

Simultaneous numerical representation of soil microsite production and consumption of carbon dioxide, methane, and nitrous oxide using probability distribution functions

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Abstract

Production and consumption of nitrous oxide (N₂O), methane (CH₄), and carbon dioxide (CO₂) are affected by complex interactions of temperature, moisture, and substrate supply, which are further complicated by spatial heterogeneity of the soil matrix. This microsite heterogeneity is often invoked to explain non-normal distributions of greenhouse gas (GHG) fluxes, also known as hot spots and hot moments. To advance numerical simulation of these belowground processes, we expanded the Dual Arrhenius and Michaelis–Menten model, to apply it consistently for all three GHGs with respect to the biophysical processes of production, consumption, and diffusion within the soil, including the contrasting effects of oxygen (O₂) as substrate or inhibitor for each process. High-frequency chamber-based measurements of all three GHGs at the Howland Forest (ME, USA) were used to parameterize the model using a multiple constraint approach. The area under a soil chamber is partitioned according to a bivariate log-normal probability distribution function (PDF) of carbon and water content across a range of microsites, which leads to a PDF of heterotrophic respiration and O₂ consumption among microsites. Linking microsite consumption of O₂ with a diffusion model generates a broad range of microsite concentrations of O₂, which then determines the PDF of microsites that produce or consume CH₄ and N₂O, such that a range of microsites occurs with both positive and negative signs for net CH₄ and N₂O flux. Results demonstrate that it is numerically feasible for microsites of N₂O reduction and CH₄ oxidation to co-occur under a single chamber, thus explaining occasional measurement of simultaneous uptake of both gases. Simultaneous simulation of all three GHGs in a parsimonious modeling framework is challenging, but it increases confidence that agreement between simulations and measurements is based on skillful numerical representation of processes across a heterogeneous environment.

KEYWORDS

CH₄, CO₂, DAMM, DAMM-GHG, greenhouse gas, N₂O, probability distribution function, soil microsite

1 | INTRODUCTION

Fluxes of greenhouse gases (GHGs) from soil to the atmosphere are likely to play a significant role as biotic feedbacks to climate change (Ciais et al., 2014; Davidson & Janssens, 2006). Soils under forest, agriculture, and other land-use classes contribute to nearly a quarter of global emissions of GHGs, including carbon dioxide (CO₂), methane (CH₄), and nitrous oxide (N₂O; IPCC, 2014). Production and consumption of these biogenic GHGs are often associated with complex processes, involving carbon (C), nitrogen (N), and oxygen (O₂) substrates and inhibitors, and environmental controllers such as temperature, moisture, and transport of solutes and gases (Conrad, 1996), which remain challenging to simulate in ecosystem and earth system models (ESMs).

In this special issue, we present an expansion of a numerical soil process model that is a logical progression of several papers published by our group in the pages of this journal. While the importance of temperature on soil heterotrophic activity has been recognized for over a century (Arrhenius, 1889; Lloyd & Taylor, 1994; Vant't Hoff & Lehfeldt, 1899), and optima at intermediate values of soil moisture have also been well described (Hurst et al., 2017; Linn & Doran, 1984; Moyano, Manzoni, & Chenu, 2013), empirical relationships with these driving factors have had limited value in revealing a mechanistic understanding of soil respiration. Davidson, Belk, and Boone (1998) demonstrated that soil temperature and moisture had opposite seasonal trends in a moist temperate forest, resulting in confounding effects on soil respiration. Drawing on a growing body of research on soil respiration in the 1990s and 2000s, Davidson, Janssens, and Luo (2006) reviewed the emerging recognized need to move beyond mostly temperature functions, such as Q₁₀s, and to mechanistically link temperature and moisture drivers to substrate supply for soil heterotrophic respiration (Rh). Those concepts formed the basis of a parsimonious numerical model that used Dual Arrhenius and Michaelis–Menten (DAMM) kinetics to link soil temperature and moisture to their effects on substrate supply for soil respiration (Davidson, Samanta, Caramori, & Savage, 2012). In the 20 year special issue of this journal, Davidson, Savage, and Finzi (2014) described a vision for how the DAMM model could be conceptually linked to related processes of soil carbon dynamics, which has since been demonstrated in the modular Millennium Model (Abramoff et al., 2018), and how it could be integrated into large ecosystem models, which was since demonstrated by Sihi et al. (2018). Davidson et al. (2014) also proposed that other soil trace gas emissions could be simulated using the DAMM approach.

Here, we offer a new version of DAMM for the greenhouse gases, CO₂, CH₄, and N₂O (hereafter, DAMM-GHG: Dual Arrhenius and Michaelis–Menten–Greenhouse Gas). We use three simultaneous data streams from chamber measurements of CO₂, CH₄, and N₂O fluxes in a New England forest to constrain the DAMM-GHG model, which has a common structure for biophysical processes of production, consumption, and diffusion within the soil, including the contrasting effects of oxygen (O₂) as substrate or inhibitor for each process. Another innovation presented here is to represent soil

microsite heterogeneity of soil carbon and moisture contents with probability distribution functions (PDFs) and to simulate the production and consumption of each gas at a microsite scale, rather than the traditional modeling approach of using bulk soil means of measured carbon and moisture as model drivers.

Thermodynamic theories suggest that CH₄ oxidation (aka methanotrophy) should proceed under aerobic conditions and CH₄ production (aka methanogenesis) should be favored under anaerobic (or reducing) conditions (Conrad, 2009; Dean et al., 2018). Production of N₂O via nitrification and denitrification processes is known to peak at an optimal intermediate soil moisture content, whereas reducing soil conditions under high water content are thought to be prerequisites for N₂O reduction to N₂ via classical denitrification (Butterbach-Bahl, Baggs, Dannenmann, Kiese, & Zechmeister-Boltenstern, 2013; Davidson, 1991; Firestone & Davidson, 1989). While a large body of literature generally supports these patterns, there are exceptions that are frequently attributed to spatial heterogeneity within soils and soil microsites.

While the highest rates of net consumption of atmospheric N₂O (i.e., N₂O reduction) is observed in wetlands, N₂O reduction in well-drained upland soils has been observed sporadically for many years (Chapuis-Lardy, Wrage, Metay, Chotte, & Bernoux, 2007; Schlesinger, 2013; Syakila, Kroeze, & Slomp, 2010). Such observations have often been discounted as measurement error or noise. The recent advent of fast response field instruments with good sensitivity and precision has permitted confirmation that upland soils can be small sinks of N₂O (Eugster et al., 2007; Savage, Phillips, & Davidson, 2014), and a modest soil sink for atmospheric N₂O is now generally accepted as plausible for some sites and times. Increasing soil sink strength of N₂O during drought events further increases perplexity, given that drought events generally facilitate soil aeration (Goldberg & Gebauer, 2009). Occasional observations of net emissions of CH₄ from well-drained upland soils, although contrary to expectations, are also common (Brewer, Calderón, Vigil, & Fischer, 2018; Cattânio, Davidson, Nepstad, Verchot, & Ackerman, 2002; Keller & Matson, 1994; Silver, Lugo, & Keller, 1999; Teh, Silver, & Conrad, 2005; Verchot, Davidson, Cattânio, & Ackerman, 2000).

Spatial heterogeneity of soil microsites is often invoked to explain net atmospheric uptake of N₂O and net emissions of CH₄ from well-drained upland soils. Soil heterogeneity at microscales can cause a wide range of microsite redox potentials and concentrations of substrates, which must be accounted for to explain highly skewed distributions of soil GHG fluxes (Parkin, 1987, 1993; Savage et al., 2014; Stoyan, De-Polli, Böhm, Robertson, & Paul, 2000). Because existing ESMs are not able to represent the underlying mechanisms that control variation in enzymatic processes at microsite scales (Tian et al., 2019; Xu et al., 2016), these models often fail to capture the dynamics of soil GHG fluxes, including the so-called hot spots and hot moments (Groffman, 2012; Groffman et al., 2009; Lurndahl, 2016; Saha et al., 2018) or control points (Bernhardt et al., 2017).

Only recently have modeling activities at ecosystem (or landscape) scales begun to shift from the classical framework based on redox strata (or water table position) and mean measured soil

moisture to emerging conceptual frameworks that consider heterogeneous environment for production and consumption of GHGs (Ebrahimi & Or, 2018; Keiluweit, Gee, Denney, & Fendorf, 2018; Or, 2019; Wang, Brewer, Shugart, Lerda, & Allison, 2019; Yang et al., 2017). However, to our knowledge, mechanistic simulation of simultaneous production, consumption, and diffusion of multiple gases (CO_2 , CH_4 , N_2O , and O_2) among multiple soil microsites has not yet been attempted. Numerical representation of microsite production and consumption of multiple GHGs is necessary to simulate concurrent N_2O reduction and CH_4 oxidation processes in well-drained upland soils (Savage et al., 2014). The overall objective of this work is to demonstrate that the qualitative explanations of microsite heterogeneity can be expressed in a mathematically consistent biophysical process model that is

numerically consistent with simultaneously measured fluxes of all three GHGs: CO_2 , CH_4 , and N_2O .

While originally developed for aerobic Rh, here we expand the original core structure of the DAMM model (Davidson et al., 2012, 2014; Figure S1) to represent methanogenesis, methanotrophy, N_2O production, and N_2O reduction reactions using the same framework and physics for simulating the availability of O_2 and other substrates and for diffusion of gases across soil-atmosphere boundary using microsite PDFs (Figure 1; Figure S1). Simultaneously constraining our GHG enzyme kinetic model (i.e., DAMM-GHG) with observations of fluxes of multiple GHGs presents large challenges, because tuning a model to agree with one data stream may cause a poorer fit to a second or third data stream. However, if all data streams can be simultaneously simulated with adequate fidelity and skill, this multiple

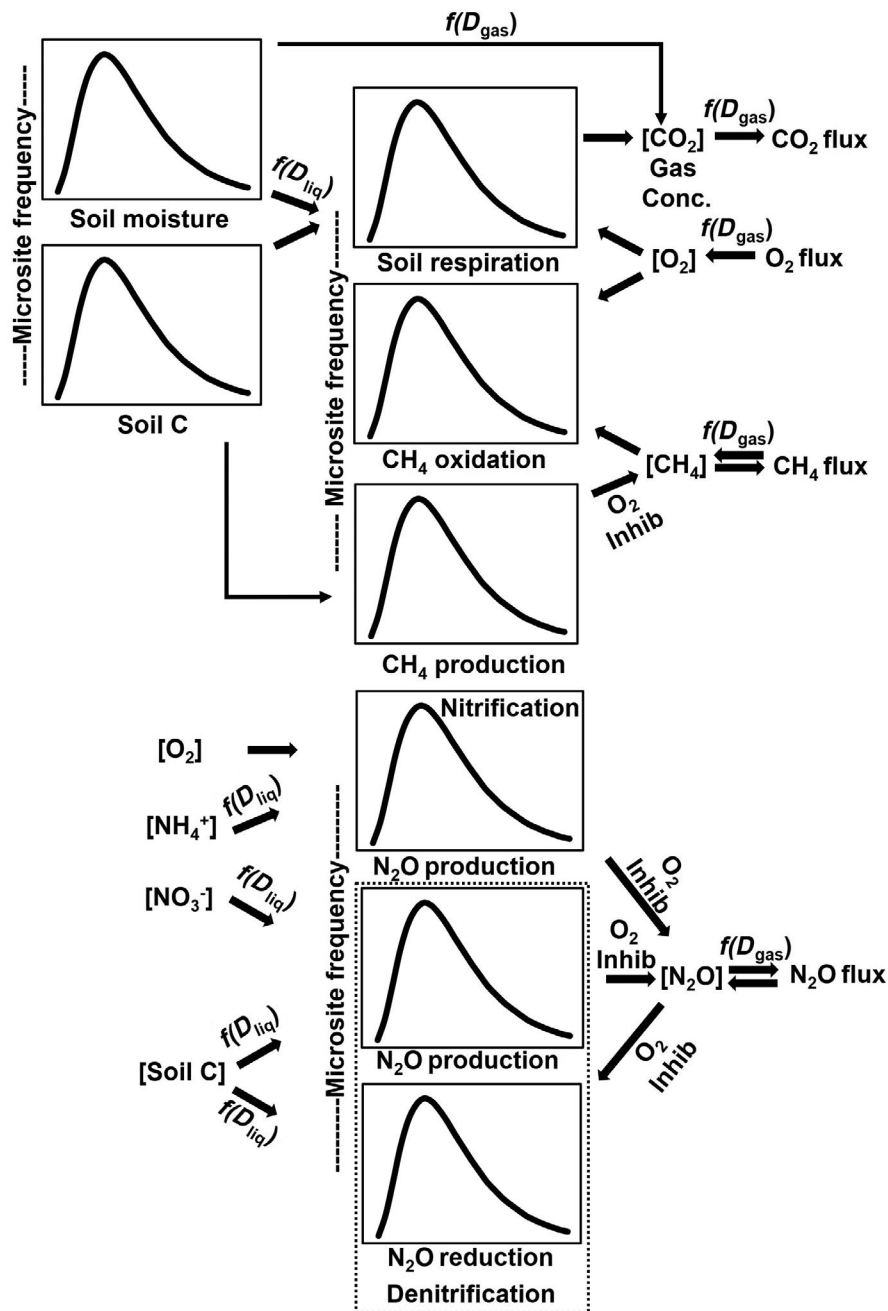


FIGURE 1 Conceptual framework for simultaneous representation of CO_2 , CH_4 , and N_2O fluxes in DAMM-GHG model. Inhib represents inhibition. $f(D_{gas})$ and $f(D_{liq})$ represent soil moisture effect on diffusion of gases and soluble substrates, respectively

constraint approach enhances the probability that the parameterization and process representations are realistic and robust. One can never be certain that a model gets the “right answer for the right reason,” but challenging a single model with multiple data streams of related but differing processes, such as CO₂, CH₄, and N₂O fluxes, confers additional credence to its structure and parameterization.

