular genes, reflecting an  
112 erosion of the soma-germline boundary.

113     As of yet, research on germline gene silencing mechanisms has focused on a handful of key genes rather  
114 than assessing germline gene suppression genome-wide, due to the lack of a comprehensive gene list.  
115 We therefore generated a list of mouse germline-enriched genes using RNA-seq datasets of *Kit*<sup>W/Wv</sup> mice  
116 that included males and females at embryonic day 12, 14, and 16<sup>32</sup> and adult male testes<sup>29</sup>. We defined  
117 genes as germline-enriched if their expression met the following criteria: 1) their expression is greater than  
118 1 FPKM in wild-type gonads 2) their expression in any non-gonadal tissue of adult wild type mice<sup>22</sup> does  
119 not exceed 20% of their maximum expression in the wild-type germline, and 3) their expression in the germ  
120 cell-depleted gonads, for any sex or time point, does not exceed 20% of their maximum expression in the  
121 wild-type germline. These criteria yielded 1,288 germline-enriched genes (Figure 2D), which was hereafter  
122 used as a resource to globally characterize germline gene misexpression with *Kdm5c* loss (Supplementary  
123 table 1).

124 ***Kdm5c*-KO epiblast-like cells aberrantly express key regulators of germline identity**

125     Germ cells are typically distinguished from somatic cells soon after the embryo implants into the uterine  
126 wall<sup>33,34</sup>, when germline genes are silenced in epiblast stem cells that will form the somatic tissues<sup>35</sup>. This  
127 developmental time point can be modeled *in vitro* through differentiation of naïve embryonic stem cells  
128 (nESCs) into epiblast-like stem cells (EpiLCs) (Figure 3A)<sup>36,37</sup>. While some germline-enriched genes are  
129 also expressed in nESCs and in the 2-cell stage<sup>38–40</sup>, they are silenced as they differentiate into EpiLCs<sup>17,18</sup>.  
130 Therefore, we tested if KDM5C was necessary for the initial silencing of germline genes in somatic lineages

131 by evaluating the impact of *Kdm5c* loss in male EpiLCs.  
132 *Kdm5c*-KO cell morpholgy during ESC to EpiLC differentiation appeared normal (Figure 3B) and EpiLCs  
133 properly expressed markers of primed pluripotency, such as *Dnmt3b*, *Fgf5*, *Pou3f1*, and *Otx2* (Figure 3C). We  
134 then identified tissue-enriched DEGs in a RNA-seq dataset of wild-type and *Kdm5c*-KO EpiLCs<sup>41</sup> (DESeq2,  
135 log2 fold change > 0.5, q < 0.1). Similar to the *Kdm5c*-KO brain, we observed general dysregulation of  
136 tissue-enriched genes, with the largest number of genes belonging to the brain and testis, although they  
137 were not significantly enriched (Figure 3D). Using our list of mouse germline-enriched genes assembled  
138 above, we found 68 germline genes were misexpressed in male *Kdm5c*-KO EpiLCs.

139 We then compared EpiLC germline DEGs to those expressed in the *Kdm5c*-KO brain to determine if  
140 germline genes are constitutively dysregulated or change over the course of development. The majority of  
141 germline DEGs were unique to either EpiLCs or the brain, with only *D1Pas1* and *Cyct* shared across all  
142 tissue/cell types (Figure 3E-F). EpiLC germline DEGs had particularly high enrichment of meiosis-related  
143 gene ontologies when compared to the brain (Figure 3G), such as meiotic cell cycle process (GO:1903046,  
144 p.adjust = 2.2e-07) and meiotic nuclear division (GO:0140013, p.adjust = 1.37e-07). While there was  
145 modest enrichment of meiotic gene ontologies in both brain regions, the *Kdm5c*-KO hippocampus primarily  
146 expressed late-stage spermatogenesis genes involved in sperm axoneme assembly (GO:0007288, p.adjust  
147 = 0.00621) and sperm motility (GO:0097722, p.adjust = 0.00612).

148 Notably, DEGs unique to *Kdm5c*-KO EpiLCs included key drivers of germline identity, such as *Stimulated*  
149 *by retinoic acid 8* (*Stra8*: log2 fold change = 3.73, q = 2.17e-39) and *Deleted in azoospermia like* (*Dazl*):  
150 log2 fold change = 3.36, q = 3.19e-12) (Figure 3H). These genes are typically expressed when primordial  
151 germ cells (PGCs) are committed to the germline fate and later in life to trigger meiotic gene expression  
152 programs<sup>42-44</sup>. Of note, some germline genes, including *Dazl*, are also expressed in the two-cell embryo<sup>39,45</sup>.  
153 However, we did not see derepression of two-cell stage-specific genes, like *Duxf3* (*Dux*) (log2 fold change  
154 = -0.282, q = 0.337) and *Zscan4d* (log2 fold change = 0.25, q = 0.381) (Figure 3H), indicating *Kdm5c*-KO  
155 EpiLCs do not revert back to a 2-cell state. Altogether, *Kdm5c*-KO EpiLCs express key drivers of germline  
156 identity and meiosis while the brain primarily expresses spermiogenesis genes, indicating germline gene  
157 misexpression mirrors germline development during the progression of somatic development.

158 **Female epiblast-like cells have increased sensitivity to germline gene misexpression  
159 with *Kdm5c* loss**

160 It is currently unknown if the misexpression of germline genes is influenced by sex, as previous studies  
161 on germline gene repressors have focused on male cells<sup>16,17,19,46,47</sup>. Sex is particularly pertinent in the case  
162 of KDM5C because it partially escapes X chromosome inactivation (XCI), resulting in a higher dosage in  
163 females<sup>48-51</sup>. We therefore explored the impact of chromosomal sex upon germline gene suppression by  
164 comparing their dysregulation in male *Kdm5c* hemizygous knockout (XY *Kdm5c*-KO, XY 5CKO), female

165 homozygous knockout (XX *Kdm5c*-KO, XX 5CKO), and female heterozygous knockout (XX *Kdm5c*-HET, XX  
166 5CHET) EpiLCs<sup>41</sup>.

167 In EpiLCs, homozygous and heterozygous *Kdm5c* knockout females expressed over double the number  
168 of germline-enriched genes than hemizygous males (Figure 4A). While the majority of germline DEGs in  
169 *Kdm5c*-KO males were also dysregulated in females (74%), many were sex-specific, such as *Tktl2* and *Esx1*  
170 (Figure 4B). We then compared the known functions of germline genes dysregulated only in females (XX  
171 only - unique to XX *Kdm5c*-KO, XX *Kdm5c*-HET, or both), only in males (XY only), or in all samples (shared)  
172 (Figure 4C). Female-specific germline DEGs were enriched for meiotic (GO:0051321 - meiotic cell cycle) and  
173 flagellar (GO:0003341 - cilium movement) functions, while male-specific DEGs had roles in mitochondrial  
174 and cell signaling (GO:0070585 - protein localization to mitochondrion). Germline transcripts expressed in  
175 both sexes were enriched for meiotic (GO:0140013 - meiotic nuclear division) and egg-specific functions  
176 (GO:0007292 - female gamete generation).

177 The majority of germline genes expressed in both sexes were more highly dysregulated in females  
178 compared to males (Figure 4D-F). This increased degree of dysregulation in females, along with the  
179 increased total number of germline genes, indicates females are more sensitive to losing KDM5C-mediated  
180 germline gene suppression. Female sensitivity could be due to impaired XCI in *Kdm5c* mutants<sup>41</sup>, as many  
181 spermatogenesis genes lie on the X chromosome<sup>52,53</sup>. However, female germline DEGs were not biased  
182 towards the X chromosome and had a similar overall proportion of X chromosome DEGs compared to  
183 males (XY *Kdm5c*-KO - 10.29%, XX *Kdm5c*-HET - 7.43%, XX *Kdm5c*-KO - 10.59%) (Figure 4G). The  
184 majority of germline DEGs instead lie on autosomes for both male and female *Kdm5c* mutants (Figure 4G).  
185 Thus, while female EpiLCs are more prone to germline gene misexpression with KDM5C loss, it is likely  
186 independent of XCI defects.

#### 187 **Germline gene misexpression in *Kdm5c* mutants is independent of germ cell sex**

188 Although many germline genes have shared functions in the male and female germline, some have  
189 unique or sex-biased expression. Therefore, we wondered if *Kdm5c* mutant males would primarily express  
190 sperm genes while mutant females would primarily express egg genes. To comprehensively assess whether  
191 germline gene sex corresponds with *Kdm5c* mutant sex, we first filtered our list of germline-enriched genes  
192 for egg and sperm-biased genes (Figure 4H). We defined germ cell sex-biased genes as those whose  
193 expression in the opposite sex, at any time point, is no greater than 20% of the gene's maximum expression  
194 in a given sex. This criteria yielded 67 egg-biased, 1,024 sperm-biased, and 197 unbiased germline-enriched  
195 genes. We found regardless of sex, egg, sperm, and unbiased germline genes were dysregulated in all  
196 *Kdm5c* mutants at similar proportions (Figure 4I-J). Furthermore, germline genes dysregulated exclusively  
197 in either male or female mutants were also not biased towards their corresponding germ cell sex (Figure  
198 4I). Altogether, these results demonstrate sex differences in germline gene dysregulation is not due to  
199 sex-specific activation of sperm or egg transcriptional programs.

200 **KDM5C binds to a subset of germline gene promoters during early embryogenesis**

201 KDM5C binds to the promoters of several germline genes in embryonic stem cells (ESCs) but its binding  
202 is absent in neurons<sup>13</sup>. However, the lack of a comprehensive list of germline-enriched genes prohibited  
203 genome-wide characterization of KDM5C binding at germline gene promoters. Thus, it is unclear if KDM5C  
204 is enriched at germline gene promoters, what types of germline genes KDM5C regulates, and if its binding is  
205 maintained at any germline genes in neurons.

206 To address these questions, we analyzed KDM5C chromatin immunoprecipitation followed by DNA  
207 sequencing (ChIP-seq) datasets in EpiLCs<sup>41</sup> and primary forebrain neuron cultures (PNCs)<sup>12</sup>. EpiLCs had a  
208 higher total number of high-confidence KDM5C peaks than PNCs (EpiLCs: 5,808, PNCs: 1,276, MACS2 q <  
209 0.1 and fold enrichment > 1). KDM5C was primarily localized to gene promoters in both cell types (EpiLCs:  
210 4,190, PNCs: 745 ± 500bp from the TSS), although PNCs showed increased localization to non-promoter  
211 regions (Figure 5A).

212 The majority of promoters bound by KDM5C in PNCs were also bound in EpiLCs (513 shared promoters),  
213 however a large portion of gene promoters were bound by KDM5C only in EpiLCs (3,677 EpiLC only  
214 promoters) (Figure 5B). Genes bound by KDM5C in both PNCs and EpiLCs were enriched for functions  
215 involving nucleic acid turnover, such as deoxyribonucleotide metabolic process (GO:0009262, p.adjust =  
216 8.28e-05) (Figure 5C). Germline-specific ontologies were enriched only in EpiLC-specific KDM5C-bound  
217 promoters, such as meiotic nuclear division (GO: 0007127 p.adjust = 6.77e-16) (Figure 5C). There were no  
218 ontologies significantly enriched for PNC-specific KDM5C target genes. Using our mouse germline gene list,  
219 we observed evident KDM5C signal around the TSS of many germline genes in EpiLCs, but not in PNCs  
220 (Figure 5D). Based on our ChIP-seq peak cut-off criteria, KDM5C was highly enriched at 211 germline gene  
221 promoters in EpiLCs (16.4% of all germline genes) (Figure 5E). Of note, KDM5C was only bound to about  
222 one third of *Kdm5c*-KO RNA-seq DEG promoters (EpiLC only DEGs: 34.92063%, Brain only DEGs: 30%)  
223 (Supplementary figure 1A-C). However, KDM5C did bind the promoter of 4 out of the 5 genes dysregulated  
224 in both the brain and EpiLCs. Representative examples of KDM5C-bound and unbound germline DEGs  
225 are *Dazl* and *Stra8*, respectively (Figure 5F). Together, these results demonstrate KDM5C is recruited to a  
226 subset of germline genes in EpiLCs, including meiotic genes, but does not directly regulate germline genes  
227 in neurons. Furthermore, the majority of germline mRNAs expressed in *Kdm5c*-KO cells are dysregulated  
228 independent of direct KDM5C binding to their promoters.

