

1 The X-linked intellectual disability gene KDM5C is a
2 sex-biased brake against germline programs in somatic
3 lineages

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11 Abstract

12 The division of labor among cellular lineages is a pivotal step in the evolution of multicellularity. In
13 mammals, the soma-germline boundary is formed during early embryogenesis, when genes that drive
14 germline identity are repressed in somatic lineages through DNA and histone modifications at promoter CpG
15 islands (CGIs). Somatic misexpression of germline genes is a signature of cancer and observed in select
16 neurodevelopmental disorders. However, it is currently unclear if all germline genes use the same repressive
17 mechanisms and if factors like development and sex influence their dysregulation. Here, we examine how
18 cellular context influences the formation of somatic tissue identity in mice lacking lysine demethylase 5c
19 (KDM5C), an X chromosome eraser of histone 3 lysine 4 di and tri-methylation (H3K4me2/3). We found male
20 *Kdm5c* knockout (-KO) mice aberrantly express many tissue-specific genes within the brain, the majority of
21 which are unique to the germline. By developing a comprehensive list of mouse germline-enriched genes,
22 we observed *Kdm5c*-KO cells aberrantly express key drivers of germline fate during early embryogenesis
23 but late-stage spermatogenesis genes within the mature brain. KDM5C binds CGIs within germline gene
24 promoters to facilitate DNA CpG methylation as embryonic stem cells differentiate into epiblast-like cells
25 (EpiLCs). However, the majority of late-stage spermatogenesis genes expressed within the *Kdm5c*-KO brain
26 did not harbor promoter CGIs. These CGI-free germline genes were not bound by KDM5C and instead
27 expressed through ectopic activation by RFX transcription factors. Furthermore, germline gene repression
28 is sexually dimorphic, as female EpiLCs require a higher dose of KDM5C to maintain germline silencing.
29 Altogether, these data revealed distinct regulatory classes of germline genes and sex-biased silencing
30 mechanisms in somatic cells.

31 Introduction

32 The separation of germline and somatic cellular identity is a pivotal step in the evolution of multicellularity
33 and sexual reproduction^{1–4}. In mammals, chromatin regulators decommission germline genes in somatic
34 lineages when the early embryo transitions from naïve to primed pluripotency. Germline gene promoters
35 initially gain repressive histone H2A lysine 119 monoubiquitination (H2AK119ub1)⁵ and histone H3 lysine
36 9 trimethylation (H3K9me3)^{5,6} in embryonic stem cells (ESCs) and are then decorated with DNA CpG
37 methylation (CpGme) at their CpG islands (CGIs) in post-implantation epiblast cells^{6–9}. While the silencing
38 mechanisms for genes that establish germline identity are well characterized, it is unclear if other types
39 of germline genes employ the same silencing mechanisms, such as those involved in the later stages
40 of oogenesis and spermatogenesis. Furthermore, because many studies have focused on the silencing
41 of key marker genes during early male embryonic development, much is unknown about how cellular
42 context (i.e. sex and tissue environment) influences the manifestation of germline gene misexpression.
43 Intriguingly, impaired soma-germline demarcation is a signature of aggressive cancers and observed in
44 select neurodevelopmental disorders (NDDs)^{10–13}. Thus, elucidating how cell context contributes to germline
45 gene dysregulation will reveal novel mechanisms governing these pathologies.

46 Here, we employed genome-wide analyses to explore the loss of tissue identity in mice lacking the
47 chromatin regulator lysine demethylase 5C (KDM5C, also known as SMCX or JARID1C). KDM5C lies on the
48 X chromosome and erases histone 3 lysine 4 di- and trimethylation (H3K4me2/3), a permissive chromatin
49 modification enriched at gene promoters¹⁴. Somatic loss of KDM5C promotes tumorigenicity in a variety of
50 cancer types^{15–17}, while pathogenic germline mutations cause the NDD Intellectual Developmental Disorder,
51 X-linked, Syndromic, Claes-Jensen Type (MRXSCJ, OMIM: 300534). MRXSCJ is more common and
52 severe in males and its neurological phenotypes include intellectual disability, seizures, aberrant aggression,
53 and autistic behaviors^{18–20}. Male *Kdm5c* knockout (-KO) mice recapitulate key MRXSCJ phenotypes,
54 including hyperaggression, increased seizure propensity, social deficits, and learning impairments^{21–23}. RNA
55 sequencing (RNA-seq) of the *Kdm5c*-KO hippocampus revealed ectopic expression of some testis germline
56 genes within the brain²². However, it is unclear if other tissue-specific genes are aberrantly transcribed with
57 KDM5C loss, at what point in development germline gene misexpression begins, what mechanisms underlie
58 their dysregulation, and how KDM5C interacts with other known germline silencing mechanisms.

59 To illuminate KDM5C's role in tissue identity, we characterized the aberrant expression of tissue-enriched
60 genes within the *Kdm5c*-KO brain and epiblast-like stem cells (EpiLCs), an *in vitro* model of the post-
61 implantation embryo. We curated a list of mouse germline-enriched genes, enabling genome-wide analysis
62 of germline gene silencing mechanisms for the first time. We identified two classes of germline genes based
63 on their promoter CpG island content, which are dysregulated with KDM5C loss by distinct mechanisms and
64 in a sex-biased manner.

65 **Results**

66 **Tissue-enriched genes are aberrantly expressed in the *Kdm5c*-KO brain**

67 Previous RNA sequencing (RNA-seq) of the adult male *Kdm5c*-KO hippocampus revealed ectopic
68 expression of some germline genes unique to the testis²². It is currently unknown if the testis is the only
69 tissue type misexpressed in the *Kdm5c*-KO brain. We first systematically tested whether other tissue-specific
70 genes are misexpressed in the male brain with constitutive knockout of *Kdm5c* (*Kdm5c*^{-Y}, 5CKO in figures)²⁴
71 by using a published list of mouse tissue-enriched genes²⁵.

72 We found a large proportion of significantly upregulated genes (DESeq2²⁶, log2 fold change > 0.5, q <
73 0.1) within the male *Kdm5c*-KO amygdala and hippocampus are non-brain, tissue-specific genes (Amygdala:
74 0/0 up DEGs, NaN% ; Hippocampus: 0/0 up DEGs, NaN%) (Figure 1A-B, Supplementary Table 1). For both
75 the amygdala and hippocampus, the majority of tissue-enriched differentially expressed genes (DEGs) were
76 testis genes (Figure 1A-B). Even though the testis has the largest total number of tissue-enriched genes
77 (2,496 genes) compared to any other tissue, testis-enriched DEGs were significantly enriched in both brain
78 regions (Amygdala p = 1.83e-05, Odds Ratio = 5.13; Hippocampus p = 4.26e-11, Odds Ratio = 4.45, Fisher's
79 Exact Test). An example of a testis-enriched gene misexpressed in the *Kdm5c*-KO brain is *FK506 binding*
80 *protein 6 (Fkbp6)*, a known regulator of PIWI-interacting RNAs (piRNAs) and meiosis^{27,28} (Figure 1C).

81 Interestingly, we also observed significant enrichment of ovary-enriched genes in both the amygdala
82 and hippocampus (Amygdala p = 0.00574, Odds Ratio = 18.7; Hippocampus p = 0.048, Odds Ratio = 5.88,
83 Fisher's Exact Test) (Figure 1A-B). Ovary-enriched DEGs included *Zygotic arrest 1 (Zar1)*, which sequesters
84 mRNAs in oocytes for meiotic maturation²⁹ (Figure 1D). Given that the *Kdm5c*-KO mice we analyzed are
85 male, these data demonstrate that the ectopic expression of gonad-enriched genes is independent of
86 organismal sex.

87 Although not consistent across brain regions, we also found significant enrichment of genes biased
88 towards two non-gonadal tissues - the liver (Amygdala p = 0.04, Odds Ratio = 6.58, Fisher's Exact Test)
89 and muscle (Hippocampus p = 0.01, Odds Ratio = 6.95, Fisher's Exact Test) (Figure 1A-B). These include
90 *Apolipoprotein C-I (Apoc1)*, a lipoprotein metabolism and transport gene³⁰ (Figure 1E, see Discussion).

91 Our analysis of oligo(dT)-primed libraries²⁴ indicates aberrantly expressed mRNAs are polyadenylated
92 and spliced into mature transcripts in the *Kdm5c*-KO brain (Figure 1C-E). Of note, we observed little to no
93 dysregulation of brain-enriched genes (Amygdala p = 1, Odds Ratio = 1.22; Hippocampus p = 0.74, Odds
94 Ratio = 1.22, Fisher's Exact Test), despite the fact these are brain samples and the brain has the second
95 highest total number of tissue-enriched genes (708 genes). Altogether, these results suggest the aberrant
96 expression of tissue-enriched genes within the brain is a major effect of KDM5C loss.

97 **Germline genes are misexpressed in the *Kdm5c*-KO brain**

98 *Kdm5c*-KO brain expresses testicular germline genes²² (Figure 1), however the testis also contains
99 somatic cells that support hormone production and germline functions. To determine if *Kdm5c*-KO results
100 in ectopic expression of testicular somatic genes, we first evaluated the known functions of testicular
101 DEGs through gene ontology. We found *Kdm5c*-KO testis-enriched DEGs had high enrichment of germline-
102 relevant ontologies, including spermatid development (GO: 0007286, p.adjust = 6.2e-12) and sperm axoneme
103 assembly (GO: 0007288, p.adjust = 2.45e-14) (Figure 2A, Supplementary Table 1).

104 We then evaluated *Kdm5c*-KO testicular DEG expression in wild-type testes versus testes with germ cell
105 depletion³¹, which was accomplished by heterozygous *W* and *Wv* mutations in the enzymatic domain of *c-Kit*
106 (*Kit*^{W/Wv})³². Almost all *Kdm5c*-KO testis-enriched DEGs lost expression with germ cell depletion (Figure 2B).
107 We then assessed testis-enriched DEG expression in a published single cell RNA-seq dataset that identified
108 cell type-specific markers within the testis³³. Some *Kdm5c*-KO testis-enriched DEGs were classified as
109 specific markers for different germ cell developmental stages (e.g. spermatogonia, spermatocytes, round
110 spermatids, and elongating spermatids), yet none marked somatic cells (Figure 2C). Together, these data
111 demonstrate that the *Kdm5c*-KO brain aberrantly expresses germline genes but not somatic testicular genes,
112 reflecting an erosion of the soma-germline boundary.