2 | MATERIALS AND METHODS

2.1 | Site description and data collection

We measured GHG fluxes in a mature boreal-transition forest with a hummock-hollow microtopography, Howland Forest research site (45.20°N, 68.74°W) from central Maine, United States. Mean annual temperature and mean annual precipitation are +5.5°C and 1,000 mm, respectively. Soils of the Howland Forest upland sites are characterized as Skerry fine sandy loam, frigid Aquic Haplorthods. More information on the Howland Forest research site can be found in Fernandez, Rustad, and Lawrence (1993).

High-frequency (sampling frequency: 1 Hz) real-time soil CH₄ and N₂O fluxes were measured using an Aerodyne quantum cascade laser integrated with soil CO₂ flux measurements by LI-COR IRGA assembly. Triplicate chambers were each sampled once every 2 hr. Chamber tops were closed for 5 min and automated fluxes were calculated by fitting a linear regression on the change in headspace GHG concentrations followed by temperature and pressure corrections. We characterized the uncertainty of measurements by the standard deviation estimates for all three GHGs. Soil temperature and soil moisture were measured at each chamber location at 10 cm depth once every hour using a Type-T thermocouple and Campbell Scientific CS616 water content reflectometer probes, respectively, and stored on a Campbell Scientific CR10X data logger (Campbell Scientific). We used daily average values of both drivers (soil temperature and soil moisture) and GHG fluxes for modeling purposes to smooth high measurement noise observed at subdaily timescale. See Savage and Davidson (2003) for more details on our chamber design and automated sampling system. Quality control protocols for soil GHG fluxes can be found in Savage, Davidson, and Richardson (2008) and Savage et al. (2014).

2.2 | Modeling scheme

Aerobic and anaerobic processes in soil are linked through heterotrophic dependence on fixed C sources for energy, but with contrasting effects of O₂ as either essential substrate or potential inhibitor (Figure 1; Figure S1). To date, most biogeochemical models use separate model versions for simulating soil organic matter decomposition resulting in CO₂ emissions and the processes affecting CH₄ and N₂O emissions, but here we simultaneously simulate biogeochemically linked multiple GHG emissions using the same biophysical framework.

For the present study, we focus primarily on Rh being the dominant source of CO₂ production and fate of O₂ consumption, with the

resulting O₂ concentrations then affecting the net fluxes of CH₄ and N₂O by methanogenesis, methanotrophy, nitrification, and denitrification processes. The logic for coupled simulation of CO₂, CH₄, and N₂O fluxes in the DAMM-GHG model, illustrated in Figure 1, is as follows:

1. The measured soil C and soil moisture can be partitioned according to a simulated log-normal PDF, such as a distribution where only a small fraction of microsites has high soil C or high soil moisture.
2. Log-normal PDFs of soil C substrates and soil moisture among microsites lead to a simulated PDF of Rh, applying the original DAMM model independently to each microsite within the PDF.
3. Simulated microsite CO₂ production is aggregated to the chamber scale to estimate Rh contributing to the chamber flux measurement. We then estimate the total soil CO₂ flux by adding the contribution of root-derived CO₂ fluxes to Rh based on previously measured ratios at the Howland Forest (Carbone et al., 2016; Savage, Davidson, Abramoff, Finzi, & Giasson, 2018; Sihi et al., 2018). A distinct seasonal pattern of the contribution of root-derived CO₂ (Ra) to total soil CO₂ fluxes (SR) increased from 0.50 in early spring to around 0.65 in early autumn, followed by a declining trend through winter (Figure S2). Total chamber-based measurements of CO₂ efflux are used as a constraint for the sum of the simulated root and heterotrophic CO₂ production rates across the simulated PDF of microsites.
4. The simulated and measured soil CO₂ efflux is a reasonable proxy for O₂ demand within the soil. The respiration quotient is not exactly unity, but is usually close enough to unity in noncalcareous soils to allow simulation of O₂ consumption within the soil based on measurement-constrained simulated CO₂ efflux (Angert et al., 2015). Knowledge of respiration quotient would be needed for the application of our DAMM-GHG model to calcareous soils. We assumed that the simulation of O₂ consumption by the original version of the DAMM model serves our purpose of estimating the O₂ demand (or consumption) here (see Figure S1).
5. Microsite PDFs of O₂ concentrations are then simulated as a function of O₂ consumption rates distributed across microsites and gaseous diffusion rates using the same DAMM functions in Figure S1 driven by air-filled porosity.
6. Next, the resulting PDF of O₂ concentrations is used to simulate methanogenesis, methanotrophy, N₂O production, and N₂O consumption at the scale of each of the distributed microsites according to similar Michaelis-Menten and diffusion equations (Figure S1), where O₂ serves as either inhibitor or substrate (Davidson et al., 2014). The net CH₄ and N₂O flux summed across simulated PDFs of microsites are constrained by observed chamber-based fluxes of CH₄ and N₂O.

We used soluble C as a proxy for the reducing power needed for methanogenesis. However, future studies may explicitly represent specific substrates for acetoclastic and hydrogenotrophic methanogenic pathways, if parameterization of that type of model structure can be constrained by the availability of data on concentrations of organic acids

(acetate, formate) and hydrogen (H_2), which was not the case for our study. We also assumed that respiration is the dominant pathway of CO_2 production in soil. Thus, we did not account for the minor contribution of acetoclastic methanogens to CO_2 production and hydrogenotrophic methanogens to CO_2 consumption. Likewise, we considered soil respiration is the major sink of O_2 and ignored the otherwise small fraction of O_2 consumed by methanotrophs.

We added a nitrification module to account for the N_2O production during nitrification using the observed seasonal dynamics of ammonium (NH_4^+) in our study area (Fernandez, Lawrence, & Son, 1995). The Howland Forest is a strongly nitrogen-limited system, with pore-water nitrate (NO_3^-) concentration always close to detection limits by inductively coupled plasma-mass spectrometry, ICP-MS (Fernandez et al., 1995). N_2O production during classical denitrification is mechanistically simulated using seasonally averaged porewater NO_3^- data along with microsite PDFs of soil C and soil moisture. Nitrous oxide is reduced to N_2 during classical denitrification following the Hole-in-the-Pipe conceptual model, including possible reduction of atmospheric N_2O that diffuses into the soil (Firestone & Davidson, 1989). See Supporting Information (Section S2) for DAMM-GHG model equations.

2.3 | Microsite probability distribution functions of soil C and soil moisture

The microsite PDFs of soil C substrate and soil moisture were generated from a bivariate truncated log-normal distribution. The PDF for soil C was truncated at 0.001 and 0.15 (g/cm^3), respectively. The soil C PDF was distributed with mean equaling the observed soil C value (see Figure S3 for more information on the PDF of soil C). Likewise, the soil moisture PDF was distributed with mean equaling the observed soil moisture value, truncations at 70% and 200% of the observed mean soil moisture values. The spatial heterogeneity of soil C and soil moisture were constrained by optimizing the parameters (standard deviation and/or coefficient of variation) that control the skewness of soil C and soil moisture PDFs by enveloping the bounds reported by Stoyan et al. (2000). If the upper truncation limit of the soil moisture PDF exceeded the soil pore volume, we reset it to 95% of the porosity value. We constructed the microsite PDF as the product of two log-normal distributions of soil C and soil moisture. The PDF was evaluated at 10×10 equally spaced quantiles for soil C and soil moisture, respectively.

Here, we focused on spatial heterogeneity across soil microsites at the mm and sub-mm scale. Within stand heterogeneity at the meter scale, such as variation in bulk density and porosity along topographical gradients, is not account for in this study.

2.4 | Parameter optimization and uncertainty analysis

We optimized model parameters within a Bayesian Markov chain Monte Carlo (MCMC) framework (see Section S1 for more details

on the optimization algorithm). We implemented the MCMC algorithm using *mcmc* and *doParallel* packages (Revolution Analytics & Weston, 2015) in the R (version 3.3.2) statistical programming language (R Core Team, 2018). We applied a posterior predictive procedure to estimate the uncertainty of the optimized parameters. We implemented the posterior predictive analyses using the *R-INLA* package (Lindgren & Rue, 2015; Rue, Martino, & Chopin, 2009). We divided daily-average soil GHG flux measurements into alternative synoptic-scale periods of 10 days, where we used one half of measured GHG fluxes for model calibration and another half for model validation.

2.5 | Sensitivity analysis

We evaluated the sensitivity of model parameters using a global variance-based sensitivity analysis and a collinearity (or parameter identifiability) analysis. We implemented a variance-based sensitivity analysis using the *R-multisensi* package, where a generalized sensitivity index (ranging between 0 and 1, extracted from the first axis of principle component analysis) was used to determine the sensitivity of multiple GHG fluxes to each model parameter value (Bidot, Lamboni, & Monod, 2018). The global sensitivity analysis quantifies the proportion of variability accounted by each of the parameters on model outputs, where a high GSI value indicates that the simulation results are highly sensitive to that parameter (Lamboni, Makowski, Lehuger, Gabrielle, & Monod, 2009).

We implemented the collinearity analysis using the *collin* function of *R-FME* package, where the collinearity index (CI) was used to determine the linear dependence of model parameters to each other (Brun, Reichert, & Künsch, 2001; Soetaert, 2016; Soetaert & Petzoldt, 2010). In general, higher values of CI indicate increased equifinality (or decreased number of identifiable parameters) of model parameters. One can compensate $(1 - 1/CI)$ % of the effect of a change in one parameter by modifying the values of other parameters. Hence, CI values can range between 1 (when all terms are orthogonal or all subsets of parameters are identifiable) and infinity (when all terms are linearly dependent, or no single subset of parameters is identifiable). The CI value of 15 is considered as a threshold above which approximate linear dependence of model parameters increases and poor identifiability can be expected (sensu Omlin, Brun, & Reichert, 2001).

3 | RESULTS

3.1 | Seasonality of soil greenhouse gas fluxes

Soil CO_2 fluxes followed the typical seasonal trend of soil temperature, where both seasonal average and peak CO_2 fluxes were comparable between 2015 (average [min-max]: 171 [73-297] $mg\ CO_2-C\ m^{-2}\ hr^{-1}$) and 2016 (average [min-max] measurement period: 168 [52-281] $mg\ CO_2-C\ m^{-2}\ hr^{-1}$; Figure 2). This is due, in part, to the comparable seasonal soil temperature ranges between 2015

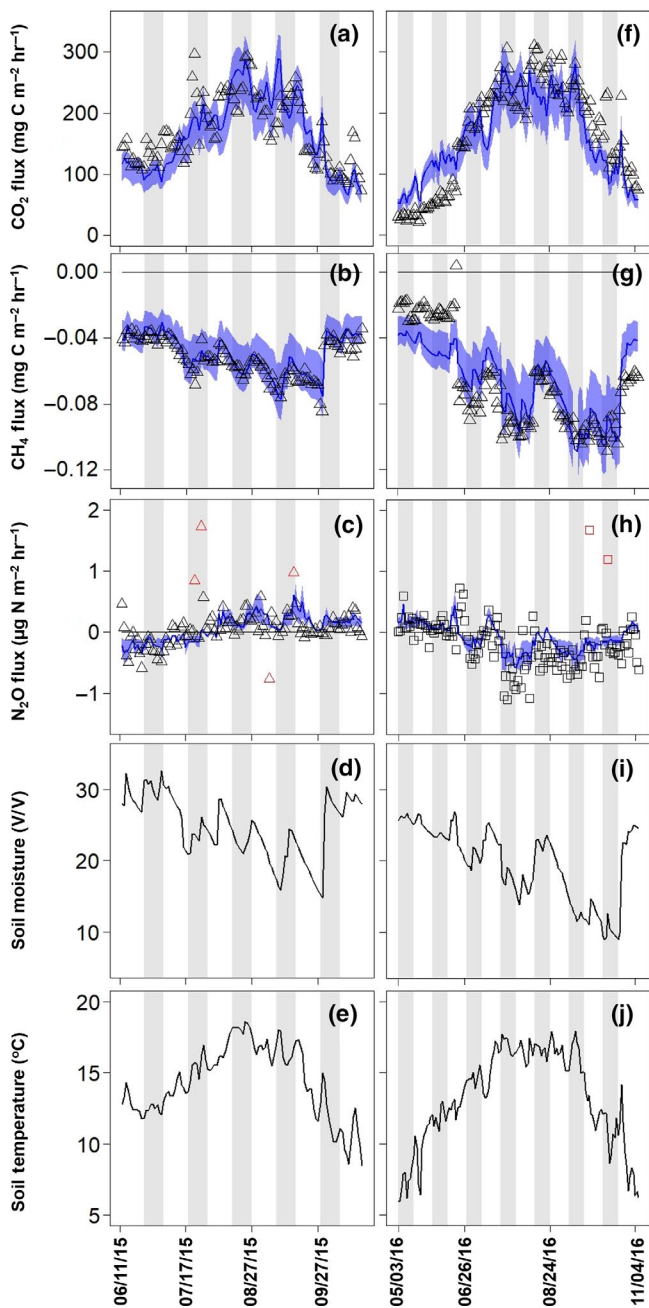


FIGURE 2 Temporal trend of greenhouse gas, GHG (CO_2 : a, f; CH_4 : b, g; and N_2O : c, h) fluxes, soil moisture (d, i), and soil temperature (e, j) during 2015 (a–e) and 2016 (f–j) growing seasons. Triangles and squares represent observed GHG fluxes in 2015 and 2016, respectively. Light gray shades in the background represent validation windows, which are interspersed throughout the observation period. Red triangles and squares represent outliers for N_2O fluxes. Blue line and shade represent median and 95% CI of simulated GHG fluxes, respectively

