Supplementary materials for the manuscript: Repeatability and intra-class correlations from time-to-event data

Kelsey B. McCune¹, Coralie Williams^{2,3}, Ned A. Dochtermann⁴, Holger Schielzeth⁵ and Shinichi Nakaga

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Affiliations:

- 1) College of Forestry, Wildlife and Environment, Auburn University, Auburn, AL, USA
- 2) Evolution & Ecology Centre and School of Biological, Earth and Environmental Sciences, University of New South Wales, Sydney, New South Wales, Australia
- 3) School of Mathematics and Statistics, University of New South Wales, Sydney, New South Wales, Australia
- 4) Department of Biological Sciences, North Dakota State University, Fargo, North Dakota, USA
- 5) Institute for Ecology and Evolution, Friedrich Schiller University Jena, Jena, Thuringia, Germany
- 6) Department of Biological Sciences, University of Alberta, CW 405, Biological Sciences Building, Edmonton, Canada

Background

In the manuscript, "Repeatability and intra-class correlations from time-to-event data: towards a standard-ized approach" we describe a method to quantify variances and intra-class correlations (repeatability) from time-to-event data. In this supplementary materials page we demonstrate the use of this method with real data in 3 worked examples. We additionally provide the data and code used in the case study section of the manuscript. Lastly, we present the methods and results for a simulation study to illustrate the relationship among model estimates (beta, random effect variance and intra-class correlation values) derived from 4 different models and implementations run on data simulated with varying random effect variance, time intervals, and amount of censored data.

Worked Examples

Time-to-event data range widely in the number of individuals (or other cluster variable) as well as the number of repeated measures on each individual. Furthermore, the presence and amount of censored data can vary. We use openly available data to present 3 worked examples that vary in model parameters, sample sizes and amount of censoring. The first example involves models that do not include fixed effects, but include one random effect. In the second example, the models included 1 fixed effect and 1 random effect, while the third example involves 1 fixed effect and 2 random effects.

Example 1: Frog aggression towards a simulated intruder

Our first example data set is from Peignier et al. 2022. They investigated whether A. femoralis frogs show repeatable aggressive behavior by measuring the latency of territorial males to approach a simulated intruder.

^{*}Corresponding author: kelseybmccune@gmail.com

Their sample size included 51 males with 3.2 ± 1.31 trials each. Censored data are present (6% of trials) because some males did not approach the simulated intruder. These individuals were given a ceiling value of 300 seconds (the longest trial time). To evaluate repeatability, the authors log-transformed this variable and used the Gaussian family in the rpt function (Stoffel et al. 2017).

In this first worked example, we give a high level of detail on the methods and model types described in the paper. In the subsequent examples, we present the data and run the analysis while elaborating less on the intermediary or alternative steps.

```
# Data from Peignier et al. 2022

fdata = read.csv("https://raw.githubusercontent.com/kelseybmccune/Time-to-Event_Repeatability/main/data
fdata = fdata[which(fdata$sex == "m"),c(1,2,9)] # Only males participated in the aggression test, we on
fdata = fdata[!is.na(fdata$jump_s),] # Unclear what the NAs represent, but the authors remove these dat
fdata$event = ifelse(fdata$jump_s==300, 0,1) # Create event column that indicates whether the event occ
head(fdata,10)
```

```
##
              ID repetition jump_s event
## 1
      i16-m-018
                           1
                                 17
## 2
      i16-m-018
                           2
                                300
                                         0
## 3 i16-m-018
                           3
                                  3
## 4 i16-m-018
                           4
                                 49
                                         1
      i16-m-059
                           1
                                  4
## 5
## 6 i18-m-001
                           1
                                  5
                                         1
## 7
      i18-m-009
                           1
                                  5
                                         1
## 8 i18-m-009
                           2
                                  4
                                         1
## 9 i18-m-009
                           3
                                 11
                                         1
## 10 i18-m-009
                                  6
                                         1
```

This is a classic (un-exploded) format for animal personality data.

