

# K-Means Spectral Unmixing for Fluorescence Microscopy

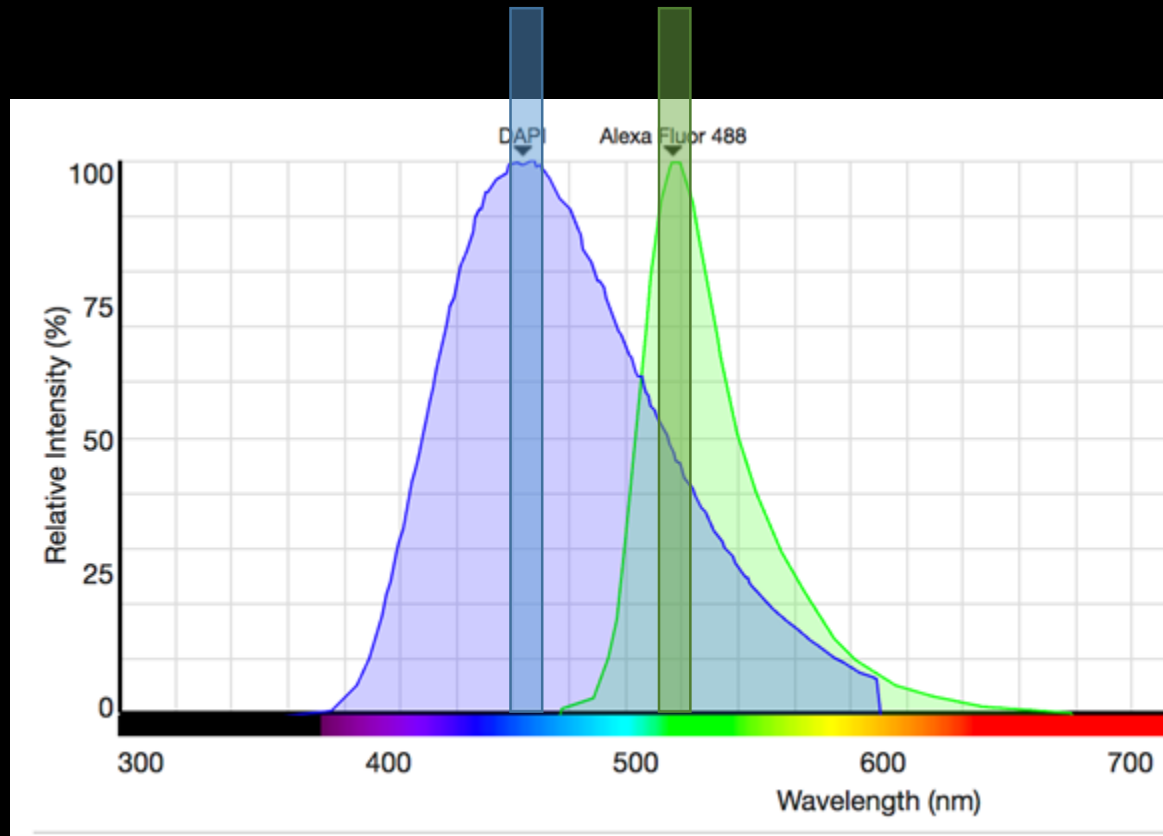
Tristan McRae

Analyst/Programmer

Multiphoton Research Core

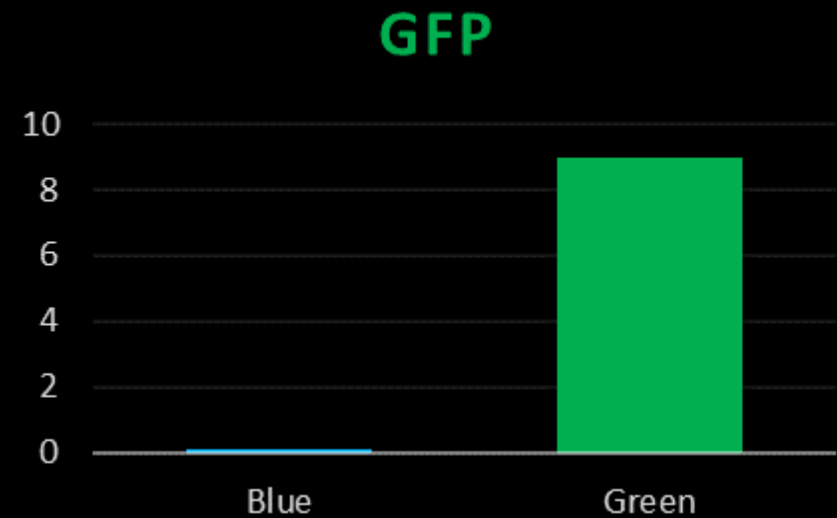
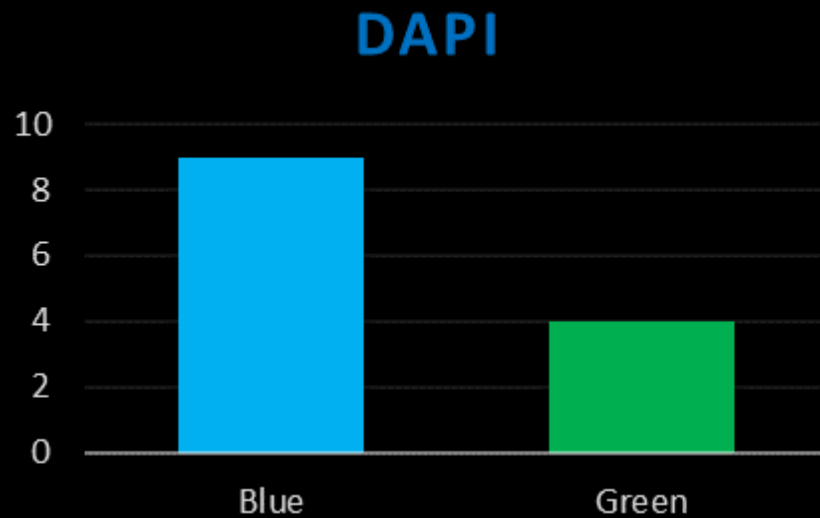
12/5/2018

# Emission spectra overlap



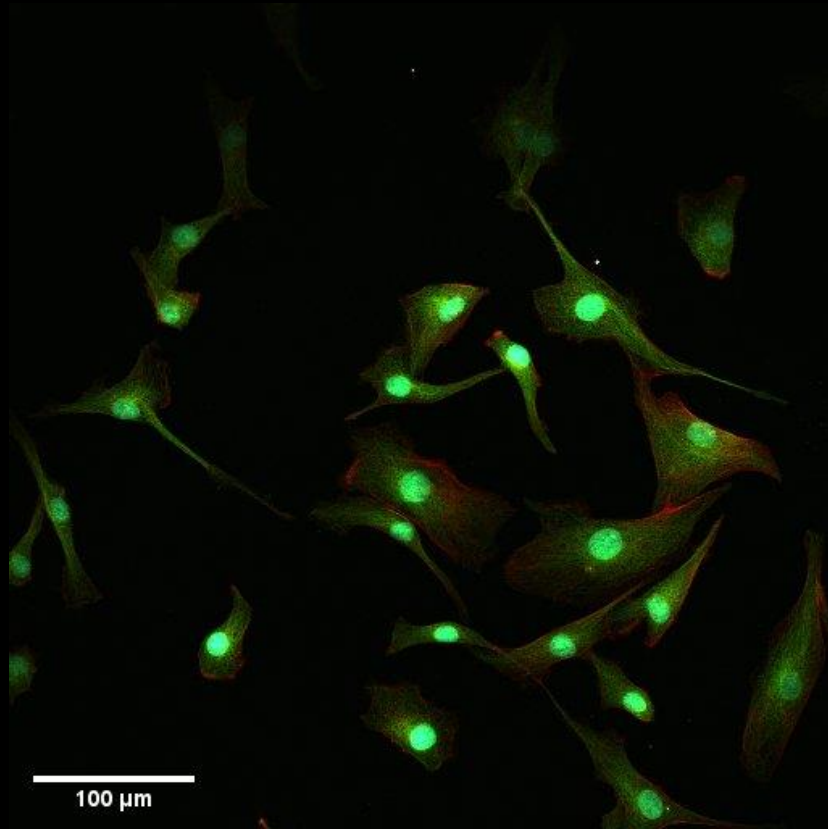
- Overlapping emission spectra of fluorophores make them show up in multiple channels.
- Different fluorophores have different intensity “signatures” in different channels.

