

Fusion-finder: Identify genomic fusion events using directed mapping and machine learning

Kez Cleal, Kate Liddiard, Duncan Baird. 2018

Manual v.1.0.1

Table of Contents

TLDR.....	1
Installation and dependencies.....	2
Commands	2
“run”	2
“find-reads”	3
“align”	3
“call-events”	3

TLDR

To install, type in a terminal:

```
$ pip install fufi
```

Run the pipeline with input bam file and a genome reference in fasta format:

```
$ fufi run ref.fa input.bam
```

This will produce a .vcf file with structural variants in the same location with the postfix .fufi.vcf

Run using multiple processors, whilst directing output to folder:

```
$ fufi run --procs 12 --dest new_folder ref.fa input.bam
```

‘exclude’ certain regions altogether, whilst limiting the ‘search’ to specified regions, but additionally to ‘limit’ final calls to some other regions of interest:

```
$ fufi run --exclude regions1.bed --search regions2.bed --limit regions3.bed ref.fa  
input.bam
```

Explore help messages:

```
$ fufi --help
```

```
$ fufi run --help
```

Overview

Fusion finder can be used to call structural variants in whole genome sequencing data aligned to a reference genome, although was specifically designed to detect structural variants in amplicons derived from repetitive regions of the genome that have characteristics such as high read-depth and mapping ambiguity.

Installation and dependencies

To install, type in a terminal:

```
$ pip install fufi
```

Alternatively download repository from <https://github.com/kcleeal> and installed by:

```
$ python setup.py install
```

Required dependencies:

python >= 2.7 and a c++11 compatible compiler

Python dependencies should be installed automatically but include:

'click', 'numpy', 'pandas', 'pysam', 'quicksect', 'pybedtools', 'natsort', 'networkx', 'scikit-learn'

Optional but recommended:

'bwa mem' <https://github.com/lh3/bwa> accessible from your path. See the align command for more information.

Commands

"run"

Runs the fufi pipeline using default setting for each tool

Options:

--include	PATH	.bed file, limit calls to regions.
--search	PATH	.bed file, limit search to regions.
--exclude	PATH	.bed file, do not search/call SVs within regions.
		Overrides include/search
--clip-length	INTEGER	Minimum soft-clip length; >= threshold are kept. [default: 21]
--map-script	PATH	External shell script for mapping. Default is to use bwa mem internally. Script must take positional arguments as: \$1 reference genome; \$2 .fastq file - must be interleaved if paired-end, otherwise single end reads are assumed; \$3 threads to use.
-p, --procs	INTEGER RANGE	Processors to use [default: 1], limited to the number of processors available on the machine
--dest	PATH	Destination folder to use/create for saving results. Defaults to directory of input bam

--help Shows the help message

Required input arguments:

REFERENCE	PATH	Genome reference in fasta format
BAM	PATH	The input .bam file to search for structural variants

Outputs:

Generates a .vcf file of structural variants in either the folder specified by the --dest option, or in the same directory as the input .bam file. The vcf is prefixed with .fufi.vcf.

Example:

```
$ fufi run -p8 ref.fa input.bam
```

“find-reads”

“align”

“call-events”