

# DIGITAL LAB NOTEBOOK OF KEVIN MURRAY

HONOUS PROJECT, 2013

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*Last updated at 17:46 on Wednesday 30<sup>th</sup> January, 2013*



# Mon 2012-12-10

## 1 Final Harvest of Keng's RIX lines

### 1.1 Aim

Harvest lines after 1 week of HL stress.

### 1.2 Method

- An Eppendorf 1.2mL deep well plate was placed on dry ice for  $\approx 10$  minutes before sampling to allow to cool.
- Whole leaves were excised and placed into 1.2mL Eppendorf 96 deep well plate.
- Where possible, the largest mature leaf was taken. In some cases, this was hard to determine, so the youngest of the fully-expanded leaves was taken (as this was generally also the largest leaf). Some plants were very small, and had only juvenile leaves, in which case the largest juvenile leaf was taken.

### 1.3 Results

The following table describes the plate layout.

Well	Line	Comments	Well	Line	Comments
A1	87	2nd plant with orange label	A7	65	NPQ4
B1	100		B7	71	
C1	92		C7	53	
D1	OSB2		D7	55	
E1	93		E7	56	
F1	94		F7	50	
G1	1*cvi		G7	51	
H1	99		H7	52	
A2	63		A8	78	
B2	98		B8	80	
C2	98		C8	85	
D2	70		D8	74	
E2	67		E8	73	
F2	99		F8	76	
G2	66		G8	83	
H2	100		H8	81	
A3	65		A9	89	
B3	72		B9	90	
C3	12		C9	91	
D3	11		D9	16	
E3	10		E9	17	
F3	12		F9	18	

Well	Line	Comments	Well	Line	Comments
G3	9		G9	21	
H3	8		H9	23	
A4	7		A10	25	
B4	6		B10	57	
C4	5		C10	28	
D4	13		D10	29	
E4	2		E10	30	
F4	1*cvi		F10	31	
G4	49		G10		
H4	47		H10		
A5	46		A11		
B5	42		B11		
C5	45		C11		
D5	71		D11		
E5	39		E11		
F5	40		F11		
G5	41		G11		
H5	43		H11		
A6	38		A12		
B6	39		B12		
C6	36		C12		
D6	33		D12		
E6	61		E12		
F6	62		F12		
G6	68		G12		
H6	58		H12		

*Attachments:*

- dec12/20121210-harvest\_photos.tar.bz2

MD5SUM:40dae2cad3babaa3c32f0d35a9d9442c

# Mon 2013-01-14

## 1 MAKE: Washed Ball Bearings

### 1.1 Method

- Aliquot approx 15mL of 3mm diameter steel ball bearings into 50mL falcon tube
- Add clean 100% ethanol
- Vortex for  $\approx$  5 minutes
- Remove ethanol, wash beads with milliQ or sterile water
- Dry in fume cupboard overnight

## 2 Tissuelyser grinding of practice samples

### 2.1 Aims

To grind tissue from the excess tissue of Keng's RIX lines collected on 3/12/12.

### 2.2 Method

- Remove pre-frozen tissuelyser blocks from -80 freezer.
- Add one cleaned bead to each eppi tube (beads were not pre-cooled)
- Pour LN<sub>2</sub> into the tissuelyser block
- Add Eppies with beads and sample, and run for 3x 1min runs at 29hz
- Replace samples in -80



# Mon 2013-01-21

## 1 Quantification of RNA samples

### 1.1 Aim

- Determine qty of RNA in previously extracted samples

### 1.2 Method

- Nanodropped RNA extraction from 15/1/13??
- Standard protocol, used sterile milliQ water as blank.

### 1.3 Result

- Of the 14 samples, 10 had reasonable amounts of RNA, and 260/280 ratios were above 1.8 in all but one case. (see ./jan/20130121-PracticeRNASamples.ods)

### 1.4 Attachements

- ./jan/20130121-PracticeRNAExtractionSamples.csv
- ./jan/20130121-PracticeRNAExtractionSamples.ndv
- ./jan/20130121-PracticeRNASamples.ods