# Fluorophores have Unique Signatures Across Channels



# Channel bleed-through contaminates image data

Nuclei (DAPI) Microtubules (BODIPY) F-actin (Texas Red)

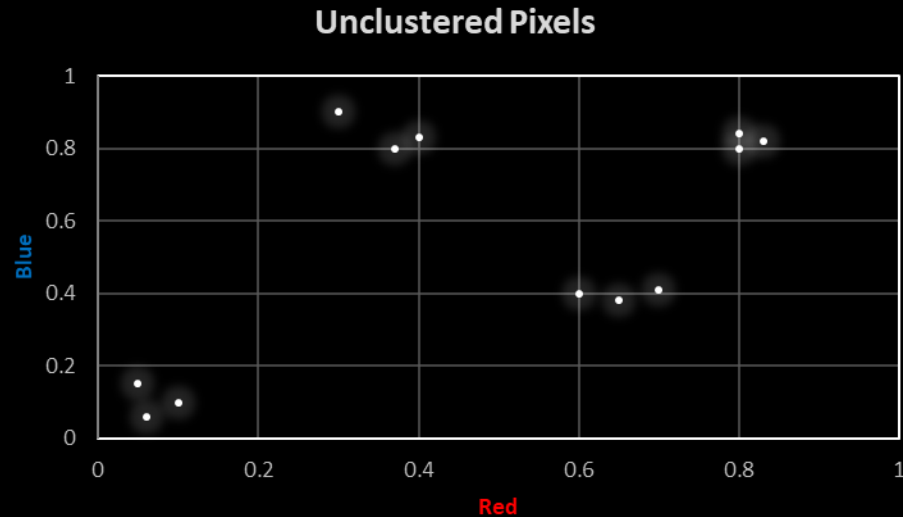


- Nuclei show up in both blue and green channels making the green channel mixes with both nuclei and microtubules information
- Blue channel only has nuclei, while green channel has both nuclei and microtubules.
- Fluorophores have different intensity “signatures” in different channels.

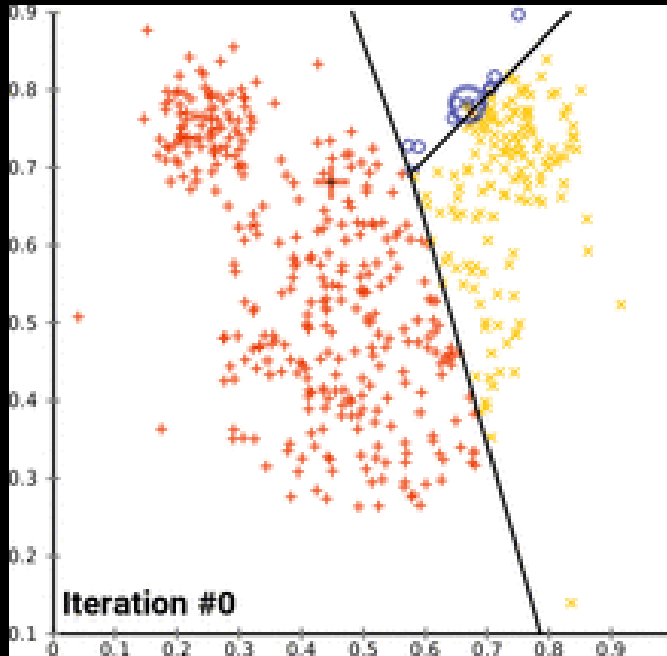
# Pixel intensities group into clusters for different fluorophores

More colors exist than channels to detect them

Individual pixels are grouped into  $K=4$  different clusters based on their intensities in each color channel



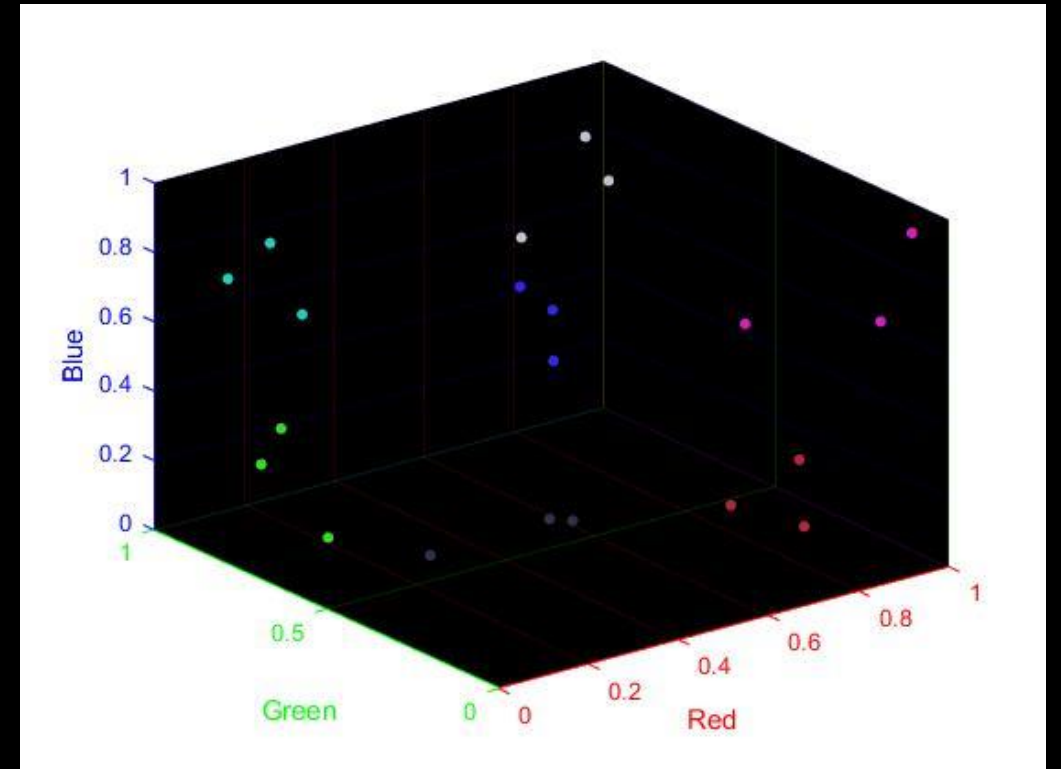
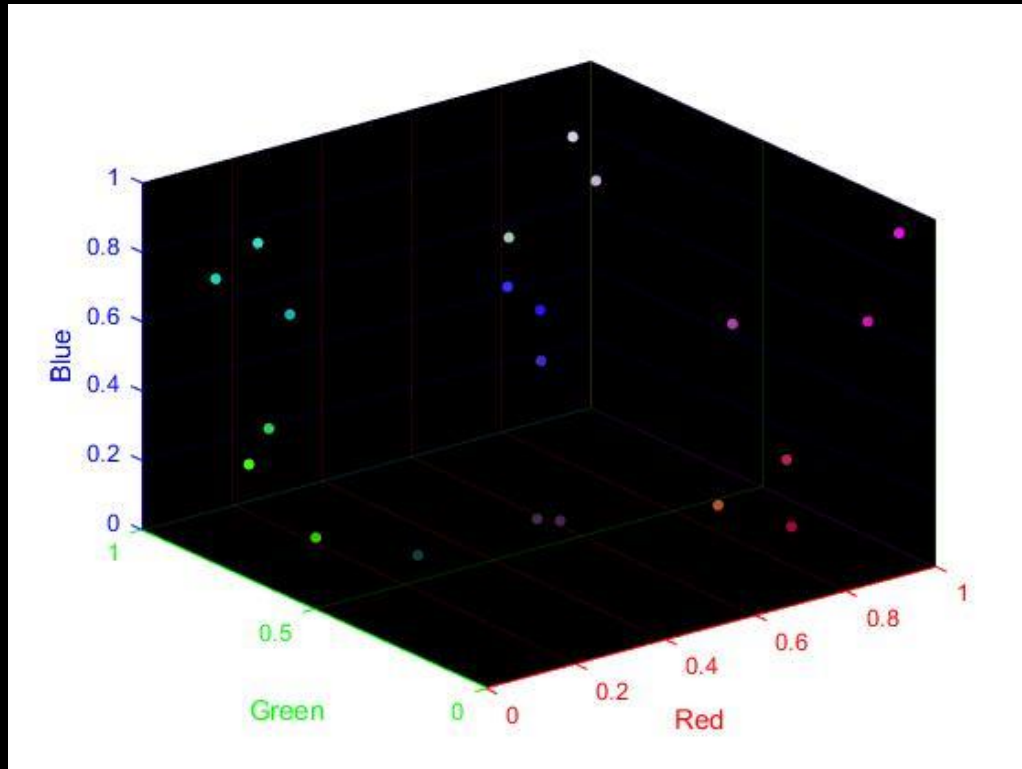
# K-Means Clustering algorithm



