# DIGITAL LAB NOTEBOOK OF KEVIN MURRAY

## Honous Project, 2013

Jointly supervised by Justin Borevitz and Barry Pogson

## 2013-01-21

### 1 Practice RNA Exraction

### 1.1 Quantification of RNA samples

#### Aim

• Deterime qty of RNA in previously extracted samples

#### Method

- Nanodropped RNA exaction from 15/1/13??
- Standard protocol, used sterile milliQ water as blank.

#### Result

• Of the 14 samples, 10 had reasonable amounts of RNA, and 260/280 ratios were above 1.8 in all but one case. (see ./jan/20130121-PracticeRNASamples.ods)

#### Attachements

- $\bullet \ ./jan/20130121-Practice RNA Extraction Samples.csv$
- ./jan/20130121-PracticeRNAExtractionSamples.ndv
- ./jan/20130121-PracticeRNASamples.ods

### 2 MADE: 10x MOPS Solution

#### Method

- Add 41.8g RNA only MOPS to beaker
- Add 450mL DEPC H2O, mix w/ stirrer bar on mag stirrer
- Add 26.6mL 3M Sodium Acetate (0.22um Filtered before use)
- Add 10mL RNA only 0.5M EDTA
- pH to 7 with 5M NaOH
- Top up to 500 mL with DEPC H2O
- Use 10ml per 100mL MOPS gel

# 3 MADE: RNA Denaturing Gel (MOPS)

### Method

- $\bullet\,$  Melt 1g RNAse-free Agarose in 72ml DEPC H2O
- Add 10mL 10x MOPS
- $\bullet$  Add 18mL 37% Formaldehyde
- $\bullet\,$  Pour in RNA-only gel tank, previously washed with 0.5% SDS and RNAse-zap