229 Many germline-specific genes are suppressed by the polycomb repressive complex 1.6 (PRC1.6), which  
230 contains transcription factor heterodimers E2F6/DP1 and MGA/MAX that respectively bind E2F and E-box  
231 motifs<sup>56</sup>. PRC1.6 members may recruit KDM5C to germline gene promoters, given their association with  
232 KDM5C in HeLa cells and ESCs<sup>45,57</sup>. We thus used HOMER<sup>58</sup> to identify transcription factor motifs enriched  
233 at KDM5C-bound or unbound germline gene promoters (TSS ± 500 bp, q-value < 0.1). MAX and E2F6 binding  
234 sites were significantly enriched at germline genes bound by KDM5C in EpiLCs (MAX q-value: 0.0068, E2F6  
235 q-value: 0.0673, E2F q-value: 0.0917), but not at germline genes unbound by KDM5C (Figure 5G). One third

236 of KDM5C-bound promoters contained the consensus sequence for either E2F6 (E2F, 5'-TCCCGC-3'), MGA  
237 (E-box, 5'-CACGTG-3'), or both, but only 17% of KDM5C-unbound genes contained these motifs (Figure  
238 5H). KDM5C-unbound germline genes were instead enriched for multiple RFX transcription factor binding  
239 sites (RFX q-value < 0.0001, RFX2 q-value < 0.0001, RFX5 q-value < 0.0001) (Figure 5I, Supplementary  
240 figure 1D). RFX transcription factors bind X-box motifs<sup>59</sup> to promote ciliogenesis<sup>60,61</sup> and among them is  
241 RFX2, a central regulator of post-meiotic spermatogenesis<sup>62,63</sup>. Interestingly, RFX2 mRNA is derepressed  
242 in *Kdm5c*-KO EpiLCs (Figure 5J), however it is also not a direct target of KDM5C (Supplementary figure  
243 1E). Thus, RFX2 is a candidate transcription factor for driving the ectopic expression of KDM5C-unbound  
244 germline genes in *Kdm5c*-KO cells.

245 **KDM5C is recruited to CpG islands at germline promoters to facilitate *de novo* DNA  
246 methylation**

247 Previous work found two germline gene promoters have a marked reduction in DNA CpG methylation  
248 (CpGme) in the *Kdm5c*-KO adult hippocampus<sup>13</sup>. Since histone 3 lysine 4 di- and trimethylation (H3K4me2/3)  
249 impede *de novo* CpGme<sup>64,65</sup>, KDM5C's removal of H3K4me2/3 may be required to suppress germline genes.  
250 However, KDM5C's catalytic activity was recently shown to be dispensable for suppressing *Dazl* in ESCs<sup>45</sup>.  
251 To reconcile these observations, we hypothesized KDM5C erases H3K4me3 to promote the initial placement  
252 of CpGme at germline gene promoters in EpiLCs.

253 To test this hypothesis, we first characterized KDM5C's expression as naïve ESCs differentiate into  
254 EpiLCs (Figure 6A). While *Kdm5c* mRNA steadily decreased from 0 to 48 hours of differentiation (Figure  
255 6B), KDM5C protein initially increased from 0 to 24 hours but then decreased to near knockout levels by 48  
256 hours (Figure 6C). We then characterized KDM5C's substrates (H3K4me2/3) at germline gene promoters  
257 with *Kdm5c* loss using published ChIP-seq datasets<sup>21,41</sup>. *Kdm5c*-KO samples showed a marked increase in  
258 H3K4me2 in EpiLCs (Figure 6D) and H3K4me3 in the amygdala (Figure 6E) around the TSS of germline  
259 genes. Together, these data suggest KDM5C acts during the transition between ESCs and EpiLCs to remove  
260 H3K4me2/3 at germline gene promoters.

261 Germline genes accumulate CpG methylation (CpGme) at CpG islands (CGIs) during the transition  
262 from naïve to primed pluripotency<sup>18,20,66</sup>. We first examined how many of our germline-enriched genes had  
263 promoter CGIs (TSS ± 500 bp) using the UCSC genome browser<sup>67</sup>. Notably, out of 1,288 germline-enriched  
264 genes, only 356 (27.64%) had promoter CGIs (Figure 6F). CGI-containing germline genes had substantial  
265 enrichment of meiotic gene ontologies compared to CGI-free genes, including meiotic nuclear division  
266 (GO:0140013, p.adjust = 2.17e-12) and meiosis I (GO:0007127, p.adjust = 3.91e-10) (Figure 6G). Germline  
267 genes with promoter CGIs were more highly expressed than CGI-free genes across spermatogenesis stages,  
268 with highest expression in meiotic spermatocytes (Figure 6H). Contrastingly, CGI-free genes only displaying  
269 substantial expression in post-meiotic round spermatids. Although only a minor portion of germline gene

270 promoters contained CGIs, CGIs strongly determined KDM5C's recruitment to germline genes, with 79.15%  
271 ( $p = 2.37e-67$ , Odds Ratio = 17.8, Fisher's exact test) of KDM5C-bound germline gene promoters harboring  
272 CGIs (Figure 6G).

273 To assess how KDM5C loss impacts initial CpGme placement at germline gene promoters, we performed  
274 whole genome bisulfite sequencing (WGBS) in male wild-type and *Kdm5c*-KO ESCs and 96-hour extend  
275 EpiLCs (exEpiLCs), when germline genes reach peak methylation level<sup>17</sup> (Figure 6I). We first identified  
276 which germline gene promoters significantly gained CpGme in wild-type cells during nESC to exEpiLCs  
277 differentiation (methylKit<sup>68</sup>,  $q < 0.01$ ,  $|methylation\ difference| \geq 25\%$ , TSS  $\pm 500$  bp). In wild-type cells,  
278 the majority of germline genes gained substantial CpGme at their promoter during differentiation (60.08%),  
279 regardless if their promoter contained a CGI (Figure 6J).

280 We then identified promoters differentially methylated in wild-type versus *Kdm5c*-KO exEpiLCs (methylKit,  
281  $q < 0.01$ ,  $|methylation\ difference| > 25\%$ , TSS  $\pm 500$  bp). Of the 48,882 promoters assessed, 274 promoters  
282 were significantly hypomethylated and 377 promoters were significantly hypermethylated with KDM5C  
283 loss (Supplementary figure 2A). Hypomethylated promoters were significantly enriched for germline gene  
284 ontologies, such as meiotic nuclear division (GO:0140013, p.adjust = 0.012)(Supplementary figure 2B), with  
285 10.22% of hypomethylated promoters belonging to germline genes. Approximately half of germline promoters  
286 hypomethylated in *Kdm5c*-KO exEpiLCs are direct targets of KDM5C in EpiLCs (13 out of 28 hypomethylated  
287 promoters). Promoters that showed the most robust loss of CpGme (lowest q-values) harbored CGIs (Figure  
288 6K). CGI promoters, but not CGI-free promoters, had a significant reduction in CpGme with KDM5C loss  
289 as a whole (Figure 6L) (Non-CGI promoters  $p = 0.0846$ , CGI promoters  $p = 0.0081$ , Mann-Whitney U test).  
290 Significantly hypomethylated promoters included germline genes consistently dysregulated across multiple  
291 *Kdm5c*-KO RNA-seq datasets<sup>13</sup>, such as *Naa11* and *D1PAs1* (methylation difference = -60.03%, q-value  
292 = 3.26e-153) (Figure 6M). Surprisingly, we found only a modest reduction in CpGme at *Dazl*'s promoter  
293 (methylation difference = -6.525%, q-value = 0.0159) (Figure 6N). Altogether, these results demonstrate  
294 KDM5C is recruited to germline gene CGIs to promote CpGme at germline gene promoters. This suggests  
295 KDM5C's catalytic activity is required for germline gene repression in EpiLCs, however some loci can  
296 compensate for KDM5C loss through other silencing mechanisms, even when retaining H3K4me around  
297 the TSS.

## 298 Discussion

299 In the above study, we demonstrate KDM5C's pivotal role in the development of tissue identity. We  
300 first characterized tissue-enriched genes expressed within the *Kdm5c*-KO brain and identified substantial  
301 derepression of testis, liver, muscle, and ovary-enriched genes. Testis genes significantly enriched within  
302 the *Kdm5c*-KO amygdala and hippocampus are specific to the germline and not expressed within somatic  
303 cells. *Kdm5c*-KO epiblast-like cells (EpiLCs) aberrantly express key drivers of germline identity and meiosis,

304 including *Dazl* and *Stra8*, while the adult brain primarily expresses genes important for late spermatogenesis.  
305 We demonstrated that although *Kdm5c* mutant sex did not influence whether sperm or egg-specific genes  
306 were misexpressed, female EpiLCs are more sensitive to germline gene de-repression. Germline genes  
307 can become aberrantly expressed in *Kdm5c*-KO cells via an indirect mechanism, such as activation via  
308 ectopic RFX transcription factors. Finally, we found KDM5C is dynamically regulated during ESC to EpiLC  
309 differentiation to promote long-term germline gene silencing through DNA methylation at CpG islands.  
310 Therefore, we propose KDM5C plays a fundamental role in the development of tissue identity during early  
311 embryogenesis, including the establishment of the soma-germline boundary. By systematically characterizing  
312 KDM5C's role in germline gene repression, we unveiled derepressive mechanisms governing distinct classes  
313 of germline gene in somatic lineages. Furthermore, these data provide molecular footholds which can be  
314 exploited to test the overarching contribution of ectopic germline gene expression upon neurodevelopment.

315 Although eggs and sperm employ the same transcriptional programs for shared functions, e.g. PGC  
316 formation, meiosis, and genome defense, some germline genes are sex specific. We found *Kdm5c* mutant  
317 males and females expressed both sperm and egg-biased genes, indicating the mechanism of derepression  
318 is independent of a given germline gene's sex. However, organismal sex did greatly influence the degree of  
319 germline gene dysregulation, as female *Kdm5c*-KO EpiLCs had over double the number of germline-enriched  
320 DEGs compared to males. The lack of X-linked gene enrichment in females suggests that this greater  
321 sensitivity to germline gene misexpress is not due to XCI defects previously reported in *Kdm5c*-KO females<sup>41</sup>.  
322 Intriguingly, females with heterozygous loss of *Kdm5c* also had over double the number of germline DEGs  
323 than hemizygous knockout males, even though their level of KDM5C should be roughly equivalent to that  
324 of wild-type males. Increased female sensitivity to germline gene de-repression may be related to females  
325 having a higher dose of KDM5C than males, due to its escape from XCI<sup>48–51</sup>. KDM5C's Y-chromosome  
326 homolog, KDM5D, exhibits weaker demethylase activity than KDM5C<sup>8</sup> and it is currently unknown to regulate  
327 germline gene expression. Altogether, these results suggests germline gene silencing mechanisms differ  
328 between males and females, which warrants further study to identify the biological implications and underlying  
329 mechanisms.

330 We found KDM5C is largely dispensable for promoting normal gene expression during development, yet is  
331 critical for suppressing ectopic developmental programs. It is important to note that while we highlighted  
332 KDM5C's regulation of germline genes, some germline-enriched genes are also expressed at the 2-cell  
333 stage and in naïve ESCs for their role in pluripotency and self-renewal<sup>40,45,69,70</sup>. Although expressed in  
334 naïve ESCs, “self-renewal” germline genes like *Dazl* are silenced during ESC differentiation into epiblast  
335 stem cells/EpiLCs<sup>17,18</sup>. We found that while *Kdm5c*-KO EpiLCs also express *Dazl*, they did not express  
336 2-cell-specific genes. These data suggest the 2-cell-like state reported in *Kdm5c*-KO ESCs<sup>45</sup> likely reflects  
337 KDM5C's primary role in germline gene repression. Germline gene misexpression in *Kdm5c*-KO EpiLCs may  
338 indicate they are differentiating into primordial germ cell-like cells (PGCLCs), rather than de-differentiating  
339 into 2-cell-like cells<sup>33,34,36</sup>. Yet, *Kdm5c*-KO EpiLCs had normal cellular morphology and properly expressed

340 markers for primed pluripotency, including *Otx2* which blocks EpiLC differentiation into PGCs/PGCLCs<sup>71</sup>.  
341 In addition to unimpaired EpiLC differentiation, *Kdm5c*-KO gross brain morphology is overall normal<sup>12</sup> and  
342 hardly any brain-specific genes were significantly dysregulated. Thus, ectopic germline gene expression  
343 occurs along with proper somatic development in *Kdm5c*-KO animals.

344 Our work provides novel insight into the cross-talk between H3K4me and CpGme. In EpiLCs, loss of  
345 KDM5C binding at a subset of germline gene promoters, e.g. *D1Pas1* and *Naa11*, strongly impaired CGI  
346 methylation, and resulted in their long-lasting de-repression into adulthood. Removal of H3K4me2/3 at CGIs  
347 is a plausible mechanism for KDM5C-mediated germline gene suppression<sup>13,72</sup>, given H3K4me2/3 primarily  
348 do not colocalize with CpGme<sup>73</sup> and can oppose DNMT3 activity<sup>64,65</sup>. However, emerging work indicates  
349 many histone-modifying enzymes have non-catalytic functions that influence gene expression, sometimes  
350 even more potently than their catalytic roles<sup>74,75</sup>. Indeed, KDM5C's catalytic activity was recently found to be  
351 dispensible for repressing *Dazl* in ESCs<sup>45</sup>. In our study, *Dazl*'s promoter still gained CpGme in *Kdm5c*-KO  
352 exEpiLCs, even with elevated H3K4me2. *Dazl* and a few other germline gene CGIs use multiple repressive  
353 mechanisms to facilitate CpGme<sup>16,17,46,47</sup>. Together, this suggests alternative silencing mechanisms are  
354 sufficient to recruit DNMT3s to some germline CGIs, while others may require KDM5C-mediated H3K4me  
355 removal to overcome promoter CGI escape from CpGme<sup>73,76</sup>. Furthermore, these results indicate the  
356 requirement for catalytic activity can change depending upon the locus and developmental stage, even for  
357 the same class of genes.

