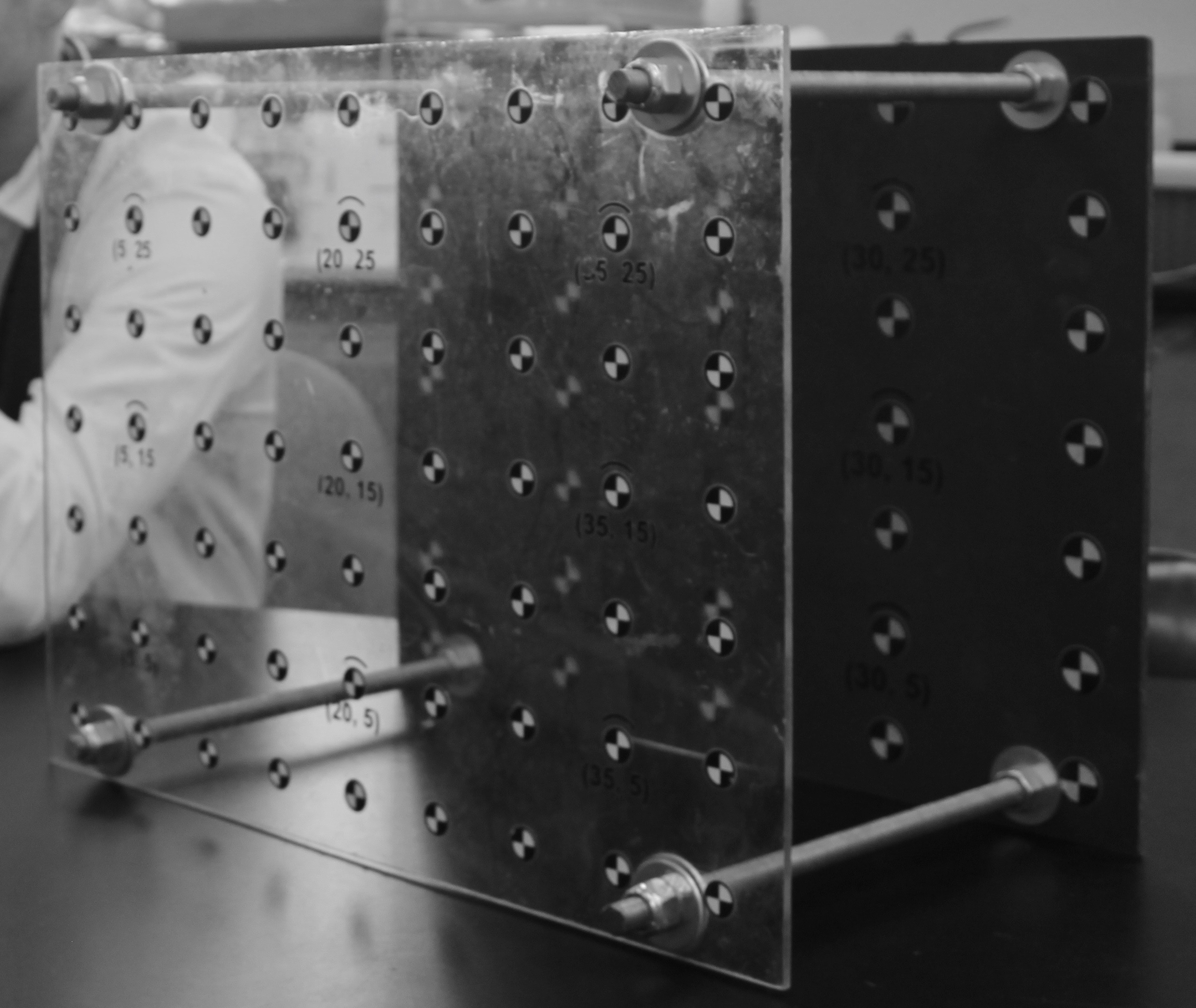
Videogrammetry Metrics for Assessing Fish Foraging Behavior: Implications for Behavioral Ecology and Management

