

# DIGITAL LAB NOTEBOOK OF KEVIN MURRAY

HONOUS PROJECT, 2013

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*Last updated at 19:14 on Tuesday 22<sup>nd</sup> January, 2013*



# 2013-01-21

## 1 Practice RNA Extraction

### 1.1 Quantification of RNA samples

#### *Aim*

- Determine qty of RNA in previously extracted samples

#### *Method*

- Nanodropped RNA extraction from 15/1/13??
- Standard protocol, used sterile milliQ water as blank.

#### *Result*

- Of the 14 samples, 10 had reasonable amounts of RNA, and 260/280 ratios were above 1.8 in all but one case. (see ./jan/20130121-PracticeRNASamples.ods)

#### *Attachments*

- ./jan/20130121-PracticeRNAExtractionSamples.csv
- ./jan/20130121-PracticeRNAExtractionSamples.ndv
- ./jan/20130121-PracticeRNASamples.ods

## 2 MADE: 10x MOPS Solution

#### *Method*

- Add 41.8g RNA only MOPS to beaker
- Add 450mL DEPC H<sub>2</sub>O, mix w/ stirrer bar on mag stirrer
- Add 26.6mL 3M Sodium Acetate (0.22um Filtered before use)
- Add 10mL RNA only 0.5M EDTA
- pH to 7 with 5M NaOH
- Top up to 500 mL with DEPC H<sub>2</sub>O
- Use 10ml per 100mL MOPS gel

## 3 MADE: RNA Denaturing Gel (MOPS)

#### *Method*

- Melt 1g RNase-free Agarose in 72ml DEPC H<sub>2</sub>O
- Add 10mL 10x MOPS
- Add 18mL 37% Formaldehyde
- Pour in RNA-only gel tank, previously washed with 0.5% SDS and RNase-zap