

1      The X-linked intellectual disability gene KDM5C is a  
2      sex-biased brake against germline programs in somatic  
3      lineages

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## 11 Abstract

12 Mutations in numerous chromatin-modifying enzymes cause neurodevelopmental disorders (NDDs). Loss  
13 of repressive chromatin regulators can lead to the aberrant transcription of tissue-specific genes outside  
14 of their intended context, however the mechanisms and consequences of their dysregulation are largely  
15 unknown. Here, we examine how the X-linked intellectual disability gene lysine demethylase 5c (KDM5C), an  
16 eraser of histone 3 lysine 4 di and tri-methylation (H3K4me2/3), contributes to tissue identity. We found male  
17 *Kdm5c* knockout (-KO) mice, which recapitulate key human neurological phenotypes, aberrantly express  
18 many liver, muscle, ovary, and testis genes within the amygdala and hippocampus. Gonad-enriched genes  
19 misexpressed in the *Kdm5c*-KO brain are unique to germ cells, indicating an erosion of the soma-germline  
20 boundary. Germline genes are typically decommissioned in somatic lineages in the post-implantation epiblast,  
21 yet *Kdm5c*-KO epiblast-like cells (EpiLCs) aberrantly expressed key regulators of germline identity and  
22 meiosis, including *Dazl* and *Stra8*. Characterizing germline gene misexpression in males and female mutants  
23 revealed germline gene repression is sexually dimorphic, with female EpiLCs requiring a higher dose of  
24 KDM5C to maintain germline gene suppression. Using a comprehensive list of mouse germline-enriched  
25 genes, we found KDM5C is selectively recruited to a subset of germline gene promoters that contain CpG  
26 islands (CGIs) to facilitate DNA CpG methylation during ESC to EpiLC differentiation. However, late-stage  
27 spermatogenesis genes devoid of promoter CGIs can become expressed in *Kdm5c*-KO cells via ectopic  
28 activation by RFX transcription factors. Together, these data demonstrate KDM5C's fundamental role in  
29 tissue identity and indicate that KDM5C acts as a brake against runaway activation of germline developmental  
30 programs in somatic lineages.

## 31 Introduction

32 A single genome holds the instructions to generate the myriad of cell types found within an organism.  
33 This is, in part, accomplished by chromatin regulators that can either promote or impede lineage-specific  
34 gene expression through DNA and histone modifications<sup>1–5</sup>. Human genetic studies revealed mutations in  
35 chromatin regulators are a major cause of neurodevelopmental disorders (NDDs)<sup>6</sup> and many studies have  
36 identified their importance for regulating brain-specific transcriptional programs. Loss of chromatin regulators  
37 can also result in the ectopic expression of tissue-specific genes outside of their target environment, such  
38 as the misexpression of liver-specific genes within adult neurons<sup>7</sup>. However, the mechanisms underlying  
39 ectopic gene expression and its impact upon neurodevelopment are poorly understood.

40 To elucidate the role of tissue identity in chromatin-linked NDDs, it is essential to first characterize the  
41 nature of the dysregulated genes and the molecular mechanisms governing their de-repression. Here we  
42 focus on the X chromosome gene lysine demethylase 5C (KDM5C, also known as SMCX or JARID1C),  
43 which erases histone 3 lysine 4 di- and trimethylation (H3K4me2/3), a permissive chromatin modification  
44 enriched at gene promoters<sup>8</sup>. Pathogenic mutations in *KDM5C* cause Intellectual Developmental Disorder,  
45 X-linked, Syndromic, Claes-Jensen Type (MRXSCJ, OMIM: 300534). MRXSCJ is more common and severe  
46 in males and its neurological phenotypes include intellectual disability, seizures, aberrant aggression, and  
47 autistic behaviors<sup>9–11</sup>. Male *Kdm5c* knockout (-KO) mice recapitulate key MRXSCJ phenotypes, including  
48 hyperaggression, increased seizure propensity, social deficits, and learning impairments<sup>12–14</sup>. RNA sequenc-  
49 ing (RNA-seq) of the *Kdm5c*-KO hippocampus revealed ectopic expression of some germline genes within  
50 the brain<sup>13</sup>. However, it is unclear if other tissue-specific genes are aberrantly transcribed with KDM5C loss,  
51 at what point in development germline gene misexpression begins, and what mechanisms underlie their  
52 dysregulation.

53 Distinguishing between germ cells and somatic cells is a key feature of multicellularity<sup>15</sup> that occurs  
54 during early embryogenesis in many metazoans<sup>16</sup>. In mammals, chromatin regulators are crucial for  
55 decommissioning germline genes during the transition from naïve to primed pluripotency. Initially, germline  
56 gene promoters gain repressive histone H2A lysine 119 monoubiquitination (H2AK119ub1)<sup>17</sup> and histone H3  
57 lysine 9 trimethylation (H3K9me3)<sup>17,18</sup> in embryonic stem cells (ESCs) and are then decorated with DNA  
58 CpG methylation (CpGme) in post-implantation epiblast cells<sup>18–21</sup>. The contribution of KDM5C to this process  
59 remains unclear. Additionally, studies on germline gene repression have primarily been conducted in males  
60 and focused on select marker genes, given the lack of a comprehensive list for germline-enriched genes.  
61 Therefore, it is unknown if the mechanism of repression differs between sexes or for different classes of  
62 germline genes, e.g. meiotic versus spermatid differentiation genes.

63 To illuminate KDM5C's role in tissue identity, here we characterized the aberrant expression of tissue-  
64 enriched genes within the *Kdm5c*-KO brain and epiblast-like stem cells (EpiLCs), an *in vitro* model of the  
65 post-implantation embryo. We curated a list of mouse germline-enriched genes, which enabled genome-wide

66 analysis of germline gene silencing mechanisms for the first time. Additionally, we characterized germline  
67 transcripts expressed in male and female *Kdm5c* mutants to illuminate the impact of sex upon germline  
68 gene suppression. Based on the data presented below, we propose KDM5C plays a fundamental, sexually  
69 dimorphic role in the development of tissue identity during early embryogenesis, including the establishment  
70 of the soma-germline boundary.

## 71 **Results**

### 72 **Tissue-enriched genes are aberrantly expressed in the *Kdm5c*-KO brain**

73 Previous RNA sequencing (RNA-seq) of the adult male *Kdm5c*-KO hippocampus revealed ectopic  
74 expression of some germline genes unique to the testis<sup>13</sup>. It is currently unknown if the testis is the only  
75 tissue type misexpressed in the *Kdm5c*-KO brain. We thus systematically tested whether other tissue-specific  
76 genes are misexpressed in the male brain with constitutive knockout of *Kdm5c* (*Kdm5c*<sup>-y</sup>, 5CKO in figures)<sup>22</sup>  
77 by using a published list of mouse tissue-enriched genes<sup>23</sup>.

78 We found a large proportion of significantly upregulated genes (DESeq2<sup>24</sup>, log2 fold change > 0.5, q <  
79 0.1) within the male *Kdm5c*-KO amygdala and hippocampus are non-brain, tissue-specific genes (Amygdala:  
80 21/59 up DEGs, 35.59% ; Hippocampus: 48/183 up DEGs, 26.23%) (Figure 1A-B, Supplementary Table  
81 1). For both the amygdala and hippocampus, the majority of tissue-enriched differentially expressed genes  
82 (DEGs) were testis genes (Figure 1A-B). Even though the testis has the largest total number of tissue-  
83 enriched genes (2,496 genes) compared to any other tissue, testis-enriched DEGs were significantly enriched  
84 in both brain regions (Amygdala p = 1.83e-05, Odds Ratio = 5.13; Hippocampus p = 4.26e-11, Odds Ratio =  
85 4.45, Fisher's Exact Test). An example of a testis-enriched gene misexpressed in the *Kdm5c*-KO brain is  
86 *FK506 binding protein 6 (Fkbp6)*, a known regulator of PIWI-interacting RNAs (piRNAs) and meiosis<sup>25,26</sup>  
87 (Figure 1C).

88 Interestingly, we also observed significant enrichment of ovary-enriched genes in both the amygdala  
89 and hippocampus (Amygdala p = 0.00574, Odds Ratio = 18.7; Hippocampus p = 0.048, Odds Ratio = 5.88,  
90 Fisher's Exact Test) (Figure 1A-B). Ovary-enriched DEGs included *Zygotic arrest 1 (Zar1)*, which sequesters  
91 mRNAs in oocytes for meiotic maturation<sup>27</sup> (Figure 1D). Given that the *Kdm5c*-KO mice we analyzed are  
92 male, these data demonstrate that the ectopic expression of gonad-enriched genes is independent of  
93 organismal sex.

94 Although not consistent across brain regions, we also found significant enrichment of genes biased  
95 towards two non-gonadal tissues - the liver (Amygdala p = 0.04, Odds Ratio = 6.58, Fisher's Exact Test)  
96 and muscle (Hippocampus p = 0.01, Odds Ratio = 6.95, Fisher's Exact Test) (Figure 1A-B). These include  
97 *Apolipoprotein C-I (Apoc1)*, a lipoprotein metabolism and transport gene<sup>28</sup> (Figure 1E, see Discussion).

98 Our analysis of oligo(dT)-primed libraries<sup>22</sup> indicates aberrantly expressed mRNAs are polyadenylated

99 and spliced into mature transcripts in the *Kdm5c*-KO brain (Figure 1C-E). Of note, we observed little to no  
100 dysregulation of brain-enriched genes (Amygdala p = 1, Odds Ratio = 1.22; Hippocampus p = 0.74, Odds  
101 Ratio = 1.22, Fisher's Exact Test), despite the fact these are brain samples and the brain has the second  
102 highest total number of tissue-enriched genes (708 genes). Altogether, these results suggest the aberrant  
103 expression of tissue-enriched genes within the brain is a major effect of KDM5C loss.

104 **Germline genes are misexpressed in the *Kdm5c*-KO brain**

105 *Kdm5c*-KO brain expresses testicular germline genes<sup>13</sup> (Figure 1), however the testis also contains  
106 somatic cells that support hormone production and germline functions. To determine if *Kdm5c*-KO results  
107 in ectopic expression of testicular somatic genes, we first evaluated the known functions of testicular  
108 DEGs through gene ontology. We found *Kdm5c*-KO testis-enriched DEGs had high enrichment of germline-  
109 relevant ontologies, including spermatid development (GO: 0007286, p.adjust = 6.2e-12) and sperm axoneme  
110 assembly (GO: 0007288, p.adjust = 2.45e-14) (Figure 2A, Supplementary Table 1).