Keane Flynn

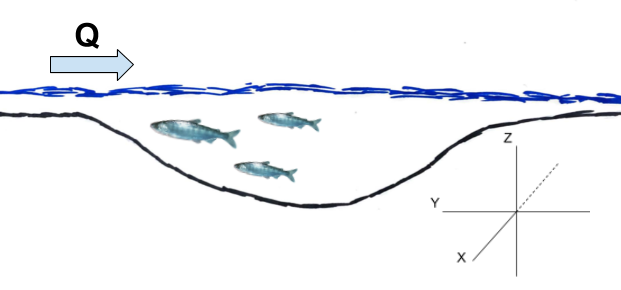
4/30/2019

# **Introduction**

Fish foraging behavior has long been a topic of natural observation, wonder, and recreational curiosity (Walton 1655). For decades, fisheries scientists have also observed foraging behavior and considered the implications of forging behavior on fish growth and energetics (Fuasch 1984; Hughes and Dill 1990; Hayes et al. 2000), adaptation to habitat variability and contexts (Dill 1983; Fausch et al. 1997; Nislow et al. 1998; Nielson 1992), niche partitioning between sympatric species (Nakano et al. 1999a) and ontogeny (Sánchez-Hernández et al. 2012). A major challenge in the scientific study of foraging behavior has been limitations associated with visual observation, including: (1) poor estimation of quantities such as distance, length, and time, (2) the inability to follow multiple focal animals at one time, (3) the inability to review a behavioral sequence or to share with multiple observers, (4) the inability to deal with quick or fleeting behaviors in real time, and (5) making observations in a non-invasive method to retain natural behavior (Altmann 1974, Neuswanger 2016). Recent advances in videogrammetry and computer programming technologies have provided technological solutions/mitigation for some of these problems and made it possible to collect precise quantitative and qualitative measurements of animal behavior and interactions in 3-dimensional space (Neuswanger 2016, Hughes and Kelly 1996; Vivancos and Closs 2015, Piccolo et al. 2007, Glass 2013).  
 The videogrammetric program Vidsync represents one of the first open source software programs to provide quantitative 3-D functionality to fish behavior ecology (Neuswanger 2016). Vidsync is a novel software program that allows the user to triangulate a 3D position, in relative space, from two or more known lines of sight (Neuswanger 2016a). The software was developed for in situ study of juvenile chinook drift-feeding behavior and growth rates (Neuswanger et al. 2014). Data collection is accomplished by calibration of two or more simultaneous in-stream videos via a dual-plane calibration frame with known, uniform distances between each point across the X, Y, Z axis (Figure 1). The primary output data from the Vidsync analysis was X, Y, Z location, time, an object (subsample and fish identification number), event (foraging or interactive behavior), or measurement (e.g. length and z-axis location of the surface) (Figure 2). These coordinates, timecodes, and observations can then be exported and processed into code-based programs to create statistical and graphical representations of the data collected.



**Figure 1** Calibration frame used in the VidSync program to calibrate the two videos to calculate positions in relative space



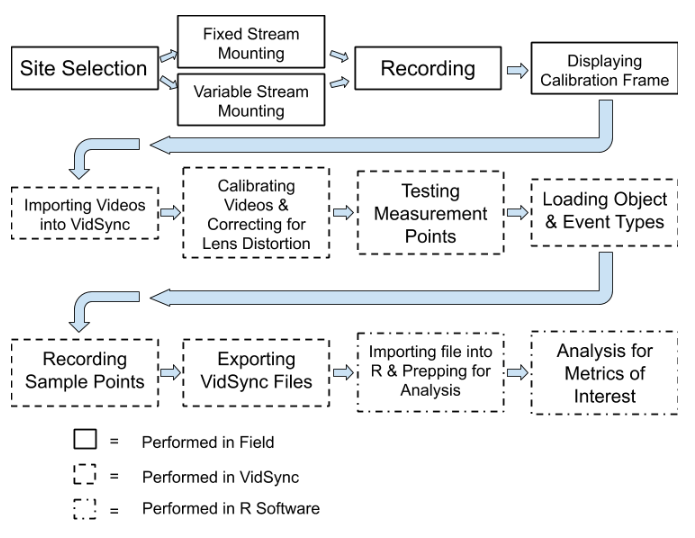
**Figure 2** Profile view of a pool-riffle sequence with X, Y, Z planes relative to a fish maintaining positive rheotaxis.