(average: 14.4°C , ranged between 8.4 and 18.6°C) and 2016 (average: 13.3°C , ranged between 5.6 and 17.9°C) measurement periods (Figures 2 and 3). Soil CO_2 fluxes exponentially increased with soil temperature ($R^2 = .75$; Figure 3a). We also observed a typical bell-shaped relationship between CO_2 flux and soil moisture ($R^2 = .16$), with the optimum for CO_2 flux at intermediate water contents

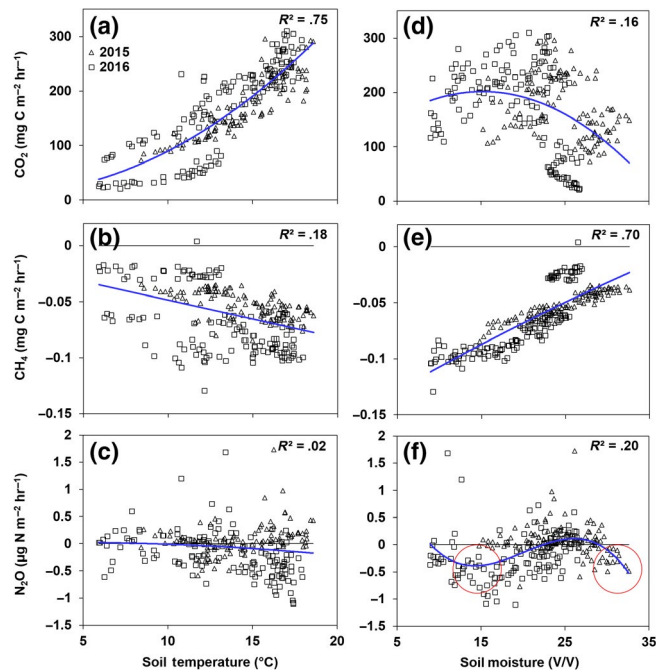


FIGURE 3 Relation of observed CO_2 (a, d), CH_4 (b, e), and N_2O (c, f) fluxes with soil temperature (a–c) and soil moisture (b–e). Triangles and squares represent observed GHG fluxes in 2015 and 2016, respectively

(Figure 3d). Although the effect of soil moisture on CO_2 fluxes was always secondary to soil temperature (Figure S5), the fit with soil moisture was better when it was more limiting in the dry summer of 2016 than the wet summer of 2015 ($R^2 = .24$ and $.52$ in 2015 and 2016, respectively; Figure S6d).

In contrast to CO_2 fluxes, soil CH_4 fluxes mimicked the seasonal trend of soil moisture for both years (Figure 2b,g). Soil moisture and net CH_4 fluxes were positively related ($R^2 = .70$), although the slope of the linear regression line was steeper during 2016 than during 2015 (Figure 3e; Figure S6e). We observed relatively smaller net CH_4 oxidation in the spring followed by higher net CH_4 oxidation in summer months, and again lower net CH_4 oxidation in the autumn. Although seasonal average values were generally similar, the range of CH_4 flux values in 2016 (average: $-0.07 \mu\text{g CH}_4\text{-C m}^{-2} \text{hr}^{-1}$, min–max range: -0.13 to $0.004 \mu\text{g CH}_4\text{-C m}^{-2} \text{hr}^{-1}$) was wider than in 2015 (average: $-0.05 \mu\text{g CH}_4\text{-C m}^{-2} \text{hr}^{-1}$, min–max range: -0.08 to $-0.03 \mu\text{g CH}_4\text{-C m}^{-2} \text{hr}^{-1}$).

Unlike CO_2 and CH_4 , the seasonal trend of soil N_2O fluxes contrasted between 2015 and 2016 (Figures 2c,h and 3f; Figure S6f). The 2015 growing season was significantly wetter than the 2016 growing season. The cumulative precipitation of the summer months (June 1 to September 30) of 2015 and 2016 was 439 mm and 279 mm, respectively (source: <https://www.ncdc.noaa.gov/crn/>). Consequently, the average soil moisture was generally higher (24.8 v/v , min–max range: 14.8 – 32.6 v/v) during the 2015 growing season (measured over June 11 to October 17) than the average soil moisture (19.7 v/v , min–max range: 8.9 – 27 v/v) during the 2016 growing season (measured over May 3 to November 6; Figures 2d,i and 3f; Figure S6f).

We observed low rates of net N_2O consumption during the spring of 2015 when soil moisture was highest, which was followed by mostly near zero net emissions and a few positive emissions throughout the 2015 summer (Figure 2c,d). In contrast, we observed net positive N_2O emission during the early spring of 2016 and net N_2O consumption throughout most of the 2016 growing season, when soil moisture was minimal (Figure 2h,i). The temperature effect was weak and inconsistent between 2 years (Figure 3c; Figure S6c). On average, we observed small net N_2O emission during the 2015 growing season (average:

$0.06 \mu\text{g N}_2\text{O-N m}^{-2} \text{hr}^{-1}$, ranged between -0.77 and $1.73 \mu\text{g N}_2\text{O-N m}^{-2} \text{hr}^{-1}$) and net N_2O consumption during the 2016 growing season (average: $-0.18 \mu\text{g N}_2\text{O-N m}^{-2} \text{hr}^{-1}$, ranged between -1.10 and $1.68 \mu\text{g N}_2\text{O-N m}^{-2} \text{hr}^{-1}$).

3.2 | Performance of the DAMM-GHG model

Overall, the model reproduced the seasonal dynamics of soil greenhouse gas fluxes (Figures 2 and 4). The model explained 72% of the variation in soil CO_2 fluxes (Figure 4a). Likewise, the 1:1 relation

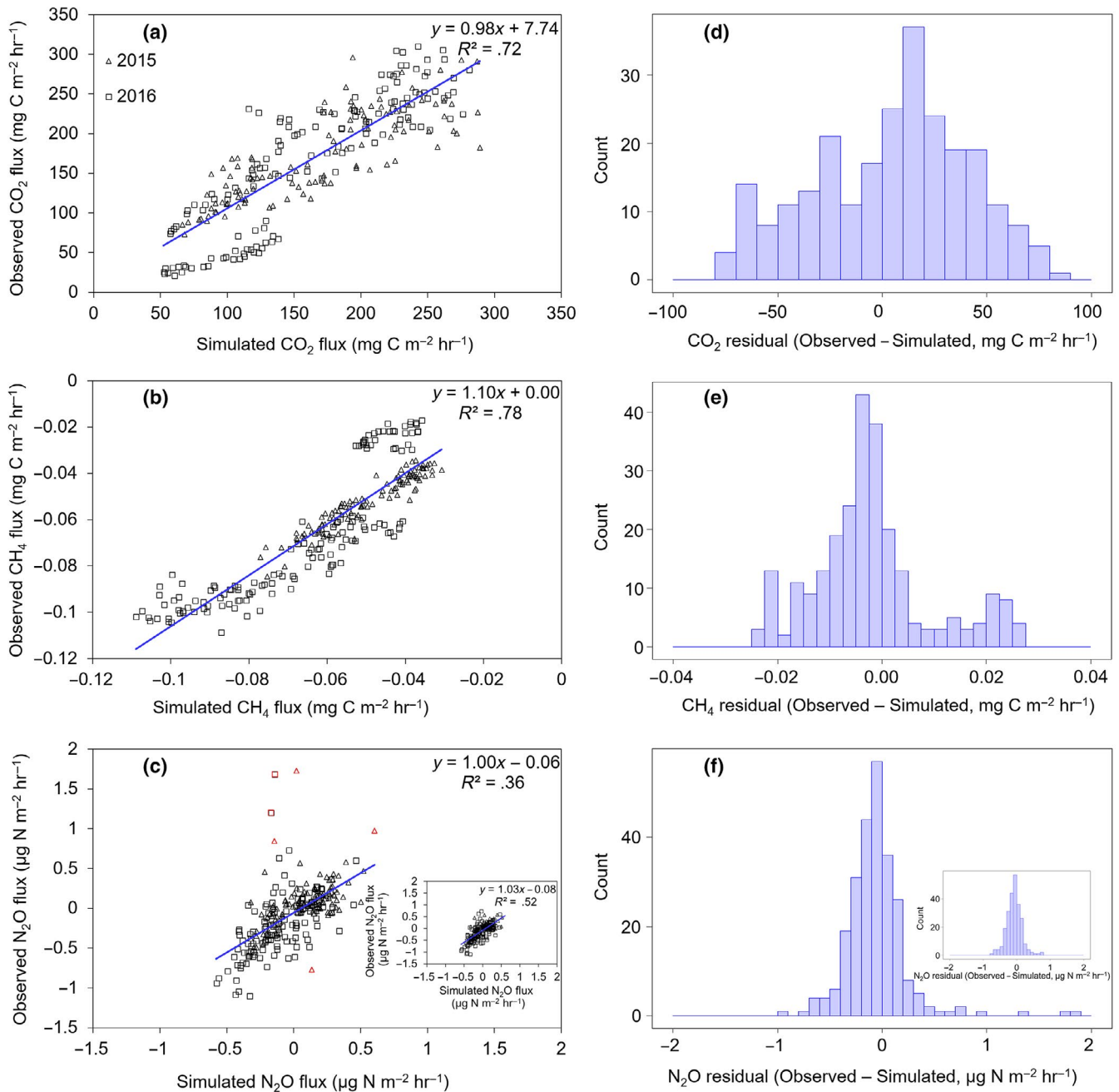


FIGURE 4 Relation between observed versus simulated greenhouse gas, GHG (CO_2 : a, d; CH_4 : b, e; and N_2O : c, f) fluxes. Triangles and squares in (a–c) represent observed GHG fluxes in 2015 and 2016, respectively. Red triangles and squares in lower left panel (c) represent outliers for N_2O fluxes. Inset figures in bottom panels (c, f) represent one-to-one relation and model residuals for N_2O after removing the outliers

TABLE 1 Parameters used for simultaneous simulation all three greenhouse gases in the DAMM-GHG model

Parameter	Description	Unit	Prior range	Posterior range	Source
<i>For CO₂ module</i>					
α_{CO_2}	Base rate for soil respiration	$\mu\text{mol CO}_2 \text{ L}^{-1} \text{ hr}^{-1}$	$2 \times 10^{10} (2 \times 10^9 - 2 \times 10^{11})$	$5 \times 10^9 (2 \times 10^9 - 9 \times 10^9)$	Abramoff et al. (2017), Davidson et al. (2018), Sihi et al. (2018)
E_{pCO_2}	Temperature sensitivity for soil respiration	kJ/mol	72 (60–80)	66 (64–72)	
$k_{\text{M}_C\text{-CO}_2}$	Half-saturation constant of C for soil respiration	$\mu\text{mol C/L}$	1 (0.1–100)	0.9 (0.7–1.6)	
$k_{\text{M}_{\text{O}_2}\text{-CO}_2}$	Half-saturation constant of O ₂ for soil respiration	$\mu\text{mol O}_2/\text{L}$	100 (3–300)	16 (4–38)	
Depth	Effective depth	cm	15 (5–30)	7 (6–11)	This study
Total microsite	Total number of microsities	unitless	$10^4 (10^3 - 10^5)$	$7 \times 10^3 (2 \times 10^3 - 9 \times 10^3)$	This study
Soil C _{SD}	Coefficient that determines skewness of soil carbon PDF	unitless	0.5 (0.1–0.9)	0.5 (0.1–0.9)	Stoyan et al. (2000)
Soil moisture _{SD}	Coefficient that determines skewness of soil moisture PDF	%	20 (5–30)	7 (6–12)	Stoyan et al. (2000)
<i>For CH₄ module</i>					
$\alpha_{\text{CH}_4\text{prod}}$	Base rate for CH ₄ production	$\mu\text{mol CH}_4 \text{ L}^{-1} \text{ hr}^{-1}$	$3 \times 10^5 (3 \times 10^4 - 3 \times 10^6)$	$3 \times 10^5 (2 \times 10^5 - 6 \times 10^5)$	This study
$E_{a_{\text{CH}_4\text{prod}}}$	Temperature sensitivity for CH ₄ production	kJ/mol	100 (50–150)	74 (68–78)	Nedwell and Watson (1995), Westermann (1993)
$k_{\text{M}_C\text{-CH}_4}$	Half-saturation constant of C for CH ₄ production	$\mu\text{mol C/L}$	1 (0.1–100)	1.2 (0.9–2.1)	This study
$k_{\text{M}_{\text{O}_2}\text{-CH}_4}$	Inhibition coefficient of O ₂ for CH ₄ production	$\mu\text{mol O}_2/\text{L}$	3 (0.3–4.3)	0.43 (0.4–0.9)	Arah and Stephen (1998)
$\alpha_{\text{CH}_4\text{ox}}$	Base rate for CH ₄ oxidation	$\mu\text{mol CH}_4 \text{ L}^{-1} \text{ hr}^{-1}$	0.07 (0.007–7)	0.1 (0.08–2)	Davidson et al. (2001)
$E_{a_{\text{CH}_4\text{ox}}}$	Temperature sensitivity for CH ₄ oxidation	kJ/mol	30 (10–50)	34 (32–37)	Crill, Martikainen, Nykanen, and Silvola (1994)
$k_{\text{M}_{\text{CH}_4}}$	Half-saturation constant of CH ₄ for CH ₄ oxidation	$\mu\text{mol CH}_4/\text{L}$	$10^{-2} (10^{-3} \text{ to } 10^{-1})$	0.005 (0.002–0.006)	Davidson et al. (2001)
$k_{\text{M}_{\text{O}_2}\text{-CH}_4}$	Half-saturation constant of O ₂ for CH ₄ oxidation	$\mu\text{mol O}_2/\text{L}$	43 (3–300)	24 (13–33)	Davidson et al. (2001)
<i>For N₂O module</i>					
$\alpha_{\text{N}_2\text{Oprod-nitrif}}$	Base rate for nitrification	$\mu\text{mol N}_2\text{O/L}$	$10^2 (10^1 - 10^3)$	282 (207–722)	This study
$E_{a_{\text{N}_2\text{Oprod-nitrif}}}$	Temperature sensitivity for N ₂ O production during nitrification	kJ/mol	60 (45–75)	62 (57–67)	Stark (1996), Stark and Firestone (1996)
$k_{\text{M}_{\text{NH}_4}}$	Half-saturation constant of NH ₄ ⁺ for N ₂ O production	$\mu\text{mol NH}_4^+/\text{L}$	15 (8–22)	9 (8–10)	Stark and Firestone (1996), Zarnetske, Matsuo, Yamazaki, and Wada (1987)