K-Means algorithm:

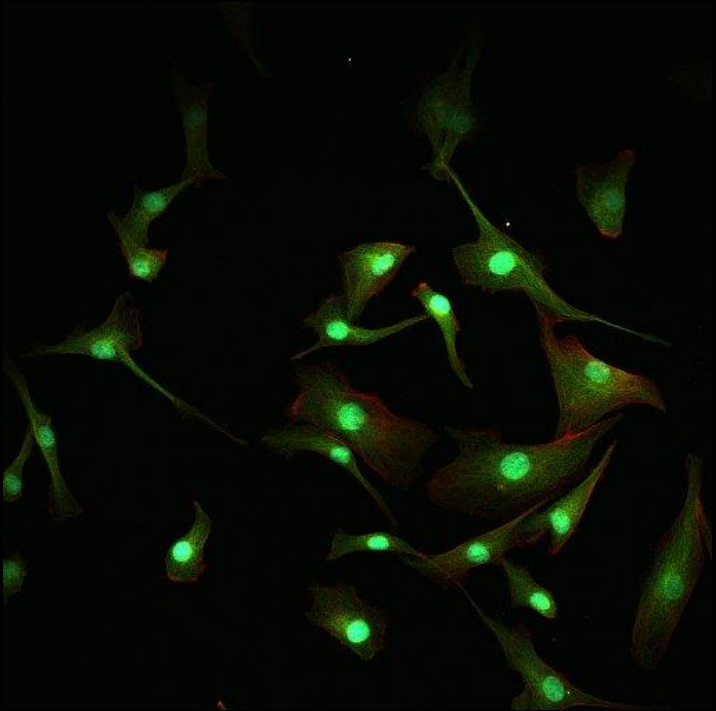
1. Initialize K semi-random cluster centroids or “means”
2. Assign data points to the cluster with the closest mean
3. Compute the new mean of each cluster
4. Iterate steps 2 and 3 until convergence

# K-Means Clustering works for any number of channels

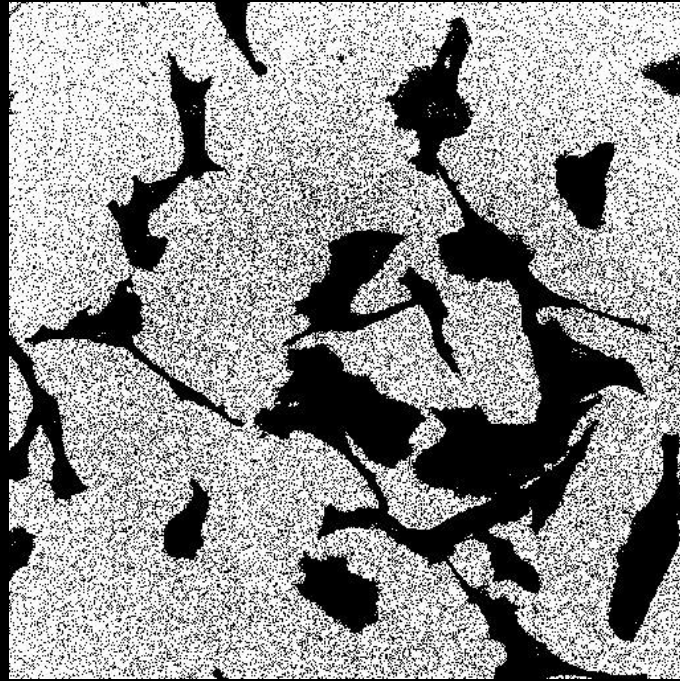


# Background as a cluster to remove background noise

Original Image



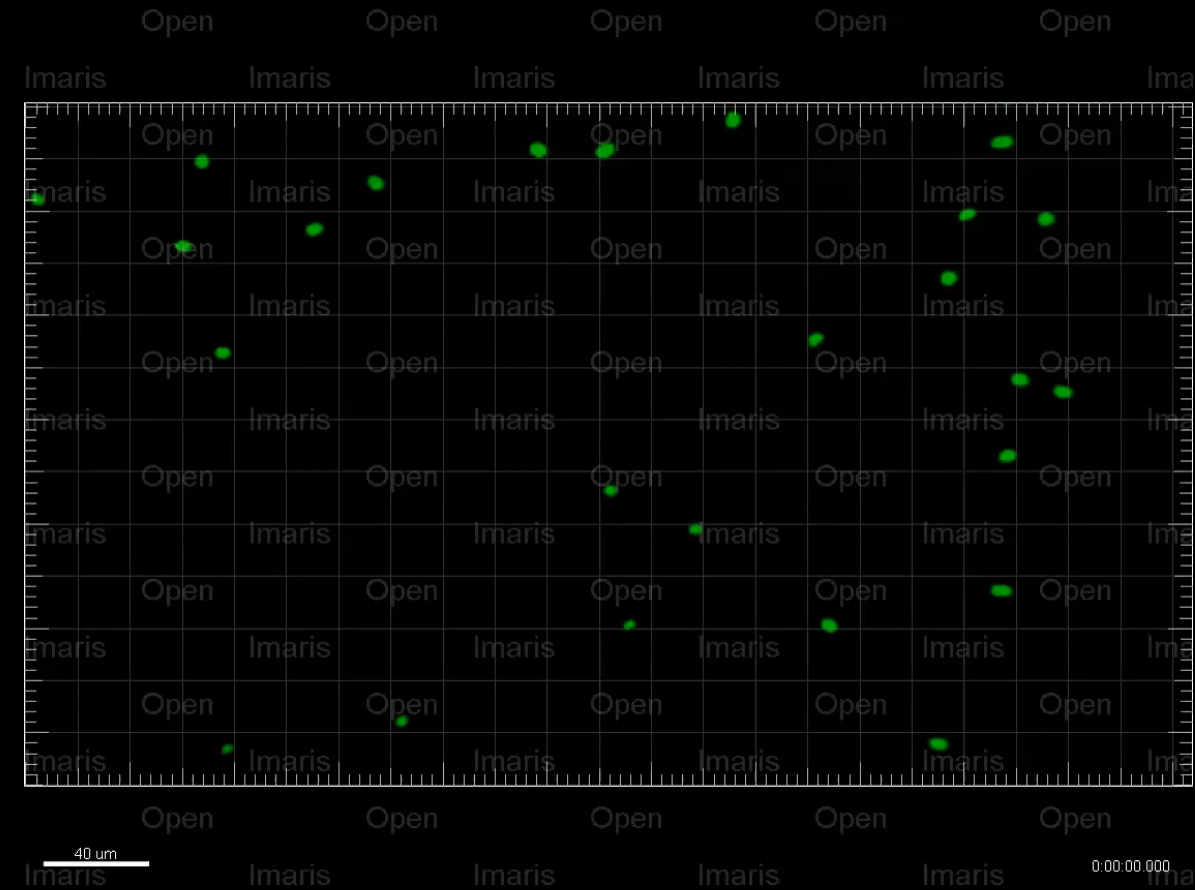
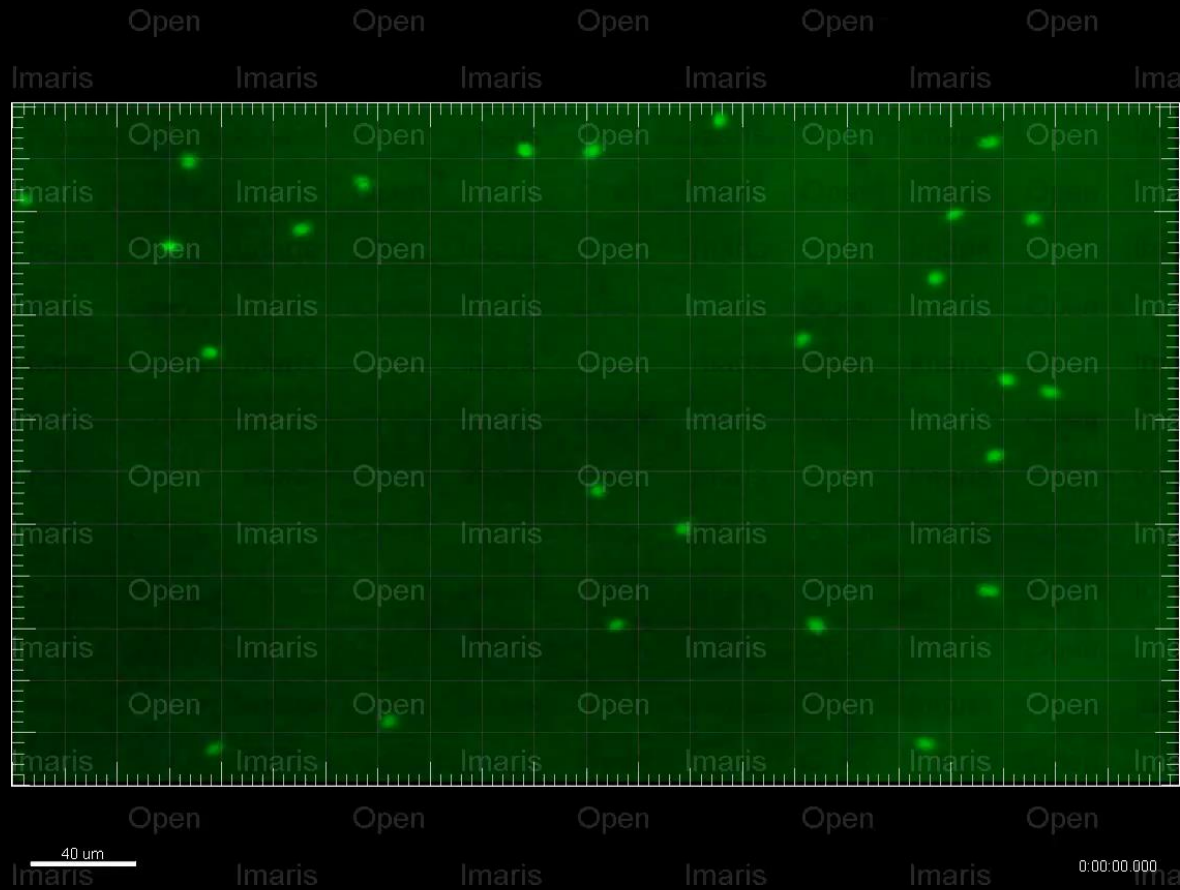
Background Cluster