Although videogrammetric programs such as Vidsync have allowed for profound improvements in the functional ability to track and quantify animal behavior, locations and movement, there has yet to be an effort to synthesize classical ethology and foraging ecology methods with output from the 3-D videogrammetric tools. In addition, the possibility of using behavioral states or changes in threshold behaviors as indicators of environmental change for resource managers is enhanced by the quantitively power of 3-D videogrammetrics (Sih et al. 2011). However, this application of videogrammetry has not been explored. In this paper we will introduce a set of specific methods to quantify behavioral states and behavioral gradients in juvenile salmonids using the 3-D program Vidsync focusing on behaviors with a long history of use in foraging ecology. We will also link the quantitative methods for evaluating these behaviors to potential management applications such as instream flow science or habitat restoration monitoring specifically targeting juvenile salmonids. Our objective is to provide a methodological and contextual basis for synthesize classical methods in juvenile foraging ecology with novel videogrammetric tools, and to consider the application of these methods to some targeted management scenarios.

# **Methods**

## *Overview*

VidSync tutorials and use have been well established for creating data from video observations (Neuswanger 2014; Neuswanger 2016b). However, its use for amalysis of specific metrics that might have ecological implications has not been well documented. We will look in-depth at 6 methods of data analysis, including: proportion of forage behaviors, fork length (FL) calculations, distance per time, nearest neightbor distance (NND), distance from surface, and occupied volume. For each of these analyses, we will look at the (1) use of each metric in behavioral ecology, (2) the significance of each calculation, (3) how these measurements are made for this application, and (4) exemplary R code for each of these calculations. To ensure that video observations can be converted into useful data, information flow must follow a logical path (Figure 3). Some of this information flow has been previously described and documented and is useful to use in conjunction with the methods below (Neuswanger 2016a; Neuswanger 2016b). Throughout this section, we will follow this information flow and provide methodology to turn observations into quantitative metrics.