(Continues)

TABLE 1 (Continued)

Parameter	Description	Unit	Prior range	Posterior range	Source
$kM_{O_2-N_2O}$	Half-saturation constant of O_2 for N_2O production	$\mu\text{mol } O_2/L$	100 (9–163)	36 (15–44)	Bodelier, Libochant, Blom, and Laanbroek (1996), Verstraete and Focht (1977), Zarnetske et al. (1987)
$\alpha_{N_2O_{prod-denit}}$	Base rate for N_2O production	$\mu\text{mol } N_2O/L$	$10^2 (10^1-10^3)$	520 (438–958)	This study
$E_{a_{N_2O_{prod-denit}}}$	Temperature sensitivity for N_2O production during denitrification	kJ/mol	60 (45–75)	67 (65–71)	Canion et al. (2014), Holtan-Hartwig, Dörsch, and Bakken (2002), Vieten (2008)
kM_{NO_3}	Half-saturation constant of NO_3^- for N_2O production	$\mu\text{mol } NO_3^-/L$	26 (15–57)	17 (16–19)	Zarnetske et al. (1987)
$kI_{N_2O-prod}$	Inhibition coefficient of O_2 for N_2O production	$\mu\text{mol } O_2/L$	14.3 (4.3–43)	41 (40–42)	Körner and Zumft (1989), Zarnetske et al. (1987)
kM_{C-N_2O}	Half-saturation constant of C for N_2O production and reduction during denitrification	$\mu\text{mol } C/L$	1 (0.1–100)	0.6 (0.3–1)	This study
$\alpha_{N_2O_{red}}$	Maximum velocity for N_2O reduction	$\mu\text{mol } N_2O/L$	$10^3 (10^2-10^4)$	3,413 (1,585–9,583)	This study
$E_{a_{N_2O_{red}}}$	Temperature sensitivity for N_2O reduction	kJ/mol	50 (45–75)	47 (46–47)	Canion et al. (2014), Holtan-Hartwig et al. (2002), Vieten (2008)
kM_{N_2O}	Half-saturation constant of N_2O for N_2O reduction	$\mu\text{mol } N_2O/L$	0.16 (0.05–0.27)	0.19 (0.13–0.26)	Holtan-Hartwig et al. (2002), Vieten (2008)
kI_{N_2O-red}	Inhibition coefficient of O_2 for N_2O reduction	$\mu\text{mol } O_2/L$	7.5 (4.3–20.1)	19.5 (18.6–19.7)	Körner and Zumft (1989), Vieten (2008)

Note: Prior range represents initial (min–max) of prior interquartile range. Posterior range represents median (95% CI) of posterior interquartile range. Units for all base rates, that is, α (s), are in μmol concentration dissolved in water.

Abbreviation: PDF, probability distribution function.

between the observed and simulated soil CH_4 fluxes was remarkable ($R^2 = .78$; Figure 4b). The model marginally overestimated CO_2 fluxes and underestimated CH_4 fluxes during early spring of 2016 (Figure 2f,g).

Soil N_2O fluxes were relatively noisier as compared to CO_2 and CH_4 fluxes with a few outliers in both years (red triangles and squares in Figures 2 and 4, respectively). The model generally explained the dynamics of N_2O fluxes ($R^2 = .36$ and $.52$ with and without outliers, respectively; Figure 4c). The model did not capture the few very low net N_2O fluxes observed in the peak season of 2016. Most importantly, the model captured the instances when net atmospheric consumption of CH_4 (i.e., net CH_4

oxidation) co-occurred with net atmospheric consumption of N_2O (i.e., net N_2O reduction) within the same soil chamber and at two extremes of the measured soil moisture, during the early wet spring of 2015 and during the driest period of the 2016 growing season (see Figure 2c,h and red circles in Figure 3f).

In general, there was little bias in the relations between the observed and simulated GHG fluxes (slope ranged between 0.98 and 1.10; Figure 4). The 95% CI of the simulated GHGs for all, CO_2 , CH_4 , and N_2O , were narrow and the model parameter values were generally well constrained. The interquartile ranges in the posterior distributions of all parameters were less than half of their respective prior interquartile ranges (Table 1). The prior interquartile ranges

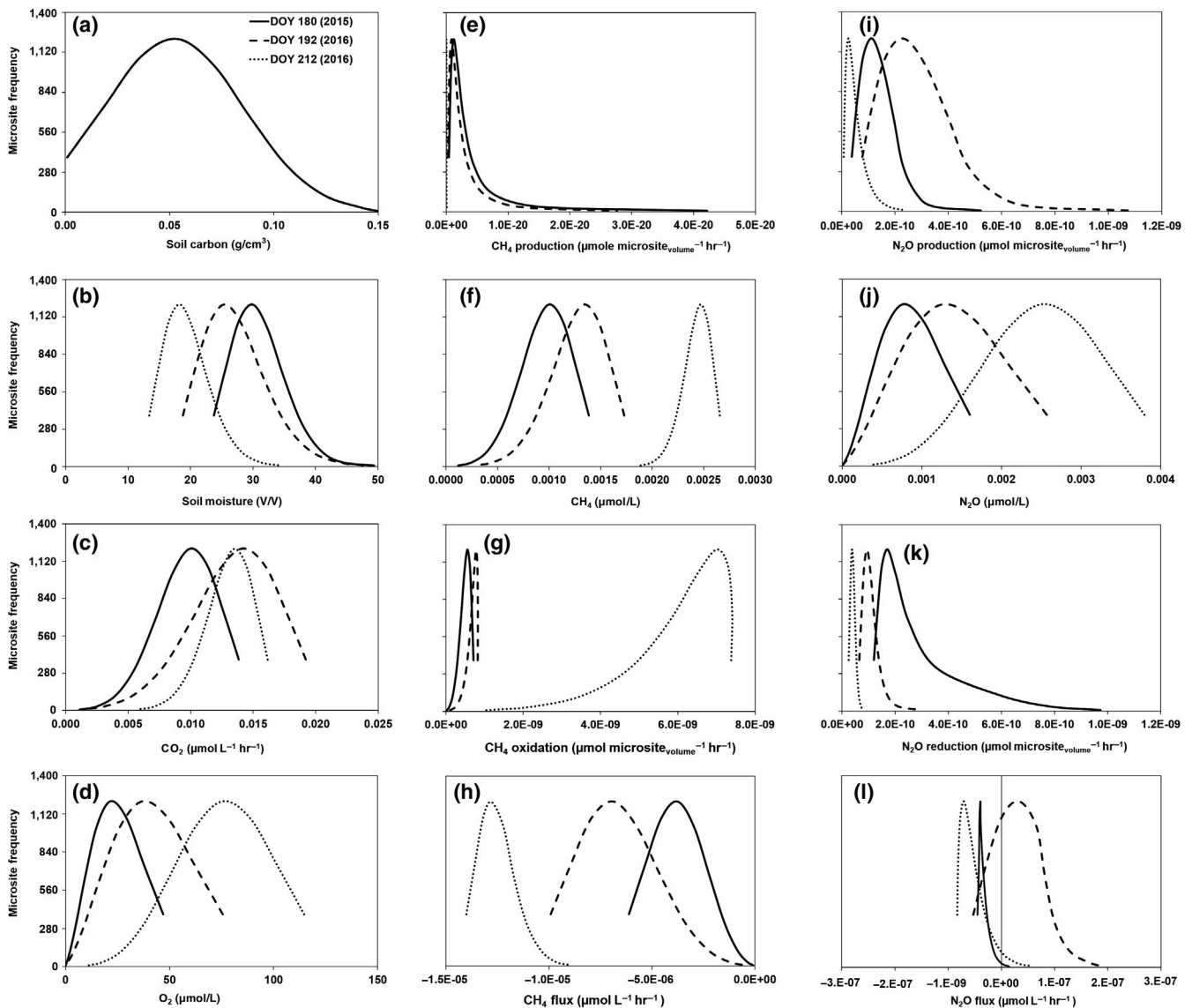


FIGURE 5 Microsite probability distribution functions (PDFs) of soil carbon (a), soil moisture (b), CO_2 flux (c), O_2 concentration (d), CH_4 production (e), CH_4 concentration (f), CH_4 oxidation (g), CH_4 flux (h), N_2O production (i), N_2O concentration (j), N_2O reduction (k), and N_2O flux (l). Solid, dashed, and dotted lines represent simulated microsite PDFs of individual processes for DOY 180, 2015 (SoilM = 32.6 v/v and SoilT = 12.0°C); DOY 192, 2016 (SoilM = 25.4 v/v and SoilT = 13.8°C); and DOY 212, 2016 (SoilM = 18.2 v/v and SoilT = 16.3°C), respectively, and correspond to three scenarios presented in the discussion section. DOY, day of year; SoilM, soil moisture; SoilT, soil temperature

represent the approximate upper and lower bounds of the measured values from the relevant literature.

The effective depth in the DAMM-GHG model was optimized to a median value of 7 cm (min-max range: 6–11 cm; Table 1), indicating that most of the important processes affecting the net GHG fluxes that we measured with chambers were occurring within the topsoil horizons at this site. The number of microsites within the 0.07 m² chamber footprint was optimized to a median value of 7,000 (min-max range: 2,000–9,000; Table 1), indicating that the simulated microsites were about 3.6 mm in diameter, which could include macroaggregates and clusters of fine roots and pockets of organic debris.

3.3 | Sensitivity analysis

Soil moisture primarily (and soil temperature secondarily) controlled the microsite PDFs of production, consumption, and diffusion processes of CO₂, CH₄, and N₂O. The net flux is the net

effect of production, consumption, and diffusion of individual gases (Figure 5; Figure S10). Of all parameters, the most sensitive ones were those that control the V_{max} terms in the Arrhenius equation (E_a and α) for production and consumption of individual GHGs, followed by the half-saturation constants (kM s) and O₂ inhibition coefficients (kI s) for each process (Figure 6). The linear dependence of the DAMM-GHG model parameters was generally low and was usually below the threshold of 15 (with a few exceptions) identified for potential equifinality issues (Figure 7).

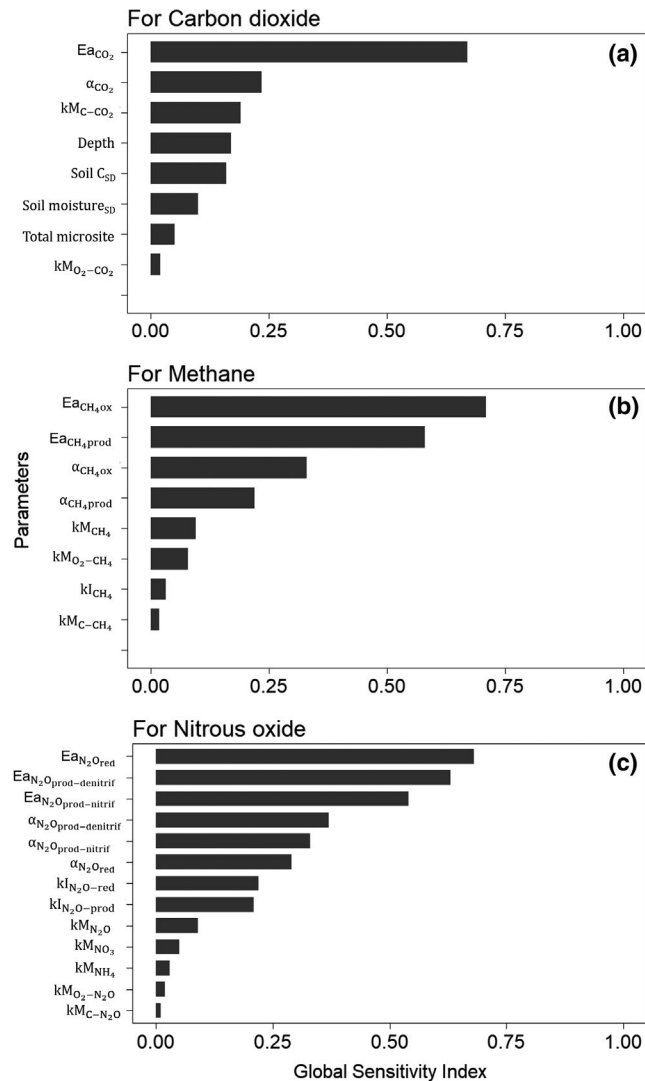


FIGURE 6 Sensitivity indices of the DAMM-GHG model parameters for CO₂ (a), CH₄ (b), and N₂O (c) modules, respectively