358 By generating a comprehensive list of mouse germline-enriched genes, we were able to reveal distinct  
359 derepressive mechanisms governing early versus late-stage germline developmental programs. Previous  
360 work on germline gene silencing has focused on genes with promoter CGIs<sup>18,73</sup>, and indeed the major-  
361 ity of KDM5C targets in EpiLCs were germ cell identity genes harboring CGIs. However, over 70% of  
362 germline-enriched gene promoters lacked CGIs, including the many KDM5C-unbound germline genes  
363 that were de-repressed in *Kdm5c*-KO cells. CGI-free, KDM5C-unbound germline genes were primarily  
364 late-stage spermatogenesis genes and significantly enriched for RFX2 binding sites, a central regulator  
365 of spermiogenesis<sup>62,63</sup>. These data suggest that once activated during early embryogenesis, drivers of  
366 germline identity like *Rfx2*, *Stra8*, and *Dazl* turn on downstream germline programs, ultimately culminating in  
367 the expression of spermiogenesis genes in the adult *Kdm5c*-KO brain. Therefore, we propose KDM5C is  
368 recruited via promoter CGIs to genes that shape germ cell formation and acts as break against runaway  
369 activation of germline-specific programs.

370 The above work provides the mechanistic foundation for KDM5C-mediated repression of tissue and  
371 germline-specific genes. However, the contribution of these ectopic, tissue-specific genes towards *Kdm5c*-  
372 KO neurological impairments is still unknown. In addition to germline genes, we also identified significant  
373 enrichment of muscle, liver, and even ovary-biased transcripts within the male *Kdm5c*-KO brain. Intriguingly,  
374 select liver and muscle-biased DEGs do have known roles within the brain, such as the liver-enriched lipid  
375 metabolism gene *Apolipoprotein C-I (Apoc1)*<sup>27</sup>. *APOC1* dysregulation is implicated in Alzheimer's disease in

376 humans<sup>28</sup> and overexpression of *Apoc1* in the mouse brain can impair learning and memory<sup>77</sup>. KDM5C may  
377 therefore be crucial for neurodevelopment by fine-tuning the expression of tissue-enriched, dosage-sensitive  
378 genes like *Apoc1*. Given germline genes have no known functions within the brain, their impact upon  
379 neurodevelopment is currently unknown. Ectopic testicular germline transcripts have been observed in a  
380 variety of cancers<sup>78,79</sup>, including brain tumors in *Drosophila* and mammals and shown to promote cancer  
381 progression<sup>80–82</sup>. Intriguingly, mouse and human models for other chromatin-linked neurodevelopmental  
382 disorders also display impaired soma-germline demarcation<sup>7,83–86</sup>, such as DNA methyltransferase 3b  
383 (DNMT3B), H3K9me1/2 methyltransferases G9A/GLP, methyl-CpG -binding protein 2 (MECP2)<sup>83</sup>. Thus,  
384 KDM5C is among a growing cohort of chromatin-linked neurodevelopmental disorders with similar erosion of  
385 the germline versus soma boundary. Further research is required to determine the impact of these germline  
386 genes and the extent to which this phenomenon occurs in humans.

## 387 Materials and Methods

### 388 Classifying tissue-enriched and germline-enriched genes

389 Tissue-enriched differentially expressed genes (DEGs) were determined by their classification in a previ-  
390 ously published dataset from 17 male and female mouse tissues<sup>22</sup>. This study defined tissue expression as  
391 greater than 1 Fragments Per Kilobase of transcript per Million mapped read (FPKM) and tissue enrichment  
392 as at least 4-fold higher expression than any other tissue.

393 We curated a list of germline-enriched genes using an RNA-seq dataset from wild-type and germline-  
394 depleted (Kit<sup>W/Wv</sup>) male and female mouse embryos from embryonic day 12, 14, and 16<sup>32</sup>, as well as adult  
395 male testes<sup>29</sup>. Germline-enriched genes met the following criteria: 1) their expression is greater than 1  
396 FPKM in wild-type germline 2) their expression in any wild-type somatic tissues<sup>22</sup> does not exceed 20%  
397 of maximum expression in wild-type germline, and 3) their expression in the germ cell-depleted (Kit<sup>W/Wv</sup>)  
398 germline, for any sex or time point, does not exceed 20% of maximum expression in wild-type germline. We  
399 defined sperm and egg-biased genes as those whose expression in the opposite sex, at any time point, is no  
400 greater than 20% of the gene's maximum expression in a given sex. Genes that did not meet this threshold  
401 for either sex were classified as 'unbiased'.

### 402 Cell culture

403 We utilized our previously established cultures of male wild-type and *Kdm5c* knockout (-KO) embryonic  
404 stem cells<sup>41</sup>. Sex was confirmed by genotyping *Uba1/Uba1y* on the X and Y chromosomes with the following  
405 primers: 5'-TGGATGGTGTGCCAATG-3', 5'-CACCTGCACGTTGCCCTT-3'. Deletion of *Kdm5c* was  
406 confirmed through the primers 5'-ATGCCCATATTAAGAGTCCTG-3', 5'-TCTGCCTTGATGGGACTGTT-3',  
407 and 5'-GGTTCTCAACACTCACATAGTG-3'.

408 Embryonic stem cells (ESCs) and epiblast-like cells were cultured using previously established  
409 methods<sup>37</sup>. Briefly, ESCs were initially cultured in primed ESC (pESC) media consisting of KnockOut  
410 DMEM (Gibco#10829–018), fetal bovine serum (Gibco#A5209501), KnockOut serum replacement  
411 (Invitrogen#10828–028), Glutamax (Gibco#35050-061), Anti-Anti (Gibco#15240-062), MEM Non-essential  
412 amino acids (Gibco#11140-050), and beta-mercaptoethanol (Sigma#M7522). They were then transitioned  
413 into ground-state, “naïve” ESCs (nESCs) by culturing for four passages in N2B27 media containing  
414 DMEM/F12 (Gibco#11330–032), Neurobasal media (Gibco#21103–049), Gluamax (Gibco#35050-061),  
415 Anti-Anti (Gibco#15240-062), N2 supplement (Invitrogen#17502048), and B27 supplement without vitamin  
416 A (Invitrogen#12587-010), and beta-mercaptoethanol. Both pESC and nESC media were supplemented  
417 with 3  $\mu$ M GSK3 inhibitor CHIR99021 (Sigma #SML1046-5MG), 1  $\mu$ M MEK inhibitor PD0325901 (Sigma  
418 #PZ0162-5MG), and 1,000 units/mL leukemia inhibitory factor (LIF, Millipore#ESG1107).

419 nESCs were differentiated into epiblast-like cells (EpiLCs, 48 hours) and extendend EpiLCs (exEpiLCs,  
420 96 hours) by culturing in N2B27 media containing DMEM/F12, Neurobasal media, Gluamax, Anti-Anti,  
421 N2 supplement, B27 supplement (Invitrogen#17504044), beta-mercaptoethanol, fibroblast growth factor 2  
422 (FGF2, R&D Biotechne 233-FB), and activin A (R&D Biotechne 338AC050CF), as previously described<sup>37</sup>.

### 423 Real time quantitative PCR (RT-qPCR)

424 nESCs were differentiated into EpiLCs as described above. Cells were lysed with Tri-reagent BD (Sigma  
425 #T3809) at 0, 24, and 48 hours of differentiation. RNA was phase separated with 0.1 uL/uL 1-bromo-3-  
426 chloropropane (Sigma #B9673) and then precipitated with isopropanol (Sigma #I9516). For each sample,  
427 2 ug of RNA was reverse transcribed using the ProtoScript II Reverse transcriptase kit from New England  
428 Biolabs (NEB #M0368S) and primed with oligo dT. Expression of *Kdm5c* was detected using the primers  
429 5'-CCCATGGAGGCCAGAGAATAAG-3' 5'-CTCAGCGATAAGAGAATTGCTAC-3' and normalized to TBP  
430 with the Power SYBR™ Green PCR Master Mix (ThermoFisher #4367659).

### 431 Western Blot

432 Total protein was extracted during nESC to EpiLC differentiation at 0, 24, and 48 hours by sonicating cells  
433 at 20% amplitude for 15 seconds in 2X SDS sample buffer, then boiling at 100 °C for 10 minutes. Proteins  
434 were separated on a 7.5% SDS page gel, transferred overnight onto a fluorescent membrane, blotted for  
435 rabbit anti-KDM5C (in house, 1:500) and mouse anti-DAXX (Santa Cruz #(H-7): sc-8043, 1:500) imaged  
436 using the LiCor Odyssey CLx system. Band intensity was quantified using ImageJ.

### 437 RNA sequencing (RNA-seq) data analysis

438 After ensuring read quality via FastQC (v0.11.8), reads were then mapped to the mm10 *Mus musculus*  
439 genome (Gencode) using STAR (v2.5.3a), during which we removed duplicates and kept only uniquely

440 mapped reads. Count files were generated by FeatureCounts (Subread v1.5.0), and BAM files were  
441 converted to bigwigs using deeptools (v3.1.3) and visualized by the UCSC genome browser. RStudio (v3.6.0)  
442 was then used to analyze counts files by DESeq2 (v1.26.0)<sup>23</sup> to identify differentially expressed genes  
443 (DEGs) with a q-value (p-adjusted via FDR/Benjamini–Hochberg correction) less than 0.1 and a log2 fold  
444 change greater than 0.5. For all DESeq2 analyses, log2 fold changes were calculated with IfcShrink using  
445 the ashr package<sup>87</sup>. MA-plots were generated by ggpahr (v0.6.0), and Eulerr diagrams were generated by  
446 eulerr (v6.1.1). Boxplots and scatterplots were generated by ggpahr (v0.6.0) and ggplot2 (v3.3.2). The Upset  
447 plot was generated via the package UpSetR (v1.4.0)<sup>88</sup>. Gene ontology (GO) analyses were performed by  
448 the R package enrichPlot (v1.16.2) using the biological processes setting and compareCluster.

#### 449 **Chromatin immunoprecipitation followed by DNA sequencing (ChIP-seq) data analysis**

450 ChIP-seq reads were aligned to mm10 using Bowtie1 (v1.1.2) allowing up to two mismatches. Only  
451 uniquely mapped reads were used for analysis. Peaks were called using MACS2 software (v2.2.9.1) using  
452 input BAM files for normalization, with filters for a q-value < 0.1 and a fold enrichment > 1. We removed  
453 “black-listed” genomic regions that often give aberrant signals. Common peak sets were obtained in R via  
454 DiffBind[estrogen@ross-innesDifferentialOestrogenReceptor2012] (v3.6.5). In the case of KDM5C ChIP-seq,  
455 *Kdm5c*-KO peaks were then subtracted from wild-type samples using bedtools (v2.25.0). Peak proximity  
456 to genome annotations was determined by ChIPSeeker<sup>89</sup> (v1.32.1). Gene ontology (GO) analyses were  
457 performed by the R package enrichPlot (v1.16.2) using the biological processes setting and compareCluster.  
458 Enriched motifs were identified using HOMER<sup>58</sup>. Average binding across the genome was visualized using  
459 deeptools (v3.1.3). Bigwigs were visualized using the UCSC genome browser.

#### 460 **CpG island (CGI) analysis**

461 Locations of CpG islands were determined through the mm10 UCSC genome browser CpG island track<sup>67</sup>,  
462 which classified CGIs as regions that have greater than 50% GC content, are larger than 200 base pairs,  
463 and have a ratio of CG dinucleotides observed over the expected amount greater than 0.6. CGI genomic  
464 coordinates were then annotated using ChIPseeker<sup>89</sup> (v1.32.1) and filtered for ones that lie within promoters  
465 of our germline-enriched genes (TSS ± 500).

#### 466 **Whole genome bisulfite sequencing (WGBS)**

467 Genomic DNA (gDNA) from naïve ESCs and extended EpiLCs was extracted using the Wizard Genomic  
468 DNA Purification Kit (Promega A1120), following the instructions for Tissue Culture Cells. gDNA from  
469 two wild-types and two *Kdm5c*-KOs of each cell type was sent to Novogene for WGBS using the Illumina  
470 NovaSeq X Plus platform and sequenced for 150bp paired-end reads (PE150). All samples had greater  
471 than 99% bisulfite conversion rates. Reads were adapter and quality trimmed with Trim Galore (v0.6.10)

472 and aligned to the mm10 genome using Bismark<sup>90</sup> (v0.22.1). Analysis of differential methylation at germline  
473 gene promoters was performed using methylKit<sup>68</sup> (v1.28.0) with a minimum coverage of 3 paired reads, a  
474 percentage cut-off of 25%, and q-value of 0.01. methylKit was also used to calculate average percentage  
475 methylation at germline gene promoters. Methylation bedgraph tracks were generated via Bismark and  
476 visualized using the UCSC genome browser.