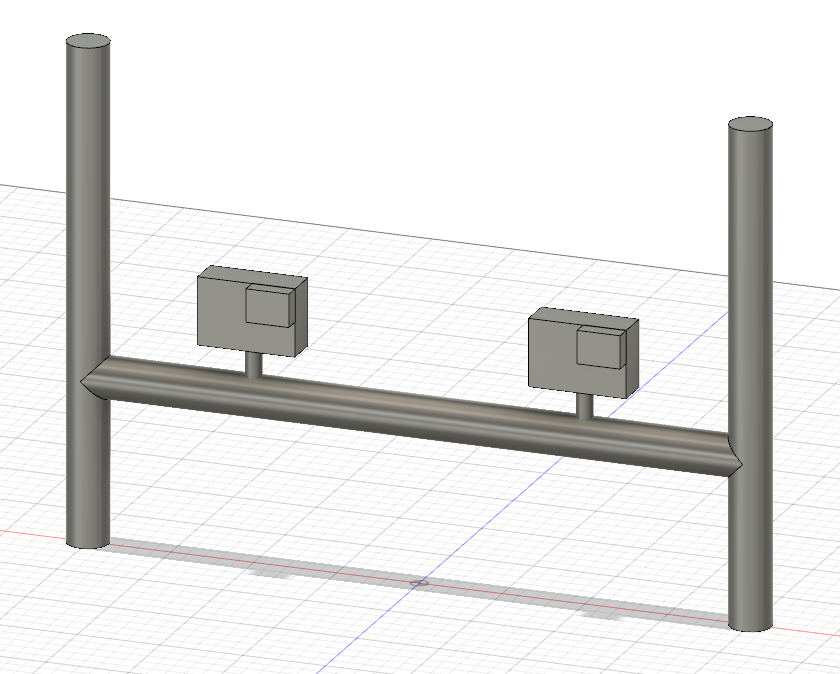


**Figure 3** Flow chart of VidSync use from field to analysis

## **Field Methods**

### *Streamside set up and use*

For underwater videography, we found that using GoPro cameras allowed for a relatively low-cost method of video capture as well as wide angle lens to capture all fish in the study area continuously. Using Eularian observational methodology, creating a fixed mount system to track behavioral observations against changes in habitat has proved an effective method (Vivancos and Closs 2015; Piccolo 2005; Rossi et al. 2019 in prep). To properly mount these cameras for repeated samples, we found that having multiple mount points to the streambed via rebar hammered into the alluvium ensures that the same habitat area is recorded each time. To fix the cameras to the mount, a simple device constructed from PVC pipe and bicycle mounts covers the widest amount of habitat while minimizing area where the camera angles do not overlap (Figure 4). When tracking specific individuals independent of changes in habitat, use of LaGrangian observations can be performed through varaible mounting throughout the habitat tracking the target population (Neuswanger 2014). Mounting cameras for this method of observation can be performed more simply using the same materials fixed in a T-shape using a single instream rebar mount for ease of adjusting for location and angle of recording.



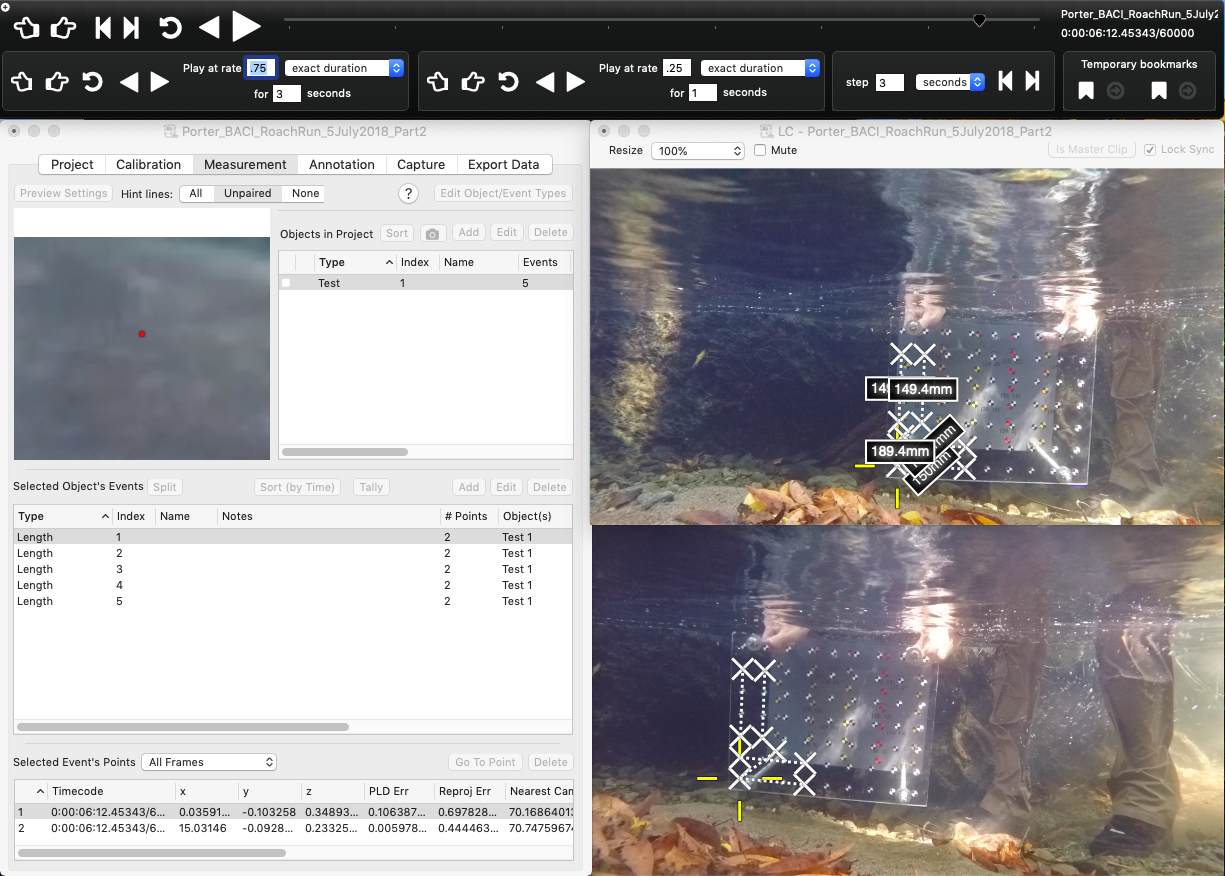
**Figure 4** Fusion 360 schematic for how to mount GoPro cameras for Eularian observation VidSync applications