113 As of yet, research on germline gene silencing mechanisms has focused on a handful of key genes rather
114 than assessing germline gene suppression genome-wide, due to the lack of a comprehensive gene list.
115 We therefore generated a list of mouse germline-enriched genes using RNA-seq datasets of *Kit*^{W/Wv} mice
116 that included males and females at embryonic day 12, 14, and 16³⁴ and adult male testes³¹. We defined
117 genes as germline-enriched if their expression met the following criteria: 1) their expression is greater than
118 1 FPKM in wild-type gonads 2) their expression in any non-gonadal tissue of adult wild type mice²⁵ does
119 not exceed 20% of their maximum expression in the wild-type germline, and 3) their expression in the germ
120 cell-depleted gonads, for any sex or time point, does not exceed 20% of their maximum expression in the
121 wild-type germline. These criteria yielded 1,288 germline-enriched genes (Figure 2D), which was hereafter
122 used as a resource to globally characterize germline gene misexpression with *Kdm5c* loss (Supplementary
123 Table 2).

124 ***Kdm5c*-KO epiblast-like cells aberrantly express key regulators of germline identity**

125 Germ cells are typically distinguished from somatic cells soon after the embryo implants into the uterine
126 wall^{35,36}, when germline genes are silenced in epiblast stem cells that will form the somatic tissues³⁷. This
127 developmental time point can be modeled *in vitro* through differentiation of naïve embryonic stem cells
128 (nESCs) into epiblast-like stem cells (EpiLCs) (Figure 3A)^{38,39}. While some germline-enriched genes are
129 also expressed in nESCs and in the 2-cell stage^{40–42}, they are silenced as they differentiate into EpiLCs^{6,7}.
130 Therefore, we tested if KDM5C was necessary for the initial silencing of germline genes in somatic lineages

131 by evaluating the impact of *Kdm5c* loss in male EpiLCs.
132 *Kdm5c*-KO cell morphology during ESC to EpiLC differentiation appeared normal (Figure 3B) and EpiLCs
133 properly expressed markers of primed pluripotency, such as *Dnmt3b*, *Fgf5*, *Pou3f1*, and *Otx2* (Figure 3C). We
134 then identified tissue-enriched DEGs in a RNA-seq dataset of wild-type and *Kdm5c*-KO EpiLCs⁴³ (DESeq2,
135 log2 fold change > 0.5, q < 0.1, Supplementary Table 3). Similar to the *Kdm5c*-KO brain, we observed
136 general dysregulation of tissue-enriched genes, with the largest number of genes belonging to the brain and
137 testis, although they were not significantly enriched (Figure 3D). Using our list of mouse germline-enriched
138 genes assembled above, we identified 68 germline genes misexpressed in male *Kdm5c*-KO EpiLCs.

139 We then compared EpiLC germline DEGs to those expressed in the *Kdm5c*-KO brain to determine if
140 germline genes are constitutively dysregulated or change over the course of development. The majority of
141 germline DEGs were unique to either EpiLCs or the brain, with only *D1Pas1* and *Cyct* shared across all
142 tissue/cell types (Figure 3E-F). EpiLC germline DEGs had particularly high enrichment of meiosis-related
143 gene ontologies when compared to the brain (Figure 3G, Supplementary Table 3), such as meiotic cell
144 cycle process (GO:1903046, p.adjust = 2.2e-07) and meiotic nuclear division (GO:0140013, p.adjust
145 = 1.37e-07). While there was modest enrichment of meiotic gene ontologies in both brain regions, the
146 *Kdm5c*-KO hippocampus primarily expressed late-stage spermatogenesis genes involved in sperm axoneme
147 assembly (GO:0007288, p.adjust = 0.00621) and sperm motility (GO:0097722, p.adjust = 0.00612).

148 Notably, DEGs unique to *Kdm5c*-KO EpiLCs included key drivers of germline identity, such as *Stimulated*
149 *by retinoic acid 8* (*Stra8*: log2 fold change = 3.73, q = 2.17e-39) and *Deleted in azoospermia like* (*Dazl*):
150 log2 fold change = 3.36, q = 3.19e-12) (Figure 3H). These genes are typically expressed when a subset
151 of epiblast stem cells become primordial germ cells (PGCs) and then again in mature germ cells to trigger
152 meiotic gene expression programs^{44–46}. Of note, some germline genes, including *Dazl*, are also expressed
153 in the two-cell embryo^{41,47}. However, we did not see derepression of two-cell stage-specific genes, like
154 *Duxf3* (*Dux*) (log2 fold change = -0.282, q = 0.337) and *Zscan4d* (log2 fold change = 0.25, q = 0.381) (Figure
155 3H, Supplementary Table 3), indicating *Kdm5c*-KO EpiLCs do not revert back to a 2-cell state. Altogether,
156 *Kdm5c*-KO EpiLCs express key drivers of germline identity and meiosis while the brain primarily expresses
157 spermiogenesis genes, indicating germline gene misexpression mirrors germline development during the
158 progression of somatic development.

159 **Female epiblast-like cells have heightened germline gene misexpression with *Kdm5c* 160 loss**

161 It is currently unknown if the misexpression of germline genes is influenced by sex, as previous studies
162 on germline gene repressors have focused on male cells^{5,6,8,48,49}. Sex is particularly pertinent in the case
163 of KDM5C because it partially escapes X chromosome inactivation (XCI), resulting in a higher dosage in
164 females^{50–53}. We therefore explored the impact of chromosomal sex upon germline gene suppression by

165 comparing their dysregulation in male *Kdm5c* hemizygous knockout (*Kdm5c*^{-/Y}, XY *Kdm5c*-KO, XY 5CKO),
166 female homozygous knockout (*Kdm5c*^{-/-}, XX *Kdm5c*-KO, XX 5CKO), and female heterozygous knockout
167 (*Kdm5c*^{+/+}, XX *Kdm5c*-HET, XX 5CHET) EpiLCs⁴³.

168 In EpiLCs, homozygous and heterozygous *Kdm5c* knockout females expressed over double the number
169 of germline-enriched genes than hemizygous males (Figure 4A, Supplementary Table 3). While the majority
170 of germline DEGs in *Kdm5c*-KO males were also dysregulated in females (74%), many were sex-specific,
171 such as *Tktl2* and *Esx1* (Figure 4B). We then compared the known functions of germline genes dysregulated
172 uniquely in males and females or misexpressed in all samples (Figure 4C, Supplementary Table 3). Female-
173 specific germline DEGs were enriched for meiotic (GO:0051321 - meiotic cell cycle, p.adjust = 7.81E-14) and
174 flagellar (GO:0003341 - cilium movement, p.adjust = 4.87E-06) functions, while male-specific DEGs had roles
175 in mitochondrial and cell signaling (GO:0070585 - protein localization to mitochondrion, p.adjust = 0.025).

176 The majority of germline genes expressed in both sexes were more highly dysregulated in females
177 compared to males (Figure 4D-F). This increased degree of dysregulation in females, along with the
178 increased total number of germline genes, indicates females are more sensitive to losing KDM5C-mediated
179 germline gene suppression. Heightened germline gene dysregulation in females could be due to impaired
180 XCI in *Kdm5c* mutants⁴³, as many spermatogenesis genes lie on the X chromosome^{54,55}. However, female
181 germline DEGs were not biased towards the X chromosome (p = 1, Odds Ratio = 0.96, Fisher's Exact Test)
182 and females had a similar overall proportion of germline DEGs belonging to the X chromosome as males
183 (XY *Kdm5c*-KO - 10.29%, XX *Kdm5c*-HET - 7.43%, XX *Kdm5c*-KO - 10.59%) (Figure 4G). The majority of
184 germline DEGs instead lie on autosomes for both male and female *Kdm5c* mutants (Figure 4G). Thus, while
185 female EpiLCs are more prone to germline gene misexpression with KDM5C loss, it is likely independent of
186 XCI defects.

187 **Germline gene misexpression in *Kdm5c* mutants is independent of germ cell sex**

188 Although many germline genes have shared functions in the male and female germline, e.g. PGC
189 formation, meiosis, and genome defense, some have unique or sex-biased expression. Therefore, we
190 wondered if *Kdm5c* mutant males would primarily express sperm genes while mutant females would primarily
191 express egg genes. To comprehensively assess whether germline gene sex corresponds with *Kdm5c*
192 mutant sex, we first filtered our list of germline-enriched genes for egg and sperm-biased genes (Figure 4,
193 Supplementary Table 2). We defined germ cell sex-biased genes as those whose expression in the opposite
194 sex, at any time point, is no greater than 20% of the gene's maximum expression in a given sex. This
195 criteria yielded 67 egg-biased, 1,024 sperm-biased, and 197 unbiased germline-enriched genes. We found
196 regardless of sex, egg, sperm, and unbiased germline genes were dysregulated in all *Kdm5c* mutants at
197 similar proportions (Figure 4I-J). Furthermore, germline genes dysregulated exclusively in either male or
198 female mutants were also not biased towards their corresponding germ cell sex (Figure 4I). Altogether, these
199 results demonstrate sex differences in germline gene dysregulation is not due to sex-specific activation of

200 sperm or egg transcriptional programs.

201 KDM5C binds to a subset of germline gene promoters during early embryogenesis

202 KDM5C binds to the promoters of several germline genes in embryonic stem cells (ESCs) but not in
203 neurons^{22,56}. However, due to the lack of a comprehensive list of germline-enriched genes, it is unclear if
204 KDM5C is enriched at germline gene promoters, what types of germline genes KDM5C regulates, and if its
205 binding is maintained at any germline genes in neurons.

206 To address these questions, we analyzed KDM5C chromatin immunoprecipitation followed by DNA
207 sequencing (ChIP-seq) datasets in EpiLCs⁴³ and primary forebrain neuron cultures (PNCs)²¹ (MACS2 q <
208 0.1, fold enrichment > 1, and removal of false-positive *Kdm5c*-KO peaks). EpiLCs had a higher total number
209 of high-confidence KDM5C peaks than PNCs (EpiLCs: 5,808, PNCs: 1,276). KDM5C was primarily localized
210 to gene promoters in both cell types (promoters = transcription start site (TSS) ± 500 bp, EpiLCs: 4,190,
211 PNCs: 745), although PNCs showed increased localization to non-promoter regions (Figure 5A).