Pixels with low intensities group together into a cluster that can be removed



# Example: Background Removal on Algae



# Example 1: BPAE Cell Fixed Slide

Original Image

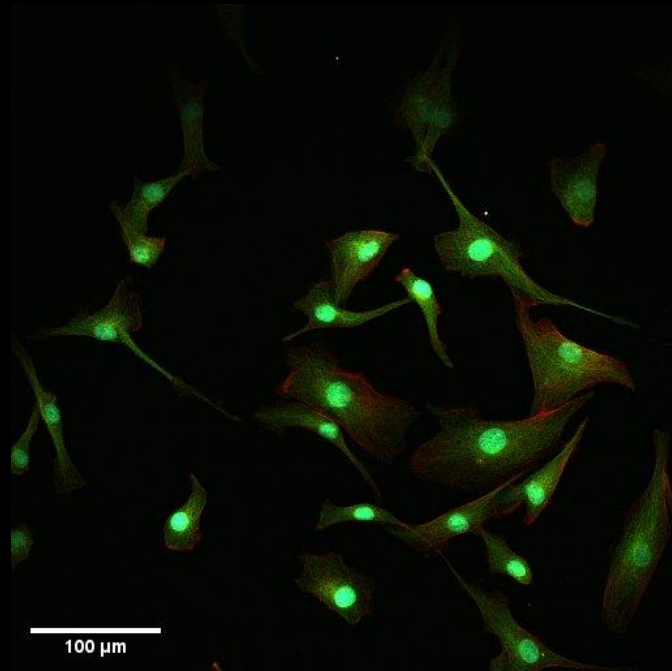
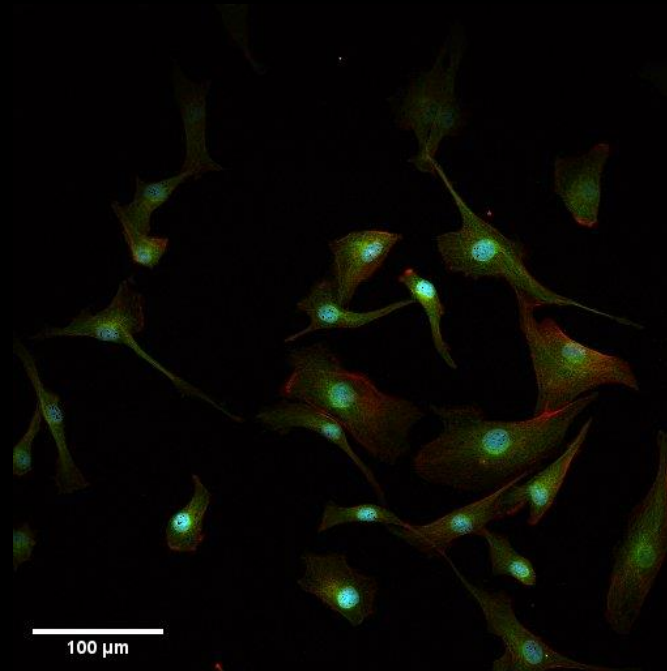


Image unmixed with K - means

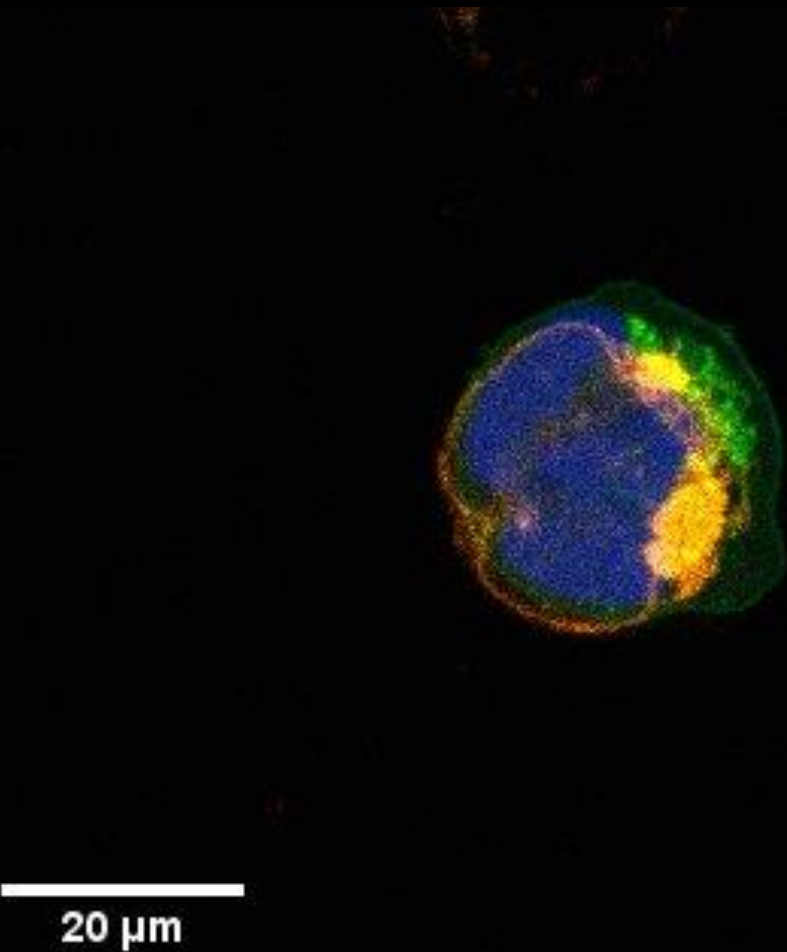


K-means is able to separate out the nuclei from the green channel.