When recording behavior, especially in low-flow situations, it is important to enter the river to mount the camera from downstream to avoid increasing turbidity in the study area. That being said, this method should not be employed in areas of high turbidity or poor lighting as behavior observations will be abysmal (Neuswanger 2016a). While it depends on the specific study, we recommend roughly 25-30 minutes of recording to ensure useful behavioral observations. Immediately following the recording period, someone must enter the creek with a calibration frame (from downstream to avoid increasing turbidity) and place it in front of the cameras where all points on the frame are visible (Figure 1).

## **VidSync Methods**

### *Test Measurements*

Once the videos have been recorded, loaded into VidSync, and calibrated, this calibrated video must be tested to ensure that the measurements are valid (Neuswanger 2014; Neuswanger 2016b). Using the known distances between points on the calibration frame, test measurements can be made along the X (front frame to rear frame), Y, and Z axes using the length function in the program to ensure that what is being measured in VidSync matches empirical measurement data (Figure 5).



**Figure 5** Test Measurements plotted on the VidSync calibration frame with known distances between points

### *VidSync Program Use*

The first 5 minutes of the imported video will be discarded to allow for an adjustment period to allow fish to resume normal foraging behavior (Nakano et al. 1999; Fausch et al. 1997). The video period will be chosen via randomly stratified selection by performing a scan sample to find the most fish-dense 10 minute period of the recording, then each 30 second interval of the video is then assigned an integer (1-20) and using a random number generator, 6 random integers (1-20) representing a unique 30 second period are selected to make up a total of 3 minutes of data supported behavioral observations (Neuswanger 2014; Rossi et al. 2019 in prep). Each of these 6 chosen integers will represent a subsample within the sample video to allow for a period of total observation similar to what has been previously used with this method, however it allows for a higher sample size while removing any potential bias in sampling period.   
 Following verification of measurement accuracy, the desired subsample intervals are selected, and the video has been calibrated and corrected for lens distortion, object and event types can be created to classify subsamples and behavioral observations to be associated with sample points (Neuswanger 2016b). The object types can be used to represent unique subsamples and will be exported into a vector named, "Object(s)" which will include the title of the object as well as the "index" which is used to represent an individual fish identification number. Additionally, the option to include a name with each object is given which should be used to identify species or specific individual fish if unique identification is possible (i.e. applications with VIE tagging). These object types should be saved as follows: Subsample\_1, Subsample\_2, Subsample\_3, Subsample\_4, Subsample\_5, Subsample\_6.  
 Following verification of measurement accuracy and the necessary object and event types are imported (see proportion of behaviors section), sample points can be recorded. Sample points are recorded in 3 second increments over the course of a 30 second subsample for a total of 11 sample points. For ease of data manipulation, each sample point for unique fish within a subsample should be taken at the same timecode (i.e. at timecode 3.000 seconds, 6.000 seconds, 9.000 seconds, etc.) until all fish in the sample are accounted for. Once all 6 subsamples have each present fish recorded, the video can then be exported from VidSync to be analyzed.

## **Data Analysis**

For data analysis, we will be using R software to compute the aforementioned metrics (RStudio Team 2015). They can be imported into other softwares such as MATLAB and Wolfram Mathematica, however for the analyses performed below, we have found that using R is the most simple method with the most amount of outside support.

### *Importing VidSync CSV File*

Data from VidSync can either be exported as an XML or CSV file, for our application we will be exporting it as the latter into R for analysis. Upon importing it, we will be cleaning up some of the data vectors using regular expression matching for ease of operation through the Stringr package in R (Whickam 2018).