111 We then evaluated *Kdm5c*-KO testicular DEG expression in wild-type testes versus testes with germ cell  
112 depletion<sup>29</sup>, which was accomplished by heterozygous *W* and *Wv* mutations in the enzymatic domain of *c-Kit*  
113 (*Kit*<sup>W/Wv</sup>)<sup>30</sup>. Almost all *Kdm5c*-KO testis-enriched DEGs lost expression with germ cell depletion (Figure 2B).  
114 We then assessed testis-enriched DEG expression in a published single cell RNA-seq dataset that identified  
115 cell type-specific markers within the testis<sup>31</sup>. Some *Kdm5c*-KO testis-enriched DEGs were classified as  
116 specific markers for different germ cell developmental stages (e.g. spermatogonia, spermatocytes, round  
117 spermatids, and elongating spermatids), yet none marked somatic cells (Figure 2C). Together, these data  
118 demonstrate that the *Kdm5c*-KO brain aberrantly expresses germline genes but not somatic testicular genes,  
119 reflecting an erosion of the soma-germline boundary.

120 As of yet, research on germline gene silencing mechanisms has focused on a handful of key genes rather  
121 than assessing germline gene suppression genome-wide, due to the lack of a comprehensive gene list.  
122 We therefore generated a list of mouse germline-enriched genes using RNA-seq datasets of *Kit*<sup>W/Wv</sup> mice  
123 that included males and females at embryonic day 12, 14, and 16<sup>32</sup> and adult male testes<sup>29</sup>. We defined  
124 genes as germline-enriched if their expression met the following criteria: 1) their expression is greater than  
125 1 FPKM in wild-type gonads 2) their expression in any non-gonadal tissue of adult wild type mice<sup>23</sup> does  
126 not exceed 20% of their maximum expression in the wild-type germline, and 3) their expression in the germ  
127 cell-depleted gonads, for any sex or time point, does not exceed 20% of their maximum expression in the  
128 wild-type germline. These criteria yielded 1,288 germline-enriched genes (Figure 2D), which was hereafter  
129 used as a resource to globally characterize germline gene misexpression with *Kdm5c* loss (Supplementary  
130 Table 2).

131 ***Kdm5c*-KO epiblast-like cells aberrantly express key regulators of germline identity**

132 Germ cells are typically distinguished from somatic cells soon after the embryo implants into the uterine  
133 wall<sup>33,34</sup>, when germline genes are silenced in epiblast stem cells that will form the somatic tissues<sup>35</sup>. This  
134 developmental time point can be modeled *in vitro* through differentiation of naïve embryonic stem cells  
135 (nESCs) into epiblast-like stem cells (EpiLCs) (Figure 3A)<sup>36,37</sup>. While some germline-enriched genes are  
136 also expressed in nESCs and in the 2-cell stage<sup>38–40</sup>, they are silenced as they differentiate into EpiLCs<sup>18,19</sup>.  
137 Therefore, we tested if KDM5C was necessary for the initial silencing of germline genes in somatic lineages  
138 by evaluating the impact of *Kdm5c* loss in male EpiLCs.

139 *Kdm5c*-KO cell morphology during ESC to EpiLC differentiation appeared normal (Figure 3B) and EpiLCs  
140 properly expressed markers of primed pluripotency, such as *Dnmt3b*, *Fgf5*, *Pou3f1*, and *Otx2* (Figure 3C). We  
141 then identified tissue-enriched DEGs in a RNA-seq dataset of wild-type and *Kdm5c*-KO EpiLCs<sup>41</sup> (DESeq2,  
142 log<sub>2</sub> fold change > 0.5, q < 0.1, Supplementary Table 3). Similar to the *Kdm5c*-KO brain, we observed  
143 general dysregulation of tissue-enriched genes, with the largest number of genes belonging to the brain and  
144 testis, although they were not significantly enriched (Figure 3D). Using our list of mouse germline-enriched  
145 genes assembled above, we identified 68 germline genes misexpressed in male *Kdm5c*-KO EpiLCs.

146 We then compared EpiLC germline DEGs to those expressed in the *Kdm5c*-KO brain to determine if  
147 germline genes are constitutively dysregulated or change over the course of development. The majority of  
148 germline DEGs were unique to either EpiLCs or the brain, with only *D1Pas1* and *Cyct* shared across all  
149 tissue/cell types (Figure 3E-F). EpiLC germline DEGs had particularly high enrichment of meiosis-related  
150 gene ontologies when compared to the brain (Figure 3G, Supplementary Table 3), such as meiotic cell  
151 cycle process (GO:1903046, p.adjust = 2.2e-07) and meiotic nuclear division (GO:0140013, p.adjust  
152 = 1.37e-07). While there was modest enrichment of meiotic gene ontologies in both brain regions, the  
153 *Kdm5c*-KO hippocampus primarily expressed late-stage spermatogenesis genes involved in sperm axoneme  
154 assembly (GO:0007288, p.adjust = 0.00621) and sperm motility (GO:0097722, p.adjust = 0.00612).

155 Notably, DEGs unique to *Kdm5c*-KO EpiLCs included key drivers of germline identity, such as *Stimulated*  
156 *by retinoic acid 8* (*Stra8*: log<sub>2</sub> fold change = 3.73, q = 2.17e-39) and *Deleted in azoospermia like* (*Dazl*:  
157 log<sub>2</sub> fold change = 3.36, q = 3.19e-12) (Figure 3H). These genes are typically expressed when a subset  
158 of epiblast stem cells become primordial germ cells (PGCs) and then again in mature germ cells to trigger  
159 meiotic gene expression programs<sup>42–44</sup>. Of note, some germline genes, including *Dazl*, are also expressed  
160 in the two-cell embryo<sup>39,45</sup>. However, we did not see derepression of two-cell stage-specific genes, like  
161 *Duxf3* (*Dux*) (log<sub>2</sub> fold change = -0.282, q = 0.337) and *Zscan4d* (log<sub>2</sub> fold change = 0.25, q = 0.381) (Figure  
162 3H, Supplementary Table 3), indicating *Kdm5c*-KO EpiLCs do not revert back to a 2-cell state. Altogether,  
163 *Kdm5c*-KO EpiLCs express key drivers of germline identity and meiosis while the brain primarily expresses  
164 spermiogenesis genes, indicating germline gene misexpression mirrors germline development during the  
165 progression of somatic development.

166 **Female epiblast-like cells have heightened germline gene misexpression with *Kdm5c***  
167 **loss**

168 It is currently unknown if the misexpression of germline genes is influenced by sex, as previous studies  
169 on germline gene repressors have focused on male cells<sup>17,18,20,46,47</sup>. Sex is particularly pertinent in the case  
170 of KDM5C because it partially escapes X chromosome inactivation (XCI), resulting in a higher dosage in  
171 females<sup>48–51</sup>. We therefore explored the impact of chromosomal sex upon germline gene suppression by  
172 comparing their dysregulation in male *Kdm5c* hemizygous knockout (*Kdm5c*<sup>-y</sup>, XY *Kdm5c*-KO, XY 5CKO),  
173 female homozygous knockout (*Kdm5c*<sup>-/-</sup>, XX *Kdm5c*-KO, XX 5CKO), and female heterozygous knockout  
174 (*Kdm5c*<sup>-/+</sup>, XX *Kdm5c*-HET, XX 5CHET) EpiLCs<sup>41</sup>.

175 In EpiLCs, homozygous and heterozygous *Kdm5c* knockout females expressed over double the number  
176 of germline-enriched genes than hemizygous males (Figure 4A, Supplementary Table 3). While the majority  
177 of germline DEGs in *Kdm5c*-KO males were also dysregulated in females (74%), many were sex-specific,  
178 such as *Tktl2* and *Esx1* (Figure 4B). We then compared the known functions of germline genes dysregulated  
179 uniquely in males and females or misexpressed in all samples (Figure 4C, Supplementary Table 3). Female-  
180 specific germline DEGs were enriched for meiotic (GO:0051321 - meiotic cell cycle) and flagellar (GO:  
181 0003341 - cilium movement) functions, while male-specific DEGs had roles in mitochondrial and cell signaling  
182 (GO:0070585 - protein localization to mitochondrion).

183 The majority of germline genes expressed in both sexes were more highly dysregulated in females  
184 compared to males (Figure 4D-F). This increased degree of dysregulation in females, along with the  
185 increased total number of germline genes, indicates females are more sensitive to losing KDM5C-mediated  
186 germline gene suppression. Heightened germline gene dysregulation in females could be due to impaired  
187 XCI in *Kdm5c* mutants<sup>41</sup>, as many spermatogenesis genes lie on the X chromosome<sup>52,53</sup>. However, female  
188 germline DEGs were not biased towards the X chromosome ( $p = 1$ , Odds Ratio = 0.96, Fisher's Exact Test)  
189 and females had a similar overall proportion of germline DEGs belonging to the X chromosome as males  
190 (XY *Kdm5c*-KO - 10.29%, XX *Kdm5c*-HET - 7.43%, XX *Kdm5c*-KO - 10.59%) (Figure 4G). The majority of  
191 germline DEGs instead lie on autosomes for both male and female *Kdm5c* mutants (Figure 4G). Thus, while  
192 female EpiLCs are more prone to germline gene misexpression with KDM5C loss, it is likely independent of  
193 XCI defects.

194 **Germline gene misexpression in *Kdm5c* mutants is independent of germ cell sex**

195 Although many germline genes have shared functions in the male and female germline, e.g. PGC  
196 formation, meiosis, and genome defense, some have unique or sex-biased expression. Therefore, we  
197 wondered if *Kdm5c* mutant males would primarily express sperm genes while mutant females would primarily  
198 express egg genes. To comprehensively assess whether germline gene sex corresponds with *Kdm5c*  
199 mutant sex, we first filtered our list of germline-enriched genes for egg and sperm-biased genes (Figure 4,

200 Supplementary Table 2). We defined germ cell sex-biased genes as those whose expression in the opposite  
201 sex, at any time point, is no greater than 20% of the gene's maximum expression in a given sex. This  
202 criteria yielded 67 egg-biased, 1,024 sperm-biased, and 197 unbiased germline-enriched genes. We found  
203 regardless of sex, egg, sperm, and unbiased germline genes were dysregulated in all *Kdm5c* mutants at  
204 similar proportions (Figure 4I-J). Furthermore, germline genes dysregulated exclusively in either male or  
205 female mutants were also not biased towards their corresponding germ cell sex (Figure 4I). Altogether, these  
206 results demonstrate sex differences in germline gene dysregulation is not due to sex-specific activation of  
207 sperm or egg transcriptional programs.

## 208 KDM5C binds to a subset of germline gene promoters during early embryogenesis

209 KDM5C binds to the promoters of several germline genes in embryonic stem cells (ESCs) but not in  
210 neurons<sup>13,54</sup>. However, due to the lack of a comprehensive list of germline-enriched genes, it is unclear if  
211 KDM5C is enriched at germline gene promoters, what types of germline genes KDM5C regulates, and if its  
212 binding is maintained at any germline genes in neurons.