212 The majority of promoters bound by KDM5C in PNCs were also bound in EpiLCs (513 shared promoters),
213 however a large portion of gene promoters were bound by KDM5C only in EpiLCs (3,677 EpiLC only
214 promoters) (Figure 5B). Genes bound by KDM5C in both PNCs and EpiLCs were enriched for functions
215 involving nucleic acid turnover, such as deoxyribonucleotide metabolic process (GO:0009262, p.adjust =
216 8.28e-05) (Figure 5C, Supplementary Table 4). Germline ontologies were enriched only in EpiLC-specific,
217 KDM5C-bound promoters, such as meiotic nuclear division (GO: 0007127 p.adjust = 6.77e-16) (Figure 5C).
218 There were no significant ontologies for PNC-specific KDM5C target genes. Using our mouse germline gene
219 list, we observed evident KDM5C signal around the TSS of many germline genes in EpiLCs, but not in PNCs
220 (Figure 5D). Based on our ChIP-seq peak cut-off criteria, KDM5C was highly enriched at 211 germline gene
221 promoters in EpiLCs (16.4% of all germline genes) (Figure 5E, Supplementary Table 2). Of note, KDM5C
222 was only bound to about one third of RNA-seq DEG promoters unique to EpiLCs or the brain (EpiLC only
223 DEGs: 34.9%, Brain only DEGs: 30%) (Supplementary Figure 1A-C). Representative examples of EpiLC
224 DEGs bound and unbound by KDM5C in EpiLCs are *Dazl* and *Stra8*, respectively (Figure 5F). However,
225 the four of the five germline genes dysregulated in both EpiLCs and the brain were bound by KDM5C in
226 EpiLCs (*D1Pas1*, *Hsf2bp*, *Cyct*, and *Stk31*) (Supplementary Figure 1A). Together, these results demonstrate
227 KDM5C is recruited to a subset of germline genes in EpiLCs, including meiotic genes, but does not directly
228 regulate germline genes in neurons. Furthermore, the majority of germline mRNAs expressed in *Kdm5c*-KO
229 cells are dysregulated independent of direct KDM5C recruitment to their gene promoters, however genes
230 dysregulated across *Kdm5c*-KO development are often direct KDM5C targets.

231 Many germline-specific genes are suppressed by the polycomb repressive complex 1.6 (PRC1.6), which
232 contains the transcription factor heterodimers E2F6/DP1 and MGA/MAX that respectively bind E2F and
233 E-box motifs within germline gene promoters^{5,6,8,42,48,49,57-59}. PRC1.6 members may recruit KDM5C to
234 germline gene promoters²², given their association with KDM5C in HeLa cells and ESCs^{47,60}. We thus

235 used HOMER⁶¹ to identify transcription factor motifs enriched at KDM5C-bound or unbound germline gene
236 promoters (TSS ± 500 bp, q-value < 0.1, Supplementary Table 4). MAX and E2F6 binding sites were
237 significantly enriched at germline genes bound by KDM5C in EpiLCs (MAX q-value: 0.0068, E2F6 q-value:
238 0.0673, E2F q-value: 0.0917), but not at germline genes unbound by KDM5C (Figure 5G). One third of
239 KDM5C-bound promoters contained the consensus sequence for either E2F6 (E2F, 5'-TCCCGC-3'), MGA
240 (E-box, 5'-CACGTG-3'), or both, but only 17% of KDM5C-unbound genes contained these motifs (Figure 5H).
241 KDM5C-unbound germline genes were instead enriched for multiple RFX transcription factor binding sites
242 (RFX q-value < 0.0001, RFX2 q-value < 0.0001, RFX5 q-value < 0.0001) (Figure 5I, Supplementary figure
243 1D). RFX transcription factors bind X-box motifs⁶² to promote ciliogenesis^{63,64} and among them is RFX2, a
244 central regulator of post-meiotic spermatogenesis^{65,66}. Although *Rfx2* is also not a direct target of KDM5C
245 (Supplementary Figure 1E), RFX2 mRNA is derepressed in *Kdm5c*-KO EpiLCs (Figure 5J). Thus, RFX2 is a
246 candidate transcription factor for driving the ectopic expression of many KDM5C-unbound germline genes in
247 *Kdm5c*-KO cells.

248 **KDM5C is recruited to CpG islands at germline promoters to facilitate *de novo* DNA
249 methylation**

250 Previous work found two germline gene promoters have a marked reduction in DNA CpG methylation
251 (CpGme) in the adult *Kdm5c*-KO hippocampus²². Since histone H3K4me2/3 impede *de novo* CpGme^{67,68},
252 KDM5C's removal of H3K4me2/3 may be required to suppress germline genes. However, KDM5C's catalytic
253 activity was recently shown to be dispensable for suppressing *Dazl* in undifferentiated ESCs⁴⁷. To reconcile
254 these observations, we hypothesized KDM5C erases H3K4me2/3 to promote the initial placement of CpGme
255 at germline gene promoters in EpiLCs.

256 To test this hypothesis, we first characterized KDM5C's expression as naïve ESCs differentiate into
257 EpiLCs (Figure 6A). While *Kdm5c* mRNA steadily decreased from 0 to 48 hours of differentiation (Figure
258 6B), KDM5C protein initially increased from 0 to 24 hours and then decreased to near knockout levels by 48
259 hours (Figure 6C). We then characterized KDM5C's substrates (H3K4me2/3) at germline gene promoters
260 with *Kdm5c* loss using published ChIP-seq datasets^{24,43}. *Kdm5c*-KO samples showed a marked increase in
261 H3K4me2 in EpiLCs (Figure 6D) and H3K4me3 in the amygdala (Figure 6E) around the TSS of germline
262 genes. Together, these data suggest KDM5C acts during the transition between ESCs and EpiLCs to remove
263 H3K4me2/3 at germline gene promoters.

264 Germline genes accumulate CpG methylation (CpGme) at CpG islands (CGIs) during the transition
265 from naïve to primed pluripotency^{7,9,69}. We first examined how many of our germline-enriched genes had
266 promoter CGIs (TSS ± 500 bp) using the UCSC genome browser⁷⁰. Notably, out of 1,288 germline-enriched
267 genes, only 356 (27.64%) had promoter CGIs (Figure 6F, Supplementary Table 2). CGI-containing germline
268 genes had higher enrichment of meiotic gene ontologies compared to CGI-free genes, including meiotic

269 nuclear division (GO:0140013, p.adjust = 2.17e-12) and meiosis I (GO:0007127, p.adjust = 3.91e-10)
270 (Figure 6G, Supplementary Table 5). Germline genes with promoter CGIs were more highly expressed than
271 CGI-free genes across spermatogenesis stages, with highest expression in meiotic spermatocytes (Figure
272 6H). Contrastingly, CGI-free genes only displayed substantial expression in post-meiotic round spermatids
273 (Figure 6H). Although only a minor portion of germline gene promoters contained CGIs, CGIs strongly
274 determined KDM5C's recruitment to germline genes ($p = 2.37e-67$, Odds Ratio = 17.8, Fisher's Exact Test),
275 with 79.15% of KDM5C-bound germline gene promoters harboring CGIs (Figure 6F).

276 To assess how KDM5C loss impacts initial CpGme placement at germline gene promoters, we performed
277 whole genome bisulfite sequencing (WGBS) in male wild-type and *Kdm5c*-KO ESCs and 96-hour extend
278 EpiLCs (exEpiLCs), when germline genes reach peak methylation levels⁶ (Figure 6I). We first identified
279 which germline gene promoters significantly gained CpGme in wild-type cells during nESC to exEpiLCs
280 differentiation (methylKit⁷¹, $q < 0.01$, $|methylation\ difference| > 25\%$, TSS ± 500 bp). In wild-type cells, the
281 majority of germline genes gained substantial CpGme at their promoter during differentiation (60.08%),
282 regardless if their promoter contained a CGI (Figure 6J, Supplementary Table 5).

283 We then identified promoters differentially methylated in wild-type versus *Kdm5c*-KO exEpiLCs (methylKit,
284 $q < 0.01$, $|methylation\ difference| > 25\%$, TSS ± 500 bp, Supplementary Table 5). Of the 48,882 promoters
285 assessed, 274 promoters were significantly hypomethylated and 377 promoters were significantly hyper-
286 methylated with KDM5C loss (Supplementary Figure 2A). Many promoters hyper- and hypomethylated
287 in *Kdm5c*-KO exEpiLCs belonged to genes with unknown functions. However, 10.22% of hypomethyl-
288 ated promoters belonged to germline genes and germline-relevant ontologies like meiotic nuclear division
289 (GO:0140013, p.adjust = 0.012) are significantly enriched (Supplementary Figure 2B, Supplementary Table
290 5). Approximately half of all germline gene promoters hypomethylated in *Kdm5c*-KO exEpiLCs are direct
291 targets of KDM5C in EpiLCs (13 out of 28 hypomethylated promoters).

292 Promoters that showed the most robust loss of CpGme in *Kdm5c*-KO exEpiLCs (lowest q-values) harbored
293 CGIs (Figure 6K). CGI promoters, but not CGI-free promoters, had a significant reduction in CpGme with
294 KDM5C loss as a whole (Figure 6L) (Non-CGI promoters $p = 0.0846$, CGI promoters $p = 0.0081$, Mann-
295 Whitney U test). Significantly hypomethylated promoters included germline genes consistently dysregulated
296 across multiple *Kdm5c*-KO RNA-seq datasets²², such as *D1Pas1* (methylation difference = -60.03%, q-value
297 = 3.26e-153) and *Naa11* (methylation difference = -42.45%, q-value = 1.44e-38) (Figure 6M). Unexpectedly,
298 we observed only a modest reduction in CpGme at *Dazl*'s promoter (methylation difference = -6.525%,
299 q-value = 0.0159) (Figure 6N). Altogether, these results demonstrate KDM5C is recruited to germline gene
300 CGIs in EpiLCs to promote CpGme at those promoters. Furthermore, our data suggest while KDM5C's
301 catalytic activity is required for the repression of some germline genes, CpGme can be placed at others even
302 with elevated H3K4me2/3 around the TSS.