## 2 MADE: 10x MOPS Solution

### *Method*

- Add 41.8g RNA only MOPS to beaker
- Add 450mL DEPC H<sub>2</sub>O, mix w/ stirrer bar on mag stirrer
- Add 26.6mL 3M Sodium Acetate (0.22um Filtered before use)
- Add 10mL RNA only 0.5M EDTA
- pH to 7 with 5M NaOH
- Top up to 500 mL with DEPC H<sub>2</sub>O
- Use 10ml per 100mL MOPS gel

*Mon 2013-01-21*

### **3 MADE: RNA Denaturing Gel (MOPS)**

#### *Method*

- Melt 1g RNase-free Agarose in 72ml DEPC H<sub>2</sub>O
- Add 10mL 10x MOPS
- Add 18mL 37% Formaldehyde
- Pour in RNA-only gel tank, previously washed with 0.5% SDS and RNase-zap



# Tue 2013-01-22

## 1 Denature RNA for RNA gels

### 1.1 Method

- Dilute RNA to 100ng/uL
- Add RNA gel loading buffer (Obtained from Pete Crisp)
- Incubate at 65 degrees for 10 minutes. The samples were incubated for 10 minutes on the evening of 2013-01-21, but the gels were not run until 2013-01-22, so they were denatured for a further 2 minutes at 65 degrees

## 2 TBE Gel

### 2.1 Aim

- To compare TBE and denaturing/MOPS gels for RNA

### 2.2 Method

- Dissolve 1g RNAase-free agarose in 90mL DEPC water
- Add 10mL RNAase-free TBE (prepared using DEPC Water, obtained from Pete Crisp)
- Pour in RNA-only gel tank, previously washed with 0.5% SDS or RNAse-zap
- Then, load denatured samples, and run in RNAse-free 1x TBE
- Run at  $\approx 80V$ ,  $\approx 40-50mA$  for  $\approx 1.75h$
- Stain gel in 0.5ug/ml Ethidium Br in DEPC water?? for 10 min on orbital shaker, and photograph.

### 2.3 Result

See Figure 1 below.

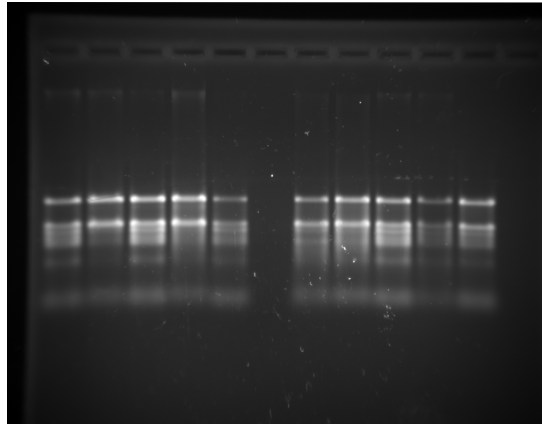
Gel indicates some degradation of RNA, however most samples are OK. Sample order is (left to right) A2, A3, A5, A6, A7, B3, B5, B7. A7 appears to have no RNA, although this is probably a misloading error. Overall, the TBE gel appears to be of more use than the MOPS gel.

## 3 MOPS gel

### 3.1 Aim

- Determine quality of RNA and Compare MOPS with TBE for RNA gels

Figure 1: TBE Gel of Practice RNA samples, 2013-01-22

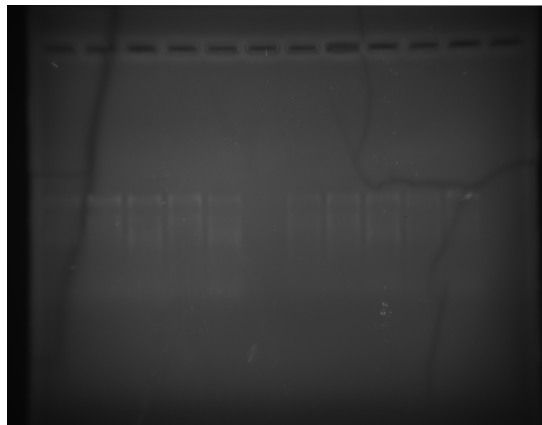


## 3.2 Method

- Load samples after denaturing as above. Sample order is (left to right) A2, A3, A5, A6, A7, B3, B5, B7.
- Run gel in RNase free 1x MOPS at  $\approx 80V$ ,  $\approx 100mA$  for  $\approx 1.75h$  as per TBE gel above.
- Stain gel in 0.5ug/ml Ethidium Br in DEPC water?? for 10 min on orbital shaker.
- Destain on orbital shaker in 1x MOPS, and photograph. Gel disintegrated whilst destaining.