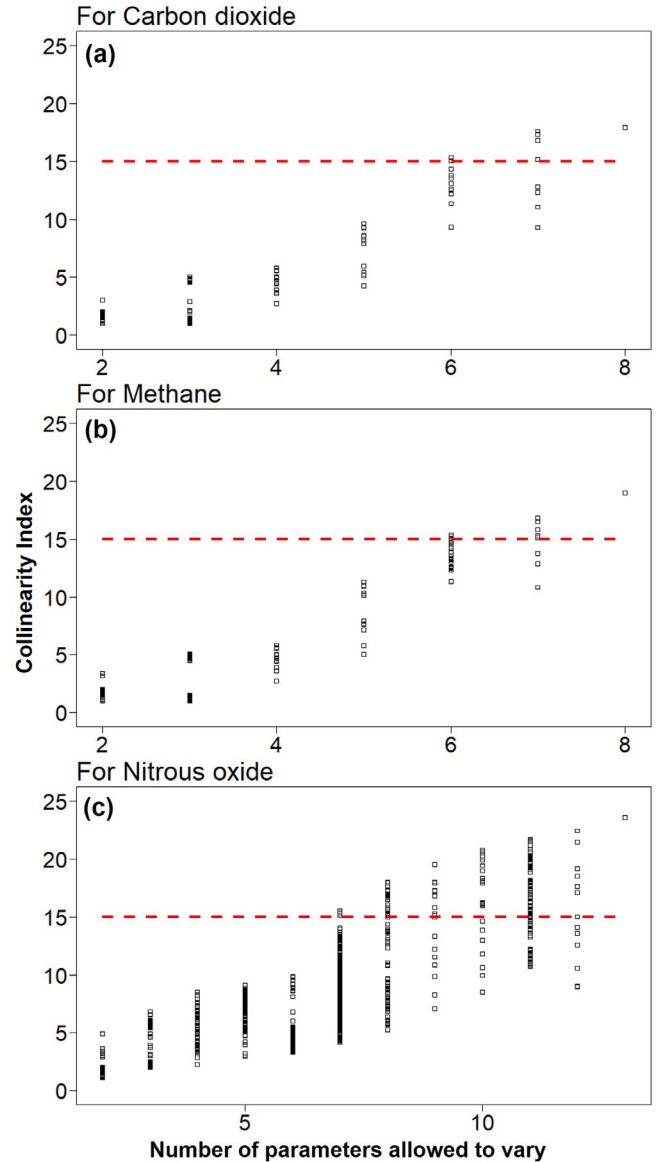


FIGURE 7 Collinearity indices of the DAMM-GHG model parameters for CO₂ (a), CH₄ (b), and N₂O (c) modules, respectively. Each point represents a unique combination of parameters allowed to vary while others are held constant. Dashed horizontal lines represent threshold value above which approximate linear dependence of model parameters increases and poor identifiability can be expected (Omlin, 1993)

4 | DISCUSSION

Our goal is a novel integration of measurement and modeling of three key greenhouse gases to improve understanding of and modeling capacity for interactions of belowground temperature, moisture, and substrate supply that control the net soil emissions of CO_2 , CH_4 , and N_2O . To this end, we built upon the DAMM model, which mechanistically simulates soil Rh using Arrhenius equations, diffusion functions, and Michaelis–Menten enzyme kinetics (Davidson et al., 2012, 2014). Our framework of the DAMM-GHG model is unique in that it represents the simultaneous production and consumption of all three GHGs within the same soil biophysical framework using microsite PDFs. Below we discuss the performance of the DAMM-GHG model and the utility of the microsite PDFs in reproducing the spatial and temporal dynamics of observed CO_2 , CH_4 , and N_2O fluxes within a multiple constraint framework.

4.1 | Microsite representation captures co-occurrence of methane oxidation and nitrous oxide reduction

Consumption of atmospheric N_2O via classical denitrification should occur only under reducing conditions. Yet, we have observed net uptake of atmospheric CH_4 (oxidation) and uptake of atmospheric N_2O (reduction) simultaneously in well-drained soils of Howland forest under both low and high soil moisture levels. With the advent of high frequency and high sensitivity flux measurement technology, we can be confident that these modest uptake rates of both CH_4 and N_2O are significantly different from zero and are not measurement errors or artifacts (Figures 2 and 4; also see Figure S4). These seemingly contradictory observations have been qualitatively explained by describing diffusional constraints of gas transport as follows: both CH_4 and N_2O can diffuse into well-drained soils; the CH_4 is oxidized at microsites where O_2 is abundant; while N_2O is reduced at other microsites where N_2O is present and Rh is sufficiently rapid to consume O_2 . Here, we demonstrate that this qualitative explanation can be expressed in a mathematically consistent biophysical process model that is numerically consistent with simultaneously measured fluxes of these gases.

The area under a soil chamber was partitioned according to a bivariate log-normal PDF of soil C and moisture across a range of microsites, which leads to a PDF of CO_2 production and O_2 consumption among microsites. The resulting broad range of microsite O_2 concentrations determines the PDF of microsites that produce or consume CH_4 and N_2O according to Michaelis–Menten and Arrhenius functions for each process (Figure 1; Figure S1). Concentrations of below ambient N_2O (hot spots of N_2O reduction) occur in microsites with simulated high C and high moisture. Net consumption and production of CH_4 and N_2O are simulated within a chamber as the average of all soil microsite simulations. To demonstrate that it is numerically feasible for microsites of N_2O reduction and CH_4 oxidation to co-occur under a single chamber, we discuss three different scenarios where mean soil moisture

levels cover the envelope of observed soil moisture in our study area (Figure 5).

Of the two growing seasons, we measured highest bulk soil moisture (32.6 v/v) on June 29 (DOY 180), 2015. Consequently, microsite PDFs of soil moisture ranged from 23.7 v/v to as high as 47.7 v/v (solid line in Figure 5b). Microsites with high soil moisture limited diffusion of gaseous O_2 through air-filled pore space. Simultaneously, soils had warmed up enough during the late spring of 2015 for soil respiration to exceed $100 \text{ mg CO}_2\text{-C m}^{-2} \text{ hr}^{-1}$, which created significant O_2 demand in microsites with high soil C. Relatively high soil respiration along with limited O_2 diffusion resulted in a large fraction of total soil microsites with low O_2 concentrations (solid line in Figure 5d). Production of N_2O was high in microsites with high soil C. However, reducing environments favored N_2O reduction more than N_2O production in microsites with low O_2 concentrations. Together, the PDFs of soil microsites resulted in a modest net negative mean flux of N_2O (solid line in Figure 5i). Classical theories of biological denitrification processes fit with our observations of net negative fluxes of N_2O under conditions of high soil moisture (and soil C) when enzymatic reduction of N_2O to N_2 under reducing environment outcompetes N_2O production rates, especially in nitrogen-limited systems like our field site (Davidson, Keller, Erickson, Verchot, & Veldkamp, 2000; Davidson et al., 1993; Firestone & Davidson, 1989). Although N_2O reduction slightly exceeded N_2O production under these conditions, net uptake rates of atmospheric N_2O were low, because diffusion of atmospheric N_2O into the soil, while occurring and thus supporting some uptake, was also limited by high water-filled pore space.

The diffusion of atmospheric N_2O into the soil increased during the drier 2016 summer, thus enabling somewhat larger net uptake of atmospheric N_2O . The observed soil moisture content of 18.2 v/v on July 30 (DOY 212), 2016, approximately represents the first quantile of observed soil moisture across 2015 and 2016 growing seasons. Rates of N_2O production during nitrification and denitrification were low in a large majority of microsites due to the oxygen inhibition effects as well as the diffusional constraints of soil C, ammonium, and nitrate substrates at low soil moisture. However, greater diffusion of atmospheric N_2O to soil microsites also increased the microsite concentrations of N_2O (dotted line in Figure 5j), including a small fraction of microsites with sufficiently low O_2 concentrations to not fully inhibit N_2O reduction (i.e., the simulated O_2 was near the kl for N_2O reduction ($kl_{\text{N}_2\text{O-red}}$) in approximately 10% of total microsites, resulting in net negative flux of N_2O (dotted line in Figure 5d; Table 1). Our observation of a net soil sink of atmospheric N_2O during summer drying events match with reports from other natural and managed ecosystems, ranging from tropical to temperate climates, where net uptake of atmospheric N_2O has been measured when mean bulk soil moisture was drier than would normally be expected for N_2O reduction (Donoso, Santana, & Sanhueza, 1993; Flechard, Neftel, Jocher, Ammann, & Fuhrer, 2005; Goldberg & Gebauer, 2009; Verchot et al., 2000; Yamulki, Goulding, Webster, & Harrison, 1995). In this case, however, we can demonstrate quantitatively that reducing

conditions in only about 10% of the microsites was sufficient to enable net uptake of atmospheric N_2O .

In contrast to the above two examples, intermediate soil moisture conditions are favorable for production to exceed consumption, resulting in modest net emissions of N_2O from soil to the atmosphere. Within this context, observed soil moisture content of 25.4 v/v on July 10, 2016 (DOY 192; dashed lines in Figure 5), approximately represents the third quantile of observed soil moisture across 2015 and 2016 growing seasons (Figure 5b). As expected, N_2O production was greatest at this intermediate soil moisture (Figure 5j), especially in microsites with high soil C. Because N_2O production was much higher than the very low N_2O reduction rates (Figure 5k) in a sufficiently large number of total soil microsites, a net positive mean N_2O flux resulted (Figure 5l).

Previous studies indicated that N_2O production during biological nitrification and denitrification often peak at 50%–80% of water-filled pore space (Davidson, 1991; Metivier, Pattey, & Grant, 2009), when soil moisture may be sufficiently high such that nitrate and nitrite are more available than O_2 as the alternate electron acceptor in many microsites, but the soil O_2 content is still high enough to mostly inhibit the reduction of N_2O to N_2 . This is the basis of the soil moisture function of the conceptual hole-in-the-pipe model (Firestone & Davidson, 1989). This relationship between soil moisture and N_2O production is often represented in models as an empirical statistical algorithm, such as a polynomial or similar function (e.g., Del Grosso et al., 2001; Potter, Matson, Vitousek, & Davidson, 1996). In contrast, the responses to soil moisture in the DAMM-GHG model produce the response pattern across simulated microsites (Figure S7) predicted by the conceptual hole-in-the-pipe model as an emergent property of the role of O_2 as substrate (for nitrification, see Figure S1) or inhibitor, as represented by different k_I values ($k_{I_{N_2O\text{-}prod}}$ and $k_{I_{N_2O\text{-}red}}$) used in DAMM mechanistic functions (Table 1).

For CH_4 , the microsite PDFs of production, consumption, and net emission were relatively straightforward. Methane production was high in a relatively few numbers of microsites with high soil moisture (and soil C). In contrast, CH_4 oxidation was high in the majority of microsites, which had low soil moisture, as the diffusion of both substrates (O_2 and CH_4) increased with decreasing soil moisture. Note that CH_4 production was several orders of magnitude lower than CH_4 oxidation across the simulated range of microsite moisture contents in our study, mainly due to significant oxygen inhibition. Therefore, microsite PDFs of CH_4 oxidation primarily dominated the net CH_4 emission, where net CH_4 emission linearly decreased with decreasing soil moisture (Figure 5e–h).

Taken together, these results demonstrate that representing production, consumption, and diffusion processes as a function of soil microsite PDFs can neatly encapsulate the factors affecting emissions of CO_2 , CH_4 , and N_2O in field studies and their underlying mechanisms. We also did a comparison of the model performance to a conventional framework, where we kept identical soil C and soil moisture values, set to the average values observed for the bulk soil, across all microsites. Although the performance of the model without microsite PDF was comparable to the one with microsite PDF

for CO_2 , model performance for CH_4 and N_2O was strongly affected by microsite variability (Figures S8 and S9). The model without microsite PDF simulated less uptake of CH_4 overall, had larger than observed peaks and valleys during in wet-up and dry-down events, and had more biased residuals (Figures S8b,g and S9b,e). For N_2O , the model with PDF representation of microsite variation also had an overall better fit to the observations and less biased residuals (Figures S8c,h and S9c,f).

The model–data fusion algorithm we used tends to reduce the overall model–data mismatch for the entire measurement window, and so it is not surprising that there are periods within that window where simulations do not match observations as well, such as the CO_2 and CH_4 fluxes of spring 2016 (Figure 2). This may also be due to other factors not included in the DAMM-GHG model, such as impacts of spring freeze–thaw cycles on C availability or phenology of root exudates. Additionally, the lower fraction of observed variability of N_2O fluxes accounted for by the model, compared to CO_2 and CH_4 fluxes, can be attributed to: (a) the low signal-to-noise ratio inherent to very low N_2O fluxes; and (b) the increased number and complexity of interactions of substrates (and inhibitors) for production (e.g., C, O_2 , NH_4^+ , and NO_3^-) and consumption (e.g., C, O_2 , and N_2O) of N_2O during nitrification and denitrification that are represented numerically in the model with additional parameters (Figures 2 and 4). We further discussed how these potential interacting processes might have increased the covariation of parameters related to N_2O dynamics than those related to CO_2 and CH_4 dynamics (see Section 4.3).

4.2 | Sensitivity analysis

4.2.1 | Sensitivity of predicted greenhouse gas fluxes to model parameters

Sensitivity indices indicated that the parameters representing the V_{max} terms in the Arrhenius equation (α and E_a) for production and consumption of each gas were the most influential for all three GHGs (Figure 6a–c), which is in line with other reports (Abramoff, Davidson, & Finzi, 2017; Zarnetske, Haggerty, Wondzell, Bokil, & González-Pinzón, 2012). Sensitivity of GHG fluxes to the parameters representing Michaelis–Menten equations was secondary to those of the $\alpha(s)$ and $E_a(s)$, where the individual ranking was associated with the importance of controlling drivers. Following $\alpha(s)$ and $E_a(s)$, some of the half-saturation constants, that is, $kM(s)$, were more important than others. For example, relatively higher sensitivity indices of the half-saturation constants of CH_4 (kM_{CH_4}) and O_2 ($kM_{O_2-CH_4}$) for CH_4 oxidation as compared to the half-saturation constants of C (kM_{C-CH_4}) and inhibition coefficient of O_2 (kI_{CH_4}) for CH_4 production can be explained by the dominant role of CH_4 oxidation in controlling net CH_4 emissions in our study. The inhibition coefficients of O_2 for N_2O reduction (kI_{N_2O-red}) and N_2O production ($kI_{N_2O-prod}$), along with the half-saturation constant of N_2O for N_2O reduction (kM_{N_2O}), had relatively higher sensitivity indices than those for the half-saturation constant of ammonium (kM_{NH_4}) and O_2 ($kM_{O_2-N_2O}$) during nitrification and nitrate (kM_{NO_3}) and C kM_{C-N_2O} during denitrification,

respectively. These results indicate the greater importance of reducing conditions and the diffusive supply of N_2O in controlling net N_2O emission at our site than either the concentrations of NH_4^+ and O_2 substrates for nitrification or NO_3^- and C substrates for denitrification. This result may be particular to our site where NO_3^- is uniformly low and the seasonal trend of NH_4^+ is much less dynamic than that of soil moisture (Fernandez et al., 1995). We speculate that kM_{NH_4} and kM_{NO_3} might be more commonly important in agricultural soils, where large temporal variations in NH_4^+ and NO_3^- are expected depending on the timing of fertilization and crop uptake.