303 **Discussion**

304 In the above study, we demonstrate KDM5C's pivotal role in the development of tissue identity. We first
305 characterized tissue-enriched genes expressed within the mouse *Kdm5c*-KO brain and identified substantial
306 derepression of testis, liver, muscle, and ovary-enriched genes. Testis genes significantly enriched within the
307 *Kdm5c*-KO amygdala and hippocampus are specific to the germline and absent in somatic cells. *Kdm5c*-
308 KO epiblast-like cells (EpiLCs) aberrantly express key drivers of germline identity and meiosis, including
309 *Dazl* and *Stra8*, while the adult brain primarily expresses genes important for late spermatogenesis. We
310 demonstrated that although sex did not influence whether sperm or egg-specific genes were misexpressed,
311 female EpiLCs have heightened germline gene de-repression with KDM5C loss. Germline genes can become
312 aberrantly expressed in *Kdm5c*-KO cells via indirect mechanisms, such as activation through ectopic RFX
313 transcription factors. Finally, we found KDM5C is dynamically regulated during ESC to EpiLC differentiation
314 to promote long-term germline gene silencing through CGI DNA methylation. Therefore, we propose KDM5C
315 plays a fundamental role in the development of tissue identity during early embryogenesis, including the
316 establishment of the soma-germline boundary. By systematically characterizing KDM5C's role in germline
317 gene repression, we unveiled divergent mechanisms governing the misexpression of distinct germline gene
318 classes in somatic lineages.

319 By comparing *Kdm5c* mutant males and females, we revealed germline gene suppression is sexually
320 dimorphic. Female EpiLCs are more severely impacted by loss of KDM5C-mediated germline gene suppression,
321 yet this difference is not due to the large number of germline genes on the X chromosome^{54,55}.
322 Heightened germline gene misexpression in females may be related to females having a higher dose of
323 KDM5C than males, due to its escape from XCI⁵⁰⁻⁵³. Intriguingly, heterozygous knockout females (*Kdm5c*^{+/−})
324 also had over double the number of germline DEGs than hemizygous knockout males (*Kdm5c*^{0/0}), even
325 though their expression of KDM5C should be roughly equivalent to that of wild-type males (*Kdm5c*^{+/+}). Males
326 could partially compensate for KDM5C's loss via the Y-chromosome homolog, KDM5D¹⁴. However, KDM5D
327 has not been reported to regulate germline gene expression. Nevertheless, these results demonstrate
328 germline gene silencing mechanisms differ between males and females, which warrants further study to
329 elucidate the biological ramifications and underlying mechanisms.

330 We found KDM5C is largely dispensable for promoting normal gene expression during development, yet
331 is critical for suppressing ectopic developmental programs. While some germline genes, such as *Dazl*, are
332 also expressed in the 2-cell stage, the inner cell mass, and naïve ESCs, they are silenced in epiblast stem
333 cells/EpiLCs^{6,42,47,72,73}. Our data suggest the 2-cell-like state reported in *Kdm5c*-KO ESCs⁴⁷ likely reflects
334 KDM5C's primary role in germline gene repression (Figure 3). Germline gene misexpression in *Kdm5c*-
335 KO EpiLCs may indicate they are differentiating into primordial germ cell-like cells (PGCLCs)^{35,36,38}. Yet,
336 *Kdm5c*-KO EpiLCs had normal cellular morphology and properly expressed markers for primed pluripotency,
337 including *Otx2* which blocks EpiLC differentiation into PGCs/PGCLCs⁷⁴. In addition to unimpaired EpiLC

338 differentiation, *Kdm5c*-KO gross brain morphology is overall normal²¹ and hardly any brain-specific genes
339 were significantly dysregulated in the amygdala and hippocampus (Figure 1). Thus, ectopic germline gene
340 expression occurs in conjunction with overall proper somatic differentiation in *Kdm5c*-KO animals.

341 Our work provides novel insight into the cross-talk between H3K4me2/3 and CpGme, which are gen-
342 erally mutually exclusive⁷⁵. In EpiLCs, loss of KDM5C binding at a subset of germline gene promoters,
343 e.g. *D1Pas1*, strongly impaired promoter CGI methylation and resulted in their long-lasting de-repression
344 into adulthood. Removal of H3K4me2/3 at CGIs is a plausible mechanism for KDM5C-mediated germline
345 gene suppression^{22,56}, given H3K4me2/3 repell DNMT3 activity^{67,68}. However, emerging work indicates
346 many histone-modifying enzymes have non-catalytic functions that influence gene expression, sometimes
347 even more potently than their catalytic roles^{76,77}. Indeed, KDM5C's catalytic activity was recently found to be
348 dispensible for repressing *Dazl* in ESCs⁴⁷. In our study, *Dazl*'s promoter still gained CpGme in *Kdm5c*-KO
349 exEpiLCs, even with elevated H3K4me2. *Dazl* and a few other germline genes employ multiple repressive
350 mechanisms to facilitate CpGme, such as DNMT3A/B recruitment via E2F6 and MGA^{5,6,48,49}. Thus, while
351 some germline CGIs require KDM5C-mediated H3K4me removal to overcome promoter CGI escape from
352 CpGme^{75,78}, others do not. These results also suggest the requirement for KDM5C's catalytic activity can
353 change depending upon the locus and developmental stage. Further experiments are required to determine
354 if catalytically inactive KDM5C can suppress germline genes at later developmental stages.

355 By generating a comprehensive list of mouse germline-enriched genes, we revealed distinct derepressive
356 mechanisms governing early versus late-stage germline programs. Previous work on germline gene silencing
357 has focused on genes with promoter CGIs^{7,75}, and indeed the majority of KDM5C targets in EpiLCs were
358 germ cell identity genes harboring CGIs. However, over 70% of germline-enriched gene promoters lacked
359 CGIs, including the many KDM5C-unbound germline genes that are de-repressed in *Kdm5c*-KO cells. CGI-
360 free, KDM5C-unbound germline genes were primarily late-stage spermatogenesis genes and significantly
361 enriched for RFX2 binding sites, a central regulator of spermiogenesis^{65,66}. These data suggest that once
362 activated during early embryogenesis, drivers of germline gene expression like *Rfx2*, *Stra8*, and *Dazl* turn
363 on downstream germline programs, ultimately culminating in the expression of spermiogenesis genes in
364 the adult *Kdm5c*-KO brain. Therefore, we propose KDM5C is recruited via promoter CGIs to act as a brake
365 against runaway activation of germline-specific programs. Future studies should address how KDM5C is
366 targeted to CGIs.

367 The above work provides the mechanistic foundation for KDM5C-mediated repression of tissue and
368 germline-specific genes. However, the contribution of these ectopic, tissue-specific genes towards neurolog-
369 ical impairments is still unknown. In addition to germline genes, we also identified significant enrichment
370 of muscle and liver-enriched transcripts within the *Kdm5c*-KO brain. Intriguingly, select liver and muscle-
371 enriched DEGs do have known roles within the brain, such as the liver-enriched lipid metabolism gene
372 *Apolipoprotein C-I (Apoc1)*³⁰. *APOC1* dysregulation is implicated in Alzheimer's disease in humans⁷⁹ and
373 overexpression of *Apoc1* in the mouse brain can impair learning and memory⁸⁰. KDM5C may therefore be

374 crucial for neurodevelopment by fine-tuning the expression of tissue-enriched, dosage-sensitive genes like
375 *Apoc1*.

376 Given that germline genes have no known functions within the brain, their impact upon neuodevelopment
377 is currently unknown. In *C. elegans*, somatic misexpression of germline genes via loss of *Retinoblastoma*
378 (*Rb*) homologs results in enhanced piRNA signaling and ectopic P granule formation in neurons^{81,82}. Ectopic
379 testicular germline transcripts have also been observed in a variety of cancers, including brain tumors in
380 *Drosophila* and mammals and shown to promote cancer progression^{10,11,83–85}. Intriguingly, mouse models
381 and human cells for other chromatin-linked NDDs also display impaired soma-germline demarcation^{13,86,87},
382 such as mutations in DNA methyltransferase 3b (DNMT3B), H3K9me1/2 methyltransferases G9A/GLP,
383 and methyl-CpG -binding protein 2 (MECP2). Recently, the transcription factor ZMYM2 (ZNF198), whose
384 mutation causes a NDD (OMIM #619522), was also shown to repress germline genes by promoting H3K4me
385 removal and CpGme⁸⁸. Thus, KDM5C is among a growing cohort of neurodevelopmental disorders with
386 erosion of the germline-soma boundary. Further research is required to determine the impact of these
387 germline genes upon neuronal functions and the extent to which this phenomenon occurs in humans.

388 Materials and Methods

389 Classifying tissue-enriched and germline-enriched genes

390 Tissue-enriched differentially expresssd genes (DEGs) were determined by their classification in a previ-
391 ously published dataset from 17 male and female mouse tissues²⁵. This study defined tissue expression as
392 greater than 1 Fragments Per Kilobase of transcript per Million mapped read (FPKM) and tissue enrichment
393 as at least 4-fold higher expression than any other tissue.

394 We curated a list of germline-enriched genes using an RNA-seq dataset from wild-type and germline-
395 depleted (Kit^{W/Wv}) male and female mouse embryos from embryonic day 12, 14, and 16³⁴, as well as adult
396 male testes³¹. Germline-enriched genes met the following criteria: 1) their expression is greater than 1
397 FPKM in wild-type germline 2) their expression in any wild-type somatic tissues²⁵ does not exceed 20%
398 of maximum expression in wild-type germline, and 3) their expression in the germ cell-depleted (Kit^{W/Wv})
399 germline, for any sex or time point, does not exceed 20% of maximum expression in wild-type germline. We
400 defined sperm and egg-biased genes as those whose expression in the opposite sex, at any time point, is no
401 greater than 20% of the gene's maximum expression in a given sex. Genes that did not meet this threshold
402 for either sex were classified as 'unbiased'.

403 Cell culture

404 We utilized our previously established cultures of male wild-type and *Kdm5c* knockout (-KO)
405 embryonic stem cells⁴³. Sex was confirmed by genotyping *Uba1/Uba1y* on the X and Y chromo-

406 somes with the following primers: 5'-TGGATGGTGTGCCAATG-3', 5'-CACCTGCACGTTGCCCTT-
407 3'. Deletion of *Kdm5c* exons 11 and 12, which destabilize KDM5C protein²¹, was confirmed
408 through the primers 5'-ATGCCCATATTAAGAGTCCCTG-3', 5'-TCTGCCTTGATGGACTGTT-3', and
409 5'-GGTTCTAACACTCACATAGTG-3'.

