# DIGITAL LAB NOTEBOOK OF KEVIN MURRAY

# Honous Project, 2013

Jointly supervised by Justin Borevitz and Barry Pogson

# Mon 2012-12-10

# 1 Final Harvest of Keng's RIX lines

### 1.1 Aim

Harvest lines after 1 week of HL stress.

### 1.2 Method

- An Eppendorf 1.2mL deep well plate was placed on dry ice for  $\approx 10$  minutes before sampling to allow to cool.
- Whole leaves were excised and placed into 1.2mL Eppendorf 96 deep well plate.
- Where possible, the largest mature leaf was taken. In some cases, this was hard to determine, so the youngest of the fully-expanded leaves was taken (as this was generally also the largest leaf). Some plants were very small, and had only juvenile leaves, in which case the largest juvenile leaf was taken.

## 1.3 Results

The following table describes the plate layout.

	Well	Line	Comments	Well	Line	Comme	ents
A1	87	'			A7	65	
B1	100				В7	71	
C1	92				C7	53	
D1	OSB2				D7	55	
E1	93				E7	56	
F1	94				F7	50	
G1	1*cvi				G7	51	
H1	99				H7	52	
A2	63				A8	78	NPQ4
B2	98				B8	80	
C2	98	2nd pl	ant with oran	ige labe	el   C8	85	
D2	70				D8	74	
E2	67				E8	73	
F2	99				F8	76	
G2	66				G8	83	
H2	100				H8	81	
A3	65				A9	89	
В3	72				B9	90	
С3	12				C9	91	
D3	11				D9	16	
E3	10				E9	17	
F3	12				F9	18	

	Well	Line	Comments	Well	Line	Comments
G3	9	•			G9	21
Н3	8				Н9	23
A4	7				A1	$0 \mid 25$
B4	6				B10	57
C4	5				C10	0 28
D4	13				D1	0 29
E4	2				E10	0 30
F4	1*cvi				F10	31
G4	49				G1	0
H4	47				H1	0   0
A5	46				A1	1
B5	42				B1	1
C5	45				C1	1
D5	71				D1	1
E5	39				E13	1
F5	40				F13	L
G5	41				G1	1
H5	43				H1	1
A6	38				A1:	2
B6	39				B1:	2
C6	36				C1:	2
D6	33				D1:	2
E6	61				E1:	2
F6	62				F12	2
G6	68				G1	2
H6	58				H1:	2

### Attachments:

 $\bullet \ dec 12/20121210 - harvest\_photos.tar.bz 2 \\ MD5SUM: 40 dae 2 cad 3 babaa 3 c 32 f 0 d 35 a 9 d 9 4 4 2 c \\ Additional of the control o$ 

# Mon 2013-01-21

# 1 Quantification of RNA samples

### 1.1 Aim

• Deterime qty of RNA in previously extracted samples

### 1.2 Method

- Nanodropped RNA exaction from 15/1/13??
- Standard protocol, used sterile milliQ water as blank.

#### 1.3 Result

• Of the 14 samples, 10 had reasonable amounts of RNA, and 260/280 ratios were above 1.8 in all but one case. (see ./jan/20130121-PracticeRNASamples.ods)

## 1.4 Attachements

- ./jan/20130121-PracticeRNAExtractionSamples.csv
- ./jan/20130121-PracticeRNAExtractionSamples.ndv
- ./jan/20130121-PracticeRNASamples.ods

## 2 MADE: 10x MOPS Solution

#### Method

- Add 41.8g RNA only MOPS to beaker
- Add 450mL DEPC H2O, mix w/ stirrer bar on mag stirrer
- Add 26.6mL 3M Sodium Acetate (0.22um Filtered before use)
- Add 10mL RNA only 0.5M EDTA
- pH to 7 with 5M NaOH
- Top up to 500 mL with DEPC H2O
- Use 10ml per 100mL MOPS gel

# 3 MADE: RNA Denaturing Gel (MOPS)

#### Method

- $\bullet\,$  Melt 1g RNAse-free Agarose in 72ml DEPC H2O
- Add 10mL 10x MOPS
- Add 18mL 37% Formaldehyde
- $\bullet$  Pour in RNA-only gel tank, previously washed with 0.5% SDS and RNAse-zap

# Tue 2013-01-22

# 1 Denature RNA for RNA gels

#### 1.1 Method

- Dilute RNA to 100ng/uL
- Add RNA gel loading buffer (Obtained from Pete Crisp)
- Incubate at 65 degrees for 10 minutes. The samples were incubated for 10 minutes on the evening of 2013-01-21, but the gels were not run until 2013-01-22, so they were denatured for a futher 2 minutes at 65 degrees

#### 2 TBE Gel

### 2.1 Aim

• To compare TBE and denaturing/MOPS gels for RNA

## 2.2 Method

- Dissolve 1g RNAase-free agarose in 90mL DEPC water
- Add 10mL RNAse-free TBE (prepared using DEPC Water, obtained from Pete Crisp)
- Pour in RNA-only gel tank, previously washed with 0.5% SDS or RNAse-zap
- Then, load denatured samples, and run in RNAse-free 1x TBE
- Run at  $\approx 80 \text{V}$ ,  $\approx 40\text{-}50 \text{mA}$  for  $\approx 1.75 \text{h}$
- Stain gel in 0.5ug/ml Ethidium Br in DEPC water?? for 10 min on orbital shaker, and photograph.