## 477 **Data availability**

### 478 **Published datasets**

479 All published datasets are available at the Gene Expression Omnibus (GEO) <https://www.ncbi.nlm.nih.gov/geo>. Published RNA-seq datasets analyzed in this study included the male wild-type and *Kdm5c*-KO  
480 adult amygdala and hippocampus<sup>21</sup> (available at GEO: GSE127722) and male wild-type and *Kdm5c*-KO  
481 EpiLCs<sup>41</sup> (available at GEO: GSE96797).

482 Previously published ChIP-seq experiments included KDM5C in wild-type and *Kdm5c*-KO EpiLCs<sup>41</sup> (avail-  
483 able at GEO: GSE96797) and mouse primary neuron cultures (PNCs) from the cortex and hippocampus<sup>12</sup>  
484 (available at GEO: GSE61036). ChIP-seq of histone 3 lysine 4 dimethylation in male wild-type and *Kdm5c*-KO  
485 EpiLCs<sup>41</sup> is also available at GEO: GSE96797. ChIP-seq of histone 3 lysine 4 trimethylation in wild-type and  
486 *Kdm5c*-KO male amygdala<sup>21</sup> are available at GEO: GSE127817.

### 488 **Data analysis**

489 Scripts used to generate the results, tables, and figures of this study are available via the GitHub  
490 repository: XXX

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503 **Author contributions**

504 K.M.B. and S.I. conceived the study and designed the experiments. I.V. generated the ESC and exEpiLC  
505 WGBS data. K.M.B performed the data analysis and all other experiments. K.M.B and S.I. wrote and edited  
506 the manuscript.

507 **References**

- 508 1. Strahl, B.D., and Allis, C.D. (2000). The language of covalent histone modifications. *Nature* **403**,  
509 41–45. <https://doi.org/10.1038/47412>.
- 510 2. Jenuwein, T., and Allis, C.D. (2001). Translating the Histone Code. *Science* **293**, 1074–1080.  
511 <https://doi.org/10.1126/science.1063127>.
- 512 3. Lewis, E.B. (1978). A gene complex controlling segmentation in *Drosophila*. *Nature* **276**, 565–570.  
513 <https://doi.org/10.1038/276565a0>.
- 514 4. Kennison, J.A., and Tamkun, J.W. (1988). Dosage-dependent modifiers of polycomb and antennapedia  
515 mutations in *Drosophila*. *Proc. Natl. Acad. Sci. U.S.A.* **85**, 8136–8140. <https://doi.org/10.1073/pnas.8>  
5.21.8136.
- 516 5. Kassis, J.A., Kennison, J.A., and Tamkun, J.W. (2017). Polycomb and Trithorax Group Genes in  
517 *Drosophila*. *Genetics* **206**, 1699–1725. <https://doi.org/10.1534/genetics.115.185116>.
- 518 6. Gabriele, M., Lopez Tobon, A., D'Agostino, G., and Testa, G. (2018). The chromatin basis of  
519 neurodevelopmental disorders: Rethinking dysfunction along the molecular and temporal axes. *Prog  
Neuropsychopharmacol Biol Psychiatry* **84**, 306–327. <https://doi.org/10.1016/j.pnpbp.2017.12.013>.
- 520 7. Schaefer, A., Sampath, S.C., Intrator, A., Min, A., Gertler, T.S., Surmeier, D.J., Tarakhovsky, A.,  
521 and Greengard, P. (2009). Control of cognition and adaptive behavior by the GLP/G9a epigenetic  
suppressor complex. *Neuron* **64**, 678–691. <https://doi.org/10.1016/j.neuron.2009.11.019>.
- 522 8. Iwase, S., Lan, F., Bayliss, P., De La Torre-Ubieta, L., Huarte, M., Qi, H.H., Whetstine, J.R., Bonni, A.,  
Roberts, T.M., and Shi, Y. (2007). The X-Linked Mental Retardation Gene SMCX/JARID1C Defines a  
Family of Histone H3 Lysine 4 Demethylases. *Cell* **128**, 1077–1088. <https://doi.org/10.1016/j.cell.200>  
7.02.017.
- 523 9. Claes, S., Devriendt, K., Van Goethem, G., Roelen, L., Meireleire, J., Raeymaekers, P., Cassiman,  
J.J., and Fryns, J.P. (2000). Novel syndromic form of X-linked complicated spastic paraparesia. *Am J  
Med Genet* **94**, 1–4.

- 526 10. Jensen, L.R., Amende, M., Gurok, U., Moser, B., Gimmel, V., Tschach, A., Janecke, A.R., Tariverdian,  
527 G., Chelly, J., Fryns, J.P., et al. (2005). Mutations in the JARID1C gene, which is involved in  
transcriptional regulation and chromatin remodeling, cause X-linked mental retardation. *Am J Hum  
Genet* *76*, 227–236. <https://doi.org/10.1086/427563>.
- 528 11. Carmignac, V., Nambot, S., Lehalle, D., Callier, P., Moortgat, S., Benoit, V., Ghoumid, J., Delobel,  
B., Smol, T., Thuillier, C., et al. (2020). Further delineation of the female phenotype with KDM5C  
529 disease causing variants: 19 new individuals and review of the literature. *Clin Genet* *98*, 43–55.  
<https://doi.org/10.1111/cge.13755>.
- 530 12. Iwase, S., Brookes, E., Agarwal, S., Badeaux, A.I., Ito, H., Vallianatos, C.N., Tomassy, G.S., Kasza, T.,  
Lin, G., Thompson, A., et al. (2016). A Mouse Model of X-linked Intellectual Disability Associated with  
Impaired Removal of Histone Methylation. *Cell Reports* *14*, 1000–1009. [https://doi.org/10.1016/j.celr.ep.2015.12.091](https://doi.org/10.1016/j.celr<br/>ep.2015.12.091).
- 531 13. Scandaglia, M., Lopez-Atalaya, J.P., Medrano-Fernandez, A., Lopez-Cascales, M.T., Del Blanco, B.,  
Lipinski, M., Benito, E., Olivares, R., Iwase, S., Shi, Y., et al. (2017). Loss of Kdm5c Causes Spurious  
533 Transcription and Prevents the Fine-Tuning of Activity-Regulated Enhancers in Neurons. *Cell Rep* *21*,  
47–59. <https://doi.org/10.1016/j.celrep.2017.09.014>.
- 534 14. Devlin, D.K., Ganley, A.R.D., and Takeuchi, N. (2023). A pan-metazoan view of germline-soma  
distinction challenges our understanding of how the metazoan germline evolves. *Current Opinion in  
535 Systems Biology* *36*, 100486. <https://doi.org/10.1016/j.coisb.2023.100486>.
- 536 15. Lehmann, R. (2012). Germline Stem Cells: Origin and Destiny. *Cell Stem Cell* *10*, 729–739.  
537 <https://doi.org/10.1016/j.stem.2012.05.016>.
- 538 16. Endoh, M., Endo, T.A., Shinga, J., Hayashi, K., Farcas, A., Ma, K.W., Ito, S., Sharif, J., Endoh, T.,  
Onaga, N., et al. (2017). PCGF6-PRC1 suppresses premature differentiation of mouse embryonic  
539 stem cells by regulating germ cell-related genes. *eLife* *6*. <https://doi.org/10.7554/eLife.21064>.
- 540 17. Mochizuki, K., Sharif, J., Shirane, K., Uranishi, K., Bogutz, A.B., Janssen, S.M., Suzuki, A., Okuda,  
A., Koseki, H., and Lorincz, M.C. (2021). Repression of germline genes by PRC1.6 and SETDB1  
in the early embryo precedes DNA methylation-mediated silencing. *Nat Commun* *12*, 7020. <https://doi.org/10.1038/s41467-021-27345-x>.
- 541 18. Borgel, J., Guibert, S., Li, Y., Chiba, H., Schübeler, D., Sasaki, H., Forné, T., and Weber, M. (2010).  
Targets and dynamics of promoter DNA methylation during early mouse development. *Nat Genet* *42*,  
543 1093–1100. <https://doi.org/10.1038/ng.708>.
- 544 19. Velasco, G., Hubé, F., Rollin, J., Neuillet, D., Philippe, C., Bouzinba-Segard, H., Galvani, A., Viegas-  
Péquignot, E., and Francastel, C. (2010). Dnmt3b recruitment through E2F6 transcriptional repressor  
mediates germ-line gene silencing in murine somatic tissues. *Proc Natl Acad Sci U S A* *107*, 9281–  
9286. <https://doi.org/10.1073/pnas.1000473107>.

- 545
- 546 20. Hackett, J.A., Reddington, J.P., Nestor, C.E., Dunican, D.S., Branco, M.R., Reichmann, J., Reik, W., Surani, M.A., Adams, I.R., and Meehan, R.R. (2012). Promoter DNA methylation couples genome-defence mechanisms to epigenetic reprogramming in the mouse germline. *Development* *139*, 3623–3632. <https://doi.org/10.1242/dev.081661>.
- 547
- 548 21. Vallianatos, C.N., Raines, B., Porter, R.S., Bonefas, K.M., Wu, M.C., Garay, P.M., Collette, K.M., Seo, Y.A., Dou, Y., Keegan, C.E., et al. (2020). Mutually suppressive roles of KMT2A and KDM5C in behaviour, neuronal structure, and histone H3K4 methylation. *Commun Biol* *3*, 278. <https://doi.org/10.1038/s42003-020-1001-6>.
- 549
- 550 22. Li, B., Qing, T., Zhu, J., Wen, Z., Yu, Y., Fukumura, R., Zheng, Y., Gondo, Y., and Shi, L. (2017). A Comprehensive Mouse Transcriptomic BodyMap across 17 Tissues by RNA-seq. *Sci Rep* *7*, 4200. <https://doi.org/10.1038/s41598-017-04520-z>.
- 551
- 552 23. Love, M.I., Huber, W., and Anders, S. (2014). Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol* *15*, 550. <https://doi.org/10.1186/s13059-014-0550-8>.
- 553
- 554 24. Crackower, M.A., Kolas, N.K., Noguchi, J., Sarao, R., Kikuchi, K., Kaneko, H., Kobayashi, E., Kawai, Y., Kozieradzki, I., Landers, R., et al. (2003). Essential Role of Fkbp6 in Male Fertility and Homologous Chromosome Pairing in Meiosis. *Science* *300*, 1291–1295. <https://doi.org/10.1126/science.1083022>.
- 555
- 556 25. Xiol, J., Cora, E., Koglgruber, R., Chuma, S., Subramanian, S., Hosokawa, M., Reuter, M., Yang, Z., Berninger, P., Palencia, A., et al. (2012). A Role for Fkbp6 and the Chaperone Machinery in piRNA Amplification and Transposon Silencing. *Molecular Cell* *47*, 970–979. <https://doi.org/10.1016/j.molcel.2012.07.019>.
- 557
- 558 26. Cheng, S., Altmeppen, G., So, C., Welp, L.M., Penir, S., Ruhwedel, T., Menelaou, K., Harasimov, K., Stützer, A., Blayney, M., et al. (2022). Mammalian oocytes store mRNAs in a mitochondria-associated membraneless compartment. *Science* *378*, eabq4835. <https://doi.org/10.1126/science.abq4835>.
- 559
- 560 27. Rouland, A., Masson, D., Lagrost, L., Vergès, B., Gautier, T., and Bouillet, B. (2022). Role of apolipoprotein C1 in lipoprotein metabolism, atherosclerosis and diabetes: A systematic review. *Cardiovasc Diabetol* *21*, 272. <https://doi.org/10.1186/s12933-022-01703-5>.
- 561
- 562 28. Leduc, V., Jasmin-Bélanger, S., and Poirier, J. (2010). APOE and cholesterol homeostasis in Alzheimer's disease. *Trends in Molecular Medicine* *16*, 469–477. <https://doi.org/10.1016/j.molmed.2010.07.008>.
- 563
- 564 29. Mueller, J.L., Skaletsky, H., Brown, L.G., Zaghlul, S., Rock, S., Graves, T., Auger, K., Warren, W.C., Wilson, R.K., and Page, D.C. (2013). Independent specialization of the human and mouse X chromosomes for the male germ line. *Nat Genet* *45*, 1083–1087. <https://doi.org/10.1038/ng.2705>.
- 565