Note the lack of background noise in the unmixed version as the background cluster is easily removed

Nuclei (DAPI) Microtubules (BODIPY) F-actin (Texas Red)

# Example 2: Cultured “Colorful” T Cells



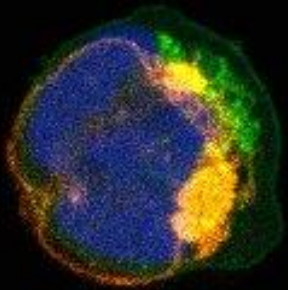
Lasers: MaiTai 840nm, Insight 1050nm,  
Sequential Scan  
Cubes: BFP/GFP, nRed/fRed Cubes

Flour	Structure	Image Color	Wavelength (Excitation, Emission, Cube)		
BFP	Nucleus	Blue	(840nm,	420-500nm,	BFP)
Cerulean	Membrane	Green	(840nm,	465-505nm,	GFP)
Azami Green	Mitochondria	Green	(1050nm,	496-536nm,	GFP)
Citrine	Golgi	Orange	(840nm,	515-565nm,	GFP)
mCherry	ER	Red	(1050nm,	605-635nm,	nRed)
iRFP670	Peroxisomes	Yellow	(1050nm,	650-675nm,	fRed)

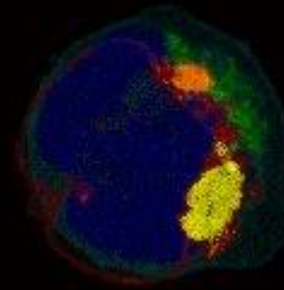
# Example 2: Cultured “Colorful” T Cells

Original

Unmixed



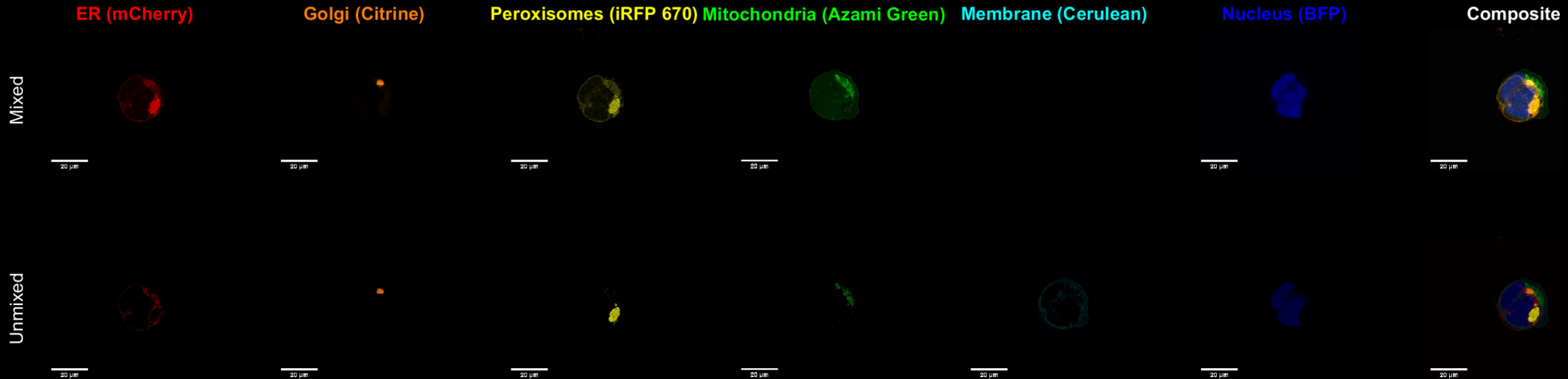
20  $\mu\text{m}$



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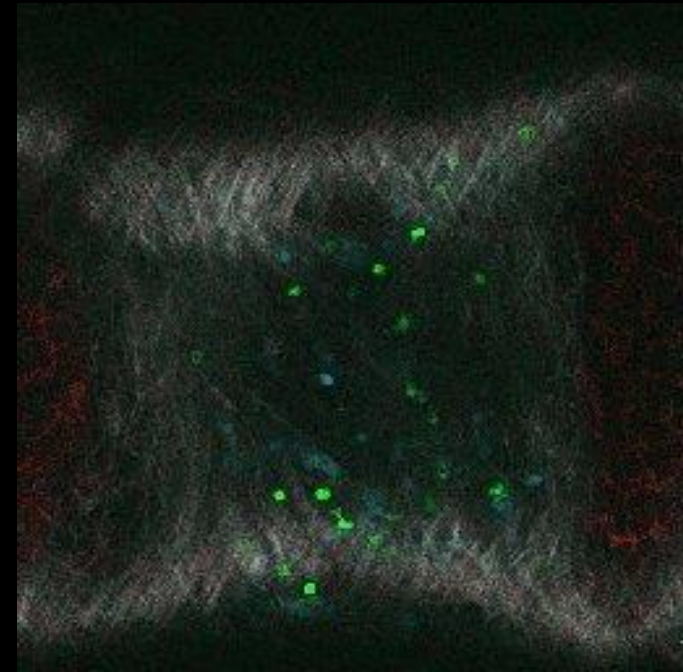
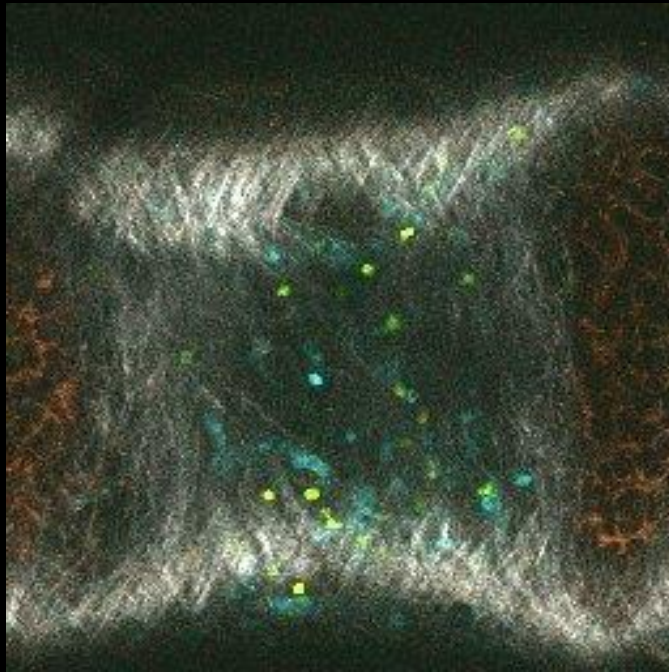
K-means generalizes well in situations with more fluorophores and channels such as this example with 6 fluorophores and 5 channels

# Cultured “Colorful” T Cells Channel Breakdown



Images courtesy of David Oleksyn, Program for Advanced Immune Bioimaging  
Colorful 840-1050 BFP-GFP cube-high qual-3