## 2.3 Result

See Figure ?? below.

Gel indicates some degradation of RNA, however most samples are OK. Sample order is (left to right) A2, A3, A5, A6, A7, B3, B5, B7. A7 appears to have no RNA, although this is probably a misloading error. Overall, the TBE gel appears to be of more use than the MOPS gel.

# 3 MOPS gel

# 3.1 Aim

• Determine qualtiy of RNA and Compare MOPS with TBE for RNA gels

Figure 1: TBE Gel of Practice RNA samples, 2013-01-22

## 3.2 Method

- Load samples after denaturing as above. Sample order is (left to right) A2, A3, A5, A6, A7, B3, B5, B7.
- Run gel in RNAse free 1x MOPS at  $\approx 80 \text{V}$ ,  $\approx 100 \text{mA}$  for  $\approx 1.75 \text{h}$  as per TBE gel above.
- Stain gel in 0.5ug/ml Ethidium Br in DEPC water?? for 10 min on orbital shaker.
- Destain on orbital shaker gel in 1x MOPS, and photograph. Gel disintegrated whilst destaining.

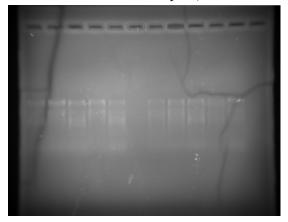
## 3.3 Results

Figure 2: MOPS Gel of Practice RNA samples, 2013-01-22

See Figures?? and??

Mops gel conifrms that the rna was of reasonable quality. The MOPS gel appears to be of less use than the TBE gel.

Figure 3: MOPS Gel of Practice RNA samples, 2013-01-22. Contrast adjusted.



# Tue 2013-01-29

# 1 Seed Stock Levels

The stocks of Joost's RIX set were checked. Seed lines were classified as having either plenty (+), limited(?) or no (-) seed. The levels of each line are shown in the table below.

	Line D	esc   Count	Line	Desc Cou	int
1	$Col-0_1915$	not in box	51	168 x 22	<u> </u>
2	$Col-0_4936$	not in box	52	$169 \times 175$	not in box
3	Cvi x Cvi	-	53	$17 \times 21$	+
4	$1 \times \text{Cvi}$	not in box	54	$170 \times 24$	_
5	$1 \times 146$	+	55	171 x 143	+
6	$10 \times 26$	+	56	$174 \times 34$	+
7	$101 \times 176$	+	57	$180 \times 157$	?
8	Ll-1	not in box	58	183 x 118	-
9	$105 \times 145$	not in box	59	$186 \times 27$	+
10	$107 \times 124$	not in box	60	$187 \times 190$	-
11	$109 \ge 185$	not in box	61	$187 \times 69$	not in box
12	$109 \ge 47$	not in box	62	189 x 133	-
13	$110 \ge 32$	?	63	$19 \times 173$	+
14	$112 \ge 30$	+	64	$19 \times 67$	+
15	$113 \times 141$	-	65	$190 \times 176$	+
16	$114 \times 3$	?	66	$191 \times 31$	not in box
17	$114 \pm 60$	+	67	$192 \times 189$	+
18	$115 \times 126$	+	68	$20 \times 138$	+
19	$117 \times 73$	?	69	$21 \times 22$	_
20	$118 \times 108$	+	70	$24 \times 171$	+
21	$118 \times 164$	+	71	$25 \times 9$	+
22	$119 \times 177$	-	72	26 x 74	+
23	$12 \times 142$	+	73	$33 \times 58$	not in box
24	$122 \ge 42$	?	74	$35 \times 120$	-
25	$125 \times 117$	+	75	$38 \times 35$	+
26	$128 \times 6$	+	76	$39 \times 27$	not in box
27	$132 \times 129$	not in box	77	40 x 74	-
28	$133 \pm 35$	+	78	npq4	not in box
29	$134 \times 29$	+	79	43 x 131	not in box
30	$135 \times 10$	+	80	$44 \times 50$	+
31	$135 \times 140$	?	81	$45 \times 23$	+
32	$136 \times 102$	+	82	46 x 29	+
33	$165 \times 137$	not in box	83	48 x 160	+
34	$139 \times 162$	-	84	49 x 158	+
35	$139 \ge 36$	+	85	5 x 172	not in box
36	14 x 4	+	86	5 x 188	not in box
37	$146 \pm 64$	not in box	87	51 x 111	+
38	$147 \pm 50$	+	88	51 x 18	+
39	$147 \times 69$	+	89	54 x 183	+

	Line D	esc Count	Line	Desc Cou	int
40	$149 \times 165$	+	90	55 x 18	+
41	$150 \times 37$	+	91	$59 \times 116$	+
42	$152 \times 42$	+	92	$6 \times 131$	+
43	$153 \times 108$	+	93	$61 \times 162$	+
44	$153 \times 20$	?	94	$63 \times 151$	not in box
45	$154 \times 144$	_	95	$7 \times 46$	-
46	$156 \times 166$	+	96	$8 \times 61$	not in box
47	16 x 4	+	97	Ler x Ler	+
48	16 x 66	+	98	Ler self	+
49	$164 \times 7$	not in box	99	Cvi x Ler	+
50	1 <u>66 x 25</u>	+	100	Ler x Cvi	+