- 566 30. Handel, M.A., and Eppig, J.J. (1979). Sertoli Cell Differentiation in the Testes of Mice Genetically  
567 Deficient in Germ Cells. *Biology of Reproduction* *20*, 1031–1038. <https://doi.org/10.1095/biolreprod20.5.1031>.
- 568 31. Green, C.D., Ma, Q., Manske, G.L., Shami, A.N., Zheng, X., Marini, S., Moritz, L., Sultan, C.,  
569 Gurczynski, S.J., Moore, B.B., et al. (2018). A Comprehensive Roadmap of Murine Spermatogenesis  
Defined by Single-Cell RNA-Seq. *Dev Cell* *46*, 651–667.e10. <https://doi.org/10.1016/j.devcel.2018.07.025>.
- 570 32. Soh, Y.Q., Junker, J.P., Gill, M.E., Mueller, J.L., van Oudenaarden, A., and Page, D.C. (2015). A  
571 Gene Regulatory Program for Meiotic Prophase in the Fetal Ovary. *PLoS Genet* *11*, e1005531.  
<https://doi.org/10.1371/journal.pgen.1005531>.
- 572 33. Magnúsdóttir, E., and Surani, M.A. (2014). How to make a primordial germ cell. *Development* *141*,  
573 245–252. <https://doi.org/10.1242/dev.098269>.
- 574 34. Günesdogan, U., Magnúsdóttir, E., and Surani, M.A. (2014). Primordial germ cell specification: A  
575 context-dependent cellular differentiation event [corrected]. *Philos Trans R Soc Lond B Biol Sci* *369*.  
<https://doi.org/10.1098/rstb.2013.0543>.
- 576 35. Bardot, E.S., and Hadjantonakis, A.-K. (2020). Mouse gastrulation: Coordination of tissue patterning,  
specification and diversification of cell fate. *Mechanisms of Development* *163*, 103617. <https://doi.org/10.1016/j.mod.2020.103617>.
- 577 36. Hayashi, K., Ohta, H., Kurimoto, K., Aramaki, S., and Saitou, M. (2011). Reconstitution of the  
578 mouse germ cell specification pathway in culture by pluripotent stem cells. *Cell* *146*, 519–532.  
<https://doi.org/10.1016/j.cell.2011.06.052>.
- 579 37. Samanta, M., and Kalantry, S. (2020). Generating primed pluripotent epiblast stem cells: A methodol-  
580 ogy chapter. *Curr Top Dev Biol* *138*, 139–174. <https://doi.org/10.1016/bs.ctdb.2020.01.005>.
- 581 38. Welling, M., Chen, H., Muñoz, J., Musheev, M.U., Kester, L., Junker, J.P., Mischerikow, N., Arbab, M.,  
582 Kuijk, E., Silberstein, L., et al. (2015). DAZL regulates Tet1 translation in murine embryonic stem cells.  
583 *EMBO Reports* *16*, 791–802. <https://doi.org/10.15252/embr.201540538>.
- 584 39. Macfarlan, T.S., Gifford, W.D., Driscoll, S., Lettieri, K., Rowe, H.M., Bonanomi, D., Firth, A., Singer, O.,  
Trono, D., and Pfaff, S.L. (2012). Embryonic stem cell potency fluctuates with endogenous retrovirus  
585 activity. *Nature* *487*, 57–63. <https://doi.org/10.1038/nature11244>.
- 586 40. Suzuki, A., Hirasaki, M., Hishida, T., Wu, J., Okamura, D., Ueda, A., Nishimoto, M., Nakachi, Y.,  
587 Mizuno, Y., Okazaki, Y., et al. (2016). Loss of MAX results in meiotic entry in mouse embryonic and  
germline stem cells. *Nat Commun* *7*, 11056. <https://doi.org/10.1038/ncomms11056>.

- 588 41. Samanta, M.K., Gayen, S., Harris, C., Maclary, E., Murata-Nakamura, Y., Malcore, R.M., Porter, R.S.,  
Garay, P.M., Vallianatos, C.N., Samollow, P.B., et al. (2022). Activation of Xist by an evolutionarily  
conserved function of KDM5C demethylase. *Nat Commun* *13*, 2602. <https://doi.org/10.1038/s41467-022-30352-1>.
- 589 42. Koubova, J., Menke, D.B., Zhou, Q., Capel, B., Griswold, M.D., and Page, D.C. (2006). Retinoic  
acid regulates sex-specific timing of meiotic initiation in mice. *Proc. Natl. Acad. Sci. U.S.A.* *103*,  
591 2474–2479. <https://doi.org/10.1073/pnas.0510813103>.
- 592 43. Lin, Y., Gill, M.E., Koubova, J., and Page, D.C. (2008). Germ Cell-Intrinsic and -Extrinsic Factors  
Govern Meiotic Initiation in Mouse Embryos. *Science* *322*, 1685–1687. <https://doi.org/10.1126/science.1166340>.
- 594 44. Endo, T., Mikedis, M.M., Nicholls, P.K., Page, D.C., and De Rooij, D.G. (2019). Retinoic Acid and Germ  
595 Cell Development in the Ovary and Testis. *Biomolecules* *9*, 775. <https://doi.org/10.3390/biom9120775>.
- 596 45. Gupta, N., Yakhou, L., Albert, J.R., Azogui, A., Ferry, L., Kirsh, O., Miura, F., Battault, S., Yamaguchi,  
K., Laisné, M., et al. (2023). A genome-wide screen reveals new regulators of the 2-cell-like cell state.  
597 *Nat Struct Mol Biol*. <https://doi.org/10.1038/s41594-023-01038-z>.
- 598 46. Pohlers, M., Truss, M., Frede, U., Scholz, A., Strehle, M., Kuban, R.-J., Hoffmann, B., Morkel, M.,  
Birchmeier, C., and Hagemeier, C. (2005). A Role for E2F6 in the Restriction of Male-Germ-Cell-  
599 Specific Gene Expression. *Current Biology* *15*, 1051–1057. <https://doi.org/10.1016/j.cub.2005.04.060>.
- 600 47. Dahlet, T., Truss, M., Frede, U., Al Adhami, H., Bardet, A.F., Dumas, M., Vallet, J., Chicher, J.,  
Hammann, P., Kotnik, S., et al. (2021). E2F6 initiates stable epigenetic silencing of germline genes  
601 during embryonic development. *Nat Commun* *12*, 3582. <https://doi.org/10.1038/s41467-021-23596-w>.
- 602 48. Agulnik, A.I., Mitchell, M.J., Mattei, M.G., Borsani, G., Avner, P.A., Lerner, J.L., and Bishop, C.E.  
(1994). A novel X gene with a widely transcribed Y-linked homologue escapes X-inactivation in mouse  
603 and human. *Hum Mol Genet* *3*, 879–884. <https://doi.org/10.1093/hmg/3.6.879>.
- 604 49. Carrel, L., Hunt, P.A., and Willard, H.F. (1996). Tissue and lineage-specific variation in inactive  
X chromosome expression of the murine Smcx gene. *Hum Mol Genet* *5*, 1361–1366. <https://doi.org/10.1093/hmg/5.9.1361>.
- 606 50. Sheardown, S., Norris, D., Fisher, A., and Brockdorff, N. (1996). The mouse Smcx gene exhibits  
developmental and tissue specific variation in degree of escape from X inactivation. *Hum Mol Genet*  
607 *5*, 1355–1360. <https://doi.org/10.1093/hmg/5.9.1355>.
- 608 51. Xu, J., Deng, X., and Disteche, C.M. (2008). Sex-Specific Expression of the X-Linked Histone  
Demethylase Gene Jarid1c in Brain. *PLoS ONE* *3*, e2553. <https://doi.org/10.1371/journal.pone.0002553>.
- 609 52. Wang, P.J., McCarrey, J.R., Yang, F., and Page, D.C. (2001). An abundance of X-linked genes  
expressed in spermatogonia. *Nat Genet* *27*, 422–426. <https://doi.org/10.1038/86927>.

- 611
- 612 53. Khil, P.P., Smirnova, N.A., Romanienko, P.J., and Camerini-Otero, R.D. (2004). The mouse X  
chromosome is enriched for sex-biased genes not subject to selection by meiotic sex chromosome  
inactivation. *Nat Genet* 36, 642–646. <https://doi.org/10.1038/ng1368>.
- 613
- 614 54. Hurlin, P.J. (1999). Mga, a dual-specificity transcription factor that interacts with Max and contains a  
T-domain DNA-binding motif. *The EMBO Journal* 18, 7019–7028. <https://doi.org/10.1093/emboj/18.2.4.7019>.
- 615
- 616 55. Tatsumi, D., Hayashi, Y., Endo, M., Kobayashi, H., Yoshioka, T., Kiso, K., Kanno, S., Nakai, Y., Maeda,  
I., Mochizuki, K., et al. (2018). DNMTs and SETDB1 function as co-repressors in MAX-mediated  
repression of germ cell-related genes in mouse embryonic stem cells. *PLoS ONE* 13, e0205969.  
<https://doi.org/10.1371/journal.pone.0205969>.
- 617
- 618 56. Stielow, B., Finkernagel, F., Stiewe, T., Nist, A., and Suske, G. (2018). MGA, L3MBTL2 and E2F6  
determine genomic binding of the non-canonical Polycomb repressive complex PRC1.6. *PLoS Genet*  
14, e1007193. <https://doi.org/10.1371/journal.pgen.1007193>.
- 619
- 620 57. Tahiliani, M., Mei, P., Fang, R., Leonor, T., Rutenberg, M., Shimizu, F., Li, J., Rao, A., and Shi, Y.  
(2007). The histone H3K4 demethylase SMCX links REST target genes to X-linked mental retardation.  
*Nature* 447, 601–605. <https://doi.org/10.1038/nature05823>.
- 621
- 622 58. Heinz, S., Benner, C., Spann, N., Bertolino, E., Lin, Y.C., Laslo, P., Cheng, J.X., Murre, C., Singh, H.,  
and Glass, C.K. (2010). Simple Combinations of Lineage-Determining Transcription Factors Prime  
cis-Regulatory Elements Required for Macrophage and B Cell Identities. *Molecular Cell* 38, 576–589.  
<https://doi.org/10.1016/j.molcel.2010.05.004>.
- 623
- 624 59. Gajiwala, K.S., Chen, H., Cornille, F., Roques, B.P., Reith, W., Mach, B., and Burley, S.K. (2000).  
Structure of the winged-helix protein hRFX1 reveals a new mode of DNA binding. *Nature* 403,  
916–921. <https://doi.org/10.1038/35002634>.
- 625
- 626 60. Swoboda, P., Adler, H.T., and Thomas, J.H. (2000). The RFX-Type Transcription Factor DAF-19  
Regulates Sensory Neuron Cilium Formation in *C. elegans*. *Molecular Cell* 5, 411–421. [https://doi.org/10.1016/S1097-2765\(00\)80436-0](https://doi.org/10.1016/S1097-2765(00)80436-0).
- 627
- 628 61. Ashique, A.M., Choe, Y., Karlen, M., May, S.R., Phamluong, K., Solloway, M.J., Ericson, J., and  
Peterson, A.S. (2009). The Rfx4 Transcription Factor Modulates Shh Signaling by Regional Control of  
Ciliogenesis. *Sci. Signal.* 2. <https://doi.org/10.1126/scisignal.2000602>.
- 629
- 630 62. Kistler, W.S., Baas, D., Lemeille, S., Paschaki, M., Seguin-Estevez, Q., Barras, E., Ma, W., Duteyrat, J.-  
L., Morlé, L., Durand, B., et al. (2015). RFX2 Is a Major Transcriptional Regulator of Spermiogenesis.  
*PLoS Genet* 11, e1005368. <https://doi.org/10.1371/journal.pgen.1005368>.
- 631