VidSync\_Test\_Data <- readr::read\_csv(file = "VidSync-TestData.csv",   
 skip = 2,  
 col\_names = c("objects", "event", "timecode", "time\_sec", "X", "Y", "Z", "pld\_error", "projection\_error", "nearest\_camera\_distance", "screen\_coordinates"),  
 col\_types = "cccdddddddd") %>%   
 mutate(subsample = as.numeric(str\_extract(objects, "\\d"))) %>% #This column will identify which of the 6 subsamples the data belongs to.  
 mutate(fish\_ID = as.numeric(str\_extract(objects, "\\h\\d{1,2}"))) %>% #A fish ID denotes a unique individual during each subsample. Unique individuals can be tracked across subsamples if desired, but must be given a unique name in the "Name" option (i.e. Omykiss-A1) when the Event type is specified while being processed in VidSync.  
 mutate(species = str\_extract(objects, "Omykiss|Okisutch")) %>% #Note that the use of regular expressions in this line are to be specified to whatever species are being recorded in your experiment and excluding any non-relevant ones.  
 select(subsample, fish\_ID, species, time\_sec, X, Y, Z, objects, event) #All other data vectors were excluded for the sake of neatness and what is useful in the metrics below.

## Nearest Neighbor Distance

Nearest neighbor distance (NND) has historically been used as a metric for understanding inter/intraspecies interactions as well as making observations about general trends regarding animal density in a study region (Clark 1954). This metric has shown to have a negative association with foraging rates and agonistic behavior in foraging juvenile salmonids (Nielsen 1992). NNDs have also been associated with relatedness in juvenile salmonids, where related fish will have closer NNDs to their kin and defend territories in groups (Brown and Brown 1993).  
 NND is calculated by finding the nearest neighbor to each unique fish over the course of a subsample. This is calculated by grouping timecodes, denoted as "time\_sec" in the example code, to show all fish present at the same timecode and then using the distance formula to find the distance between present fish. This is calculated using the three dimensional distance formula derived from Pythagorean's Theorem: sqrt((X1-X2)^2+(Y1-Y2)^2+(Z1-Z2)^2). The final NND column will return a single NND value for each fish in a subsample and represent the closest nearest neighbor for each fish and exclude any larger distances (in cm) so as to not skew the data away from any values that are the absolute nearest neighbor point over the course of the subsample. It is important to note that subsamples with only one observed fish will return an NND that is infinite and should be dealt with accordingly when graphing/calculating statistics of these values.  
The exemplary code is as follows:

NND\_Data <- VidSync\_Test\_Data %>%   
 filter(grepl("^Subsample.\*", objects)) %>% #In the following two lines of code, we will be removing any extraneous data points that are not relevant to the regular 3 second interval sample points taken.  
 filter(!grepl("^Length.\*", event)) %>%  
 na.omit() %>% #To remove any additional rows from the data set not associated with fish (i.e. surface shots).  
 arrange(time\_sec) %>%  
 group\_by(time\_sec) %>%  
 mutate(distance\_between\_X = X - lag(X, default = first(X))) %>%  
 mutate(distance\_between\_Y = Y - lag(Y, default = first(Y))) %>%  
 mutate(distance\_between\_Z = Z - lag(Z, default = first(Z))) %>%  
 mutate(nnd\_cm = sqrt((distance\_between\_X)^2 +   
 (distance\_between\_Y)^2 +  
 (distance\_between\_Z)^2)) %>%  
 filter(!nnd\_cm == 0) %>%  
 filter(nnd\_cm == min(nnd\_cm)) %>%  
 ungroup() %>%   
 distinct(fish\_ID, .keep\_all = TRUE) %>%   
 arrange(subsample, fish\_ID, time\_sec) %>%  
 select(subsample, fish\_ID, nnd\_cm)

## Distance per Time

Travel speeds were originally used as a variable of energy expedature when considering optimal foraging theory (Werner and Hall 1974). Recently, this idea has expanded into bioenergetics modeling, of which swimming speeds are considered into the negative flux of energy in net rate of energy intake (NREI) calculations (Fausch 1984; Hughes 1990; Hayes et al. 2000; Rosenfeld and Taylor 2009). These energy expenditures, when combined with energy intake via calculations of available prey and capture success can be converted into overall NREI values which tell us whether or not the biotic and abiotic conditions allow for potential energy profit (Fausch 1984; Hughes 1990).  
 Calculating distance per time is done by tracking a specific individual over the course of a subsample and dividing the distance travelled between each sample point by the time between each point. The distance between each point is calculated by emploring the use of the three dimensional distance formula (see NND calculation).   
The exemplary code is as follows:

DistancePerTime\_Data <- VidSync\_Test\_Data %>%   
 filter(!grepl("Surface\_Shots.\*", objects)) %>%  
 filter(!grepl("^Length.\*", event)) %>%  
 group\_by(fish\_ID) %>%   
 mutate(distance\_travelled\_X\_cm = X - lag(X, default = first(X))) %>%  
 mutate(distance\_travelled\_Y\_cm = Y - lag(Y, default = first(Y))) %>%  
 mutate(distance\_travelled\_Z\_cm = Z - lag(Z, default = first(Z))) %>%  
 mutate(fish\_distance\_travelled\_cm = sqrt((distance\_travelled\_X\_cm)^2  
 + (distance\_travelled\_Y\_cm)^2  
 + (distance\_travelled\_Z\_cm)^2)) %>%  
 ungroup() %>%   
 group\_by(fish\_ID) %>%  
 mutate(distance\_cm\_per\_sec = fish\_distance\_travelled\_cm /(time\_sec - lag(time\_sec, default = first(time\_sec)))) %>%  
 filter(!distance\_cm\_per\_sec == Inf) %>%   
 arrange(subsample, fish\_ID, time\_sec) %>%  
 select(subsample, fish\_ID, time\_sec, distance\_cm\_per\_sec)

## Proportion of Behaviors

The event types will be represented by the behavioral classifications assigned to each 3 second sample increment in each subsample, this length of time allows for classification of unique behaviors as well as tracking movement accurately with small temporal increments (Rossi et al. 2019 in prep). The behavior classifications, as well as additional points to be recorded, and their definitions have been used or adopted from prior behavioral studies with some modifications (Kalleberg 1958; Nielsen 1992; Nakano et al. 1999a). They are as follows:  
  
\*Benthic Forage\*: A distinct forage event targeted at the stream benthos.  
\*Drift Forage\*: A fish maintaining positive rheotaxis to target prey carried through the water column in a sit-and-wait fashion.   
\*Search Forage\*: A fish covering large foraging patches in search of non-benthic prey.  
\*Surface Strike\*: A fish making contact with the water surface to obtain prey.  
\*Aspirating\*: A fish visibly flaring their gills while minimally moving and making no forage attempts.   
\*Attack\*: A fish making contact with another fish in an act of territoriality.  
\*Feint\*: Adopted from a fencing term, a fish will charge another in an act of territoriality, but make no contact and quickly resume foraging thereafter.  
\*Movement\*: Any movement throughout the pool not focused on foraging.  
\*Length\*: A fork length measurement taken by creating a point at the nose and the fork of the caudal fin and using the three-dimensional distance formula to calculate length.  
\*Surface Shots\*: Points taken on the surface (usually of an easily identifiable floating object) to record a Z-plane value for use in determining the distance fish are holding from the surface.  
  
To assign a behavior type we use regular expressions to identify the behavior from the predetermined object type as specified above in the behavioral classification section. After each sample point has been identified and placed in a bin of behavior types using a nested "if-else" chain, specific behaviors can then be filtered out depending upon the target variables (i.e. only foraging behavior types).  
The exemplary code is as follows:

ForageBehaviors\_Data <- VidSync\_Test\_Data %>%   
 filter(!grepl("Surface\_Shots.\*", objects)) %>%  
 filter(!grepl("^Length.\*", event)) %>%  
 mutate(Behaviors = if\_else(grepl("^Drift\_Forage", event), "Drift Forage",   
 if\_else(grepl("^Search\_Forage", event), "Search Forage",   
 if\_else(grepl("^Search\_Forage", event), "Search Forage",   
 if\_else(grepl("^Benthic\_Forage", event), "Benthic Forage",   
 if\_else(grepl("^Feint", event), "Feint",   
 if\_else(grepl("^Attack", event), "Attack",  
 if\_else(grepl("^Surface\_Strike", event), "Surface Strike",   
 if\_else(grepl("^Movement", event), "Movement", "NA"))))))))) %>% #Note that "nip" has since been changed to the definition of "attack" and attack to feint  
 arrange(subsample, fish\_ID, time\_sec) %>%   
 select(subsample, fish\_ID, time\_sec, Behaviors)