4.2.2 | Sensitivity of predicted greenhouse gas fluxes to model drivers

Soil moisture primarily (and soil temperature secondarily) controlled the skewness of the microsite PDFs of production, consumption, and diffusion processes of each gas (Figure S10). Within this context, it is important to note that even though the parameters representing the V_{max} terms in the Arrhenius equation (E_a and α) had higher sensitivity indices than the Michaelis–Menten parameters that represent the influence of soil moisture (Figure 6a–c), most of the temporal variations in CH_4 and N_2O fluxes were nevertheless explained by soil moisture rather than temperature. The correlation matrix in Figure S5 also indicates that soil moisture, rather than soil temperature, controlled the temporal variation of CH_4 and N_2O fluxes. In other words, if the E_a value is changed, the average simulated flux for the entire time period increases or decreases significantly, but the within-season variation in CH_4 and N_2O fluxes is still dominantly influenced by variation in soil moisture (Figures S6 and S10).

4.3 | Collinearity analysis

As expected, the CI increases as the number of variable parameters increases (Figure 7). Given the parsimonious model structure, we had few problems of identifiability of the processes for CO_2 and CH_4 module, and the most parameter combinations remain below the threshold value of 15 (Brun, Kühni, Siegrist, Gujer, & Reichert, 2002; Omlin et al., 2001). However, we do have some parameter combinations with $CI > 15$ for the N_2O module, due to probable ambiguity of whether N_2O fluxes are affected more by production via nitrification (affected by NH_4^+ substrate), production via denitrification (affected by NO_3^- substrates), or consumption of N_2O (affected by N_2O diffusion). Trade-offs of processes within models can account for inflation of CI values (Keenan, Carbone, Reichstein, & Richardson, 2011; Richardson et al., 2010), which is likely the case for the N_2O module relative to the CO_2 and CH_4 modules. Overall, however, collinearity analysis indicates that the chances of having equifinality issues characterized by biologically improbable process representations were generally low in the DAMM-GHG model.

We believe that this success is largely due to the multiple constraints imposed by simultaneously modeling data streams of three different gases, which enabled us to capture the influence of

temporally and spatially varying drivers on GHG fluxes (Myrgiotis, Williams, Topp, & Rees, 2018). For example, when simulating only CH_4 flux, a better fit of the model to the data might be achieved by adjusting either the Michaelis–Menten parameters or the diffusion parameters, but it may be impossible to know which the “correct” adjustment is. However, if N_2O and CO_2 are also being simultaneously simulated, then adjusting the diffusion parameters will affect simulations of all three gases, whereas adjusting the Michaelis–Menten parameters for CH_4 oxidation should have little effect on the other two gases. Hence, the additional constraints help identify which model parameterizations are consistent with all three data streams of flux measurements.

4.4 | Microsite probability distribution functions permit model parsimony

Simulating a 3-D array of soil aggregates or pore networks (Arah & Vinten, 1995; Ebrahimi & Or, 2014; Yan et al., 2016) is another approach for representing spatial heterogeneity of soil matrix in biogeochemical models. However, explicit representation of soil spatial variability requires detailed information on soil structure, involving high-throughput instrumentations such as X-ray CT scan (see Carducci, Zinn, Rossoni, Heck, & Oliveira, 2017) and may require greater computational power than our PDF approach. Because of the limited measurements on the spatial variability of soil aggregates (or pores) at a plot scale, let alone at larger scales, and because of the increased computational complexity in spatially explicit model structures, scaling up of 3-D soil aggregate (or pore network) models to the ecosystem models and ESMs is still challenging.

Statistically representing microsite variation as PDFs in the DAMM-GHG model offers a relatively computationally efficient, yet mechanistically consistent, alternative way of simulating soil heterogeneity and maintaining model parsimony, as in the original DAMM model (Davidson et al., 2012, 2014; Sihi et al., 2018). We believe that our framework could be used to simulate fluxes of GHGs from other natural and managed systems as well as be scaled up to ecosystem models and ESMs.

4.5 | Opportunities for future improvement of the DAMM-GHG model

We represented all microsite-scale processes by optimizing an equivalent depth for R_h , where most of the biological reactions appear to happen in the soil of this study site (posterior range: 6–11 cm) and fixed that depth for simulating processes related to production and consumption of CH_4 and N_2O . This simplification allowed us to use unitless diffusion constants for gaseous and dissolved substrates and to avoid needing to know exact diffusion path lengths (see Davidson et al., 2012 for details). Exploring heterogeneity of diffusivity within and among soil horizons could be an appropriate next step.

Including more complex models of gas diffusion that includes variable diffusivity between intra-aggregate and inter-aggregate

pore spaces may be more appropriate for aggregated media like soil (Millington & Shearer, 1971; Resurreccion et al., 2010). Representing soil microsite PDFs in more than one vertically stratified soil horizon by differentiating between the organic and various mineral horizons may be needed for application to other sites (e.g., wetland with a seasonally variable depth to the water table), such that transport of gases between soil horizons and across soil-air boundary can be estimated using Fick's law. Measured vertical concentration profiles of soil gases could serve as additional data constraints for soil gas concentration profiles that become emergent simulated properties of this modeling approach. It would also be useful to have a data stream of heterogeneity of O₂ or redox potentials across microsites, but that would require new generations of microprobes.

Additionally, techniques that disentangle gross production and gross consumption rates of CH₄ and N₂O under field conditions could increase the predictive power of the dynamics of soil GHGs fluxes. For example, Chanton, Powelson, Abichou, and Hater (2008) reported that measuring stable carbon isotope of emitted CH₄ (¹³CH₄) is a feasible way to quantify gross CH₄ oxidation in situ. Likewise, Wen et al. (2016) demonstrated that ¹⁵N₂O pool dilution method can be effective to measure atmospheric N₂O uptake in soil under field conditions. In situ quantification of microbial activities pertaining to gross production and consumption of CH₄ and N₂O can also be pursued following the gas push-pull method (Urmann, Gonzalez-Gil, Schroth, Hofer, & Zeyer, 2005). Quantifying gross nitrogen transformations using stable isotope tracing could help constrain the sources of N₂O emissions (Morse & Bernhardt, 2013; Müller, Rütting, Kattge, Laughlin, & Stevens, 2007; Myrold & Tiedje, 1986).

In addition to nitrification and biological denitrification, other bacterial (Jensen & Burris, 1986; Yamazaki, Yoshida, Wada, & Matsuo, 1987) and fungal (Hayatsu, Tago, & Saito, 2008; Shoun, Kim, Uchiyama, & Sugiyama, 1992) contributions, as well as abiotic (Davidson, Chorover, & Dail, 2003; Vieten, 2008) sinks of N₂O in soil could be explored if warranted. Adding other controlling factors, such as pH effects, temporal dynamics of enzyme synthesis, and root exudation, could improve model performance for some sites (Butterbach-Bahl et al., 2013; Zheng & Doskey, 2015). In all of these cases, however, the potential additional explanatory value of more parameters and model complexity must be balanced with the availability of data to constrain them and with the advantages of model structure parsimony.

5 | CONCLUSIONS

Representing microsite heterogeneity as PDFs related to predictive processes offers a new approach for numerical representation of methanogenesis, methane oxidation, nitrification, and denitrification and other spatially and temporally variable microbial processes in soil. Our ability to accurately measure and skillfully model rates of these processes has been hampered by highly variable soil microsite conditions, which are difficult

to measure and simulate, but our use of PDFs to represent that variability offers a promising and computationally efficient approach. In addition, by measuring and modeling all three greenhouse gases (CO₂, CH₄, and N₂O), we have mechanistically and quantitatively explained the apparent paradox of observed simultaneous aerobic respiration that produces CO₂, CH₄ uptake (oxidation), CH₄ production, and N₂O uptake (reduction) in the same soil profile. Skillful representations of multiple ecologically relevant processes increase confidence of getting the right answers for the right reasons. This relatively parsimonious process modeling framework has the potential to be implemented within ecosystem models and ESMs to better capture the dynamics of soil-based greenhouse gases at landscape, regional, and global scales.

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REFERENCES

- Abramoff, R. Z., Davidson, E. A., & Finzi, A. C. (2017). A parsimonious modular approach to building a mechanistic belowground carbon and nitrogen model. *Journal of Geophysical Research: Biogeosciences*, 122(9), 2418–2434. <https://doi.org/10.1002/2017JG003796>
- Abramoff, R., Xu, X., Hartman, M., O'Brien, S., Feng, W., Davidson, E., ... Mayes, M. A. (2018). The Millennial model: In search of measurable pools and transformations for modeling soil carbon in the new century. *Biogeochemistry*, 137(1), 51–71. <https://doi.org/10.1007/s10533-017-0409-7>
- Angert, A., Yakir, D., Rodeghiero, M., Preisler, Y., Davidson, E. A., & Weiner, T. (2015). Using O₂ to study the relationships between soil CO₂ efflux and soil respiration. *Biogeosciences*, 12, 2089–2099. <https://doi.org/10.5194/bg-12-2089-2015>
- Arah, J. R. M., & Stephen, K. D. (1998). A model of the processes leading to methane emission from peatland. *Atmospheric Environment*, 32(19), 3257–3264. [https://doi.org/10.1016/S1352-2310\(98\)00052-1](https://doi.org/10.1016/S1352-2310(98)00052-1)
- Arah, J. R. M., & Vinten, A. J. A. (1995). Simplified models of anoxia and denitrification in aggregated and simple-structured soils. *European Journal of Soil Science*, 46(4), 507–517. <https://doi.org/10.1111/j.1365-2389.1995.tb01347.x>