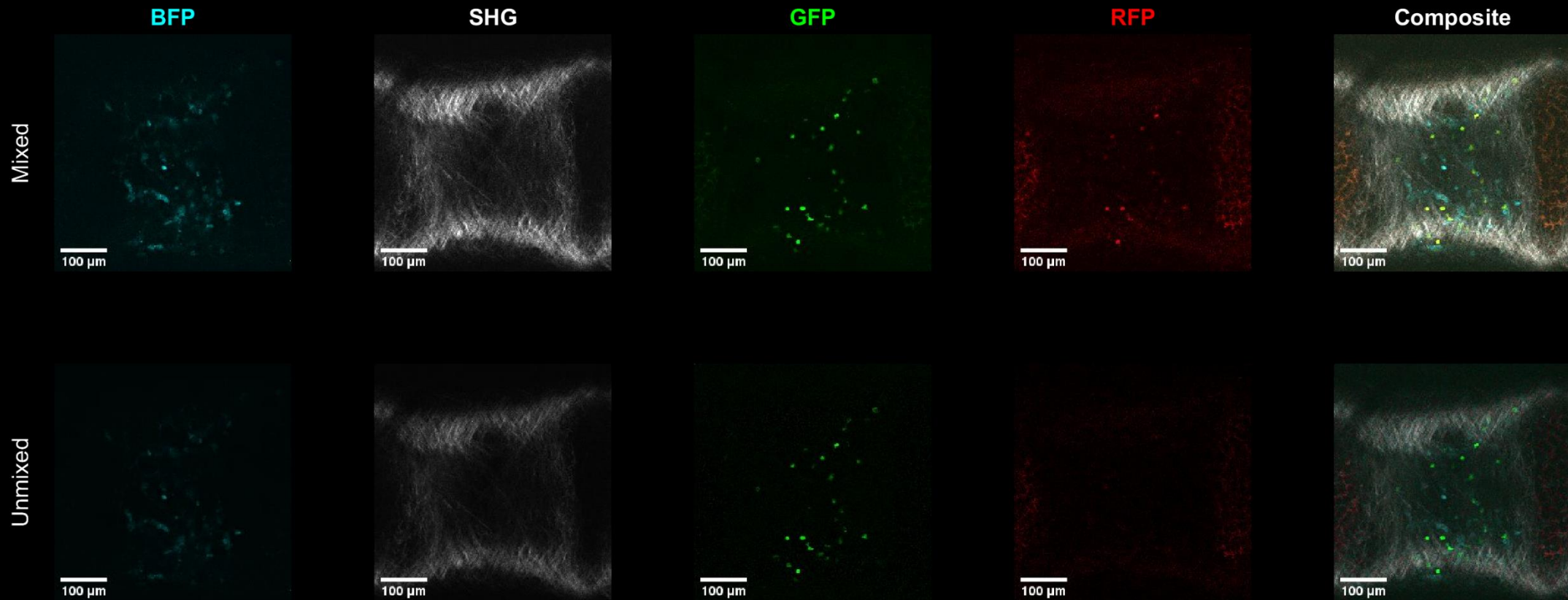
# Example 3: In Vivo Mouse Trachea Sample



Images courtesy of Kris Lambert and Emma Reilly, Program for Advanced Immune Bioimaging  
090618 D7 Rex3 MHC NT-1

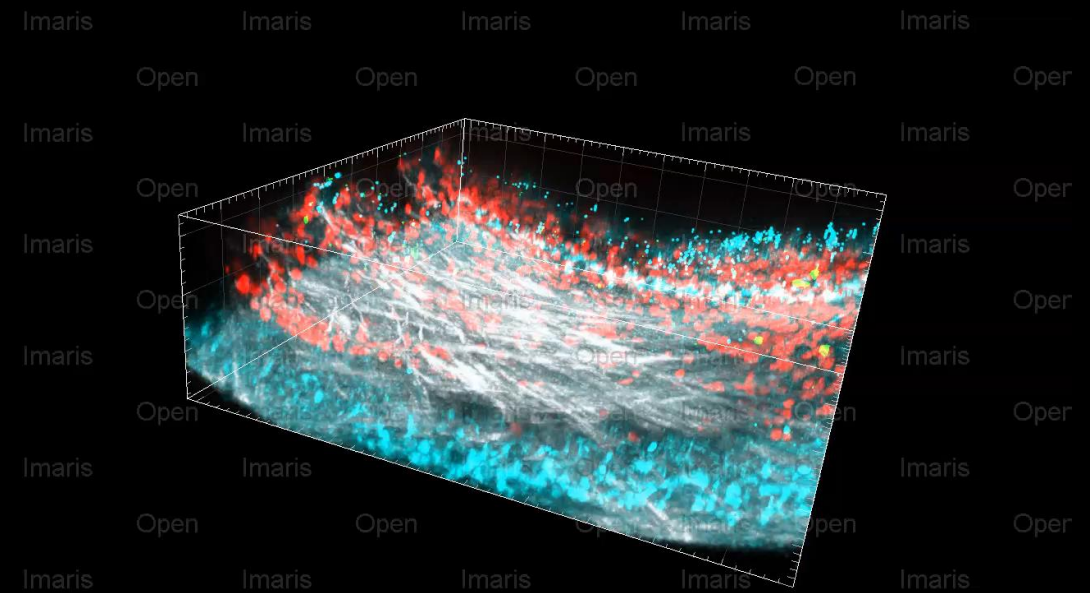
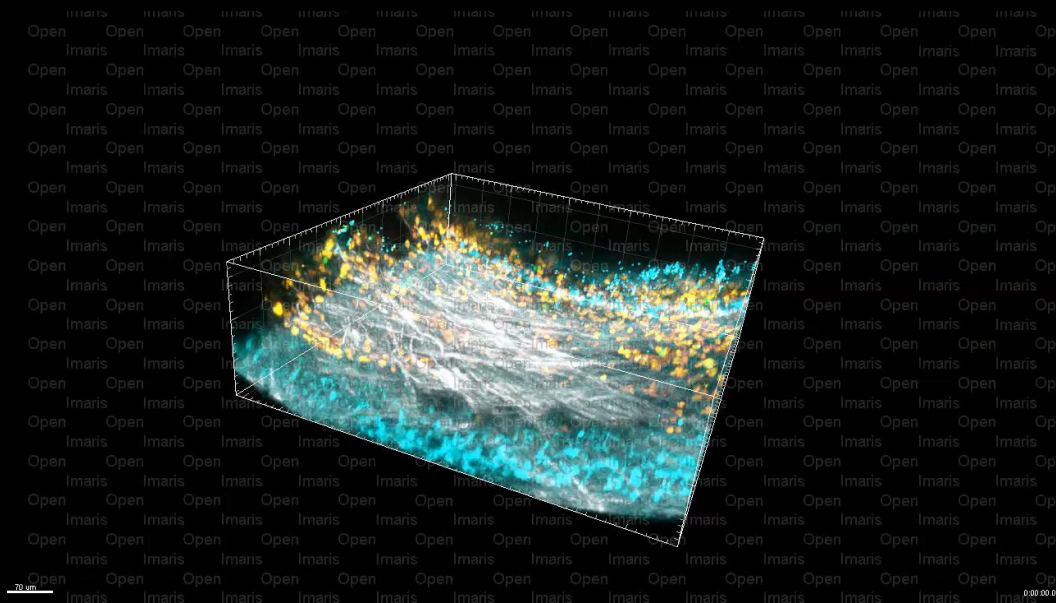


# In Vivo Mouse Trachea Channel Breakdown



Images courtesy of Kris Lambert and Emma Reilly, Program for Advanced Immune Bioimaging  
090618 D7 Rex3 MHC NT-1