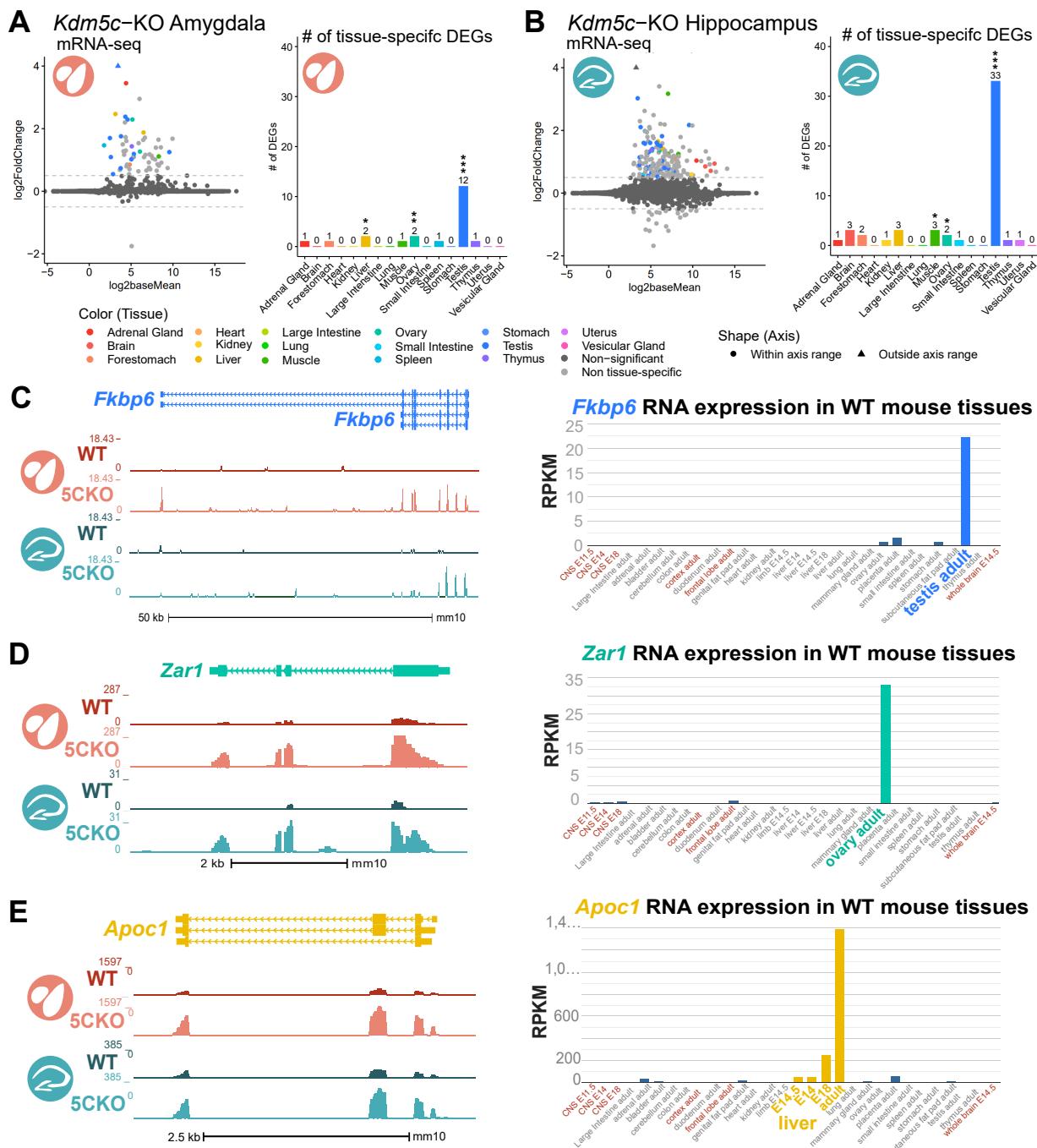
- 632 63. Wu, Y., Hu, X., Li, Z., Wang, M., Li, S., Wang, X., Lin, X., Liao, S., Zhang, Z., Feng, X., et al.  
(2016). Transcription Factor RFX2 Is a Key Regulator of Mouse Spermiogenesis. *Sci Rep* 6, 20435.  
633 <https://doi.org/10.1038/srep20435>.
- 634 64. Otani, J., Nankumo, T., Arita, K., Inamoto, S., Ariyoshi, M., and Shirakawa, M. (2009). Structural basis  
for recognition of H3K4 methylation status by the DNA methyltransferase 3A ATRX–DNMT3–DNMT3L  
635 domain. *EMBO Reports* 10, 1235–1241. <https://doi.org/10.1038/embor.2009.218>.
- 636 65. Guo, X., Wang, L., Li, J., Ding, Z., Xiao, J., Yin, X., He, S., Shi, P., Dong, L., Li, G., et al. (2015).  
Structural insight into autoinhibition and histone H3-induced activation of DNMT3A. *Nature* 517,  
637 640–644. <https://doi.org/10.1038/nature13899>.
- 638 66. Meissner, A., Mikkelsen, T.S., Gu, H., Wernig, M., Hanna, J., Sivachenko, A., Zhang, X., Bernstein,  
B.E., Nusbaum, C., Jaffe, D.B., et al. (2008). Genome-scale DNA methylation maps of pluripotent and  
639 differentiated cells. *Nature* 454, 766–770. <https://doi.org/10.1038/nature07107>.
- 640 67. Nassar, L.R., Barber, G.P., Benet-Pagès, A., Casper, J., Clawson, H., Diekhans, M., Fischer, C.,  
Gonzalez, J.N., Hinrichs, A.S., Lee, B.T., et al. (2023). The UCSC Genome Browser database: 2023  
641 update. *Nucleic Acids Research* 51, D1188–D1195. <https://doi.org/10.1093/nar/gkac1072>.
- 642 68. Akalin, A., Kormaksson, M., Li, S., Garrett-Bakelman, F.E., Figueiroa, M.E., Melnick, A., and Mason,  
C.E. (2012). methylKit: A comprehensive R package for the analysis of genome-wide DNA methylation  
643 profiles. *Genome Biol* 13, R87. <https://doi.org/10.1186/gb-2012-13-10-r87>.
- 644 69. Torres-Padilla, M.-E. (2020). On transposons and totipotency. *Philos Trans R Soc Lond B Biol Sci*  
645 375, 20190339. <https://doi.org/10.1098/rstb.2019.0339>.
- 646 70. Yang, M., Yu, H., Yu, X., Liang, S., Hu, Y., Luo, Y., Izsvák, Z., Sun, C., and Wang, J. (2022). Chemical-  
induced chromatin remodeling reprograms mouse ESCs to totipotent-like stem cells. *Cell Stem Cell*  
647 29, 400–418.e13. <https://doi.org/10.1016/j.stem.2022.01.010>.
- 648 71. Zhang, J., Zhang, M., Acampora, D., Vojtek, M., Yuan, D., Simeone, A., and Chambers, I. (2018).  
OTX2 restricts entry to the mouse germline. *Nature* 562, 595–599. [018-0581-5](https://doi.org/10.1038/s41586-<br/>649 018-0581-5).
- 650 72. Al Adhami, H., Vallet, J., Schaal, C., Schumacher, P., Bardet, A.F., Dumas, M., Chicher, J., Hammann,  
P., Daujat, S., and Weber, M. (2023). Systematic identification of factors involved in the silencing  
of germline genes in mouse embryonic stem cells. *Nucleic Acids Research* 51, 3130–3149. <https://doi.org/10.1093/nar/gkad071>.
- 652 73. Weber, M., Hellmann, I., Stadler, M.B., Ramos, L., Pääbo, S., Rebhan, M., and Schübeler, D. (2007).  
Distribution, silencing potential and evolutionary impact of promoter DNA methylation in the human  
653 genome. *Nat Genet* 39, 457–466. <https://doi.org/10.1038/ng1990>.

- 654 74. Aubert, Y., Egolf, S., and Capell, B.C. (2019). The Unexpected Noncatalytic Roles of Histone Modifiers  
in Development and Disease. *Trends in Genetics* 35, 645–657. <https://doi.org/10.1016/j.tig.2019.06.004>.
- 655 75. Morgan, M.A.J., and Shilatifard, A. (2020). Reevaluating the roles of histone-modifying enzymes  
and their associated chromatin modifications in transcriptional regulation. *Nat Genet* 52, 1271–1281.  
<https://doi.org/10.1038/s41588-020-00736-4>.
- 656 76. Long, H.K., King, H.W., Patient, R.K., Odom, D.T., and Klose, R.J. (2016). Protection of CpG  
islands from DNA methylation is DNA-encoded and evolutionarily conserved. *Nucleic Acids Res* 44,  
659 6693–6706. <https://doi.org/10.1093/nar/gkw258>.
- 660 77. Abildayeva, K., Berbée, J.F.P., Blokland, A., Jansen, P.J., Hoek, F.J., Meijer, O., Lütjohann, D., Gautier,  
T., Pillot, T., De Vente, J., et al. (2008). Human apolipoprotein C-I expression in mice impairs learning  
and memory functions. *Journal of Lipid Research* 49, 856–869. <https://doi.org/10.1194/jlr.M700518JLR200>.
- 661 78. Nielsen, A.Y., and Gjerstorff, M.F. (2016). Ectopic Expression of Testis Germ Cell Proteins in Cancer  
663 and Its Potential Role in Genomic Instability. *Int J Mol Sci* 17. <https://doi.org/10.3390/ijms17060890>.
- 664 79. Adebayo Babatunde, K., Najafi, A., Salehipour, P., Modarressi, M.H., and Mobasher, M.B. (2017).  
Cancer/Testis genes in relation to sperm biology and function. *Iranian Journal of Basic Medical  
665 Sciences* 20. <https://doi.org/10.22038/ijbms.2017.9259>.
- 666 80. Janic, A., Mendizabal, L., Llamazares, S., Rossell, D., and Gonzalez, C. (2010). Ectopic expression  
of germline genes drives malignant brain tumor growth in *Drosophila*. *Science* 330, 1824–1827.  
667 <https://doi.org/10.1126/science.1195481>.
- 668 81. Ghafouri-Fard, S., and Modarressi, M.-H. (2012). Expression of Cancer–Testis Genes in Brain Tumors:  
669 Implications for Cancer Immunotherapy. *Immunotherapy* 4, 59–75. <https://doi.org/10.2217/imt.11.145>.
- 670 82. Nin, D.S., and Deng, L.-W. (2023). Biology of Cancer-Testis Antigens and Their Therapeutic Implica-  
671 tions in Cancer. *Cells* 12, 926. <https://doi.org/10.3390/cells12060926>.
- 672 83. Bonefas, K.M., and Iwase, S. (2021). Soma-to-germline transformation in chromatin-linked neurode-  
673 velopmental disorders? *FEBS J.* <https://doi.org/10.1111/febs.16196>.
- 674 84. Velasco, G., Walton, E.L., Sterlin, D., Héduouin, S., Nitta, H., Ito, Y., Fouyssac, F., Mégarbané, A.,  
Sasaki, H., Picard, C., et al. (2014). Germline genes hypomethylation and expression define a  
molecular signature in peripheral blood of ICF patients: Implications for diagnosis and etiology.  
675 *Orphanet J Rare Dis* 9, 56. <https://doi.org/10.1186/1750-1172-9-56>.
- 676 85. Walton, E.L., Francastel, C., and Velasco, G. (2014). Dnmt3b Prefers Germ Line Genes and Cen-  
677 tromeric Regions: Lessons from the ICF Syndrome and Cancer and Implications for Diseases. *Biology  
(Basel)* 3, 578–605. <https://doi.org/10.3390/biology3030578>.

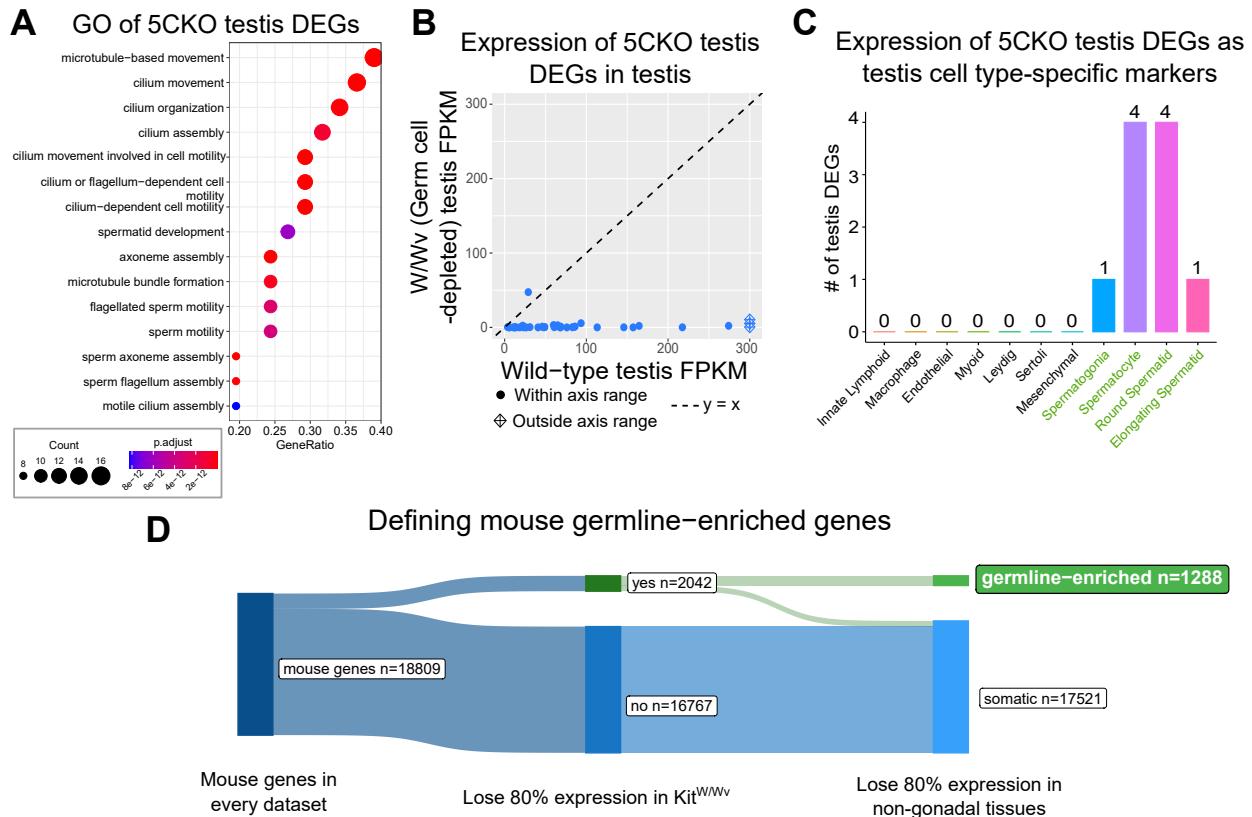
- 678 86. Samaco, R.C., Mandel-Brehm, C., McGraw, C.M., Shaw, C.A., McGill, B.E., and Zoghbi, H.Y. (2012).  
Crh and Oprm1 mediate anxiety-related behavior and social approach in a mouse model of MECP2  
679 duplication syndrome. *Nat Genet* 44, 206–211. <https://doi.org/10.1038/ng.1066>.
- 680 87. Stephens, M. (2016). False discovery rates: A new deal. *Biostat*, kxw041. <https://doi.org/10.1093/biostatistics/kxw041>.
- 682 88. Conway, J.R., Lex, A., and Gehlenborg, N. (2017). UpSetR: An R package for the visualization of  
intersecting sets and their properties. *Bioinformatics* 33, 2938–2940. <https://doi.org/10.1093/bioinformatics/btx364>.
- 684 89. Yu, G., Wang, L.G., and He, Q.Y. (2015). ChIPseeker: An R/Bioconductor package for ChIP peak  
annotation, comparison and visualization. *Bioinformatics* 31, 2382–2383. <https://doi.org/10.1093/bioinformatics/btv145>.
- 686 90. Krueger, F., and Andrews, S.R. (2011). Bismark: A flexible aligner and methylation caller for Bisulfite-  
687 Seq applications. *Bioinformatics* 27, 1571–1572. <https://doi.org/10.1093/bioinformatics/btr167>.