213 To address these questions, we analyzed KDM5C chromatin immunoprecipitation followed by DNA  
214 sequencing (ChIP-seq) datasets in EpiLCs<sup>41</sup> and primary forebrain neuron cultures (PNCs)<sup>12</sup> (MACS2 q <  
215 0.1, fold enrichment > 1, and removal of false-positive *Kdm5c*-KO peaks). EpiLCs had a higher total number  
216 of high-confidence KDM5C peaks than PNCs (EpiLCs: 5,808, PNCs: 1,276). KDM5C was primarily localized  
217 to gene promoters in both cell types (promoters = transcription start site (TSS) ± 500 bp, EpiLCs: 4,190,  
218 PNCs: 745), although PNCs showed increased localization to non-promoter regions (Figure 5A).

219 The majority of promoters bound by KDM5C in PNCs were also bound in EpiLCs (513 shared promoters),  
220 however a large portion of gene promoters were bound by KDM5C only in EpiLCs (3,677 EpiLC only  
221 promoters) (Figure 5B). Genes bound by KDM5C in both PNCs and EpiLCs were enriched for functions  
222 involving nucleic acid turnover, such as deoxyribonucleotide metabolic process (GO:0009262, p.adjust =  
223 8.28e-05) (Figure 5C, Supplementary Table 4). Germline ontologies were enriched only in EpiLC-specific,  
224 KDM5C-bound promoters, such as meiotic nuclear division (GO: 0007127 p.adjust = 6.77e-16) (Figure 5C).  
225 There were no significant ontologies for PNC-specific KDM5C target genes. Using our mouse germline gene  
226 list, we observed evident KDM5C signal around the TSS of many germline genes in EpiLCs, but not in PNCs  
227 (Figure 5D). Based on our ChIP-seq peak cut-off criteria, KDM5C was highly enriched at 211 germline gene  
228 promoters in EpiLCs (16.4% of all germline genes) (Figure 5E, Supplementary Table 2). Of note, KDM5C  
229 was only bound to about one third of RNA-seq DEG promoters unique to EpiLCs or the brain (EpiLC only  
230 DEGs: 34.9%, Brain only DEGs: 30%) (Supplementary Figure 1A-C). Representative examples of EpiLC  
231 DEGs bound and unbound by KDM5C in EpiLCs are *Dazl* and *Stra8*, respectively (Figure 5F). However,  
232 the four of the five germline genes dysregulated in both EpiLCs and the brain were bound by KDM5C in  
233 EpiLCs (*D1Pas1*, *Hsf2bp*, *Cyct*, and *Stk31*) (Supplementary Figure 1A). Together, these results demonstrate  
234 KDM5C is recruited to a subset of germline genes in EpiLCs, including meiotic genes, but does not directly

235 regulate germline genes in neurons. Furthermore, the majority of germline mRNAs expressed in *Kdm5c*-KO  
236 cells are dysregulated independent of direct KDM5C recruitment to their gene promoters, however genes  
237 dysregulated across *Kdm5c*-KO development are often direct KDM5C targets.

238 Many germline-specific genes are suppressed by the polycomb repressive complex 1.6 (PRC1.6), which  
239 contains the transcription factor heterodimers E2F/DP1 and MGA/MAX that respectively bind E2F and  
240 E-box motifs within germline gene promoters<sup>17,18,20,40,46,47,55-57</sup>. PRC1.6 members may recruit KDM5C to  
241 germline gene promoters<sup>13</sup>, given their association with KDM5C in HeLa cells and ESCs<sup>45,58</sup>. We thus  
242 used HOMER<sup>59</sup> to identify transcription factor motifs enriched at KDM5C-bound or unbound germline gene  
243 promoters (TSS ± 500 bp, q-value < 0.1, Supplementary Table 4). MAX and E2F6 binding sites were  
244 significantly enriched at germline genes bound by KDM5C in EpiLCs (MAX q-value: 0.0068, E2F6 q-value:  
245 0.0673, E2F q-value: 0.0917), but not at germline genes unbound by KDM5C (Figure 5G). One third of  
246 KDM5C-bound promoters contained the consensus sequence for either E2F6 (E2F, 5'-TCCCGC-3'), MGA  
247 (E-box, 5'-CACGTG-3'), or both, but only 17% of KDM5C-unbound genes contained these motifs (Figure 5H).  
248 KDM5C-unbound germline genes were instead enriched for multiple RFX transcription factor binding sites  
249 (RFX q-value < 0.0001, RFX2 q-value < 0.0001, RFX5 q-value < 0.0001) (Figure 5I, Supplementary figure  
250 1D). RFX transcription factors bind X-box motifs<sup>60</sup> to promote ciliogenesis<sup>61,62</sup> and among them is RFX2, a  
251 central regulator of post-meiotic spermatogenesis<sup>63,64</sup>. Although *Rfx2* is also not a direct target of KDM5C  
252 (Supplementary Figure 1E), RFX2 mRNA is derepressed in *Kdm5c*-KO EpiLCs (Figure 5J). Thus, RFX2 is a  
253 candidate transcription factor for driving the ectopic expression of many KDM5C-unbound germline genes in  
254 *Kdm5c*-KO cells.

255 **KDM5C is recruited to CpG islands at germline promoters to facilitate *de novo* DNA  
256 methylation**

257 Previous work found two germline gene promoters have a marked reduction in DNA CpG methylation  
258 (CpGme) in the adult *Kdm5c*-KO hippocampus<sup>13</sup>. Since histone H3K4me2/3 impede *de novo* CpGme<sup>65,66</sup>,  
259 KDM5C's removal of H3K4me2/3 may be required to suppress germline genes. However, KDM5C's catalytic  
260 activity was recently shown to be dispensable for suppressing *Dazl* in undifferentiated ESCs<sup>45</sup>. To reconcile  
261 these observations, we hypothesized KDM5C erases H3K4me2/3 to promote the initial placement of CpGme  
262 at germline gene promoters in EpiLCs.

263 To test this hypothesis, we first characterized KDM5C's expression as naïve ESCs differentiate into  
264 EpiLCs (Figure 6A). While *Kdm5c* mRNA steadily decreased from 0 to 48 hours of differentiation (Figure  
265 6B), KDM5C protein initially increased from 0 to 24 hours and then decreased to near knockout levels by 48  
266 hours (Figure 6C). We then characterized KDM5C's substrates (H3K4me2/3) at germline gene promoters  
267 with *Kdm5c* loss using published ChIP-seq datasets<sup>22,41</sup>. *Kdm5c*-KO samples showed a marked increase in  
268 H3K4me2 in EpiLCs (Figure 6D) and H3K4me3 in the amygdala (Figure 6E) around the TSS of germline

269 genes. Together, these data suggest KDM5C acts during the transition between ESCs and EpiLCs to remove  
270 H3K4me2/3 at germline gene promoters.

271 Germline genes accumulate CpG methylation (CpGme) at CpG islands (CGIs) during the transition  
272 from naïve to primed pluripotency<sup>19,21,67</sup>. We first examined how many of our germline-enriched genes had  
273 promoter CGIs (TSS ± 500 bp) using the UCSC genome browser<sup>68</sup>. Notably, out of 1,288 germline-enriched  
274 genes, only 356 (27.64%) had promoter CGIs (Figure 6F, Supplementary Table 2). CGI-containing germline  
275 genes had higher enrichment of meiotic gene ontologies compared to CGI-free genes, including meiotic  
276 nuclear division (GO:0140013, p.adjust = 2.17e-12) and meiosis I (GO:0007127, p.adjust = 3.91e-10)  
277 (Figure 6G, Supplementary Table 5). Germline genes with promoter CGIs were more highly expressed than  
278 CGI-free genes across spermatogenesis stages, with highest expression in meiotic spermatocytes (Figure  
279 6H). Contrastingly, CGI-free genes only displayed substantial expression in post-meiotic round spermatids  
280 (Figure 6H). Although only a minor portion of germline gene promoters contained CGIs, CGIs strongly  
281 determined KDM5C's recruitment to germline genes ( $p = 2.37e-67$ , Odds Ratio = 17.8, Fisher's Exact Test),  
282 with 79.15% of KDM5C-bound germline gene promoters harboring CGIs (Figure 6F).

283 To assess how KDM5C loss impacts initial CpGme placement at germline gene promoters, we performed  
284 whole genome bisulfite sequencing (WGBS) in male wild-type and *Kdm5c*-KO ESCs and 96-hour extend  
285 EpiLCs (exEpiLCs), when germline genes reach peak methylation levels<sup>18</sup> (Figure 6I). We first identified  
286 which germline gene promoters significantly gained CpGme in wild-type cells during nESC to exEpiLCs  
287 differentiation (methylKit<sup>69</sup>,  $q < 0.01$ ,  $|methylation\ difference| > 25\%$ , TSS ± 500 bp). In wild-type cells, the  
288 majority of germline genes gained substantial CpGme at their promoter during differentiation (60.08%),  
289 regardless if their promoter contained a CGI (Figure 6J, Supplementary Table 5).

290 We then identified promoters differentially methylated in wild-type versus *Kdm5c*-KO exEpiLCs (methylKit,  
291  $q < 0.01$ ,  $|methylation\ difference| > 25\%$ , TSS ± 500 bp, Supplementary Table 5). Of the 48,882 promoters  
292 assessed, 274 promoters were significantly hypomethylated and 377 promoters were significantly hyper-  
293 methylated with KDM5C loss (Supplementary Figure 2A). Many promoters hyper- and hypomethylated  
294 in *Kdm5c*-KO exEpiLCs belonged to genes with unknown functions. However, 10.22% of hypomethyl-  
295 ated promoters belonged to germline genes and germline-relevant ontologies like meiotic nuclear division  
296 (GO:0140013, p.adjust = 0.012) are significantly enriched (Supplementary Figure 2B, Supplementary Table  
297 5). Approximately half of all germline gene promoters hypomethylated in *Kdm5c*-KO exEpiLCs are direct  
298 targets of KDM5C in EpiLCs (13 out of 28 hypomethylated promoters).

299 Promoters that showed the most robust loss of CpGme in *Kdm5c*-KO exEpiLCs (lowest q-values) harbored  
300 CGIs (Figure 6K). CGI promoters, but not CGI-free promoters, had a significant reduction in CpGme with  
301 KDM5C loss as a whole (Figure 6L) (Non-CGI promoters  $p = 0.0846$ , CGI promoters  $p = 0.0081$ , Mann-  
302 Whitney U test). Significantly hypomethylated promoters included germline genes consistently dysregulated  
303 across multiple *Kdm5c*-KO RNA-seq datasets<sup>13</sup>, such as *D1Pas1* (methylation difference = -60.03%, q-value  
304 = 3.26e-153) and *Naa11* (methylation difference = -42.45%, q-value = 1.44e-38) (Figure 6M). Unexpectedly,

305 we observed only a modest reduction in CpGme at *Dazl*'s promoter (methylation difference = -6.525%,  
306 q-value = 0.0159) (Figure 6N). Altogether, these results demonstrate KDM5C is recruited to germline gene  
307 CGIs in EpiLCs to promote CpGme at those promoters. Furthermore, our data suggest while KDM5C's  
308 catalytic activity is required for the repression of some germline genes, CpGme can be placed at others even  
309 with elevated H3K4me2/3 around the TSS.