410 Embryonic stem cells (ESCs) and epiblast-like cells were cultured using previously established
411 methods³⁹. Briefly, ESCs were initially cultured in primed ESC (pESC) media consisting of KnockOut
412 DMEM (Gibco#10829-018), fetal bovine serum (Gibco#A5209501), KnockOut serum replacement
413 (Invitrogen#10828-028), Glutamax (Gibco#35050-061), Anti-Anti (Gibco#15240-062), MEM Non-essential
414 amino acids (Gibco#11140-050), and beta-mercaptoethanol (Sigma#M7522). They were then transitioned
415 into ground-state, “naïve” ESCs (nESCs) by culturing for four passages in N2B27 media containing
416 DMEM/F12 (Gibco#11330-032), Neurobasal media (Gibco#21103-049), Gluamax (Gibco#35050-061),
417 Anti-Anti (Gibco#15240-062), N2 supplement (Invitrogen#17502048), and B27 supplement without vitamin
418 A (Invitrogen#12587-010), and beta-mercaptoethanol (Sigma#M7522). Both pESC and nESC media
419 were supplemented with 3 μ M GSK3 inhibitor CHIR99021 (Sigma #SML1046-5MG), 1 μ M MEK inhibitor
420 PD0325901 (Sigma #PZ0162-5MG), and 1,000 units/mL leukemia inhibitory factor (LIF, Millipore#ESG1107).
421 nESCs were differentiated into epiblast-like cells (EpiLCs, 48 hours) and extendend EpiLCs (exEpiLCs,
422 96 hours) by culturing in N2B27 media containing DMEM/F12, Neurobasal media, Gluamax, Anti-Anti, N2
423 supplement, B27 supplement (Invitrogen#17504044), and beta-mercaptoethanol supplemented with 10
424 ng/mL fibroblast growth factor 2 (FGF2, R&D Biotechne 233-FB), and 20 ng/mL activin A (R&D Biotechne
425 338AC050CF), as previously described³⁹.

426 Real time quantitative PCR (RT-qPCR)

427 nESCs were differentiated into EpiLCs as described above. Cells were lysed with Tri-reagent BD (Sigma
428 #T3809) at 0, 24, and 48 hours of differentiation. RNA was phase separated with 0.1 μ L/ μ L 1-bromo-3-
429 chloropropane (Sigma #B9673) and then precipitated with isopropanol (Sigma #I9516) and ethanol puri-
430 fied. For each sample, 2 μ g of RNA was reverse transcribed using the ProtoScript II Reverse transcriptase kit
431 from New England Biolabs (NEB #M0368S) and primed with oligo dT. Expression of *Kdm5c* was detected us-
432 ing the primers 5'-CCCATGGAGGCCAGAGAATAAG-3' 5'-CTCAGCGATAAGAGAATTGCTAC-3' and nor-
433 malized to TBP using the primers 5'-TTCAGAGGATGCTCTAGGAAGA-3' 5'-CTGTGGAGTAAGTCCTGTGCC-
434 3' with the Power SYBR™ Green PCR Master Mix (ThermoFisher #4367659).

435 Western Blot

436 Total protein was extracted during nESC to EpiLC differentiation at 0, 24, and 48 hours by sonicating cells
437 at 20% amplitude for 15 seconds in 2X SDS sample buffer, then boiling at 100 °C for 10 minutes. Proteins
438 were separated on a 7.5% SDS page gel, transferred overnight onto a fluorescent membrane, blotted for

439 rabbit anti-KDM5C (in house, 1:500) and mouse anti-DAXX (Santa Cruz #(H-7): sc-8043, 1:500), and then
440 imaged using the LiCor Odyssey CLx system. Band intensity was quantified using ImageJ.

441 **RNA sequencing (RNA-seq) data analysis**

442 After ensuring read quality via FastQC (v0.11.8), reads were then mapped to the mm10 *Mus musculus*
443 genome (Gencode) using STAR (v2.5.3a), during which we removed duplicates and kept only uniquely
444 mapped reads. Count files were generated by FeatureCounts (Subread v1.5.0), and BAM files were
445 converted to bigwigs using deeptools (v3.1.3) and visualized by the UCSC genome browser⁷⁰. RStudio
446 (v3.6.0) was then used to analyze counts files by DESeq2 (v1.26.0)²⁶ to identify differentially expressed
447 genes (DEGs) with a q-value (p-adjusted via FDR/Benjamini–Hochberg correction) less than 0.1 and a log2
448 fold change greater than 0.5. For all DESeq2 analyses, log2 fold changes were calculated with IfcShrink
449 using the ashr package⁸⁹. MA-plots were generated by ggpubr (v0.6.0), and Eulerr diagrams were generated
450 by eulerr (v6.1.1). Boxplots and scatterplots were generated by ggpubr (v0.6.0) and ggplot2 (v3.3.2). The
451 Upset plot was generated via the package UpSetR (v1.4.0)⁹⁰. Gene ontology (GO) analyses were performed
452 by the R package enrichPlot (v1.16.2) using the biological processes setting and compareCluster.

453 **Chromatin immunoprecipitation followed by DNA sequencing (ChIP-seq) data analysis**

454 ChIP-seq reads were aligned to mm10 using Bowtie1 (v1.1.2) allowing up to two mismatches. Only
455 uniquely mapped reads were used for analysis. Peaks were called using MACS2 software (v2.2.9.1) using
456 input BAM files for normalization, with filters for a q-value < 0.1 and a fold enrichment > 1. We removed
457 “black-listed” genomic regions that often give aberrant signals. Common peak sets were obtained in R via
458 DiffBind⁹¹ (v3.6.5). In the case of KDM5C ChIP-seq, *Kdm5c*-KO false-positive peaks were then removed from
459 wild-type samples using bedtools (v2.25.0). Peak proximity to genomic loci was determined by ChIPSeeker⁹²
460 (v1.32.1). Gene ontology (GO) analyses were performed by the R package enrichPlot (v1.16.2) using the
461 biological processes setting and compareCluster. Enriched motifs were identified using HOMER⁶¹ to search
462 for known motifs within 500 base pairs up and downstream of the transcription start site. Average binding
463 across genes was visualized using deeptools (v3.1.3). Bigwigs were visualized using the UCSC genome
464 browser⁷⁰.

465 **CpG island (CGI) analysis**

466 Locations of CpG islands were determined through the mm10 UCSC genome browser CpG island track⁷⁰,
467 which classified CGIs as regions that have greater than 50% GC content, are larger than 200 base pairs,
468 and have a ratio of CG dinucleotides observed over the expected amount greater than 0.6. CGI genomic
469 coordinates were then annotated using ChIPseeker⁹² (v1.32.1) and filtered for ones that lie within promoters
470 of germline-enriched genes (TSS ± 500).

471 **Whole genome bisulfite sequencing (WGBS)**

472 Genomic DNA (gDNA) from male naïve ESCs and extended EpiLCs was extracted using the Wizard
473 Genomic DNA Purification Kit (Promega A1120), following the instructions for Tissue Culture Cells. gDNA
474 from two wild-types and two *Kdm5c*-KOs of each cell type was sent to Novogene for WGBS using the
475 Illumina NovaSeq X Plus platform and sequenced for 150 bp paired-end reads (PE150). All samples had
476 greater than 99% bisulfite conversion rates. Reads were adapter and quality trimmed with Trim Galore
477 (v0.6.10) and aligned to the mm10 genome using Bismark⁹³ (v0.22.1). Analysis of differential methylation at
478 gene promoters was performed using methylKit⁷¹ (v1.28.0) with a minimum coverage of 3 paired reads, a
479 percentage greater than 25% or less than -25%, and q-value less than 0.01. methylKit was also used to
480 calculate average percentage methylation at germline gene promoters. Methylation bedgraph tracks were
481 generated via Bismark and visualized using the UCSC genome browser⁷⁰.

482 **Data availability**

483 **WGBS in wild-type and *Kdm5c*-KO ESCs and exEpiLCs**

484 Raw fastq files are deposited in the Sequence Read Archive (SRA) <https://www.ncbi.nlm.nih.gov/sra>
485 under the bioProject PRJNA1165148. <https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1165148>

486 **Published datasets**

487 All published datasets are available at the Gene Expression Omnibus (GEO) <https://www.ncbi.nlm.nih.gov/geo>. Published RNA-seq datasets analyzed in this study included the male wild-type and *Kdm5c*-KO
488 adult amygdala and hippocampus²⁴, available at GEO: GSE127722. Male and female wild-type, *Kdm5c*-KO,
489 and *Kdm5c*-HET EpiLCs⁴³ are available at GEO: GSE96797.

491 Previously published ChIP-seq experiments included KDM5C binding in wild-type and *Kdm5c*-KO
492 EpiLCs⁴³ (available at GEO: GSE96797) and mouse primary neuron cultures (PNCs) from the cortex
493 and hippocampus²¹ (available at GEO: GSE61036). ChIP-seq of histone 3 lysine 4 dimethylation (H3K4me2)
494 in male wild-type and *Kdm5c*-KO EpiLCs⁴³ is also available at GEO: GSE96797. ChIP-seq of histone 3 lysine
495 4 trimethylation (H3K4me3) in wild-type and *Kdm5c*-KO male amygdala²⁴ are available at GEO: GSE127817.

496 **Data analysis**

497 Scripts used to generate the results, tables, and figures of this study are available via the GitHub
498 repository: https://github.com/kbonefas/KDM5C_Germ_Mechanism

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514 **Author Contributions**

515 K.M.B. and S.I. conceived the study and designed the experiments. I.V. generated the ESC and exEpiLC
516 WGBS data. K.M.B performed all data analysis and all other experiments. The manuscript was written by
517 K.M.B and S.I. and edited by K.M.B, S.I., and I.V.

518 **Declaration of Interest**

519 S.I. is a member of the Scientific Advisory Board of KDM5C Advocacy, Research, Education & Support
520 (KARES). Other authors declare no conflict of interest.