688 **Figures and Tables**

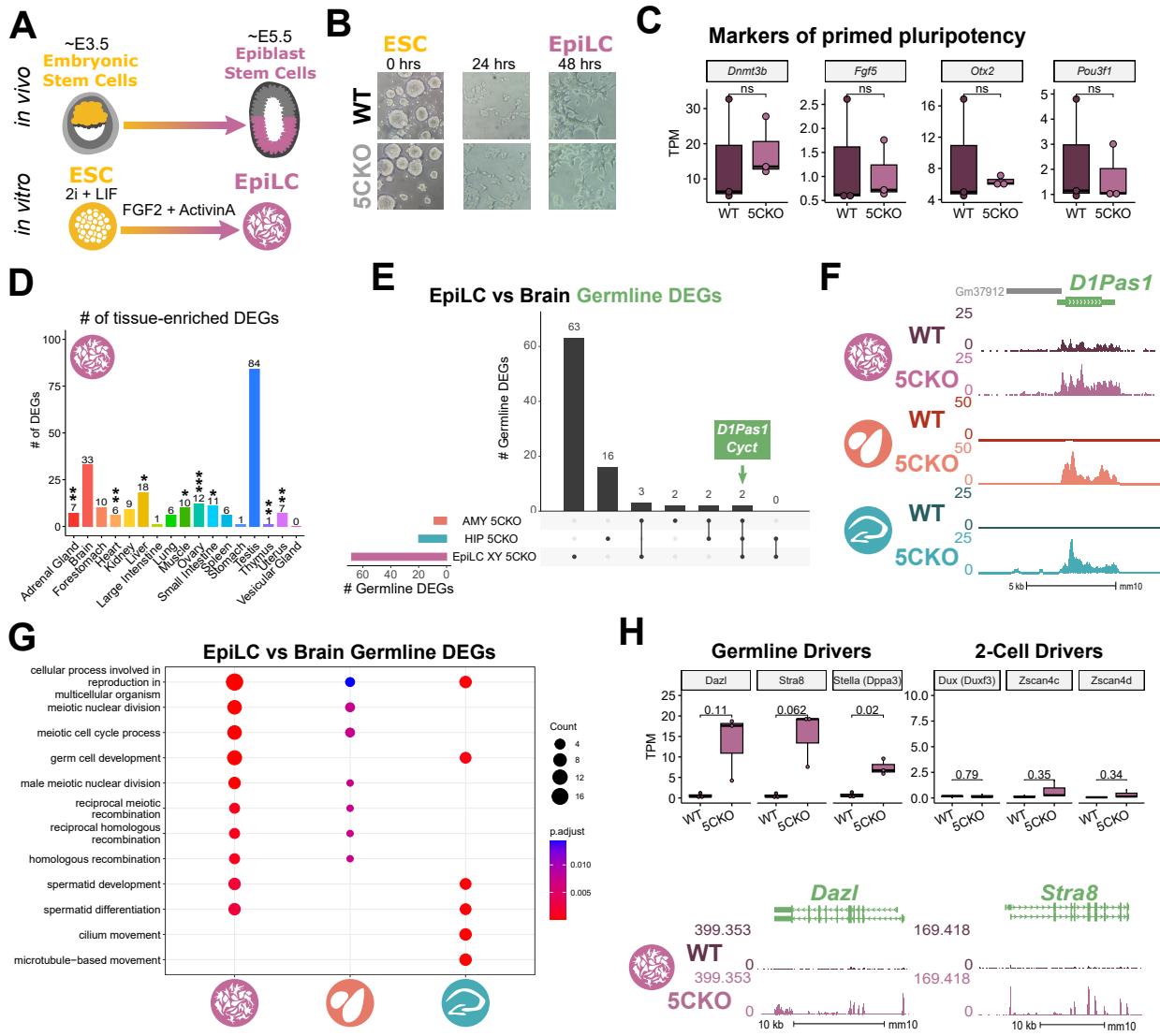
- 689     • Supplementary table 1: list of all germline genes.
- 690       – Columns to include:
- 691           \* KDM5C bound vs not
- 692           \* Log2fc in EpiLC, brain (separate columns?)
- 693       – CGI vs non



**Figure 1: Tissue-enriched genes are misexpressed in the *Kdm5c*-KO brain.** **A-B.** Expression of tissue-enriched genes (Li et al 2017) in the male *Kdm5c*-KO amygdala (A) and hippocampus (B). Left - MA plot of mRNA-sequencing. Right - Number of tissue-enriched differentially expressed genes (DEGs). \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , Fisher's exact test. **C.** Left - UCSC browser view of an example aberrantly expressed testis-enriched DEG, *FK506 binding protein 6* (*Fkbp6*) in the wild-type (WT) and *Kdm5c*-KO (5CKO) amygdala (red) and hippocampus (teal) (Average,  $n = 4$ ). Right - Expression of *Cyct* in wild-type tissues from NCBI Gene, with testis highlighted in blue and brain tissues highlighted in red. **D.** Left - UCSC browser view of an example ovary-enriched DEG, *Zygotic arrest 1* (*Zar1*). Right - Expression of *Zar1* in wild-type tissues from NCBI Gene, with ovary highlighted in teal and brain tissues highlighted in red. **E.** Left - UCSC browser view of an example liver-enriched DEG, *Apolipoprotein C-I* (*Apoc1*). Right - Expression of *Apoc1* in wild-type tissues from NCBI Gene, with liver highlighted in orange and brain tissues highlighted in red.



**Figure 2: Aberrant transcription of germline genes in the *Kdm5c*-KO in the brain.** **A.** enrichPlot gene ontology (GO) of *Kdm5c*-KO amygdala and hippocampus testis-enriched DEGs **B.** Expression of testis DEGs in wild-type (WT) testis versus germ cell-depleted (W/Wv) testis (Mueller et al 2013). Expression is in Fragments Per Kilobase of transcript per Million mapped reads (FPKM). **C.** Number of testis DEGs that were classified as cell-type specific markers in a single cell RNA-seq dataset of the testis (Green et al 2018). Germline cell types are highlighted in green, somatic cell types in black. **D.** Sankey diagram of mouse genes filtered for germline enrichment based on their expression in wild-type and W/Wv mice and in adult mouse non-gonadal tissues (Li et al 2017).



**Figure 3: Kdm5c-KO epiblast-like cells express key drivers of germline identity**

**A.** Top - Diagram of *in vivo* differentiation of embryonic stem cells (ESCs) of the inner cell mass into epiblast stem cells. Bottom - *in vitro* differentiation of ESCs into epiblast-like cells (EpiLCs).

**B.** Representative images of male wild-type (WT) and *Kdm5c*-KO (5CKO) cells during ESC to EpiLC differentiation. Brightfield images taken at 20X.

**C.** No significant difference in primed pluripotency marker expression in wild-type versus *Kdm5c*-KO EpiLCs. Welch's t-test, expression in transcripts per million (TPM).

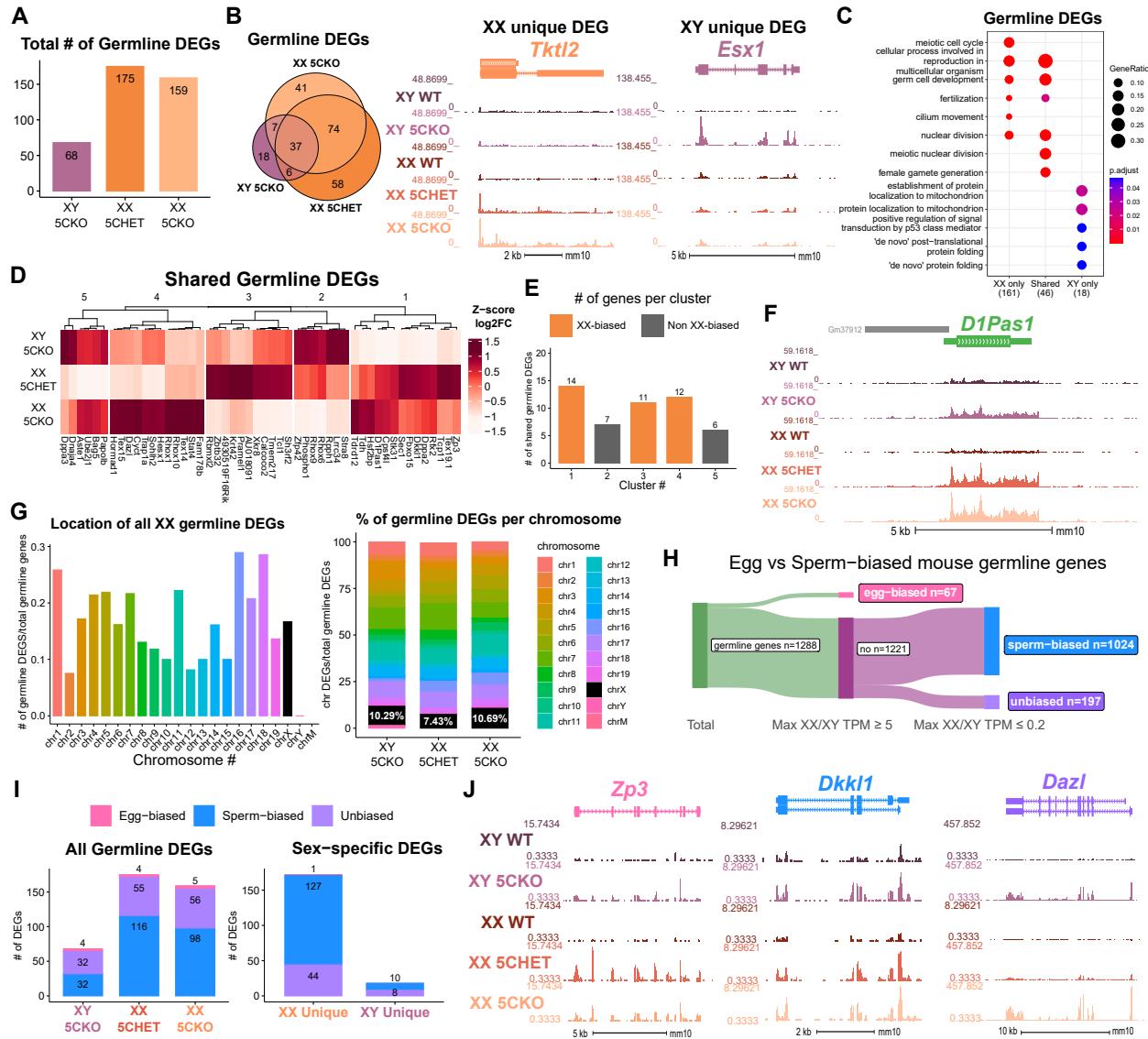
**D.** Number of tissue-enriched differentially expressed genes (DEGs) in *Kdm5c*-KO EpiLCs, amygdala (AMY), and hippocampus (HIP) RNA-seq datasets. \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , Fisher's exact test.

**E.** Upset plot displaying the overlap of germline DEGs expressed in *Kdm5c*-KO EpiLCs, amygdala (AMY), and hippocampus (HIP) RNA-seq datasets.