- Arrhenius, S. (1889). Über die Reaktionsgeschwindigkeit bei der Inversion von Rohrzucker durch Säuren. *Zeitschrift für Physikalische Chemie*, 4U(1), 226. <https://doi.org/10.1515/zpch-1889-0416>
- Bernhardt, E. S., Blaszczak, J. R., Ficken, C. D., Fork, M. L., Kaiser, K. E., & Seybold, E. C. (2017). Control points in ecosystems: Moving beyond the hot spot hot moment concept. *Ecosystems*, 20(4), 665–682. <https://doi.org/10.1007/s10021-016-0103-y>
- Bidot, C., Lamboni, M., & Monod, H. (2018). *_multisensi: Multivariate Sensitivity Analysis_*. R package version 2.1-1. Retrieved from <https://CRAN.R-project.org/package=multisensi>
- Bodelier, P., Libochant, J. A., Blom, C., & Laanbroek, H. J. (1996). Dynamics of nitrification and denitrification in root-oxygenated sediments and adaptation of ammonia-oxidizing bacteria to low-oxygen or anoxic habitats. *Applied and Environment Microbiology*, 62(11), 4100–4107.
- Brewer, P., Calderón, F., Vigil, M., & von Fischer, J. (2018). Impacts of moisture, soil respiration, and agricultural practices on methanogenesis in upland soils as measured with stable isotope pool dilution. *Soil Biology and Biochemistry*, 127, 239–251. <https://doi.org/10.1016/j.soilbio.2018.09.014>
- Brun, R., Kühni, M., Siegrist, H., Gujer, W., & Reichert, P. (2002). Practical identifiability of ASM2d parameters—Systematic selection and tuning of parameter subsets. *Water Research*, 36(16), 4113–4127. [https://doi.org/10.1016/S0043-1354\(02\)00104-5](https://doi.org/10.1016/S0043-1354(02)00104-5)
- Brun, R., Reichert, P., & Künsch, H. R. (2001). Practical identifiability analysis of large environmental simulation models. *Water Resources Research*, 37(4), 1015–1030. <https://doi.org/10.1029/2000WR900350>
- Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R., & Zechmeister-Boltenstern, S. (2013). Nitrous oxide emissions from soils: How well do we understand the processes and their controls? *Philosophical Transactions of the Royal Society B: Biological Sciences*, 368(1621), 20130122. <https://doi.org/10.1098/rstb.2013.0122>
- Canion, A., Kostka, J. E., Gihring, T. M., Huettel, M., van Beusekom, J. E. E., Gao, H., ... Kuypers, M. M. M. (2014). Temperature response of denitrification and anammox reveals the adaptation of microbial communities to in situ temperatures in permeable marine sediments that span 50° in latitude. *Biogeosciences*, 11(2), 309–320. <https://doi.org/10.5194/bg-11-309-2014>
- Carbone, M. S., Richardson, A. D., Chen, M., Davidson, E. A., Hughes, H., Savage, K. E., & Hollinger, D. Y. (2016). Constrained partitioning of autotrophic and heterotrophic respiration reduces model uncertainties of forest ecosystem carbon fluxes but not stocks. *Journal of Geophysical Research: Biogeosciences*, 121(9), 2476–2492. <https://doi.org/10.1002/2016JG003386>
- Carducci, C. E., Zinn, Y. L., Rossoni, D. F., Heck, R. J., & Oliveira, G. C. (2017). Visual analysis and X-ray computed tomography for assessing the spatial variability of soil structure in a cultivated Oxisol. *Soil and Tillage Research*, 173, 15–23. <https://doi.org/10.1016/j.still.2016.03.006>
- Cattânio, J. H., Davidson, E. A., Nepstad, D. C., Verchot, L. V., & Ackerman, I. L. (2002). Unexpected results of a pilot throughfall exclusion experiment on soil emissions of CO₂, CH₄, N₂O, and NO in eastern Amazonia. *Biology and Fertility of Soils*, 36(2), 102–108. <https://doi.org/10.1007/s00374-002-0517-x>
- Chanton, J. P., Powelson, D. K., Abichou, T., & Hater, G. (2008). Improved field methods to quantify methane oxidation in landfill cover materials using stable carbon isotopes. *Environmental Science & Technology*, 42(3), 665–670. <https://doi.org/10.1021/es0710757>
- Chapuis-Lardy, L., Wrage, N., Metay, A., Chotte, J. L., & Bernoux, M. (2007). Soils, a sink for N₂O? A review. *Global Change Biology*, 13(1), 1–17. <https://doi.org/10.1111/j.1365-2486.2006.01280.x>
- Ciais, P., Sabine, C., Bala, G., Bopp, L., Brovkin, V., Canadell, J., ... Heimann, M. (2014). Carbon and other biogeochemical cycles. In T. F. Stocker, D. Qin, G.-K. Plattner, M. Tignor, S. K. Allen, J. Boschung, ... P. M. Midgley (Eds.), *Climate change 2013: The physical science basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change* (pp. 465–570). Cambridge, UK: Cambridge University Press.
- Conrad, R. (1996). Soil microorganisms as controllers of atmospheric trace gases (H₂, CO, CH₄, OCS, N₂O, and NO). *Microbiological Reviews*, 60(4), 609–640.
- Conrad, R. (2009). The global methane cycle: Recent advances in understanding the microbial processes involved. *Environmental Microbiology Reports*, 1(5), 285–292. <https://doi.org/10.1111/j.1758-2229.2009.00038.x>
- Crill, P., Martikainen, P., Nykanen, H., & Silvola, J. (1994). Temperature and N fertilization effects on methane oxidation in a drained peatland soil. *Soil Biology and Biochemistry*, 26(10), 1331–1339. [https://doi.org/10.1016/0038-0717\(94\)90214-3](https://doi.org/10.1016/0038-0717(94)90214-3)
- Davidson, E. A. (1991). Fluxes of nitrous oxide and nitric oxide from terrestrial ecosystems. In J. E. Rogers & W. B. Whitman (Eds.), *Microbial production and consumption of greenhouse gases: Methane, nitrous oxide, and halomethanes* (pp. 219–235). Washington, DC: American Society for Microbiology.
- Davidson, E. A., Belk, E., & Boone, R. D. (1998). Soil water content and temperature as independent or confounded factors controlling soil respiration in a temperate mixed hardwood forest. *Global Change Biology*, 4(2), 217–227. <https://doi.org/10.1046/j.1365-2486.1998.00128.x>
- Davidson, E. A., Chorover, J., & Dail, D. B. (2003). A mechanism of abiotic immobilization of nitrate in forest ecosystems: The ferrous wheel hypothesis. *Global Change Biology*, 9(2), 228–236. <https://doi.org/10.1046/j.1365-2486.2003.00592.x>
- Davidson, E. A., & Janssens, I. A. (2006). Temperature sensitivity of soil carbon decomposition and feedbacks to climate change. *Nature*, 440, 165. <https://doi.org/10.1038/nature04514>
- Davidson, E. A., Janssens, I. A., & Luo, Y. (2006). On the variability of respiration in terrestrial ecosystems: Moving beyond Q₁₀. *Global Change Biology*, 12(2), 154–164. <https://doi.org/10.1111/j.1365-2486.2005.01065.x>
- Davidson, E. A., Keller, M., Erickson, H. E., Verchot, L. V., & Veldkamp, E. (2000). Testing a conceptual model of soil emissions of nitrous and nitric oxides: Using two functions based on soil nitrogen availability and soil water content, the hole-in-the-pipe model characterizes a large fraction of the observed variation of nitric oxide and nitrous oxide emissions from soils. *BioScience*, 50(8), 667–680. [https://doi.org/10.1641/0006-3568\(2000\)050\[0667:Tacmos\]2.0.Co;2](https://doi.org/10.1641/0006-3568(2000)050[0667:Tacmos]2.0.Co;2)
- Davidson, E. A., Matson, P. A., Vitousek, P. M., Riley, R., Dunkin, K., Garcia-Mendez, G., & Maass, J. M. (1993). Processes regulating soil emissions of NO and N₂O in a seasonally dry tropical forest. *Ecology*, 74(1), 130–139. <https://doi.org/10.2307/1939508>
- Davidson, E. A., Samanta, S., Caramori, S. S., & Savage, K. (2012). The Dual Arrhenius and Michaelis-Menten kinetics model for decomposition of soil organic matter at hourly to seasonal time scales. *Global Change Biology*, 18(1), 371–384. <https://doi.org/10.1111/j.1365-2486.2011.02546.x>
- Davidson, E. A., Savage, K. E., & Finzi, A. C. (2014). A big-microsite framework for soil carbon modeling. *Global Change Biology*, 20(12), 3610–3620. <https://doi.org/10.1111/gcb.12718>
- Dean, J. F., Middelburg, J. J., Röckmann, T., Aerts, R., Blauw, L. G., Egger, M., ... Dolman, A. J. (2018). Methane feedbacks to the global climate system in a warmer world. *Reviews of Geophysics*, 56(1), 207–250. <https://doi.org/10.1002/2017RG000559>
- Del Grosso, S. J., Parton, W. J., Mosier, A. R., Hartman, M. D., Keough, C. A., Peterson, G. A., ... Schimel, D. S. (2001). Simulated effects of land use, soil texture, and precipitation on N gas emissions using DAYCENT. In R. F. Follett & J. L. Hatfield (Eds.), *Nitrogen in the environment: Sources, problems, and management* (pp. 413–431). Amsterdam, The Netherlands: Elsevier Science Publ.
- Donoso, L., Santana, R., & Sanhueza, E. (1993). Seasonal variation of N₂O fluxes at a tropical savannah site: Soil consumption of N₂O during the dry season. *Geophysical Research Letters*, 20(13), 1379–1382. <https://doi.org/10.1029/93GL01537>
- Ebrahimi, A. N., & Or, D. (2014). Microbial dispersal in unsaturated porous media: Characteristics of motile bacterial cell motions in unsaturated

- angular pore networks. *Water Resources Research*, 50(9), 7406–7429. <https://doi.org/10.1002/2014wr015897>
- Ebrahimi, A., & Or, D. (2018). On upscaling of soil microbial processes and biogeochemical fluxes from aggregates to landscapes. *Journal of Geophysical Research: Biogeosciences*, 123(5), 1526–1547. <https://doi.org/10.1029/2017JG004347>
- Eugster, W., Zeyer, K., Zeeman, M., Michna, P., Zingg, A., Buchmann, N., & Emmenegger, L. (2007). Methodical study of nitrous oxide eddy covariance measurements using quantum cascade laser spectrometry over a Swiss forest. *Biogeosciences*, 4(5), 927–939. <https://doi.org/10.5194/bg-4-927-2007>
- Fernandez, I. J., Lawrence, G. B., & Son, Y. (1995). Soil-solution chemistry in a low-elevation spruce-fir ecosystem, Howland, Maine. *Water, Air, and Soil Pollution*, 84(1), 129–145. <https://doi.org/10.1007/bf00479593>
- Fernandez, I. J., Rustad, L. E., & Lawrence, G. B. (1993). Estimating total soil mass, nutrient content, and trace metals in soils under a low elevation spruce-fir forest. *Canadian Journal of Soil Science*, 73(3), 317–328. <https://doi.org/10.4141/cjss93-034>
- Firestone, M. K., & Davidson, E. A. (1989). Microbiological basis of NO and N₂O production and consumption in soil. In M. O. Andreae & D. S. Schimel (Eds.), *Exchange of trace gases between terrestrial ecosystems and the atmosphere* (pp. 7–21). New York, NY: John Wiley & Sons Inc.
- Flechar, C. R., Neftel, A., Jocher, M., Ammann, C., & Fuhrer, J. (2005). Bidirectional soil/atmosphere N₂O exchange over two mown grassland systems with contrasting management practices. *Global Change Biology*, 11(12), 2114–2127. <https://doi.org/10.1111/j.1365-2486.2005.01056.x>
- Goldberg, S. D., & Gebauer, G. (2009). Drought turns a Central European Norway spruce forest soil from an N₂O source to a transient N₂O sink. *Global Change Biology*, 15(4), 850–860. <https://doi.org/10.1111/j.1365-2486.2008.01752.x>
- Groffman, P. M. (2012). Terrestrial denitrification: Challenges and opportunities. *Ecological Processes*, 1(1), 11. <https://doi.org/10.1186/2192-1709-1-11>
- Groffman, P. M., Butterbach-Bahl, K., Fulweiler, R. W., Gold, A. J., Morse, J. L., Stander, E. K., ... Vidon, P. (2009). Challenges to incorporating spatially and temporally explicit phenomena (hotspots and hot moments) in denitrification models. *Biogeochemistry*, 93(1), 49–77. <https://doi.org/10.1007/s10533-008-9277-5>
- Hayatsu, M., Tago, K., & Saito, M. (2008). Various players in the nitrogen cycle: Diversity and functions of the microorganisms involved in nitrification and denitrification. *Soil Science and Plant Nutrition*, 54(1), 33–45. <https://doi.org/10.1111/j.1747-0765.2007.00195.x>
- Holtan-Hartwig, L., Dörsch, P., & Bakken, L. R. (2002). Low temperature control of soil denitrifying communities: Kinetics of N₂O production and reduction. *Soil Biology and Biochemistry*, 34(11), 1797–1806. [https://doi.org/10.1016/S0038-0717\(02\)00169-4](https://doi.org/10.1016/S0038-0717(02)00169-4)
- Hursh, A., Ballantyne, A., Cooper, L., Maneta, M., Kimball, J., & Watts, J. (2017). The sensitivity of soil respiration to soil temperature, moisture, and carbon supply at the global scale. *Global Change Biology*, 23(5), 2090–2103. <https://doi.org/10.1111/gcb.13489>
- IPCC. (2014). *Climate Change 2014: Synthesis Report. Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change* [Core Writing Team, R. K. Pachauri & L. A. Meyer (Eds.)]. Geneva, Switzerland: IPCC, 151 pp.
- Jensen, B. B., & Burris, R. H. (1986). Nitrous oxide as a substrate and as a competitive inhibitor of nitrogenase. *Biochemistry*, 25(5), 1083–1088. <https://doi.org/10.1021/bi00353a021>
- Keenan, T. F., Carbone, M. S., Reichstein, M., & Richardson, A. D. (2011). The model–data fusion pitfall: Assuming certainty in an uncertain world. *Oecologia*, 167(3), 587. <https://doi.org/10.1007/s00442-011-2106-x>
- Keiluweit, M., Gee, K., Denney, A., & Fendorf, S. (2018). Anoxic microsites in upland soils dominantly controlled by clay content. *Soil Biology and Biochemistry*, 118, 42–50. <https://doi.org/10.1016/j.soilbio.2017.12.002>
- Keller, M., & Matson, P. A. (1994). Biosphere–atmosphere exchanges of trace gases in the tropics: Evaluating the effects of land use changes. In R. G. Prinn (Ed.), *Global atmospheric biospheric chemistry* (pp. 103–117). New York, NY: Plenum Press.
- Körner, H., & Zumft, W. G. (1989). Expression of denitrification enzymes in response to the dissolved oxygen level and respiratory substrate in continuous culture of *Pseudomonas stutzeri*. *Applied and Environmental Microbiology*, 55(7), 1670–1676.
- Lamboni, M., Makowski, D., Lehuger, S., Gabrielle, B., & Monod, H. (2009). Multivariate global sensitivity analysis for dynamic crop models. *Field Crops Research*, 113(3), 312–320. <https://doi.org/10.1016/j.fcr.2009.06.007>
- Lindgren, F., & Rue, H. (2015). Bayesian spatial modelling with R-INLA. *Journal of Statistical Software*, 63(19), 1–25. <https://doi.org/10.18637/jss.v063.i19>
- Linn, D. M., & Doran, J. W. (1984). Effect of water-filled pore space on carbon dioxide and nitrous oxide production in tilled and nontilled soils 1. *Soil Science Society of America Journal*, 48(6), 1267–1272. <https://doi.org/10.2136/sssaj1984.03615995004800060013x>
- Lloyd, J., & Taylor, J. A. (1994). On the temperature dependence of soil respiration. *Functional Ecology*, 8(3), 315–323. <https://doi.org/10.2307/2389824>
- Lurndahl, N. (2016). *Temporal and spatial trends in the abundance of functional denitrification genes and observed soil moisture and potential denitrification rates*. MS thesis, University of Minnesota, Minneapolis.
- Metivier, K. A., Pattey, E., & Grant, R. F. (2009). Using the ecosys mathematical model to simulate temporal variability of nitrous oxide emissions from a fertilized agricultural soil. *Soil Biology and Biochemistry*, 41(12), 2370–2386. <https://doi.org/10.1016/j.soilbio.2009.03.007>
- Millington, R. J., & Shearer, R. C. (1971). Diffusion in aggregated porous media. *Soil Science*, 111(6), 372–378. <https://doi.org/10.1097/00010694-197106000-00007>
- Morse, J. L., & Bernhardt, E. S. (2013). Using ¹⁵N tracers to estimate N₂O and N₂ emissions from nitrification and denitrification in coastal plain wetlands under contrasting land-uses. *Soil Biology and Biochemistry*, 57, 635–643. <https://doi.org/10.1016/j.soilbio.2012.07.025>
- Moyano, F. E., Manzoni, S., & Chenu, C. (2013). Responses of soil heterotrophic respiration to moisture availability: An exploration of processes and models. *Soil Biology and Biochemistry*, 59, 72–85. <https://doi.org/10.1016/j.soilbio.2013.01.002>
- Müller, C., Rütting, T., Kattge, J., Laughlin, R. J., & Stevens, R. J. (2007). Estimation of parameters in complex ¹⁵N tracing models by Monte Carlo sampling. *Soil Biology and Biochemistry*, 39(3), 715–726. <https://doi.org/10.1016/j.soilbio.2006.09.021>
- Myrgiotis, V., Williams, M., Topp, C. F., & Rees, R. M. (2018). Improving model prediction of soil N₂O emissions through Bayesian calibration. *Science of the Total Environment*, 624, 1467–1477. <https://doi.org/10.1016/j.scitotenv.2017.12.202>
- Myrold, D. D., & Tiedje, J. M. (1986). Simultaneous estimation of several nitrogen cycle rates using ¹⁵N: Theory and application. *Soil Biology and Biochemistry*, 18(6), 559–568. [https://doi.org/10.1016/0038-0717\(86\)90076-3](https://doi.org/10.1016/0038-0717(86)90076-3)
- Nedwell, D. B., & Watson, A. (1995). CH₄ production, oxidation and emission in a U.K. ombrotrophic peat bog: Influence of SO₄²⁻ From acid rain. *Soil Biology and Biochemistry*, 27(7), 893–903. [https://doi.org/10.1016/0038-0717\(95\)00018-A](https://doi.org/10.1016/0038-0717(95)00018-A)
- Omlin, M., Brun, R., & Reichert, P. (2001). Biogeochemical model of Lake Zürich: Sensitivity, identifiability and uncertainty analysis. *Ecological Modelling*, 141(1), 105–123. [https://doi.org/10.1016/S0304-3800\(01\)00257-5](https://doi.org/10.1016/S0304-3800(01)00257-5)