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708 **Figures and Tables**

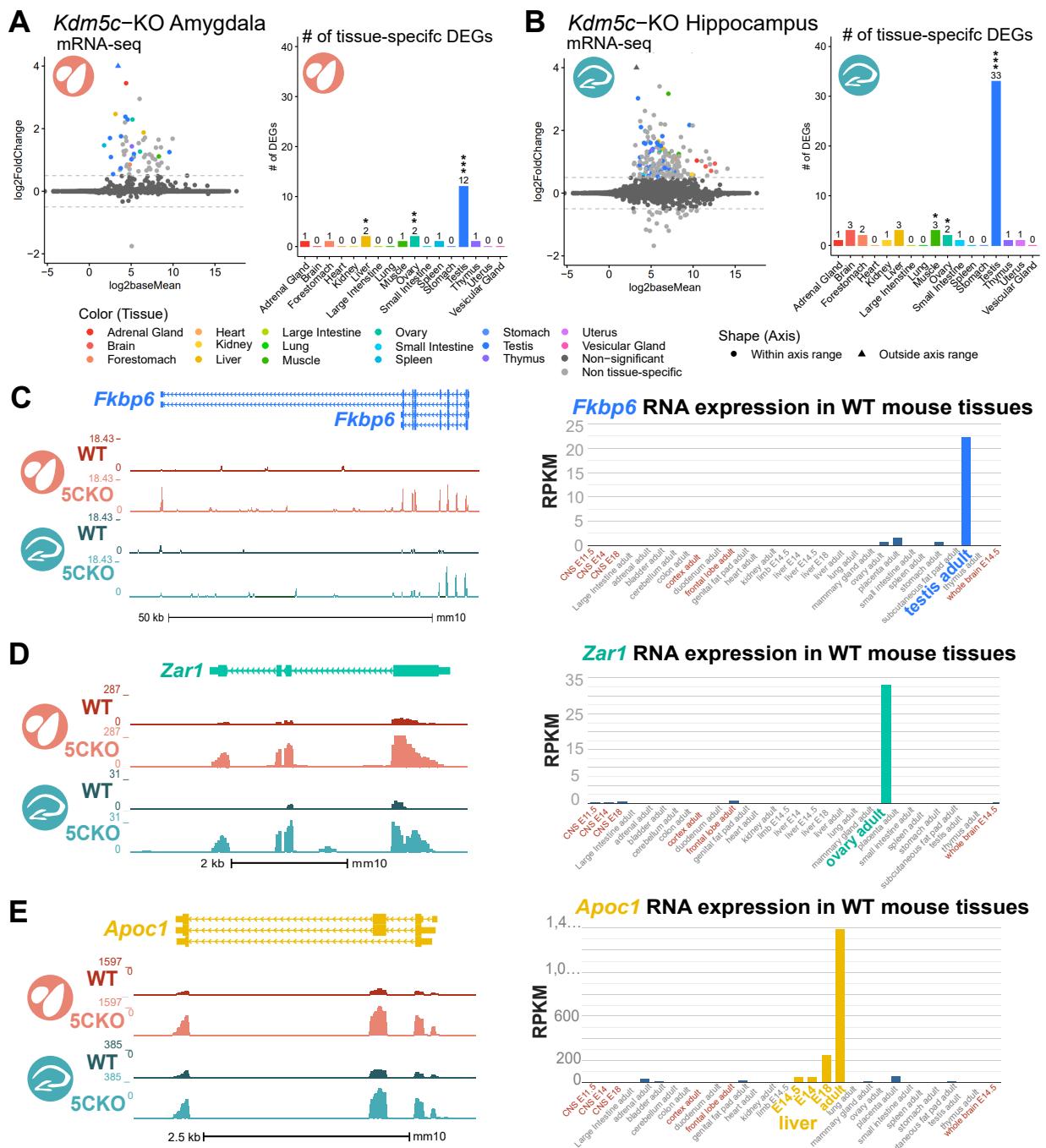


Figure 1: Tissue-enriched genes are misexpressed in the *Kdm5c*-KO brain. **A-B.** Expression of tissue-enriched genes (Li et al 2017) in the male *Kdm5c*-KO amygdala (A) and hippocampus (B). Left - MA plot of mRNA-sequencing. Right - Number of tissue-enriched differentially expressed genes (DEGs). * p<0.05, ** p<0.01, *** p<0.001, Fisher's Exact Test. **C.** Left - UCSC browser view of an example aberrantly expressed testis-enriched DEG, *FK506 binding protein 6* (*Fkbp6*) in the wild-type (WT) and *Kdm5c*-KO (5CKO) amygdala (red) and hippocampus (teal) (Average, n = 4). Right - Expression of *Cyct* in wild-type tissues from NCBI Gene, with testis highlighted in blue and brain tissues highlighted in red. **D.** Left - UCSC browser view of an example ovary-enriched DEG, *Zygotic arrest 1* (*Zar1*). Right - Expression of *Zar1* in wild-type tissues from NCBI Gene, with ovary highlighted in teal and brain tissues highlighted in red. **E.** Left - UCSC browser view of an example liver-enriched DEG, *Apolipoprotein C-I* (*Apoc1*). Right - Expression of *Apoc1* in wild-type tissues from NCBI Gene, with liver highlighted in orange and brain tissues highlighted in red.

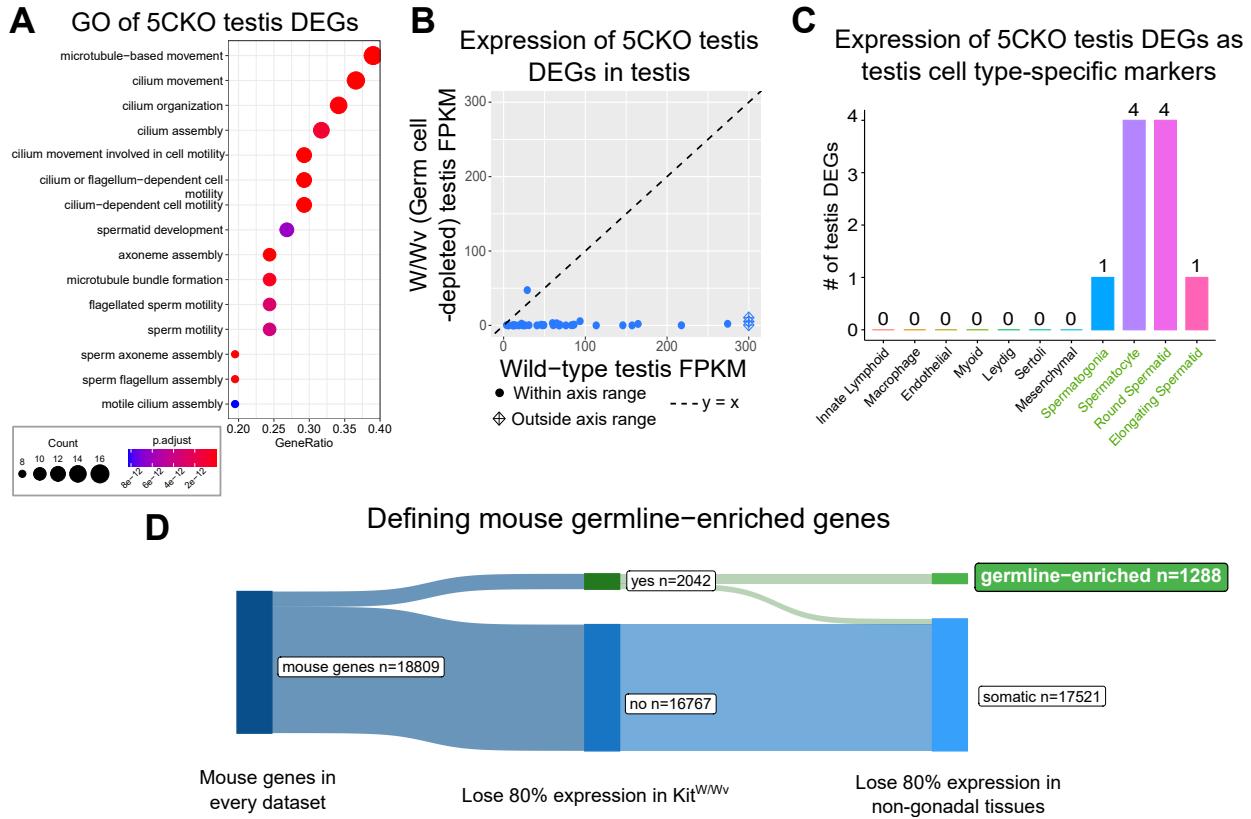


Figure 2: Aberrant transcription of germline genes in the *Kdm5c*-KO in the brain. **A.** enrichPlot gene ontology (GO) of *Kdm5c*-KO amygdala and hippocampus testis-enriched DEGs **B.** Expression of testis DEGs in wild-type (WT) testis versus germ cell-depleted (W/Wv) testis (Mueller et al 2013). Expression is in Fragments Per Kilobase of transcript per Million mapped reads (FPKM). **C.** Number of testis DEGs that were classified as cell-type specific markers in a single cell RNA-seq dataset of the testis (Green et al 2018). Germline cell types are highlighted in green, somatic cell types in black. **D.** Sankey diagram of mouse genes filtered for germline enrichment based on their expression in wild-type and W/Wv mice and in adult mouse non-gonadal tissues (Li et al 2017).