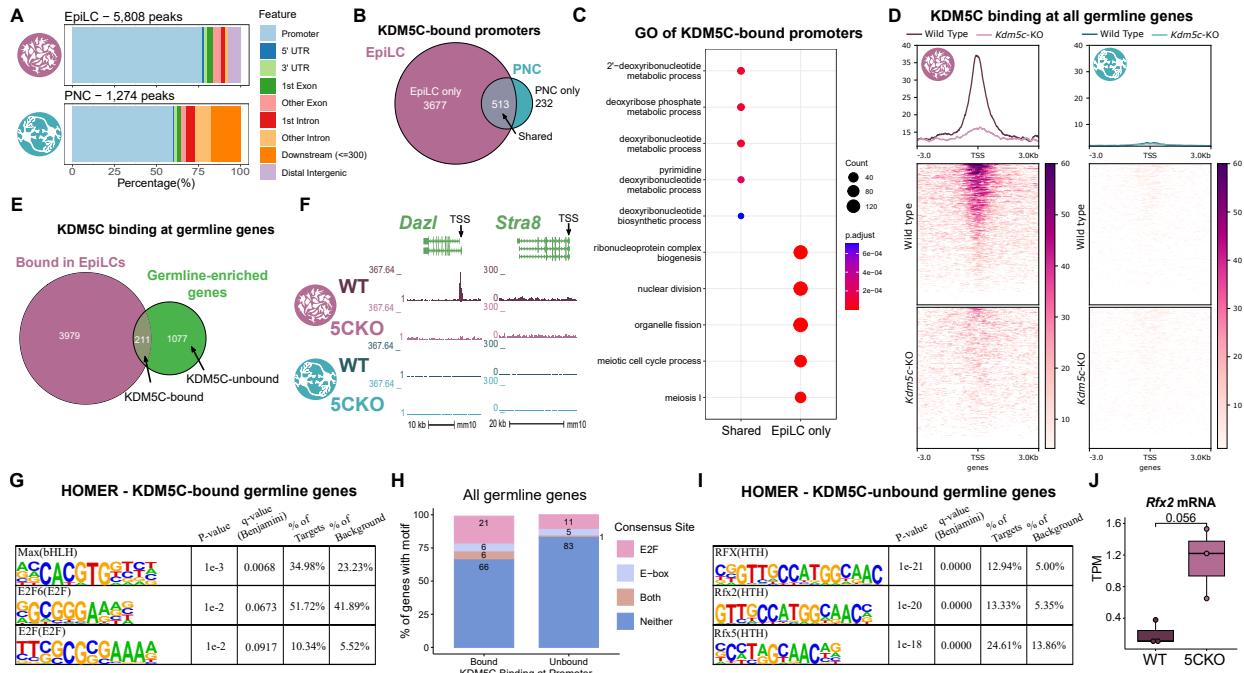
**F.** UCSC browser view of an example germline gene, *D1Pas1*, that is dysregulated *Kdm5c*-KO EpiLCs (top, purple. Average,  $n = 3$ ), amygdala (middle, red. Average,  $n = 4$ ), and hippocampus (bottom, blue. Average,  $n = 4$ ).

**G.** enrichPlot gene ontology analysis comparing enriched biological processes for *Kdm5c*-KO EpiLC, amygdala, and hippocampus germline DEGs.

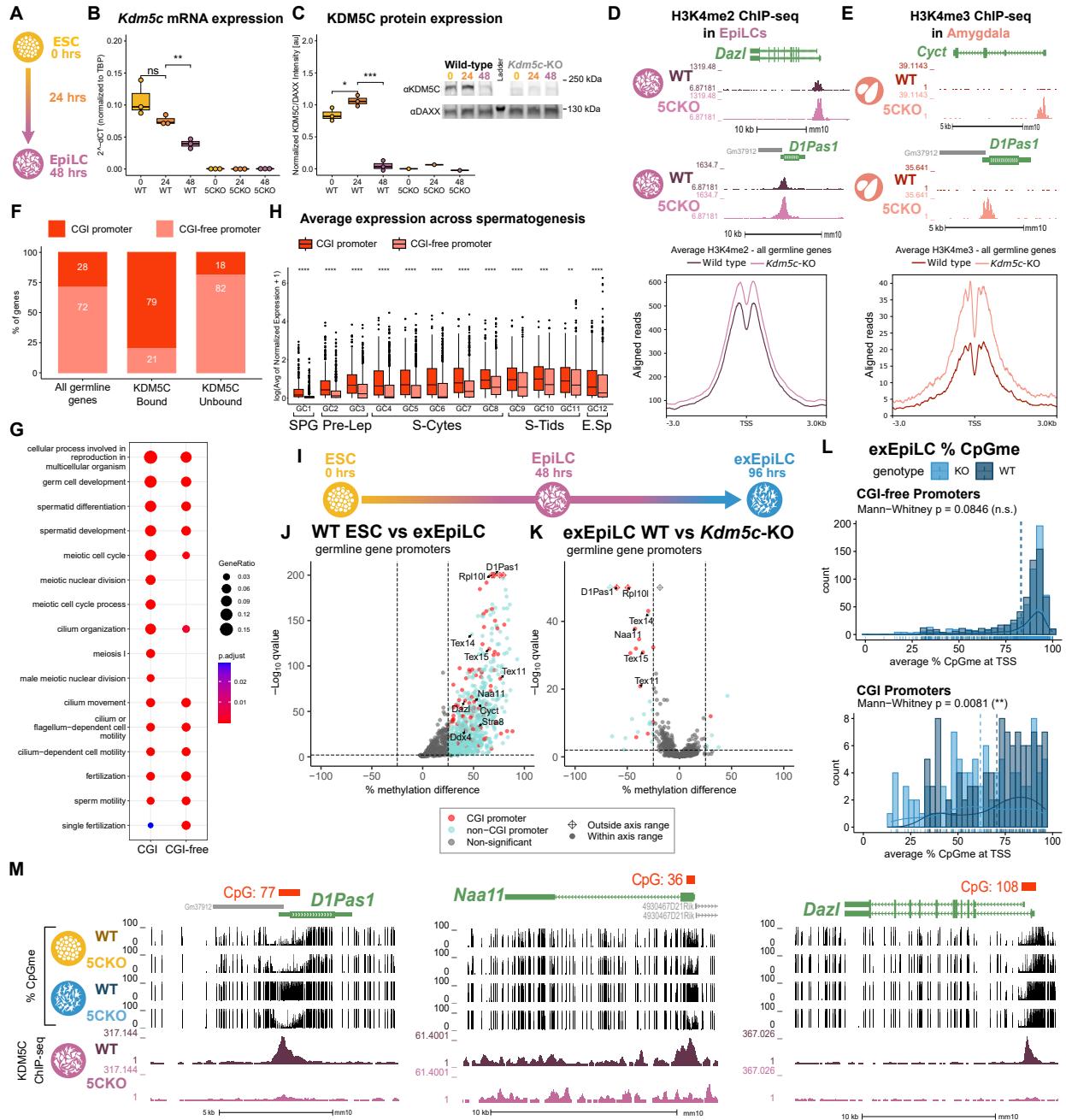
**H.** Top left - Example germline identity DEGs unique to EpiLCs. Top right - Example 2-cell genes that are not dysregulated in *Kdm5c*-KO EpiLCs. p-values for Welch's t-test. Bottom - UCSC browser view of *Dazl* and *Stra8* expression in wild-type and *Kdm5c*-KO EpiLCs (Average,  $n = 3$ ).



**Figure 4: Chromosomal sex influences *Kdm5c*-KO germline gene misexpression.** **A.** Total number of germline-enriched RNA-seq DEGs for male hemizygous *Kdm5c* knockout EpilCs (XY 5CKO, purple), female heterozygous *Kdm5c* knockout (XX 5CHET, orange), female homozygous *Kdm5c* knockout (XX 5CKO, light orange) EpilCs. **B.** Left - Eulerr overlap of *Kdm5c* mutant male and female EpilC germline DEGs. Right - Example of germline DEGs unique to females or males, *Tktl2* and *Esx1*. **C.** enrichPlot gene ontology analysis comparing enriched biological processes for germline DEGs shared between *Kdm5c* mutant males and females, or unique to one sex (XX only or XY only). **D.** Heatmap of germline DEGs shared between male and female mutants. Color is the average log 2 fold change from sex-matched wild-type. **E.** Number of genes within each cluster from D. Clusters with higher expression in females compared to males (XX-biased) highlighted in orange. **F.** UCSC browser view of a male and female shared germline DEG *D1Pas1* that is more highly expressed in female mutants (Average, n = 3). **G.** Left - Number of all female germline DEGs located on each chromosome over the total number of germline-enriched genes on that chromosome. Right - Percentage of germline DEGs that lie on each chromosome for each *Kdm5c* mutant. X chromosome highlighted in black. **H.** Sankey diagram classifying egg-biased (pink) and sperm-biased (blue) and unbiased (purple) mouse germline-enriched genes. **I.** Number of egg, sperm, or unbiased germline DEGs for male and female *Kdm5c* mutants. **J.** Example bigwigs of egg-biased (*Zp3*), sperm-biased (*Dkk1*), and unbiased (*Dazl*) germline genes dysregulated in both male and female *Kdm5c* mutants.



**Figure 5: KDM5C binds to a subset of germline gene promoters during early embryogenesis.** **A.** ChIPseeker localization of KDM5C peaks at different genomic regions in EpiLCs (top) and hippocampal and cortex primary neuron cultures (PNCs, bottom). **B.** Overlap of genes with KDM5C bound to their promoters ( $TSS \pm 500$ ) in EpiLCs (purple) and PNCs (blue). **C.** Gene ontology (GO) comparison of genes with KDM5C bound to their promoter in EpiLCs and PNCs. Genes were classified as either bound in EpiLCs only (EpiLC only), unique to PNCs (PNC only, no significant ontologies) or bound in both PNCs and EpiLCs (Shared). **D.** Average KDM5C binding around the transcription start site (TSS) of all germline-enriched genes in EpiLCs (left) and PNCs (right). **E.** Eulerr overlap of germline-enriched genes (green) with significant KDM5C binding at their promoter in EpiLCs (purple). **F.** Example KDM5C ChIP-seq signal around the *Dazl* TSS but not *Stra8* in EpiLCs. **G.** HOMER motif analysis of all KDM5C-bound germline gene promoters, highlighting significant enrichment of MAX, E2F6, and E2F motifs. **H.** Number of all gene promoters bound or unbound by KDM5C with instances of the E2F or E-box consensus sequence. **I.** HOMER motif analysis of all KDM5C-unbound germline gene promoters, highlighting significant enrichment of RFX family transcription factor motifs. **J.** Expression of RNA-seq DEG *Rfx2* in wild-type and *Kdm5c*-KO EpiLCs. P-value of Welch's t-test, expression in transcripts per million (TPM).



**Figure 6: KDM5C promotes long-term silencing of germline genes via DNA methylation at CpG islands.** **A.** Diagram of embryonic stem cell (ESC) to epiblast-like cell (EpiLC) differentiation and collection time points. **B.** Real time quantitative PCR (RT-qPCR) of *Kdm5c* mRNA expression in wild-type (WT) and *Kdm5c*-KO (5CKO) ESCs at 0, 24, and 48 hours of differentiation into EpiLCs. Expression calculated in comparison to TBP mRNA expression ( $2^{-\Delta\Delta CT}$ ). \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , Welch's t-test. **C.** KDM5C protein expression normalized to DAXX. Quantified intensity using ImageJ (artificial units - au). Right - representative lanes of Western blot for KDM5C and DAXX. \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , Welch's t-test. **D.** Top - Representative UCSC browser view of histone 3 lysine 4 dimethylation (H3K4me2) ChIP-seq signal at two germline genes in wild-type and *Kdm5c*-KO EpiLCs. Bottom - Average H3K4me2 signal at the TSS of all germline-enriched genes in wild-type (dark purple) and *Kdm5c*-KO (light purple) EpiLCs. **E.** Top - Representative UCSC browser view of histone 3 lysine 4 trimethylation (H3K4me3) ChIP-seq signal at two germline genes in the wild-type (WT) and *Kdm5c*-KO (5CKO) adult amygdala. Bottom - Average H3K4me3 signal at the transcription start site (TSS) of all germline-enriched genes in wild-type (dark red) and *Kdm5c*-KO (light red) amygdala. **F.** Percentage of germline genes that harbor CpG islands (CGIs) in their promoters (CGI promoter, red), based on UCSC annotation. Left - percentage of all germline-enriched genes, middle - KDM5C-bound germline genes, and right - KDM5C-unbound germline genes. (Legend continued on next page.)

**Figure 6: KDM5C promotes long-term silencing of germline genes via DNA methylation at CpG islands.** (Legend continued.) **G.** enrichPlot gene ontology analysis of germline genes with (CGI-promoter) or without (CGI-free) CGIs in their promoter. **H.** Expression of germline genes with (CGI-promoter, red) or without (CGI-free, salmon) CGIs in their promoter across stages of spermatogenesis from Green et al 2018. SPG - spermatogonia, Pre-Lep - preleptotene spermatocytes, S-Cytes - meiotic spermatocytes, S-Tids - post-meiotic haploid round spermatids, E.Sp - elongating spermatids. Wilcoxon test, \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , \*\*\*\*  $p < 0.0001$ . **I.** Diagram of ESC to extended EpiLC (exEpiLC) differentiation. **J.** Volcano plot of whole genome bisulfite sequencing (WGBS) comparing CpG methylation (CpGme) at germline gene promoters ( $TSS \pm 500$ ) in wild-type (WT) ESCs versus exEpiLCs. Significantly differentially methylated promoters ( $q < 0.01$ ,  $|methyl\text{ation difference}| > 25\%$ ) with CGIs in red, CGI-free promoters in light blue **K.** Volcano plot of WGBS of wild-type (WT) versus *Kdm5c*-KO exEpiLCs for germline gene promoters. Promoters with CGIs in red, CGI-free promoters in light blue. **L.** Histogram of average percent CpGme at the promoter of germline genes with or without CGIs. Wild-type in navy and *Kdm5c*-KO (KO) in light blue. Dashed lines are average methylation for each genotype, p-values for Mann-Whitney U test. **M.** UCSC browser view of germline genes, showing UCSC annotated CGI with number of CpGs, representative CpGme in wild-type (WT) and *Kdm5c*-KO (5CKO) ESCs and exEpiLCs, and KDM5C ChIP-seq signal in EpiLCs.