## 310 Discussion

311 In the above study, we demonstrate KDM5C's pivotal role in the development of tissue identity. We first  
312 characterized tissue-enriched genes expressed within the mouse *Kdm5c*-KO brain and identified substantial  
313 derepression of testis, liver, muscle, and ovary-enriched genes. Testis genes significantly enriched within the  
314 *Kdm5c*-KO amygdala and hippocampus are specific to the germline and absent in somatic cells. *Kdm5c*-  
315 KO epiblast-like cells (EpiLCs) aberrantly express key drivers of germline identity and meiosis, including  
316 *Dazl* and *Stra8*, while the adult brain primarily expresses genes important for late spermatogenesis. We  
317 demonstrated that although sex did not influence whether sperm or egg-specific genes were misexpressed,  
318 female EpiLCs have heightened germline gene de-repression with KDM5C loss. Germline genes can become  
319 aberrantly expressed in *Kdm5c*-KO cells via indirect mechanisms, such as activation through ectopic RFX  
320 transcription factors. Finally, we found KDM5C is dynamically regulated during ESC to EpiLC differentiation  
321 to promote long-term germline gene silencing through CGI DNA methylation. Therefore, we propose KDM5C  
322 plays a fundamental role in the development of tissue identity during early embryogenesis, including the  
323 establishment of the soma-germline boundary. By systematically characterizing KDM5C's role in germline  
324 gene repression, we unveiled distinct mechanisms governing the misexpression of distinct germline gene  
325 classes in somatic lineages. Ultimately, these data provide molecular footholds which can be exploited to  
326 test the overarching contribution of ectopic germline gene expression upon neurodevelopment.

327 By comparing *Kdm5c* mutant males and females, we revealed germline gene suppression is sexually  
328 dimorphic. Female EpiLCs are more severely impacted by loss of KDM5C-mediated germline gene sup-  
329 pression, yet this difference is not due to the large number of germline genes on the X chromosome<sup>52,53</sup>.  
330 Heightened germline gene misexpression in females may be related to females having a higher dose of  
331 KDM5C than males, due to its escape from XCI<sup>48–51</sup>. Intriguingly, heterozygous knockout females (*Kdm5c*<sup>-/+</sup>)  
332 also had over double the number of germline DEGs than hemizygous knockout males (*Kdm5c*<sup>+/Y</sup>), even  
333 though their expression of KDM5C should be roughly equivalent to that of wild-type males (*Kdm5c*<sup>+/Y</sup>). Males  
334 could partially compensate for KDM5C's loss via the Y-chromosome homolog, KDM5D<sup>8</sup>. However, KDM5D  
335 has not been reported to regulate germline gene expression. Nevertheless, these results demonstrate  
336 germline gene silencing mechanisms differ between males and females, which warrants further study to  
337 elucidate the biological ramifications and underlying mechanisms.

338 We found KDM5C is largely dispensable for promoting normal gene expression during development, yet

339 is critical for suppressing ectopic developmental programs. While some germline genes, such as *Dazl*, are  
340 also expressed in the 2-cell stage, the inner cell mass, and naïve ESCs, they are silenced in epiblast stem  
341 cells/EpiLCs<sup>18,40,45,70,71</sup>. Our data suggest the 2-cell-like state reported in *Kdm5c*-KO ESCs<sup>45</sup> likely reflects  
342 KDM5C's primary role in germline gene repression (Figure 3). Germline gene misexpression in *Kdm5c*-  
343 KO EpiLCs may indicate they are differentiating into primordial germ cell-like cells (PGCLCs)<sup>33,34,36</sup>. Yet,  
344 *Kdm5c*-KO EpiLCs had normal cellular morphology and properly expressed markers for primed pluripotency,  
345 including *Otx2* which blocks EpiLC differentiation into PGCs/PGCLCs<sup>72</sup>. In addition to unimpaired EpiLC  
346 differentiation, *Kdm5c*-KO gross brain morphology is overall normal<sup>12</sup> and hardly any brain-specific genes  
347 were significantly dysregulated in the amygdala and hippocampus (Figure 1). Thus, ectopic germline gene  
348 expression occurs in conjunction with overall proper somatic differentiation in *Kdm5c*-KO animals.

349 Our work provides novel insight into the cross-talk between H3K4me2/3 and CpGme, which are gen-  
350 erally mutually exclusive<sup>73</sup>. In EpiLCs, loss of KDM5C binding at a subset of germline gene promoters,  
351 e.g. *D1Pas1*, strongly impaired promoter CGI methylation and resulted in their long-lasting de-repression  
352 into adulthood. Removal of H3K4me2/3 at CGIs is a plausible mechanism for KDM5C-mediated germline  
353 gene suppression<sup>13,54</sup>, given H3K4me2/3 repell DNMT3 activity<sup>65,66</sup>. However, emerging work indicates  
354 many histone-modifying enzymes have non-catalytic functions that influence gene expression, sometimes  
355 even more potently than their catalytic roles<sup>74,75</sup>. Indeed, KDM5C's catalytic activity was recently found to be  
356 dispensable for repressing *Dazl* in ESCs<sup>45</sup>. In our study, *Dazl*'s promoter still gained CpGme in *Kdm5c*-KO  
357 exEpiLCs, even with elevated H3K4me2. *Dazl* and a few other germline genes employ multiple repressive  
358 mechanisms to facilitate CpGme, such as DNMT3A/B recruitment via E2F6 and MGA<sup>17,18,46,47</sup>. Thus, while  
359 some germline CGIs require KDM5C-mediated H3K4me removal to overcome promoter CGI escape from  
360 CpGme<sup>73,76</sup>, others do not. These results also suggest the requirement for KDM5C's catalytic activity can  
361 change depending upon the locus and developmental stage. Further experiments are required to determine  
362 if catalytically inactive KDM5C can suppress germline genes at later developmental stages.

363 By generating a comprehensive list of mouse germline-enriched genes, we revealed distinct derepressive  
364 mechanisms governing early versus late-stage germline programs. Previous work on germline gene silencing  
365 has focused on genes with promoter CGIs<sup>19,73</sup>, and indeed the majority of KDM5C targets in EpiLCs were  
366 germ cell identity genes harboring CGIs. However, over 70% of germline-enriched gene promoters lacked  
367 CGIs, including the many KDM5C-unbound germline genes that are de-repressed in *Kdm5c*-KO cells. CGI-  
368 free, KDM5C-unbound germline genes were primarily late-stage spermatogenesis genes and significantly  
369 enriched for RFX2 binding sites, a central regulator of spermiogenesis<sup>63,64</sup>. These data suggest that once  
370 activated during early embryogenesis, drivers of germline gene expression like *Rfx2*, *Stra8*, and *Dazl* turn  
371 on downstream germline programs, ultimately culminating in the expression of spermiogenesis genes in  
372 the adult *Kdm5c*-KO brain. Therefore, we propose KDM5C is recruited via promoter CGIs to act as a brake  
373 against runaway activation of germline-specific programs. Future studies should address how KDM5C is  
374 targeted to CGIs.

375 The above work provides the mechanistic foundation for KDM5C-mediated repression of tissue and  
376 germline-specific genes. However, the contribution of these ectopic, tissue-specific genes towards neurolog-  
377 ical impairments is still unknown. In addition to germline genes, we also identified significant enrichment  
378 of muscle and liver-enriched transcripts within the *Kdm5c*-KO brain. Intriguingly, select liver and muscle-  
379 enriched DEGs do have known roles within the brain, such as the liver-enriched lipid metabolism gene  
380 *Apolipoprotein C-I (Apoc1)*<sup>28</sup>. *APOC1* dysregulation is implicated in Alzheimer's disease in humans<sup>77</sup> and  
381 overexpression of *Apoc1* in the mouse brain can impair learning and memory<sup>78</sup>. KDM5C may therefore be  
382 crucial for neurodevelopment by fine-tuning the expression of tissue-enriched, dosage-sensitive genes like  
383 *Apoc1*.

384 Given that germline genes have no known functions within the brain, their impact upon neurodevelopment  
385 is currently unknown. In *C. elegans*, somatic misexpression of germline genes via loss of *Retinoblastoma*  
386 (*Rb*) homologs results in enhanced piRNA signaling and ectopic P granule formation in neurons<sup>79,80</sup>. Ectopic  
387 testicular germline transcripts have also been observed in a variety of cancers, including brain tumors in  
388 *Drosophila* and mammals<sup>81,82</sup> and shown to promote cancer progression<sup>83–85</sup>. Intriguingly, mouse models  
389 and human cells for other chromatin-linked NDDs also display impaired soma-germline demarcation<sup>86–88</sup>,  
390 such as mutations in DNA methyltransferase 3b (DNMT3B), H3K9me1/2 methyltransferases G9A/GLP,  
391 and methyl-CpG -binding protein 2 (MECP2). Recently, the transcription factor ZMYM2 (ZNF198), whose  
392 mutation causes a NDD (OMIM #619522), was also shown to repress germline genes by promoting H3K4me  
393 removal and CpGme<sup>89</sup>. Thus, KDM5C is among a growing cohort of neurodevelopmental disorders with  
394 erosion of the germline-soma boundary. Further research is required to determine the impact of these  
395 germline genes upon neuronal functions and the extent to which this phenomenon occurs in humans.

## 396 Materials and Methods

### 397 Classifying tissue-enriched and germline-enriched genes

398 Tissue-enriched differentially expresssd genes (DEGs) were determined by their classification in a previ-  
399 ously published dataset from 17 male and female mouse tissues<sup>23</sup>. This study defined tissue expression as  
400 greater than 1 Fragments Per Kilobase of transcript per Million mapped read (FPKM) and tissue enrichment  
401 as at least 4-fold higher expression than any other tissue.

402 We curated a list of germline-enriched genes using an RNA-seq dataset from wild-type and germline-  
403 depleted (Kit<sup>W/W<sup>v</sup></sup>) male and female mouse embryos from embryonic day 12, 14, and 16<sup>32</sup>, as well as adult  
404 male testes<sup>29</sup>. Germline-enriched genes met the following criteria: 1) their expression is greater than 1  
405 FPKM in wild-type germline 2) their expression in any wild-type somatic tissues<sup>23</sup> does not exceed 20%  
406 of maximum expression in wild-type germline, and 3) their expression in the germ cell-depleted (Kit<sup>W/W<sup>v</sup></sup>)  
407 germline, for any sex or time point, does not exceed 20% of maximum expression in wild-type germline. We

408 defined sperm and egg-biased genes as those whose expression in the opposite sex, at any time point, is no  
409 greater than 20% of the gene's maximum expression in a given sex. Genes that did not meet this threshold  
410 for either sex were classified as 'unbiased'.