## 3.3 Results

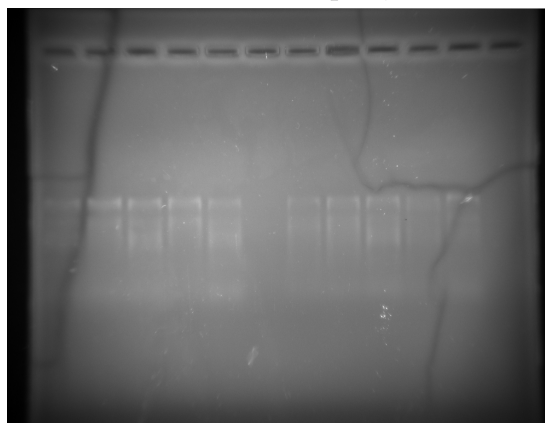
Figure 2: MOPS Gel of Practice RNA samples, 2013-01-22



See Figures 2 and 3

Mops gel confirms that the rna was of reasonable quality. The MOPS gel appears to be of less use than the TBE gel.

Figure 3: MOPS Gel of Practice RNA samples, 2013-01-22. Contrast adjusted.





# Tue 2013-01-29

## 1 Seed Stock Levels

The stocks of Joost's RIX set were checked. Seed lines were classified as having either plenty (+), limited(?) or no (-) seed. The levels of each line are shown in the table below.

Line	Desc	Count	Line	Desc	Count
1	Col-0 <sub>1</sub> 915	not in box	51	168 x 22	+
2	Col-0 <sub>4</sub> 936	not in box	52	169 x 175	not in box
3	Cvi x Cvi	-	53	17 x 21	+
4	1 x Cvi	not in box	54	170 x 24	-
5	1 x 146	+	55	171 x 143	+
6	10 x 26	+	56	174 x 34	+
7	101 x 176	+	57	180 x 157	?
8	Ll-1	not in box	58	183 x 118	-
9	105 x 145	not in box	59	186 x 27	+
10	107 x 124	not in box	60	187 x 190	-
11	109 x 185	not in box	61	187 x 69	not in box
12	109 x 47	not in box	62	189 x 133	-
13	110 x 32	?	63	19 x 173	+
14	112 x 30	+	64	19 x 67	+
15	113 x 141	-	65	190 x 176	+
16	114 x 3	?	66	191 x 31	not in box
17	114 x 60	+	67	192 x 189	+
18	115 x 126	+	68	20 x 138	+
19	117 x 73	?	69	21 x 22	-
20	118 x 108	+	70	24 x 171	+
21	118 x 164	+	71	25 x 9	+
22	119 x 177	-	72	26 x 74	+
23	12 x 142	+	73	33 x 58	not in box
24	122 x 42	?	74	35 x 120	-
25	125 x 117	+	75	38 x 35	+
26	128 x 6	+	76	39 x 27	not in box
27	132 x 129	not in box	77	40 x 74	-
28	133 x 35	+	78	npq4	not in box
29	134 x 29	+	79	43 x 131	not in box
30	135 x 10	+	80	44 x 50	+
31	135 x 140	?	81	45 x 23	+
32	136 x 102	+	82	46 x 29	+
33	165 x 137	not in box	83	48 x 160	+
34	139 x 162	-	84	49 x 158	+
35	139 x 36	+	85	5 x 172	not in box
36	14 x 4	+	86	5 x 188	not in box
37	146 x 64	not in box	87	51 x 111	+
38	147 x 50	+	88	51 x 18	+
39	147 x 69	+	89	54 x 183	+

Line	Desc	Count	Line	Desc	Count
40	149 x 165	+	90	55 x 18	+
41	150 x 37	+	91	59 x 116	+
42	152 x 42	+	92	6 x 131	+
43	153 x 108	+	93	61 x 162	+
44	153 x 20	?	94	63 x 151	not in box
45	154 x 144	-	95	7 x 46	-
46	156 x 166	+	96	8 x 61	not in box
47	16 x 4	+	97	Ler x Ler	+
48	16 x 66	+	98	Ler self	+
49	164 x 7	not in box	99	Cvi x Ler	+
50	166 x 25	+	100	Ler x Cvi	+