## Distance from Surface

The exemplary code is as follows:

DFS\_Mean <- VidSync\_Test\_Data %>%   
 filter(grepl("Surface\_Shots.\*", objects)) %>%   
 summarise(mean(Z)) %>%   
 as.matrix()  
  
DistFromSurface\_Data <- VidSync\_Test\_Data %>%  
 filter(!grepl("^Length.\*", event),   
 !grepl("Surface\_Shots.\*", objects)) %>%  
 select(subsample, fish\_ID, time\_sec, Z, event) %>%   
 group\_by(fish\_ID) %>%  
 mutate(mean = mean(Z)) %>%  
 mutate(DFS\_cm = abs(mean - DFS\_Mean)) %>%   
 distinct(fish\_ID, .keep\_all = TRUE) %>%   
 select(subsample, fish\_ID, DFS\_cm)

## Fork Length Calculations

The exemplary code is as follows:

Length\_Data <- VidSync\_Test\_Data %>%   
 filter(grepl("^Length.\*", event)) %>%   
 na.omit() %>%   
 group\_by(event) %>%   
 mutate(length\_mm = 10\*sqrt((X - lag(X, default = first(X)))^2 +   
 (Y - lag(Y, default = first(Y)))^2) +   
 (Z - lag(Z, default = first(Z)))^2) %>%   
 filter(!length\_mm == 0) %>%   
 group\_by(subsample, fish\_ID) %>%   
 summarise(length\_mm = mean(length\_mm)) %>%   
 arrange(subsample, fish\_ID)

## Occupied Volumes

Vivancos and Class looked into area use, but not volume. Glass 2013 looked into volume but not very in depth.  
  
This method makes use of delaunay triangulation to calculate the volumes of a three-dimensional point cloud using the Geometry package in R ()  
The exemplary code is as follows:

Volume\_Data <- VidSync\_Test\_Data %>%   
 filter(!grepl("Surface\_Shots.\*", objects)) %>%  
 filter(!grepl("^Length.\*", event)) %>%  
 select(subsample, fish\_ID, X, Y, Z) %>%  
 arrange(subsample, fish\_ID)  
  
Test\_Volume <- Volume\_Data %>%   
 filter(fish\_ID == 1) %>% #Using a single volume output from a subsample as an example data point  
 select(X, Y, Z) %>%  
 as.matrix() %>%  
 geometry::convhulln("FA")   
 print(Test\_Volume$vol) #Displayed in cubic centimeters

## [1] 76.84104

#This can also be run in a for loop over the entire dataset from the sample to occupy a data frame with the collected volume data  
Vol\_forloop <-   
for (i in 1:33) {  
 Volume\_Data %>%  
 filter(fish\_ID == i) %>%  
 select(X, Y, Z) %>%  
 as.matrix() %>%  
 geometry::convhulln("FA")   
}

# Discussion

challenges of using

This study involved both Lagrangian and Eulerian sampling of organisms in the stream environment. The Lagrangian approach involves tracking an organism through time and space, while the Eulerian approach involves repeated sampling of animal distribution at fixed points in space and time (Smouse et el. 2010, Price 2006). In Elder Creek video of fish behavior was used to track individuals through time and space (Lagrangian) and to take scan samples of fish behavior over a fixed time interval (Eulerian).

However, Lagrangian analysis in vidsync can also be done by moving the cameras to track fish (as Jason did in his Chinook feeding studies) rather than having the cameras in a fixed point.

Previously the habitat use of a foraging fish has been defined as a two dimensional area made from position estimations of fish from snorkel observations.   
hypervolume definition  
recent hypervolume work -- Blonder et al 2013

# Acknowledgements

“You better stay away from Copperhead Road” - Steve Earl

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