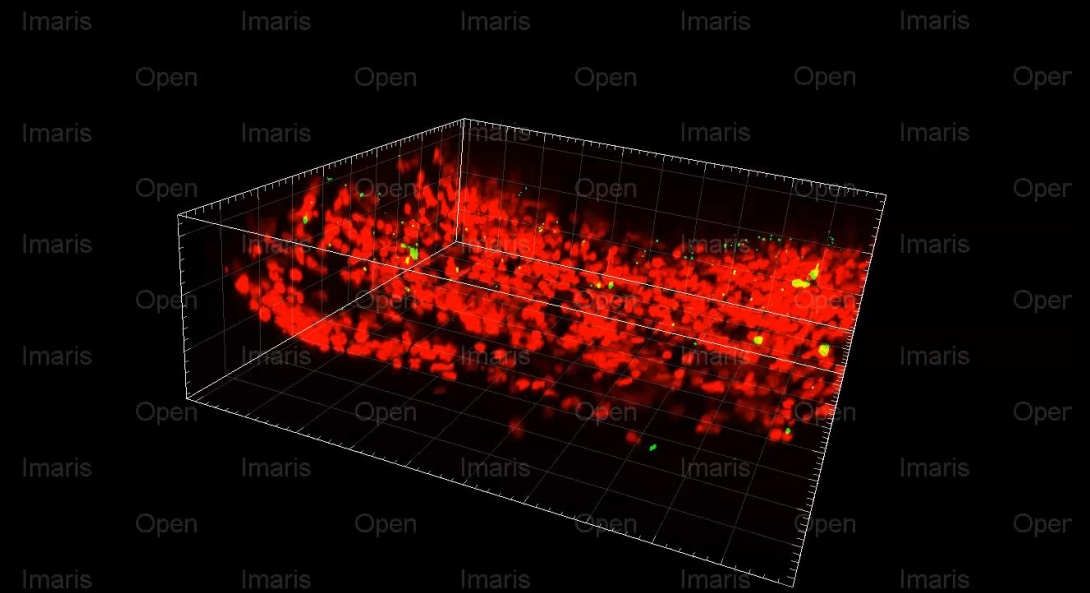
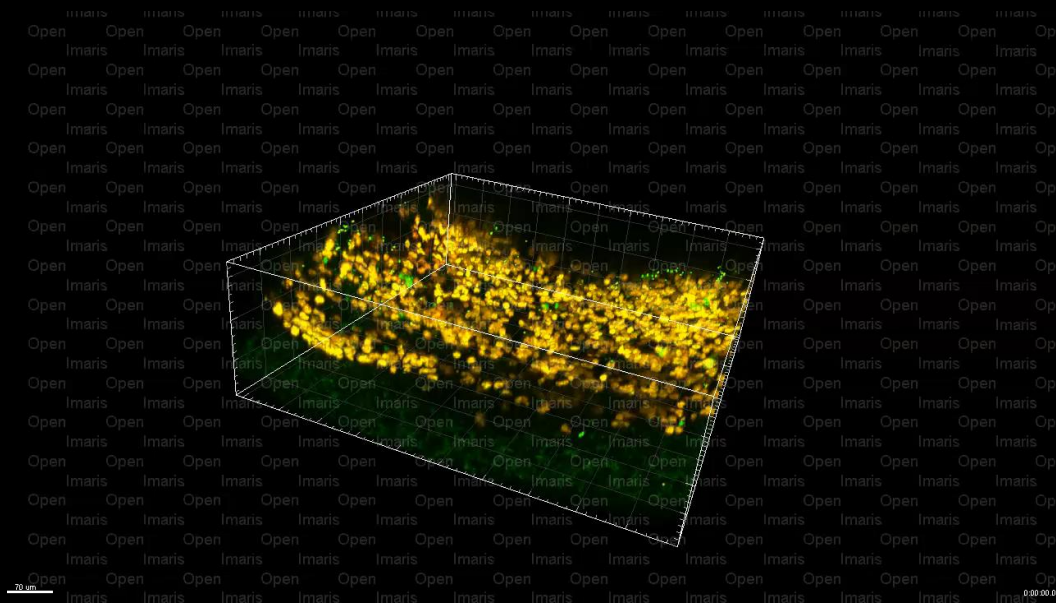
# Unmixing Works for 3D Videos



Images courtesy of Kris Lambert and Emma Reilly, Program for Advanced Immune Bioimaging  
181106\_D7\_OTI TdOTII GFP\_Ecad\_L\_0001

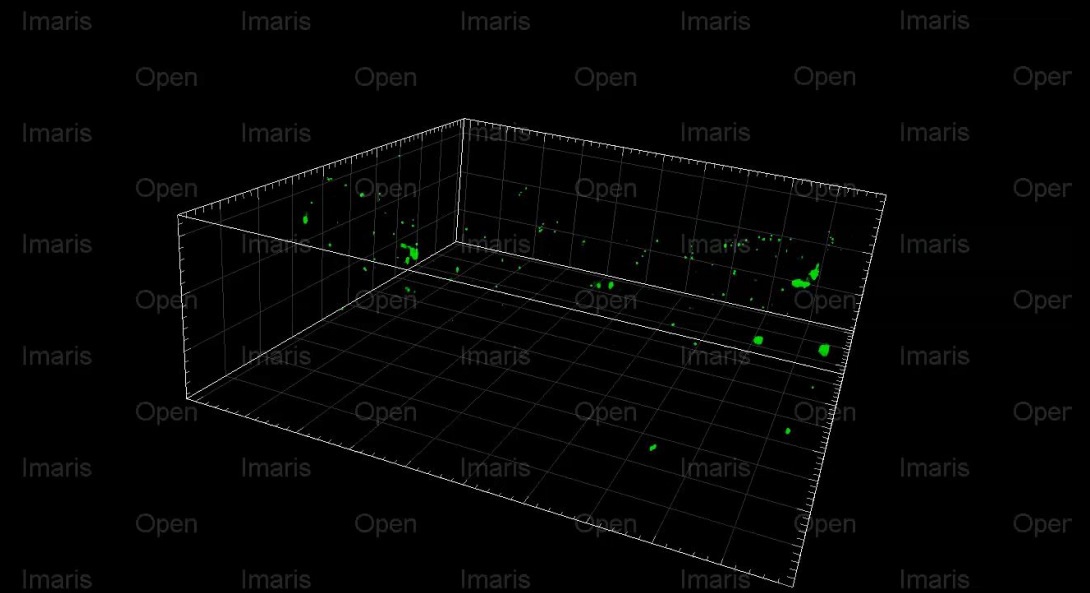
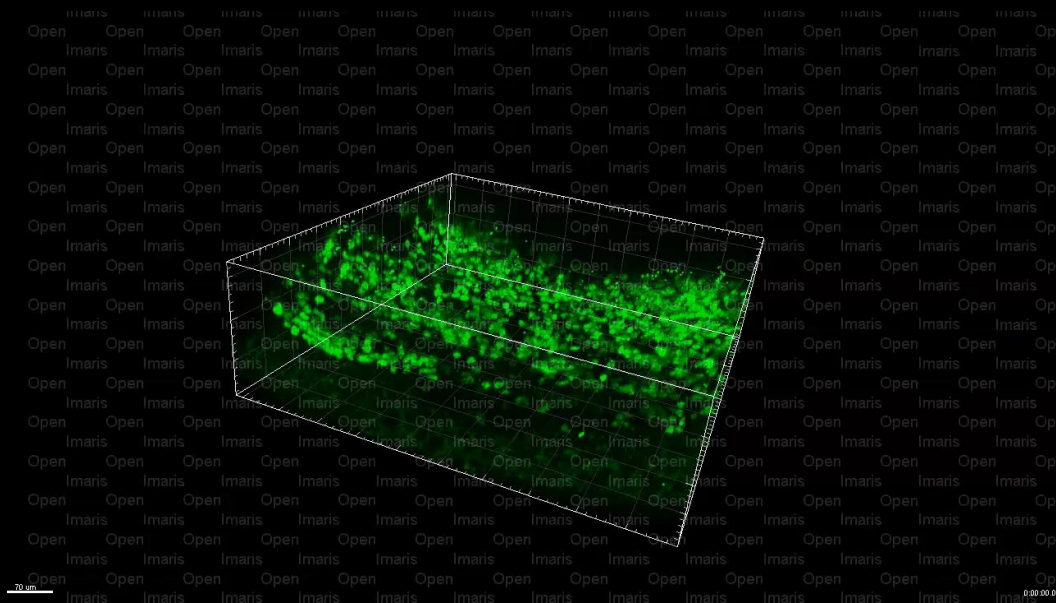


# Unmixing Works for 3D Videos



Images courtesy of Kris Lambert and Emma Reilly, Program for Advanced Immune Bioimaging  
181106\_D7\_OTI TdOTII GFP\_Ecad\_L\_0001