## 411 **Cell culture**

412 We utilized our previously established cultures of male wild-type and *Kdm5c* knockout (-KO)  
413 embryonic stem cells<sup>41</sup>. Sex was confirmed by genotyping *Uba1/Uba1y* on the X and Y chromo-  
414 somes with the following primers: 5'-TGGATGGTGTGCCAATG-3', 5'-CACCTGCACGTTGCCCTT-  
415 3'. Deletion of *Kdm5c* exons 11 and 12, which destabilize KDM5C protein<sup>12</sup>, was confirmed  
416 through the primers 5'-ATGCCCATATTAAGAGTCCCTG-3', 5'-TCTGCCTTGATGGACTGTT-3', and  
417 5'-GGTTCTAACACTCACATAGTG-3'.

418 Embryonic stem cells (ESCs) and epiblast-like cells were cultured using previously established  
419 methods<sup>37</sup>. Briefly, ESCs were initially cultured in primed ESC (pESC) media consisting of KnockOut  
420 DMEM (Gibco#10829–018), fetal bovine serum (Gibco#A5209501), KnockOut serum replacement  
421 (Invitrogen#10828–028), Glutamax (Gibco#35050-061), Anti-Anti (Gibco#15240-062), MEM Non-essential  
422 amino acids (Gibco#11140-050), and beta-mercaptoethanol (Sigma#M7522). They were then transitioned  
423 into ground-state, "naïve" ESCs (nESCs) by culturing for four passages in N2B27 media containing  
424 DMEM/F12 (Gibco#11330–032), Neurobasal media (Gibco#21103–049), Gluamax (Gibco#35050-061),  
425 Anti-Anti (Gibco#15240-062), N2 supplement (Invitrogen#17502048), and B27 supplement without vitamin  
426 A (Invitrogen#12587-010), and beta-mercaptoethanol (Sigma#M7522). Both pESC and nESC media  
427 were supplemented with 3 µM GSK3 inhibitor CHIR99021 (Sigma #SML1046-5MG), 1 µM MEK inhibitor  
428 PD0325901 (Sigma #PZ0162-5MG), and 1,000 units/mL leukemia inhibitory factor (LIF, Millipore#ESG1107).

429 nESCs were differentiated into epiblast-like cells (EpiLCs, 48 hours) and extendend EpiLCs (exEpiLCs,  
430 96 hours) by culturing in N2B27 media containing DMEM/F12, Neurobasal media, Gluamax, Anti-Anti, N2  
431 supplement, B27 supplement (Invitrogen#17504044), and beta-mercaptoethanol supplemented with 10  
432 ng/mL fibroblast growth factor 2 (FGF2, R&D Biotechne 233-FB), and 20 ng/mL activin A (R&D Biotechne  
433 338AC050CF), as previously described<sup>37</sup>.

## 434 **Real time quantitative PCR (RT-qPCR)**

435 nESCs were differentiated into EpiLCs as described above. Cells were lysed with Tri-reagent BD (Sigma  
436 #T3809) at 0, 24, and 48 hours of differentiation. RNA was phase separated with 0.1 µL/µL 1-bromo-3-  
437 chloropropane (Sigma #B9673) and then precipitated with with isopropanol (Sigma #I9516) and ethanol puri-  
438 fied. For each sample, 2 µg of RNA was reverse transcribed using the ProtoScript II Reverse transcriptase kit  
439 from New England Biolabs (NEB #M0368S) and primed with oligo dT. Expression of *Kdm5c* was detected us-  
440 ing the primers 5'-CCCATGGAGGCCAGAGAATAAG-3' 5'-CTCAGCGGATAAGAGAATTGCTAC-3' and nor-

441 malized to TBP using the primers 5'-TTCAGAGGATGCTCTAGGAAAGA-3' 5'-CTGTGGAGTAAGTCCTGTGCC-  
442 3' with the Power SYBR™ Green PCR Master Mix (ThermoFisher #4367659).

443 **Western Blot**

444 Total protein was extracted during nESC to EpiLC differentiation at 0, 24, and 48 hours by sonicating cells  
445 at 20% amplitude for 15 seconds in 2X SDS sample buffer, then boiling at 100 °C for 10 minutes. Proteins  
446 were separated on a 7.5% SDS page gel, transferred overnight onto a fluorescent membrane, blotted for  
447 rabbit anti-KDM5C (in house, 1:500) and mouse anti-DAXX (Santa Cruz #(H-7): sc-8043, 1:500), and then  
448 imaged using the LiCor Odyssey CLx system. Band intensity was quantified using ImageJ.

449 **RNA sequencing (RNA-seq) data analysis**

450 After ensuring read quality via FastQC (v0.11.8), reads were then mapped to the mm10 *Mus musculus*  
451 genome (Gencode) using STAR (v2.5.3a), during which we removed duplicates and kept only uniquely  
452 mapped reads. Count files were generated by FeatureCounts (Subread v1.5.0), and BAM files were  
453 converted to bigwigs using deeptools (v3.1.3) and visualized by the UCSC genome browser<sup>68</sup>. RStudio  
454 (v3.6.0) was then used to analyze counts files by DESeq2 (v1.26.0)<sup>24</sup> to identify differentially expressed  
455 genes (DEGs) with a q-value (p-adjusted via FDR/Benjamini–Hochberg correction) less than 0.1 and a log2  
456 fold change greater than 0.5. For all DESeq2 analyses, log2 fold changes were calculated with IfcShrink  
457 using the ashR package<sup>90</sup>. MA-plots were generated by ggpubr (v0.6.0), and Eulerr diagrams were generated  
458 by eulerr (v6.1.1). Boxplots and scatterplots were generated by ggpubr (v0.6.0) and ggplot2 (v3.3.2). The  
459 Upset plot was generated via the package UpSetR (v1.4.0)<sup>91</sup>. Gene ontology (GO) analyses were performed  
460 by the R package enrichPlot (v1.16.2) using the biological processes setting and compareCluster.

461 **Chromatin immunoprecipitation followed by DNA sequencing (ChIP-seq) data analysis**

462 ChIP-seq reads were aligned to mm10 using Bowtie1 (v1.1.2) allowing up to two mismatches. Only  
463 uniquely mapped reads were used for analysis. Peaks were called using MACS2 software (v2.2.9.1) using  
464 input BAM files for normalization, with filters for a q-value < 0.1 and a fold enrichment > 1. We removed  
465 “black-listed” genomic regions that often give aberrant signals. Common peak sets were obtained in R via  
466 DiffBind<sup>92</sup> (v3.6.5). In the case of KDM5C ChIP-seq, *Kdm5c*-KO false-positive peaks were then removed from  
467 wild-type samples using bedtools (v2.25.0). Peak proximity to genomic loci was determined by ChIPSeeker<sup>93</sup>  
468 (v1.32.1). Gene ontology (GO) analyses were performed by the R package enrichPlot (v1.16.2) using the  
469 biological processes setting and compareCluster. Enriched motifs were identified using HOMER<sup>59</sup> to search  
470 for known motifs within 500 base pairs upstream and downstream of the transcription start site. Average binding  
471 across genes was visualized using deeptools (v3.1.3). Bigwigs were visualized using the UCSC genome  
472 browser<sup>68</sup>.

473 **CpG island (CGI) analysis**

474 Locations of CpG islands were determined through the mm10 UCSC genome browser CpG island track<sup>68</sup>,  
475 which classified CGIs as regions that have greater than 50% GC content, are larger than 200 base pairs,  
476 and have a ratio of CG dinucleotides observed over the expected amount greater than 0.6. CGI genomic  
477 coordinates were then annotated using ChIPseeker<sup>93</sup> (v1.32.1) and filtered for ones that lie within promoters  
478 of germline-enriched genes (TSS ± 500).

479 **Whole genome bisulfite sequencing (WGBS)**

480 Genomic DNA (gDNA) from male naïve ESCs and extended EpiLCs was extracted using the Wizard  
481 Genomic DNA Purification Kit (Promega A1120), following the instructions for Tissue Culture Cells. gDNA  
482 from two wild-types and two *Kdm5c*-KOs of each cell type was sent to Novogene for WGBS using the  
483 Illumina NovaSeq X Plus platform and sequenced for 150 bp paired-end reads (PE150). All samples had  
484 greater than 99% bisulfite conversion rates. Reads were adapter and quality trimmed with Trim Galore  
485 (v0.6.10) and aligned to the mm10 genome using Bismark<sup>94</sup> (v0.22.1). Analysis of differential methylation at  
486 gene promoters was performed using methylKit<sup>69</sup> (v1.28.0) with a minimum coverage of 3 paired reads, a  
487 percentage greater than 25% or less than -25%, and q-value less than 0.01. methylKit was also used to  
488 calculate average percentage methylation at germline gene promoters. Methylation bedgraph tracks were  
489 generated via Bismark and visualized using the UCSC genome browser<sup>68</sup>.

490 **Data availability**

491 **WGBS in wild-type and *Kdm5c*-KO ESCs and exEpiLCs**

492 Raw fastq files are deposited in the Sequence Read Archive (SRA) <https://www.ncbi.nlm.nih.gov/sra>  
493 under the bioProject PRJNA1165148.

494 **Published datasets**

495 All published datasets are available at the Gene Expression Omnibus (GEO) <https://www.ncbi.nlm.nih.gov/geo/>. Published RNA-seq datasets analyzed in this study included the male wild-type and *Kdm5c*-KO  
496 adult amygdala and hippocampus<sup>22</sup>, available at GEO: GSE127722. Male and female wild-type, *Kdm5c*-KO,  
497 and *Kdm5c*-HET EpiLCs<sup>41</sup> are available at GEO: GSE96797.

499 Previously published ChIP-seq experiments included KDM5C binding in wild-type and *Kdm5c*-KO  
500 EpiLCs<sup>41</sup> (available at GEO: GSE96797) and mouse primary neuron cultures (PNCs) from the cortex  
501 and hippocampus<sup>12</sup> (available at GEO: GSE61036). ChIP-seq of histone 3 lysine 4 dimethylation (H3K4me2)  
502 in male wild-type and *Kdm5c*-KO EpiLCs<sup>41</sup> is also available at GEO: GSE96797. ChIP-seq of histone 3 lysine  
503 4 trimethylation (H3K4me3) in wild-type and *Kdm5c*-KO male amygdala<sup>22</sup> are available at GEO: GSE127817.

504 **Data analysis**

505 Scripts used to generate the results, tables, and figures of this study are available via the GitHub  
506 repository: [https://github.com/kbonefas/KDM5C\\_Germ\\_Mechanism](https://github.com/kbonefas/KDM5C_Germ_Mechanism)

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522 **Author contributions**

523 K.M.B. and S.I. conceived the study and designed the experiments. I.V. generated the ESC and exEpiLC  
524 WGBS data. K.M.B performed all data analysis and all other experiments. The manuscript was written by  
525 K.M.B and S.I., and edited by K.M.B, S.I., and I.V.