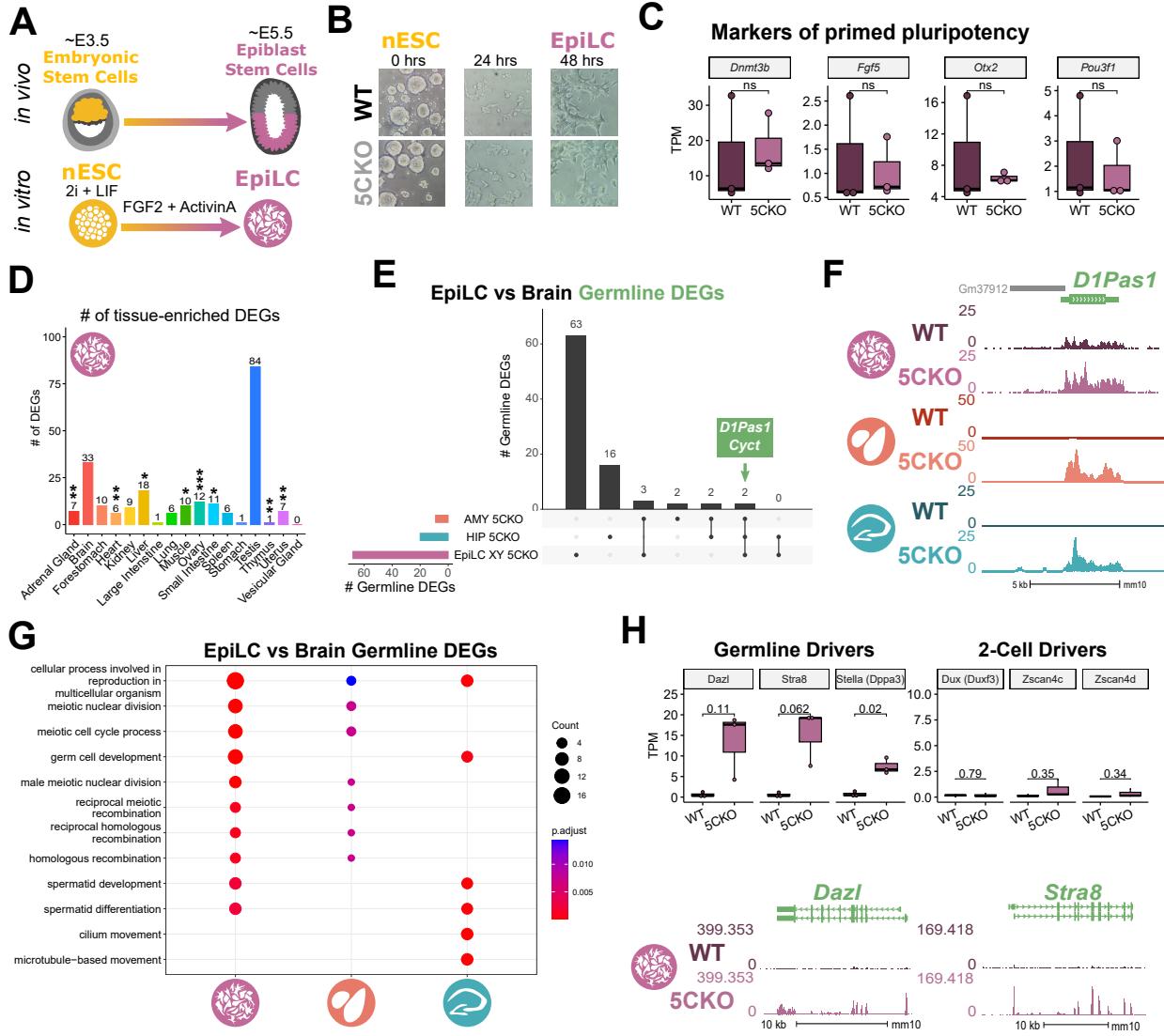


Figure 3: Kdm5c-KO epiblast-like cells express key drivers of germline identity **A.** Top - Diagram of *in vivo* differentiation of embryonic stem cells (ESCs) of the inner cell mass into epiblast stem cells. Bottom - *in vitro* differentiation of ESCs into epiblast-like cells (EpiLCs). **B.** Representative images of male wild-type (WT) and *Kdm5c*-KO (5CKO) cells during ESC to EpiLC differentiation. Brightfield images taken at 20X. **C.** No significant difference in primed pluripotency marker expression in wild-type versus *Kdm5c*-KO EpiLCs. Welch's t-test, expression in transcripts per million (TPM). **D.** Number of tissue-enriched differentially expressed genes (DEGs) in *Kdm5c*-KO EpiLCs. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, Fisher's Exact Test. **E.** Upset plot displaying the overlap of germline DEGs expressed in *Kdm5c*-KO EpiLCs, amygdala (AMY), and hippocampus (HIP) RNA-seq datasets. **F.** UCSC browser view of an example germline gene, *D1Pas1*, that is dysregulated *Kdm5c*-KO EpiLCs (top, purple. Average, $n = 3$), amygdala (middle, red. Average, $n = 4$), and hippocampus (bottom, blue. Average, $n = 4$). **G.** enrichPlot gene ontology analysis comparing enriched biological processes for *Kdm5c*-KO EpiLC, amygdala, and hippocampus germline DEGs. **H.** Top left - Example germline identity DEGs unique to EpiLCs. Top right - Example 2-cell genes that are not dysregulated in *Kdm5c*-KO EpiLCs. p-values for Welch's t-test. Bottom - UCSC browser view of *Dazl* and *Stra8* expression in wild-type and *Kdm5c*-KO EpiLCs (Average, $n = 3$).

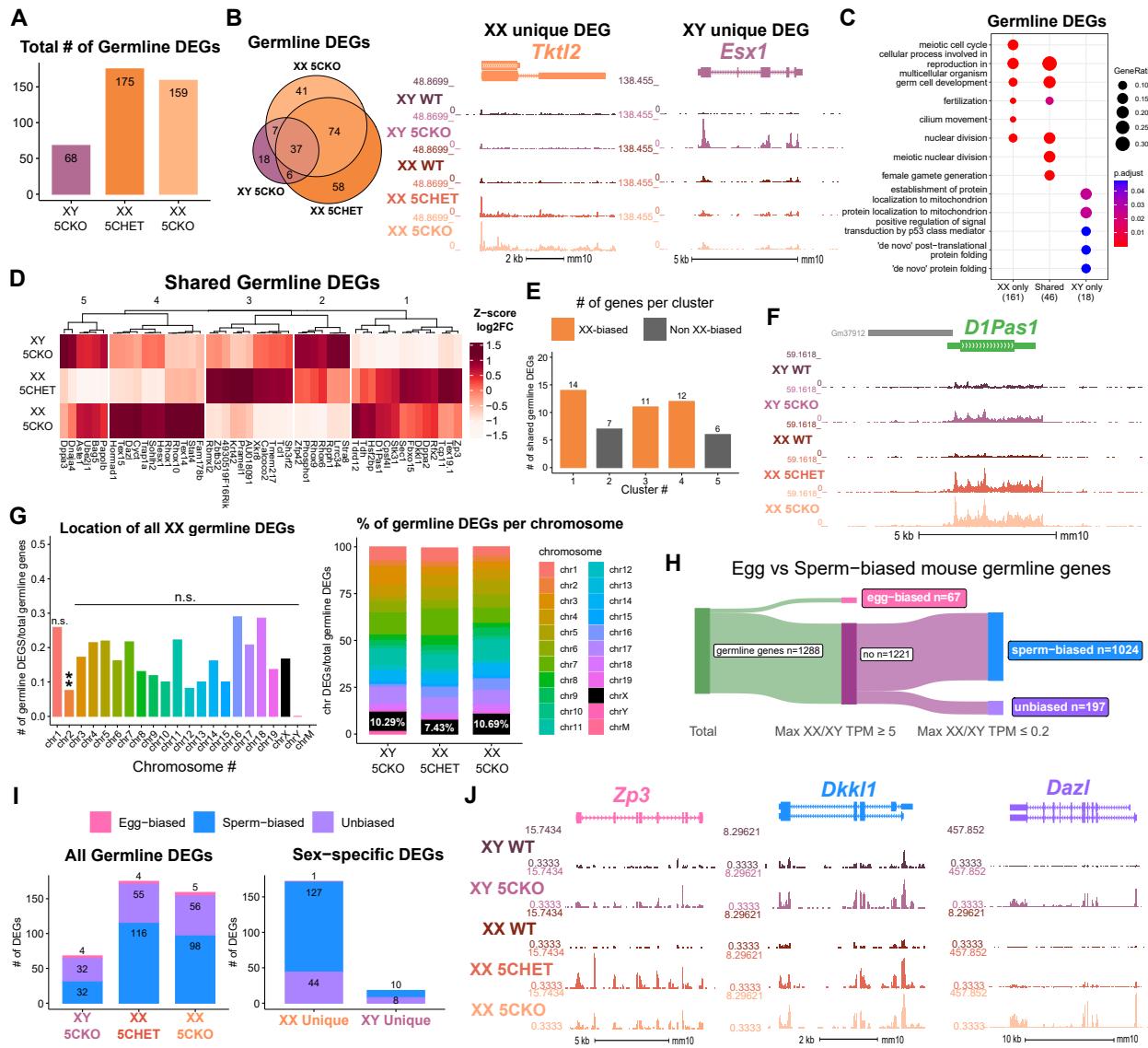


Figure 4: Chromosomal sex influences *Kdm5c*-KO germline gene misexpression. **A.** Total number of germline-enriched RNA-seq DEGs for male hemizygous *Kdm5c* knockout EpiLCs (XY 5CKO, purple), female heterozygous *Kdm5c* knockout (XX 5CHET, orange), female homozygous *Kdm5c* knockout (XX 5CKO, light orange) EpiLCs. **B.** Left - Eulerr overlap of *Kdm5c* mutant male and female EpiLC germline DEGs. Right - Example of germline DEGs unique to females or males, *Tktl2* and *Esx1*. **C.** enrichPlot gene ontology analysis comparing enriched biological processes for germline DEGs shared between *Kdm5c* mutant males and females (Shared), or unique to one sex (XX only or XY only). **D.** Heatmap of germline DEGs shared between male and female mutants. Color is the average log 2 fold change from sex-matched wild-type, z-scored across rows. **E.** Number of genes within each cluster from D. Clusters with higher expression in females compared to males (XX-biased) highlighted in orange. **F.** UCSC browser view of a male and female shared germline DEG *D1Pas1* that is more highly expressed in female mutants (Average, n = 3). **G.** Left - Number of all female germline DEGs located on each chromosome over the total number of germline-enriched genes on that chromosome. P-values for Fisher Exact Test, ** p < 0.01, n.s. non-significant. Germline DEGs were only significant for chromosome 2, in which they were significantly depleted. Right - Percentage of germline DEGs that lie on each chromosome for each *Kdm5c* mutant. X chromosome highlighted in black. **H.** Sankey diagram classifying egg-biased (pink) and sperm-biased (blue) and unbiased (purple) mouse germline-enriched genes. **I.** Number of egg, sperm, or unbiased germline DEGs for male and female *Kdm5c* mutants. **J.** UCSC browser view of egg-biased (*Zp3*), sperm-biased (*Dkk1*), and unbiased (*Dazl*) germline genes dysregulated in both male and female *Kdm5c* mutants (Average of n = 3).