# Unmixing Works for 3D Videos



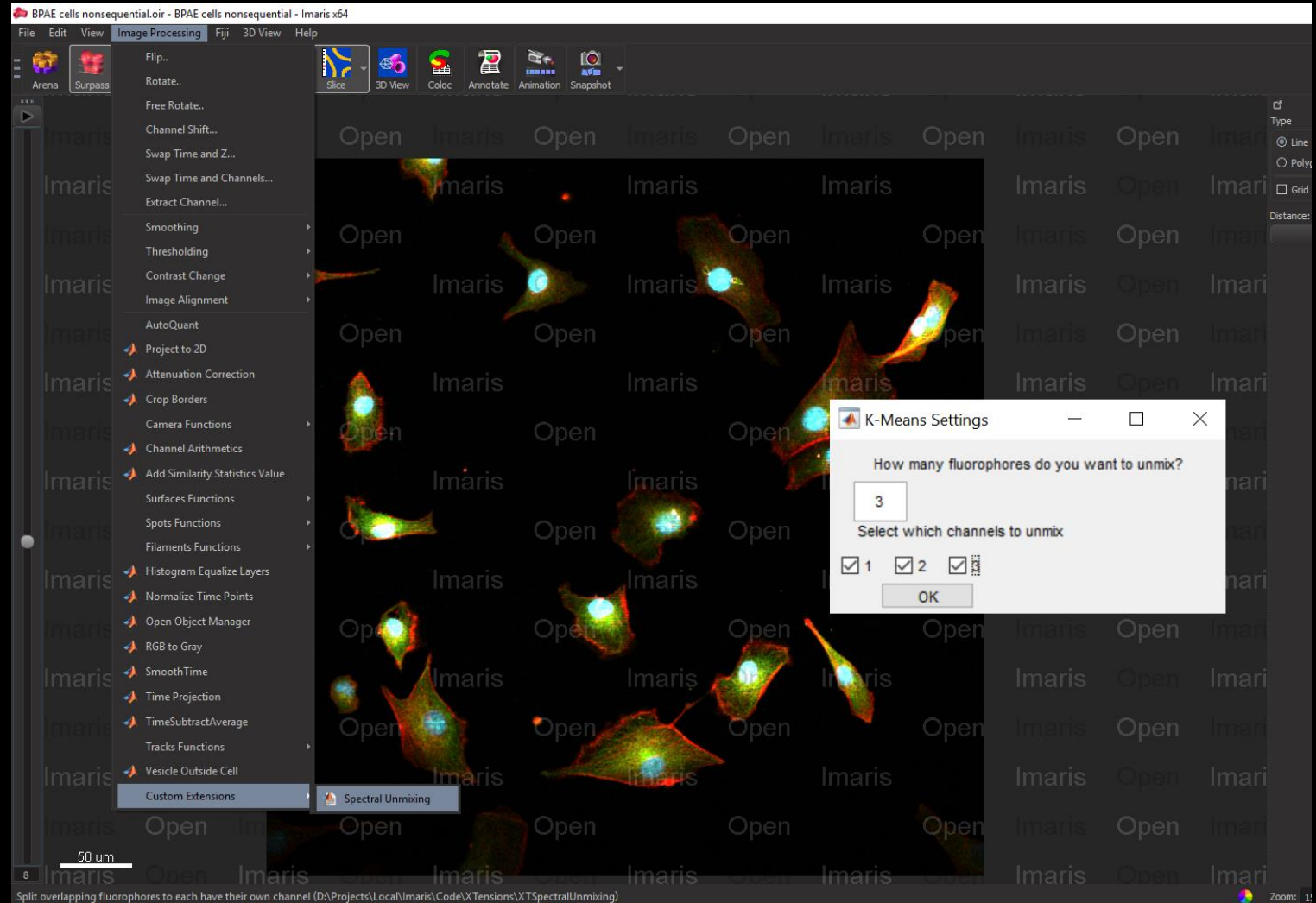
Images courtesy of Kris Lambert and Emma Reilly, Program for Advanced Immune Bioimaging  
181106\_D7\_OTI TdOTII GFP\_Ecad\_L\_0001

# Imaris XTension

We made an XTension to perform k-means unmixing inside of Imaris

[Instructions](#) on running this and other XTensions are available in the [resources](#) section of the Multiphoton Core Website

The XTension is available on [GitHub](#)



# Overview of Unmixing Methods

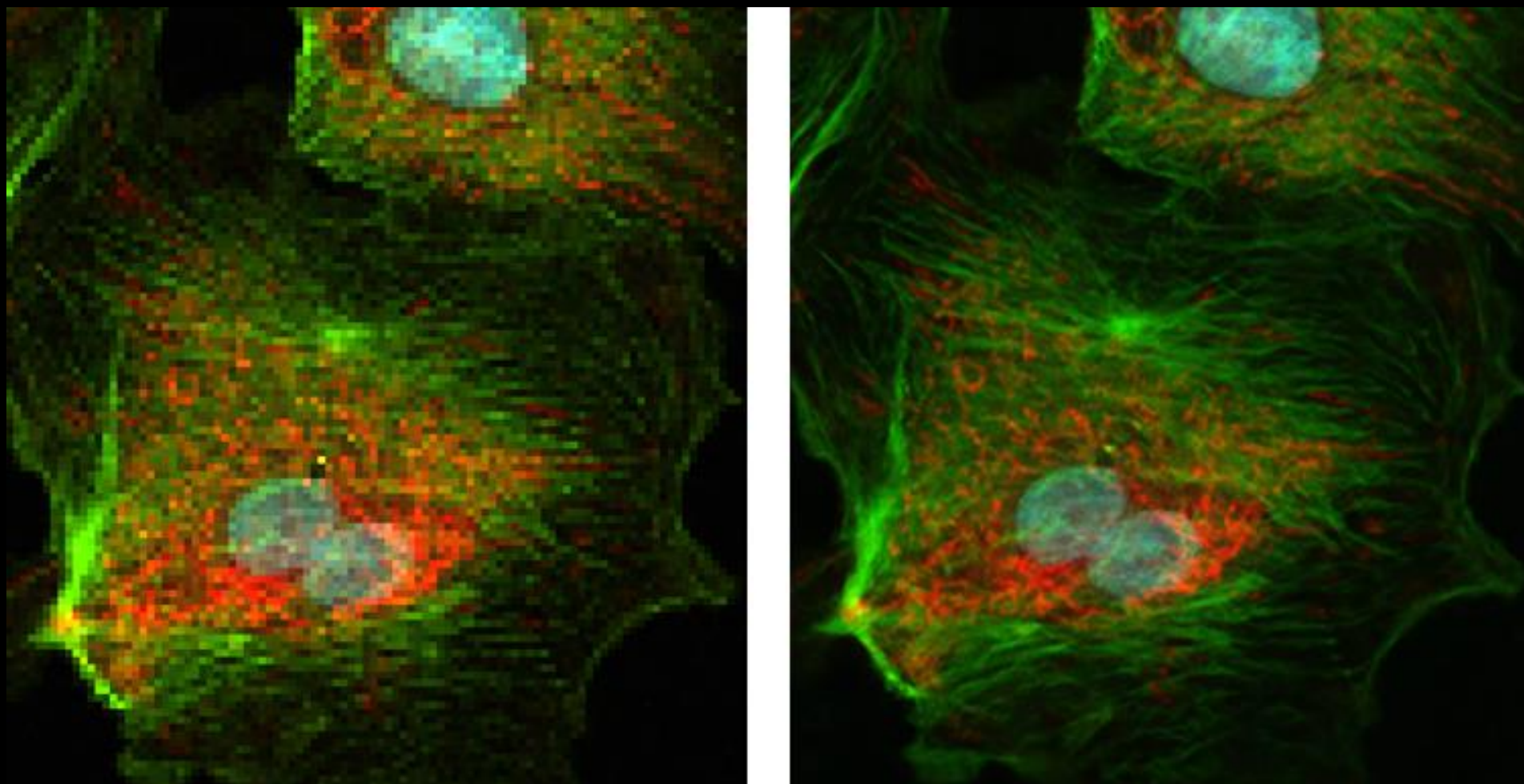
Method	Blind?	Uses Spatial Relations?	Key Limitations
K-means	Yes	No	May lose effectiveness with very high dimensional data (8+ fluorophores)
SIMI	No	No	Needs emission spectra, assumes one fluorophore per pixel
Cornell	No	No	Needs excitation and emission spectra (particularly hard for multiphoton)
Non-negative Matrix Factorization	Yes	No	No solution when fluorophores outnumber channels, struggles to converge with more than 3 fluorophores
In-Silico Labeling	No	Yes	Requires labeled datasets
Autoencoder Clustering	Yes	Optional	Requires training time

# Future Work

- Improved performance and functionality of k-means XTension
- Addition of other spectral unmixing methods
- Requests accepted
- Super resolution through machine learning



# Super Resolution through Machine Learning



Questions?