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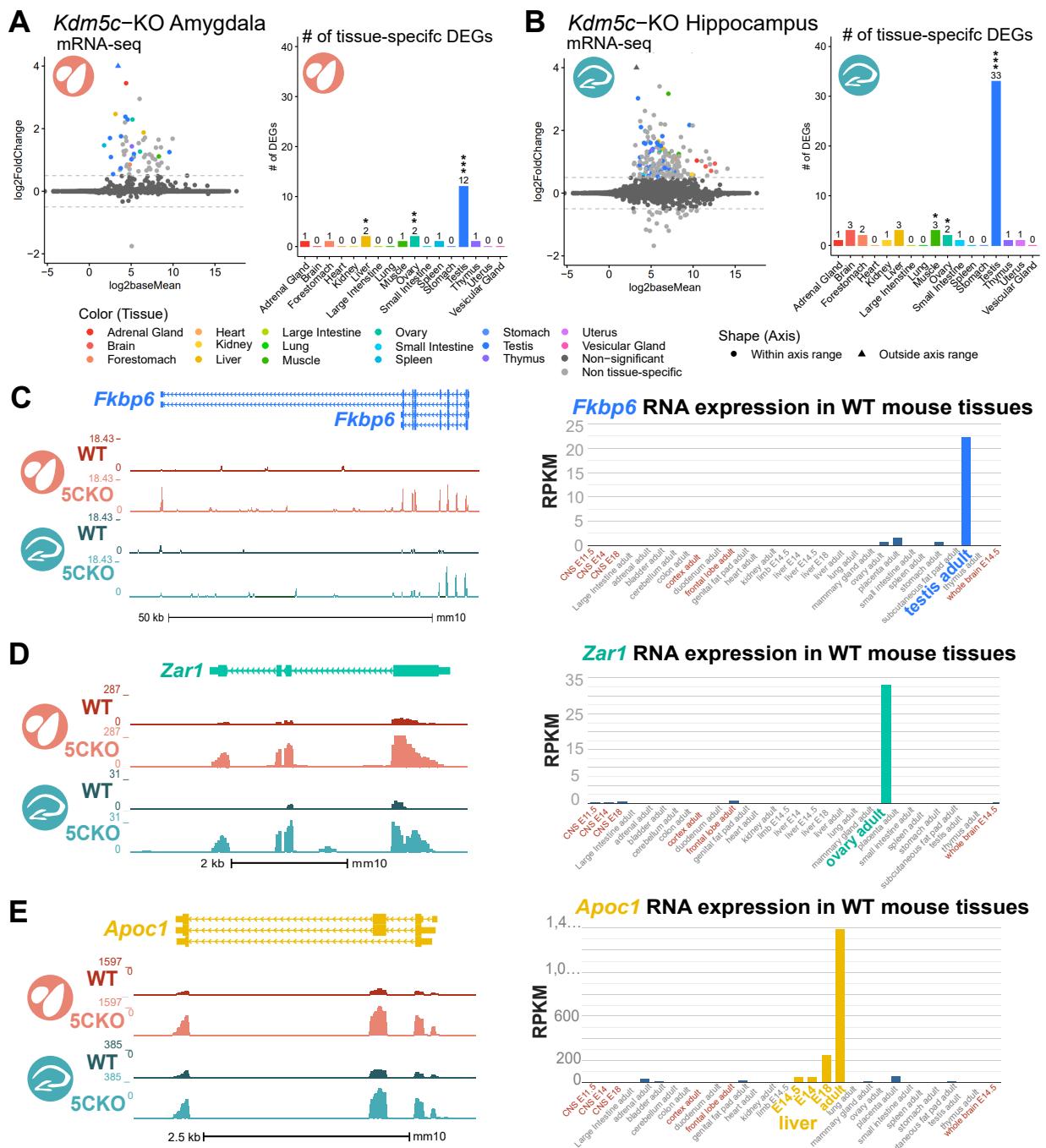
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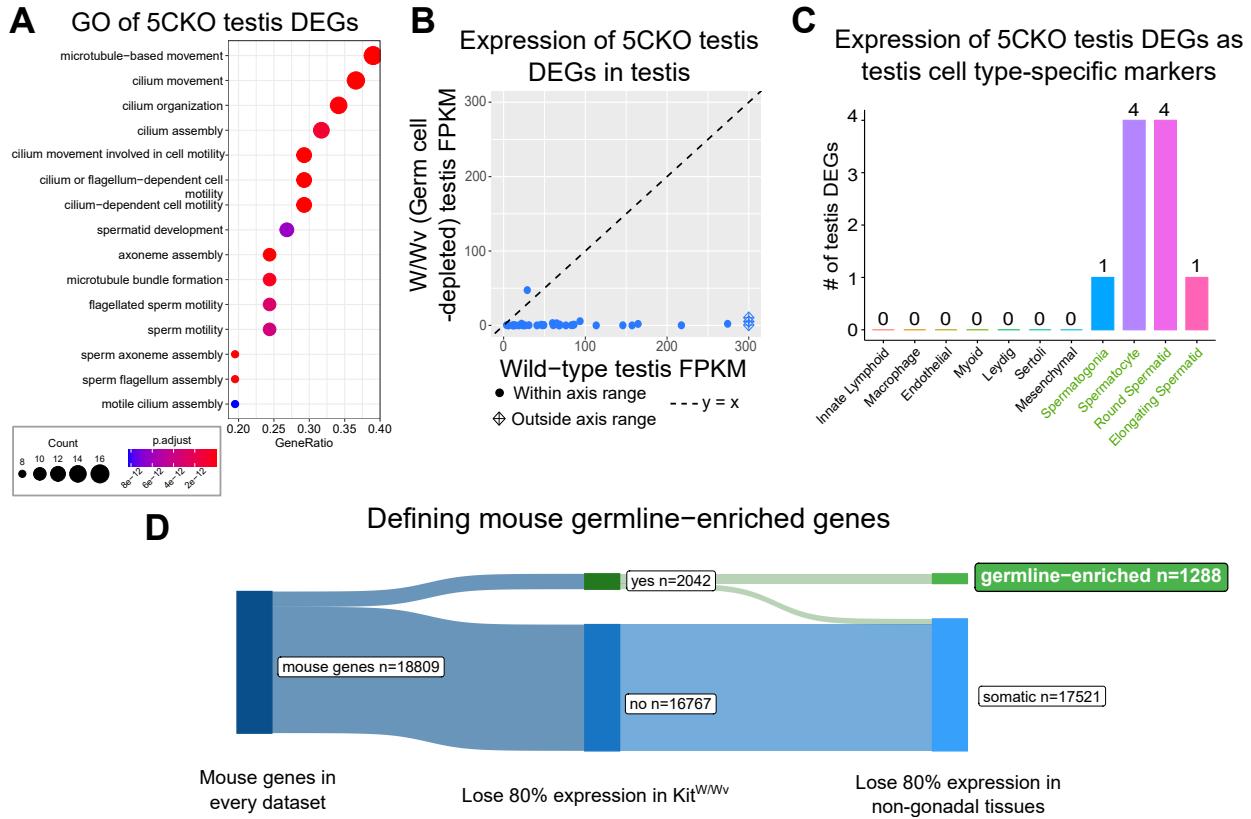
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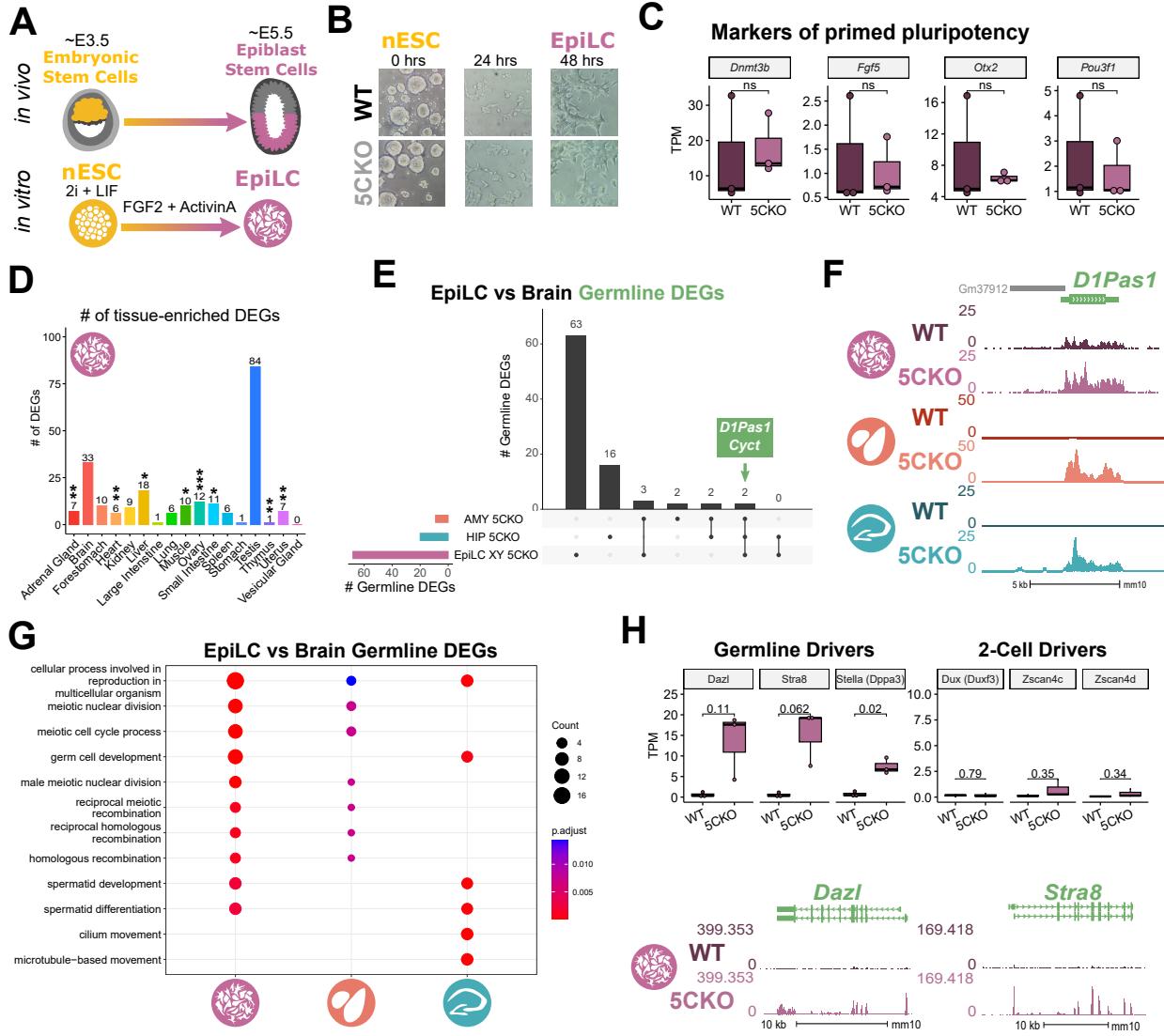
715 **Figures and Tables**