- Or, D. (2019). The tyranny of small scales – on representing soil processes in global land surface models. *Water Resources Research*. <https://doi.org/10.1029/2019wr024846>
- Parkin, T. B. (1987). Soil microsites as a source of denitrification variability. *Soil Science Society of America Journal*, 51(5), 1194–1199. <https://doi.org/10.2136/sssaj1987.03615995005100050019x>
- Parkin, T. B. (1993). Spatial variability of microbial processes in soil—A review. *Journal of Environmental Quality*, 22(3), 409–417. <https://doi.org/10.2134/jeq1993.00472425002200030004x>
- Potter, C. S., Matson, P. A., Vitousek, P. M., & Davidson, E. A. (1996). Process modeling of controls on nitrogen trace gas emissions from soils worldwide. *Journal of Geophysical Research: Atmospheres*, 101(D1), 1361–1377. <https://doi.org/10.1029/95jd02028>
- R Core Team. (2018). *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. Retrieved from <https://www.R-project.org/>
- Resurreccion, A. C., Moldrup, P., Kawamoto, K., Hamamoto, S., Rolston, D. E., & Komatsu, T. (2010). Hierarchical, bimodal model for gas diffusivity in aggregated, unsaturated soils. *Soil Science Society of America Journal*, 74(2), 481–491. <https://doi.org/10.2136/sssaj2009.0055>
- Revolution Analytics, & Weston, S. (2015). doParallel: Foreach Parallel Adaptor for the 'parallel' Package. R package version 1.0.10. Retrieved from <https://CRAN.R-project.org/package=doParallel>
- Richardson, A. D., Hollinger, D. Y., Shoemaker, J. K., Hughes, H., Savage, K., & Davidson, E. A. (2019). Six years of ecosystem-atmosphere greenhouse gas fluxes measured in a sub-boreal forest. *Scientific Data*, 6(1), 117. <https://doi.org/10.1038/s41597-019-0119-1>
- Richardson, A. D., Williams, M., Hollinger, D. Y., Moore, D. J. P., Dail, D. B., Davidson, E. A., ... Savage, K. (2010). Estimating parameters of a forest ecosystem C model with measurements of stocks and fluxes as joint constraints. *Oecologia*, 164(1), 25–40. <https://doi.org/10.1007/s00442-010-1628-y>
- Rue, H., Martino, S., & Chopin, N. (2009). Approximate Bayesian inference for latent Gaussian models by using integrated nested Laplace approximations. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)*, 71(2), 319–392. <https://doi.org/10.1111/j.1467-9868.2008.00700.x>
- Saha, D., Kemanian, A. R., Montes, F., Gall, H., Adler, P. R., & Rau, B. M. (2018). Lorenz curve and gini coefficient reveal hot spots and hot moments for nitrous oxide emissions. *Journal of Geophysical Research: Biogeosciences*, 123(1), 193–206. <https://doi.org/10.1002/2017JG004041>
- Savage, K. E., & Davidson, E. A. (2003). A comparison of manual and automated systems for soil CO₂ flux measurements: Trade-offs between spatial and temporal resolution. *Journal of Experimental Botany*, 54(384), 891–899. <https://doi.org/10.1093/jxb/erg121>
- Savage, K. E., Davidson, E. A., Abramoff, R. Z., Finzi, A. C., & Giasson, M.-A. (2018). Partitioning soil respiration: Quantifying the artifacts of the trenching method. *Biogeochemistry*, 140(1), 53–63. <https://doi.org/10.1007/s10533-018-0472-8>
- Savage, K. E., Davidson, E. A., & Richardson, A. D. (2008). A conceptual and practical approach to data quality and analysis procedures for high-frequency soil respiration measurements. *Functional Ecology*, 22(6), 1000–1007. <https://doi.org/10.1111/j.1365-2435.2008.01414.x>
- Savage, K. E., Phillips, R., & Davidson, E. (2014). High temporal frequency measurements of greenhouse gas emissions from soils. *Biogeosciences*, 11(10), 2709–2720. <https://doi.org/10.5194/bg-11-2709-2014>
- Schlesinger, W. H. (2013). An estimate of the global sink for nitrous oxide in soils. *Global Change Biology*, 19(10), 2929–2931. <https://doi.org/10.1111/gcb.12239>
- Shoun, H., Kim, D.-H., Uchiyama, H., & Sugiyama, J. (1992). Denitrification by fungi. *FEMS Microbiology Letters*, 94(3), 277–281. <https://doi.org/10.1111/j.1574-6968.1992.tb05331.x>
- Sihi, D., Davidson, E. A., Chen, M., Savage, K. E., Richardson, A. D., Keenan, T. F., & Hollinger, D. Y. (2018). Merging a mechanistic enzymatic model of soil heterotrophic respiration into an ecosystem model in two AmeriFlux sites of northeastern USA. *Agricultural and Forest Meteorology*, 252, 155–166. <https://doi.org/10.1016/j.agrfor.2018.01.026>
- Silver, W. L., Lugo, A. E., & Keller, M. (1999). Soil oxygen availability and biogeochemistry along rainfall and topographic gradients in upland wet tropical forest soils. *Biogeochemistry*, 44(3), 301–328. <https://doi.org/10.1007/bf00996995>
- Soetaert, K. (2016). R Package FME: Inverse modelling, sensitivity, Monte Carlo—Applied to a dynamic simulation model (CRAN Vignette 2). Retrieved from <https://cran.r-project.org/web/packages/FME/vignettes/FMEdyna.pdf>
- Soetaert, K., & Petzoldt, T. (2010). Inverse modelling, sensitivity and Monte Carlo analysis in R using package FME. *Journal of Statistical Software*, 33(3), 1–28.
- Stark, J. M. (1996). Modeling the temperature response of nitrification. *Biogeochemistry*, 35(3), 433–445. <https://doi.org/10.1007/BF02183035>
- Stark, J. M., & Firestone, M. K. (1996). Kinetic characteristics of ammonium-oxidizer communities in a California oak woodland-annual grassland. *Soil Biology and Biochemistry*, 28(10–11), 1307–1317. [https://doi.org/10.1016/S0038-0717\(96\)00133-2](https://doi.org/10.1016/S0038-0717(96)00133-2)
- Stoyan, H., De-Polli, H., Böhm, S., Robertson, G. P., & Paul, E. A. (2000). Spatial heterogeneity of soil respiration and related properties at the plant scale. *Plant and Soil*, 222(1), 203–214. <https://doi.org/10.1023/a:1004757405147>
- Syakila, A., Kroeze, C., & Slomp, C. P. (2010). Neglecting sinks for N₂O at the earth's surface: Does it matter? *Journal of Integrative Environmental Sciences*, 7(S1), 79–87. <https://doi.org/10.1080/1943815x.2010.497492>
- Teh, Y. A., Silver, W. L., & Conrad, M. E. (2005). Oxygen effects on methane production and oxidation in humid tropical forest soils. *Global Change Biology*, 11(8), 1283–1297. <https://doi.org/10.1111/j.1365-2486.2005.00983.x>
- Tian, H., Yang, J., Xu, R., Lu, C., Canadell, J. G., Davidson, E. A., ... Zhang, B. (2019). Global soil nitrous oxide emissions since the preindustrial era estimated by an ensemble of terrestrial biosphere models: Magnitude, attribution, and uncertainty. *Global Change Biology*, 25(2), 640–659. <https://doi.org/10.1111/gcb.14514>
- Urmann, K., Gonzalez-Gil, G., Schroth, M. H., Hofer, M., & Zeyer, J. (2005). New field method: Gas push-pull test for the in-situ quantification of microbial activities in the vadose zone. *Environmental Science & Technology*, 39(1), 304–310. <https://doi.org/10.1021/es0495720>
- Vant't Hoff, J. H., & Lehfeldt, R. A. (1899). *Lectures on theoretical and physical chemistry. Part 1. Chemical dynamics*. London: Edward Arnold.
- Verchot, L. V., Davidson, E. A., Cattânio, J. H., & Ackerman, I. L. (2000). Land-use change and biogeochemical controls of methane fluxes in soils of eastern Amazonia. *Ecosystems*, 3(1), 41–56. <https://doi.org/10.1007/s100210000009>
- Verstraete, W., & Focht, D. (1977). Biochemical ecology of nitrification and denitrification. In M. Alexander (Ed.), *Advances in microbial ecology* (Vol. 1, pp. 135–214). Boston, MA: Springer.
- Vieten, B. (2008). *N₂O reduction in soils*. PhD thesis, University of Basel, Switzerland.
- Wang, B., Brewer, P. E., Shugart, H. H., Lerdau, M. T., & Allison, S. D. (2019). Soil aggregates as biogeochemical reactors and implications for soil-atmosphere exchange of greenhouse gases—A concept. *Global Change Biology*, 25(2), 373–385. <https://doi.org/10.1111/gcb.14515>
- Wen, Y., Chen, Z., Dannenmann, M., Carminati, A., Willibald, G., Kiese, R., ... Corre, M. D. (2016). Disentangling gross N₂O production and consumption in soil. *Scientific Reports*, 6, 36517. <https://doi.org/10.1038/srep36517>

- Westermann, P. (1993). Temperature regulation of methanogenesis in wetlands. *Chemosphere*, 26(1–4), 321–328. [https://doi.org/10.1016/0045-6535\(93\)90428-8](https://doi.org/10.1016/0045-6535(93)90428-8)
- Xu, X., Yuan, F., Hanson, P. J., Wullschleger, S. D., Thornton, P. E., Riley, W. J., ... Tian, H. (2016). Reviews and syntheses: Four decades of modeling methane cycling in terrestrial ecosystems. *Biogeosciences*, 13(12), 3735–3755. <https://doi.org/10.5194/bg-13-3735-2016>
- Yamazaki, T., Yoshida, N., Wada, E., & Matsuo, S. (1987). N₂O reduction by *Azotobacter vinelandii* with emphasis on kinetic nitrogen isotope effects. *Plant and Cell Physiology*, 28(2), 263–271. <https://doi.org/10.1093/oxfordjournals.pcp.a077292>
- Yamulki, S., Goulding, K. W. T., Webster, C. P., & Harrison, R. M. (1995). Studies on NO and N₂O fluxes from a wheat field. *Atmospheric Environment*, 29(14), 1627–1635. [https://doi.org/10.1016/1352-2310\(95\)00059-8](https://doi.org/10.1016/1352-2310(95)00059-8)
- Yan, Z., Liu, C., Todd-Brown, K. E., Liu, Y., Bond-Lamberty, B., & Bailey, V. L. (2016). Pore-scale investigation on the response of heterotrophic respiration to moisture conditions in heterogeneous soils. *Biogeochemistry*, 131(1), 121–134. <https://doi.org/10.1007/s10533-016-0270-0>
- Yang, W. H., McNicol, G., Teh, Y. A., Estera-Molina, K., Wood, T. E., & Silver, W. L. (2017). Evaluating the classical versus an emerging conceptual model of peatland methane dynamics. *Global Biogeochemical Cycles*, 31(9), 1435–1453. <https://doi.org/10.1002/2017GB005622>
- Yoshida, N., Matsuo, S., Yamazaki, T., & Wada, E. (1987). N₂O Reduction by *Azotobacter vinelandii* with emphasis on kinetic nitrogen isotope effects. *Plant and Cell Physiology*, 28(2), 263–271. <https://doi.org/10.1093/oxfordjournals.pcp.a077292>
- Zarnetske, J. P., Haggerty, R., Wondzell, S. M., Bokil, V. A., & González-Pinzón, R. (2012). Coupled transport and reaction kinetics control the nitrate source-sink function of hyporheic zones. *Water Resources Research*, 48(11), 1–15. <https://doi.org/10.1029/2012WR011894>
- Zheng, J., & Doskey, P. V. (2015). Modeling Nitrous oxide production and reduction in soil through explicit representation of denitrification enzyme kinetics. *Environmental Science & Technology*, 49(4), 2132–2139. <https://doi.org/10.1021/es504513v>

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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