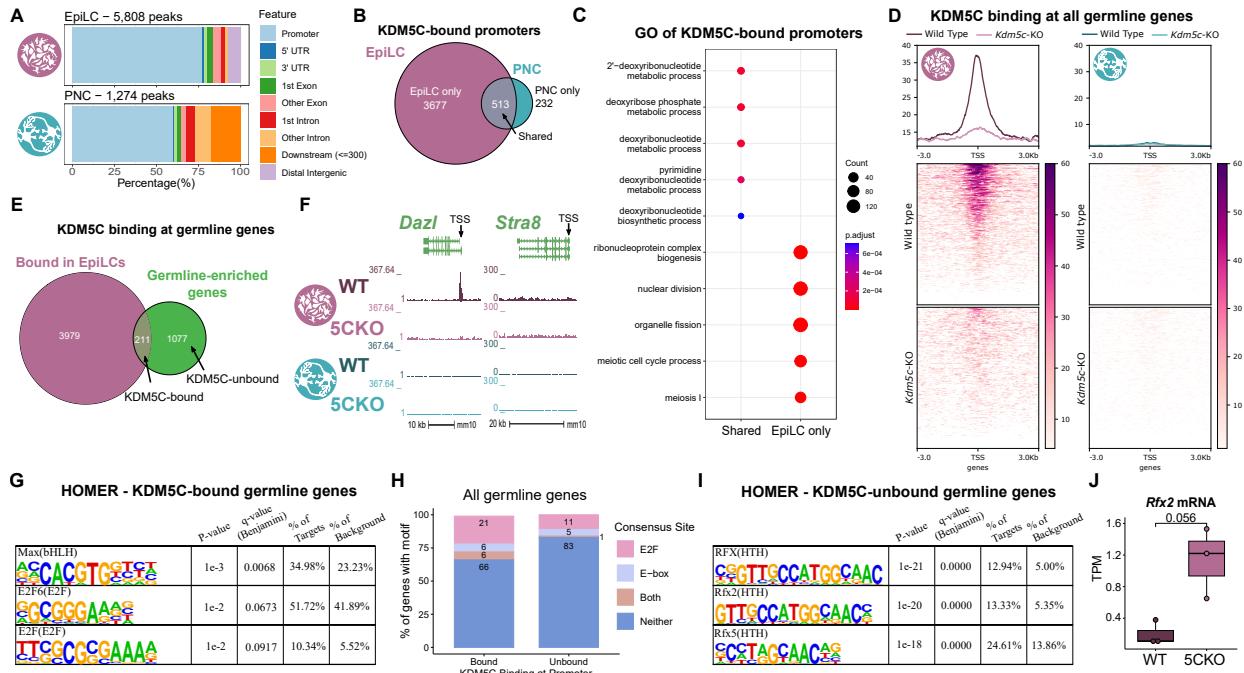


Figure 5: KDM5C binds to a subset of germline gene promoters during early embryogenesis. **A.** ChIPseeker localization of KDM5C peaks at different genomic regions in EpiLCs (top) and hippocampal and cortex primary neuron cultures (PNCs, bottom). **B.** Overlap of genes with KDM5C bound to their promoters ($TSS \pm 500$) in EpiLCs (purple) and PNCs (blue). **C.** Gene ontology (GO) comparison of genes with KDM5C bound to their promoter in EpiLCs and PNCs. Genes were classified as either bound in EpiLCs only (EpiLC only), unique to PNCs (PNC only, no significant ontologies) or bound in both PNCs and EpiLCs (Shared). **D.** Average KDM5C binding around the transcription start site (TSS) of all germline-enriched genes in EpiLCs (left) and PNCs (right). **E.** Eulerr overlap of germline-enriched genes (green) with significant KDM5C binding at their promoter in EpiLCs (purple). **F.** Example KDM5C ChIP-seq signal around the *Dazl* TSS but not *Stra8* in EpiLCs. **G.** HOMER motif analysis of all KDM5C-bound germline gene promoters, highlighting significant enrichment of MAX, E2F6, and E2F motifs. **H.** Number of all gene promoters bound or unbound by KDM5C with instances of the E2F or E-box consensus sequence. **I.** HOMER motif analysis of all KDM5C-unbound germline gene promoters, highlighting significant enrichment of RFX family transcription factor motifs. **J.** Expression of RNA-seq DEG *Rfx2* in wild-type and *Kdm5c*-KO EpiLCs. P-value of Welch's t-test, expression in transcripts per million (TPM).

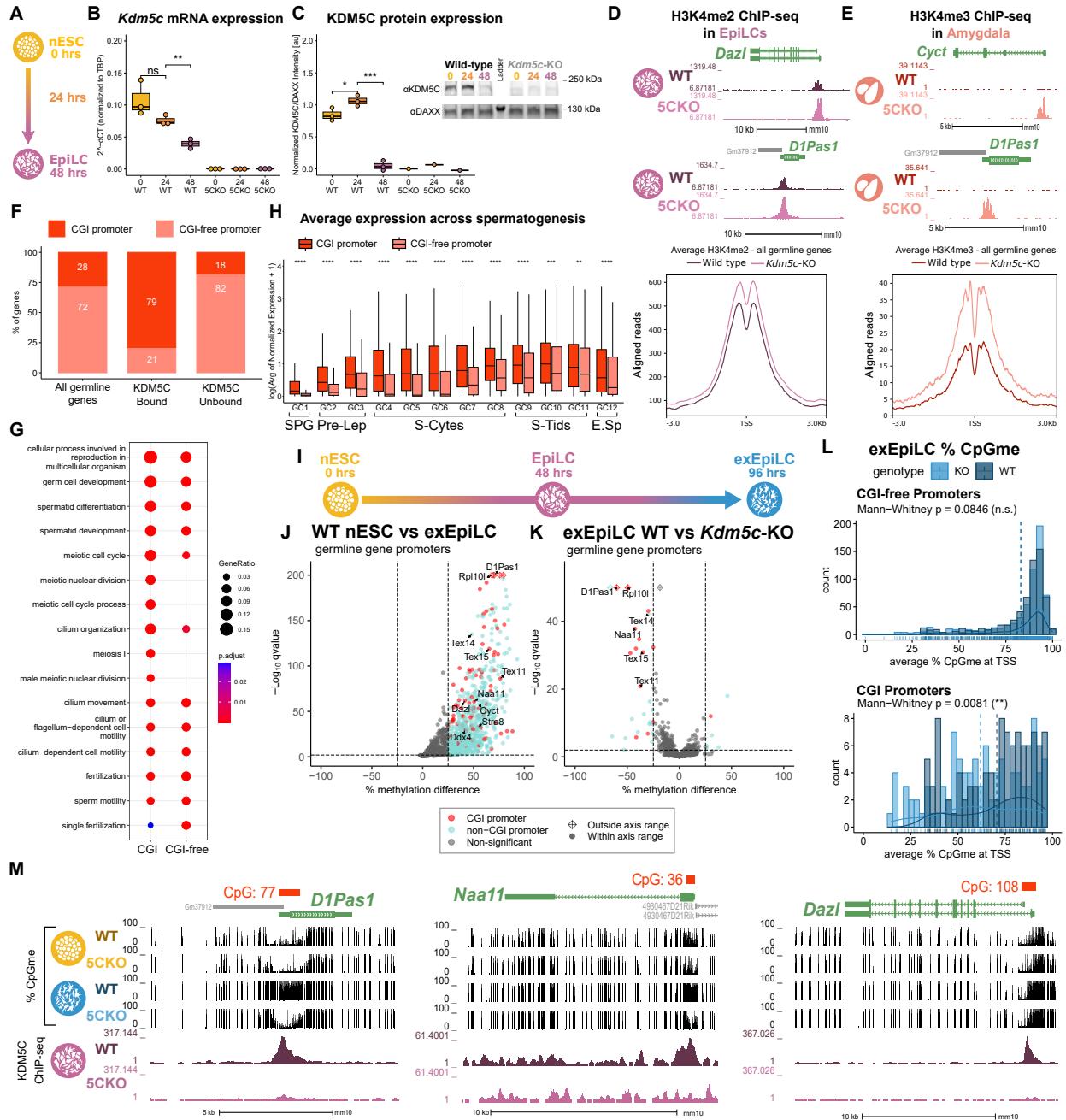


Figure 6: KDM5C promotes long-term silencing of germline genes via DNA methylation at CpG islands. **A.** Diagram of embryonic stem cell (ESC) to epiblast-like cell (EpiLC) differentiation and collection time points. **B.** Real time quantitative PCR (RT-qPCR) of *Kdm5c* mRNA expression in wild-type (WT) and *Kdm5c*-KO (5CKO) ESCs at 0, 24, and 48 hours of differentiation into EpiLCs. Expression calculated in comparison to TBP mRNA expression ($2^{-\Delta\Delta CT}$). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, Welch's t-test. **C.** KDM5C protein expression normalized to DAXX. Quantified intensity using ImageJ (artificial units - au). Right - representative lanes of Western blot for KDM5C and DAXX. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, Welch's t-test. **D.** Top - Representative UCSC browser view of histone 3 lysine 4 dimethylation (H3K4me2) ChIP-seq signal at two germline genes in wild-type and *Kdm5c*-KO EpiLCs. Bottom - Average H3K4me2 signal at the TSS of all germline-enriched genes in wild-type (dark purple) and *Kdm5c*-KO (light purple) EpiLCs. **E.** Top - Representative UCSC browser view of histone 3 lysine 4 trimethylation (H3K4me3) ChIP-seq signal at two germline genes in the wild-type (WT) and *Kdm5c*-KO (5CKO) adult amygdala. Bottom - Average H3K4me3 signal at the transcription start site (TSS) of all germline-enriched genes in wild-type (dark red) and *Kdm5c*-KO (light red) amygdala. **F.** Percentage of germline genes that harbor CpG islands (CGIs) in their promoters (CGI promoter, red), based on UCSC annotation. Left - percentage of all germline-enriched genes, middle - KDM5C-bound germline genes, and right - KDM5C-unbound germline genes. (Legend continued on next page.)

Figure 6: KDM5C promotes long-term silencing of germline genes via DNA methylation at CpG islands. (Legend continued.) **G.** enrichPlot gene ontology analysis of germline genes with (CGI-promoter) or without (CGI-free) CGIs in their promoter. **H.** Expression of germline genes with (CGI-promoter, red) or without (CGI-free, salmon) CGIs in their promoter across stages of spermatogenesis from Green et al 2018. SPG - spermatogonia, Pre-Lep - preleptotene spermatocytes, S-Cytes - meiotic spermatocytes, S-Tids - post-meiotic haploid round spermatids, E.Sp - elongating spermatids. Wilcoxon test, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$. **I.** Diagram of ESC to extended EpiLC (exEpiLC) differentiation. **J.** Volcano plot of whole genome bisulfite sequencing (WGBS) comparing CpG methylation (CpGme) at germline gene promoters ($TSS \pm 500$) in wild-type (WT) ESCs versus exEpiLCs. Significantly differentially methylated promoters ($q < 0.01$, $|methylated difference| > 25\%$) with CGIs in red, CGI-free promoters in light blue **K.** Volcano plot of WGBS of wild-type (WT) versus *Kdm5c*-KO exEpiLCs for germline gene promoters. Promoters with CGIs in red, CGI-free promoters in light blue. **L.** Histogram of average percent CpGme at the promoter of germline genes with or without CGIs. Wild-type in navy and *Kdm5c*-KO (KO) in light blue. Dashed lines are average methylation for each genotype, p-values for Mann-Whitney U test. **M.** UCSC browser view of germline genes, showing UCSC annotated CGI with number of CpGs, representative CpGme in wild-type (WT) and *Kdm5c*-KO (5CKO) ESCs and exEpiLCs, and KDM5C ChIP-seq signal in EpiLCs.