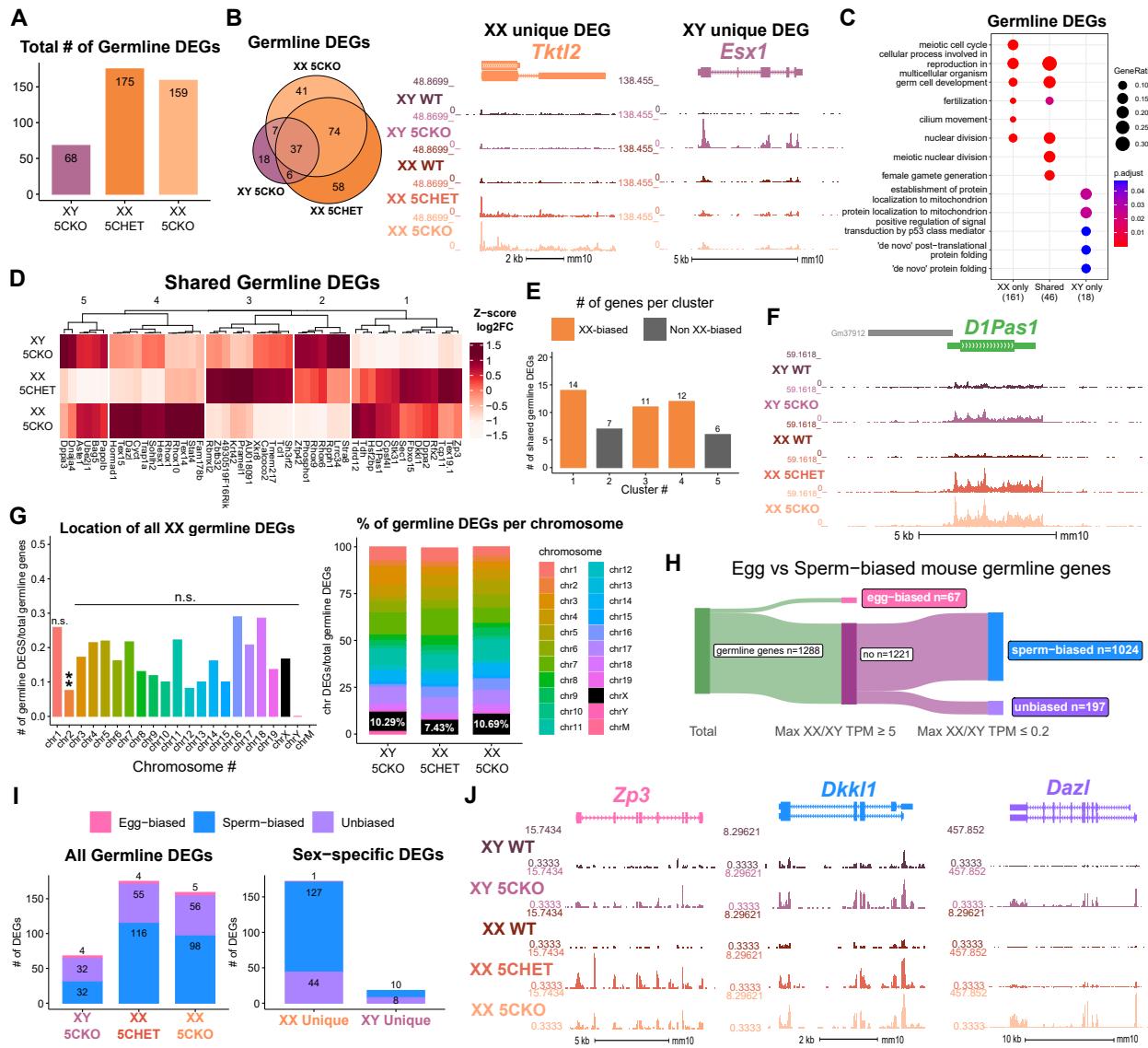
**Figure 1: Tissue-enriched genes are misexpressed in the *Kdm5c*-KO brain.** **A-B.** Expression of tissue-enriched genes (Li et al 2017) in the male *Kdm5c*-KO amygdala (A) and hippocampus (B). Left - MA plot of mRNA-sequencing. Right - Number of tissue-enriched differentially expressed genes (DEGs). \* p<0.05, \*\* p<0.01, \*\*\* p<0.001, Fisher's Exact Test. **C.** Left - UCSC browser view of an example aberrantly expressed testis-enriched DEG, *FK506 binding protein 6* (*Fkbp6*) in the wild-type (WT) and *Kdm5c*-KO (5CKO) amygdala (red) and hippocampus (teal) (Average, n = 4). Right - Expression of *Cyct* in wild-type tissues from NCBI Gene, with testis highlighted in blue and brain tissues highlighted in red. **D.** Left - UCSC browser view of an example ovary-enriched DEG, *Zygotic arrest 1* (*Zar1*). Right - Expression of *Zar1* in wild-type tissues from NCBI Gene, with ovary highlighted in teal and brain tissues highlighted in red. **E.** Left - UCSC browser view of an example liver-enriched DEG, *Apolipoprotein C-I* (*Apoc1*). Right - Expression of *Apoc1* in wild-type tissues from NCBI Gene, with liver highlighted in orange and brain tissues highlighted in red.



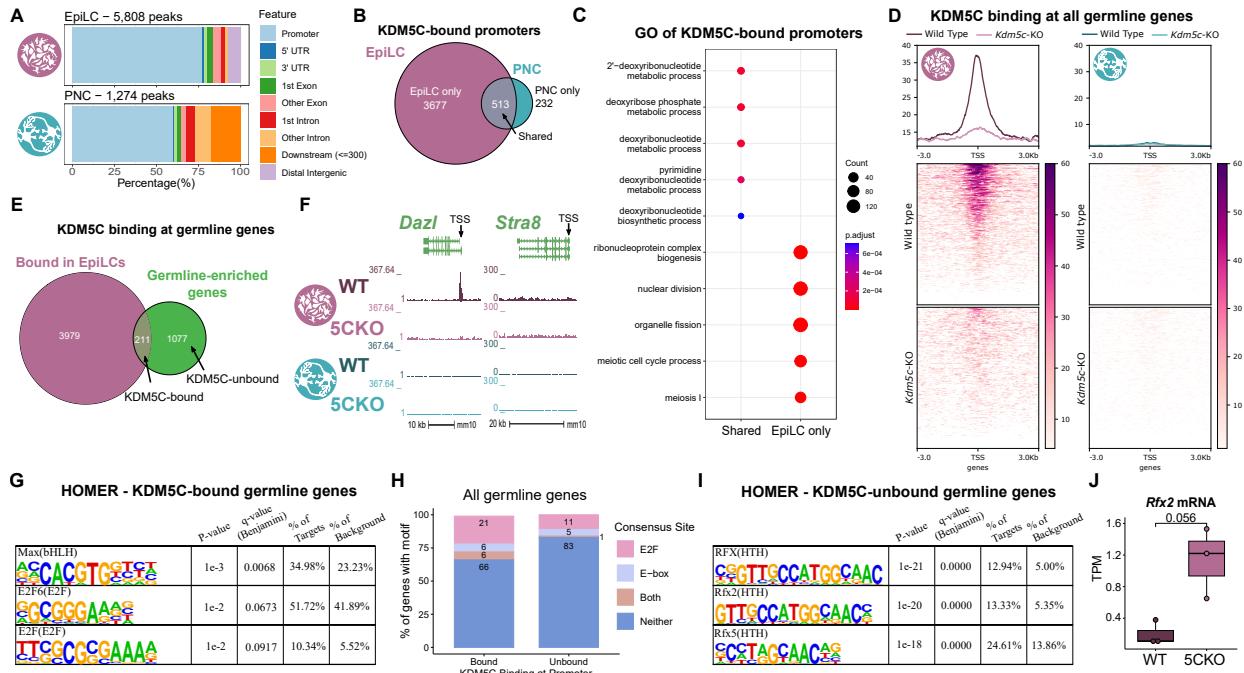
**Figure 2: Aberrant transcription of germline genes in the *Kdm5c*-KO in the brain.** **A.** enrichPlot gene ontology (GO) of *Kdm5c*-KO amygdala and hippocampus testis-enriched DEGs **B.** Expression of testis DEGs in wild-type (WT) testis versus germ cell-depleted (W/Wv) testis (Mueller et al 2013). Expression is in Fragments Per Kilobase of transcript per Million mapped reads (FPKM). **C.** Number of testis DEGs that were classified as cell-type specific markers in a single cell RNA-seq dataset of the testis (Green et al 2018). Germline cell types are highlighted in green, somatic cell types in black. **D.** Sankey diagram of mouse genes filtered for germline enrichment based on their expression in wild-type and W/Wv mice and in adult mouse non-gonadal tissues (Li et al 2017).



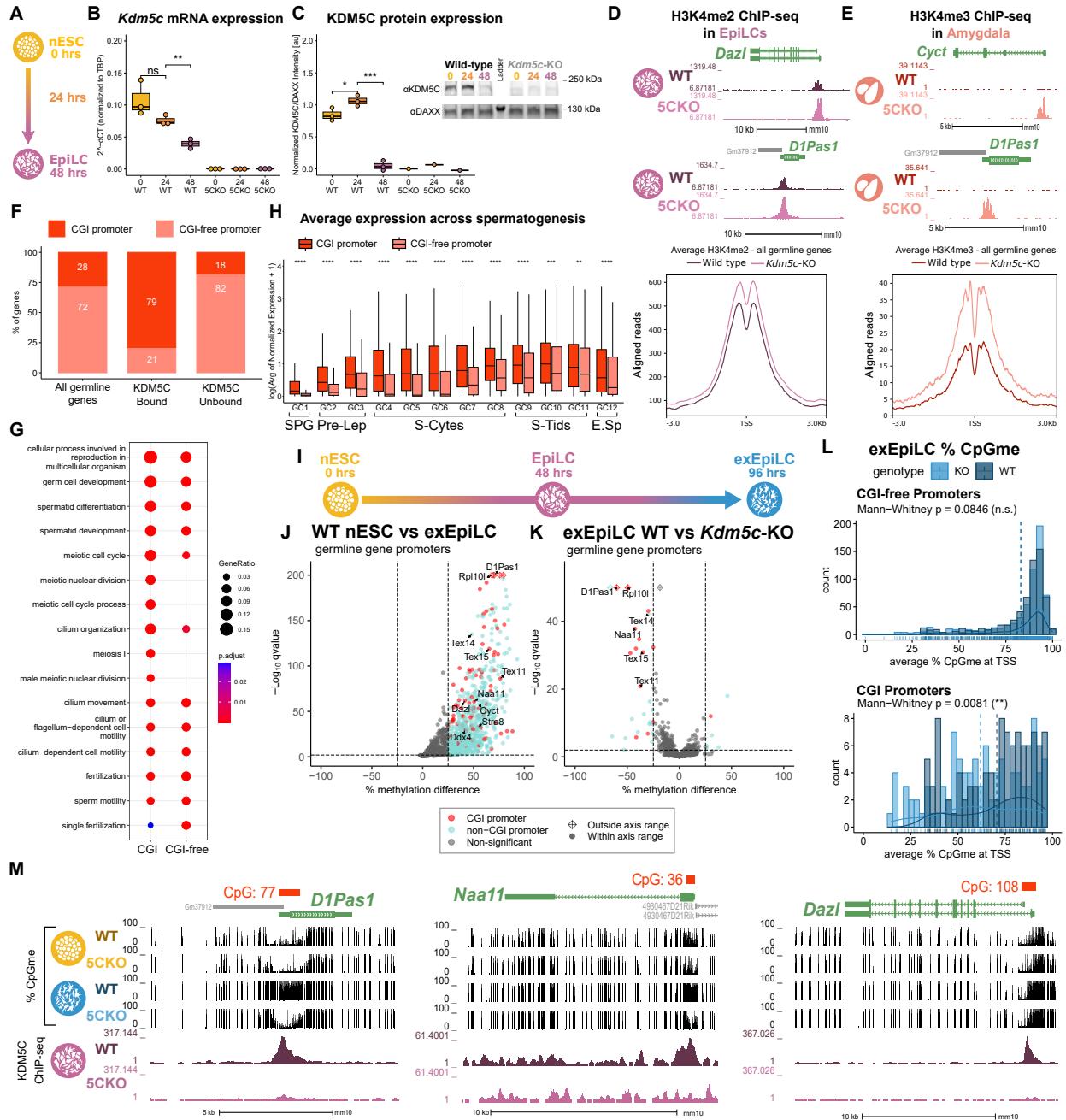
**Figure 3: Kdm5c-KO epiblast-like cells express key drivers of germline identity** **A.** Top - Diagram of *in vivo* differentiation of embryonic stem cells (ESCs) of the inner cell mass into epiblast stem cells. Bottom - *in vitro* differentiation of ESCs into epiblast-like cells (EpiLCs). **B.** Representative images of male wild-type (WT) and *Kdm5c*-KO (5CKO) cells during ESC to EpiLC differentiation. Brightfield images taken at 20X. **C.** No significant difference in primed pluripotency marker expression in wild-type versus *Kdm5c*-KO EpiLCs. Welch's t-test, expression in transcripts per million (TPM). **D.** Number of tissue-enriched differentially expressed genes (DEGs) in *Kdm5c*-KO EpiLCs. \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , Fisher's Exact Test. **E.** Upset plot displaying the overlap of germline DEGs expressed in *Kdm5c*-KO EpiLCs, amygdala (AMY), and hippocampus (HIP) RNA-seq datasets. **F.** UCSC browser view of an example germline gene, *D1Pas1*, that is dysregulated *Kdm5c*-KO EpiLCs (top, purple. Average,  $n = 3$ ), amygdala (middle, red. Average,  $n = 4$ ), and hippocampus (bottom, blue. Average,  $n = 4$ ). **G.** enrichPlot gene ontology analysis comparing enriched biological processes for *Kdm5c*-KO EpiLC, amygdala, and hippocampus germline DEGs. **H.** Top left - Example germline identity DEGs unique to EpiLCs. Top right - Example 2-cell genes that are not dysregulated in *Kdm5c*-KO EpiLCs. p-values for Welch's t-test. Bottom - UCSC browser view of *Dazl* and *Stra8* expression in wild-type and *Kdm5c*-KO EpiLCs (Average,  $n = 3$ ).



**Figure 4: Chromosomal sex influences *Kdm5c*-KO germline gene misexpression.** **A.** Total number of germline-enriched RNA-seq DEGs for male hemizygous *Kdm5c* knockout EpiLCs (XY 5CKO, purple), female heterozygous *Kdm5c* knockout (XX 5CHET, orange), female homozygous *Kdm5c* knockout (XX 5CKO, light orange) EpiLCs. **B.** Left - Eulerr overlap of *Kdm5c* mutant male and female EpiLC germline DEGs. Right - Example of germline DEGs unique to females or males, *Tktl2* and *Esx1*. **C.** enrichPlot gene ontology analysis comparing enriched biological processes for germline DEGs shared between *Kdm5c* mutant males and females (Shared), or unique to one sex (XX only or XY only). **D.** Heatmap of germline DEGs shared between male and female mutants. Color is the average log 2 fold change from sex-matched wild-type, z-scored across rows. **E.** Number of genes within each cluster from D. Clusters with higher expression in females compared to males (XX-biased) highlighted in orange. **F.** UCSC browser view of a male and female shared germline DEG *D1Pas1* that is more highly expressed in female mutants (Average, n = 3). **G.** Left - Number of all female germline DEGs located on each chromosome over the total number of germline-enriched genes on that chromosome. P-values for Fisher Exact Test, \*\* p < 0.01, n.s. non-significant. Germline DEGs were only significant for chromosome 2, in which they were significantly depleted. Right - Percentage of germline DEGs that lie on each chromosome for each *Kdm5c* mutant. X chromosome highlighted in black. **H.** Sankey diagram classifying egg-biased (pink) and sperm-biased (blue) and unbiased (purple) mouse germline-enriched genes. **I.** Number of egg, sperm, or unbiased germline DEGs for male and female *Kdm5c* mutants. **J.** UCSC browser view of egg-biased (*Zp3*), sperm-biased (*Dkk1*), and unbiased (*Dazl*) germline genes dysregulated in both male and female *Kdm5c* mutants (Average of n = 3).



**Figure 5: KDM5C binds to a subset of germline gene promoters during early embryogenesis.** **A.** ChIPseeker localization of KDM5C peaks at different genomic regions in EpiLCs (top) and hippocampal and cortex primary neuron cultures (PNCs, bottom). **B.** Overlap of genes with KDM5C bound to their promoters ( $TSS \pm 500$ ) in EpiLCs (purple) and PNCs (blue). **C.** Gene ontology (GO) comparison of genes with KDM5C bound to their promoter in EpiLCs and PNCs. Genes were classified as either bound in EpiLCs only (EpiLC only), unique to PNCs (PNC only, no significant ontologies) or bound in both PNCs and EpiLCs (Shared). **D.** Average KDM5C binding around the transcription start site (TSS) of all germline-enriched genes in EpiLCs (left) and PNCs (right). **E.** Eulerr overlap of germline-enriched genes (green) with significant KDM5C binding at their promoter in EpiLCs (purple). **F.** Example KDM5C ChIP-seq signal around the *Dazl* TSS but not *Stra8* in EpiLCs. **G.** HOMER motif analysis of all KDM5C-bound germline gene promoters, highlighting significant enrichment of MAX, E2F6, and E2F motifs. **H.** Number of all gene promoters bound or unbound by KDM5C with instances of the E2F or E-box consensus sequence. **I.** HOMER motif analysis of all KDM5C-unbound germline gene promoters, highlighting significant enrichment of RFX family transcription factor motifs. **J.** Expression of RNA-seq DEG *Rfx2* in wild-type and *Kdm5c*-KO EpiLCs. P-value of Welch's t-test, expression in transcripts per million (TPM).



**Figure 6: KDM5C promotes long-term silencing of germline genes via DNA methylation at CpG islands.** **A.** Diagram of embryonic stem cell (ESC) to epiblast-like cell (EpiLC) differentiation and collection time points. **B.** Real time quantitative PCR (RT-qPCR) of *Kdm5c* mRNA expression in wild-type (WT) and *Kdm5c*-KO (5CKO) ESCs at 0, 24, and 48 hours of differentiation into EpiLCs. Expression calculated in comparison to TBP mRNA expression ( $2^{-\Delta\Delta CT}$ ). \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , Welch's t-test. **C.** KDM5C protein expression normalized to DAXX. Quantified intensity using ImageJ (artificial units - au). Right - representative lanes of Western blot for KDM5C and DAXX. \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , Welch's t-test. **D.** Top - Representative UCSC browser view of histone 3 lysine 4 dimethylation (H3K4me2) ChIP-seq signal at two germline genes in wild-type and *Kdm5c*-KO EpiLCs. Bottom - Average H3K4me2 signal at the TSS of all germline-enriched genes in wild-type (dark purple) and *Kdm5c*-KO (light purple) EpiLCs. **E.** Top - Representative UCSC browser view of histone 3 lysine 4 trimethylation (H3K4me3) ChIP-seq signal at two germline genes in the wild-type (WT) and *Kdm5c*-KO (5CKO) adult amygdala. Bottom - Average H3K4me3 signal at the transcription start site (TSS) of all germline-enriched genes in wild-type (dark red) and *Kdm5c*-KO (light red) amygdala. **F.** Percentage of germline genes that harbor CpG islands (CGIs) in their promoters (CGI promoter, red), based on UCSC annotation. Left - percentage of all germline-enriched genes, middle - KDM5C-bound germline genes, and right - KDM5C-unbound germline genes. (Legend continued on next page.)

**Figure 6: KDM5C promotes long-term silencing of germline genes via DNA methylation at CpG islands.** (Legend continued.) **G.** enrichPlot gene ontology analysis of germline genes with (CGI-promoter) or without (CGI-free) CGIs in their promoter. **H.** Expression of germline genes with (CGI-promoter, red) or without (CGI-free, salmon) CGIs in their promoter across stages of spermatogenesis from Green et al 2018. SPG - spermatogonia, Pre-Lep - preleptotene spermatocytes, S-Cytes - meiotic spermatocytes, S-Tids - post-meiotic haploid round spermatids, E.Sp - elongating spermatids. Wilcoxon test, \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , \*\*\*\*  $p < 0.0001$ . **I.** Diagram of ESC to extended EpiLC (exEpiLC) differentiation. **J.** Volcano plot of whole genome bisulfite sequencing (WGBS) comparing CpG methylation (CpGme) at germline gene promoters ( $TSS \pm 500$ ) in wild-type (WT) ESCs versus exEpiLCs. Significantly differentially methylated promoters ( $q < 0.01$ ,  $|methylated difference| > 25\%$ ) with CGIs in red, CGI-free promoters in light blue **K.** Volcano plot of WGBS of wild-type (WT) versus *Kdm5c*-KO exEpiLCs for germline gene promoters. Promoters with CGIs in red, CGI-free promoters in light blue. **L.** Histogram of average percent CpGme at the promoter of germline genes with or without CGIs. Wild-type in navy and *Kdm5c*-KO (KO) in light blue. Dashed lines are average methylation for each genotype, p-values for Mann-Whitney U test. **M.** UCSC browser view of germline genes, showing UCSC annotated CGI with number of CpGs, representative CpGme in wild-type (WT) and *Kdm5c*-KO (5CKO) ESCs and exEpiLCs, and KDM5C ChIP-seq signal in EpiLCs.