ICC from the coxme function with the classic data set We will first conduct a Cox proportional hazards model using the coxme package in R (Therneau, 2022). The coxme function is able to estimate multiple random effects, as well as random slopes. We included a random effect for individual identity as in Equation 3 of the main text, except that there are no fixed effects in this frog model. In this model specification, the response variable is a "survival" object that combines latency to jump ("jump_s") and the event indicator.

library(coxme)

```
## Loading required package: survival
## Loading required package: bdsmatrix
##
## Attaching package: 'bdsmatrix'
## The following object is masked from 'package:base':
##
## backsolve
```

```
library(survival)
c1 = coxme(Surv(jump_s,event)~1+(1|ID), data = fdata)
var <- VarCorr(c1)$ID[[1]] # Random effect variance</pre>
```

We can now estimate the ICC using Equation 7 from the main text. Because of the equivalence of a mixed-effects Cox proportional hazards model with the binomial generalized linear mixed-effects model with the clog-log link, Equation 7 uses the random effect variance from the Cox model and the binomial GLMM distribution-specific residual variance estimator (Nakagawa et al. 2017).

```
var/(var + pi^2/6)
## [1] 0.1948825
# Original paper reports a repeatabiltiy of 0.24 (CI: 0.07-0.40)
```

This yields an ICC value of 0.19.

We created two R functions to estimate the p-value and a confidence interval for time-to-event ICCs calculated using coxme.

```
# Define the function
# missing values ignored
#' @title comxe_pval
#' @description This function calculates the p-value of the effect of the random effect in a coxme mode
#' Oparam model A coxme model object
#' Oparam data The original data used to fit the model
#' @param boot Number of simulations to run to produce 95 percent confidence intervals for I2. Default
#' @return A vector of p-values
#' @author Shinichi Nakagawa - s.nakagawa@unsw.edu.au
#' @author etc
coxme_pval <- function(model, data, boot = NULL) {</pre>
  # Get the original data
  if(all(class(model) %in% c("coxme")) == FALSE) {stop("Sorry, you need to fit a coxme model of class c
  # I think we need to use get the dimension of the data
  response <- as.data.frame(model$y[,1:2])
  fixed_formula <- as.formula(model$formulaList$fixed)</pre>
  # fit the model without any random effects
  fit <- survival::coxph(as.formula(fixed_formula), data = data)</pre>
  # loglikelihood ratio test
  # this is p value of effect of taking all random effects
  pval<- anova(fit, model)$P[-1]</pre>
  names(pval) <- "liklihood_ratio_test"</pre>
  if(!is.null(boot)){
```

```
# we need to use replicate to create many vectors of these - randomize the data
orders <- replicate(boot, sample(1:nrow(response)))</pre>
fixed formula <- as.character(fixed formula)</pre>
random_formula <- as.vector(as.character(model$formulaList$random))</pre>
formula <- as.formula(paste("Surv(new_time, new_status)",</pre>
                          fixed formula[3],
                          0+0.
                          paste(random_formula, collapse = "+")))
data2 <- data
# randomizaton/permutation tests
pb <- progress::progress_bar$new(total = boot,</pre>
                                 format = "Bootstrapping [:bar] :percent ETA: :eta",
                                 show_after = 0)
# loop
num <- length(summary(model)$random$variance)</pre>
store <- matrix(NA, nrow = num, ncol = boot)</pre>
# Loop over the number of bootstraps
for (i in 1:boot) {
# Permute the data
data2$new_time <- response$time[orders[ ,i]]</pre>
data2$new_status <- response$status[orders[ ,i]]</pre>
# Fit the original coxme model
temp <- tryCatch(coxme(formula, data = data2))</pre>
# get variance component
store[ ,i] <- summary(temp)$random$variance</pre>
pb$tick()
Sys.sleep(1 / boot)
  }
# getting the p value
pval2 <- sapply(1:num, function(x) {</pre>
   sum(store[x,] > summary(model)$random$variance[x])/boot}
names(pval2) <- paste(rep("bootstrapped_pval", num), 1:num, sep = "_")</pre>
if(exists("pval2")) {
res <- c(pval, pval2)
return(res)
```

```
} else {
  res <- pval
  return(res)
}
#' @title coxme_icc_ci
#' @description This function calculates the 95 percent confidence interval for the intraclass correlat
#' @param model A coxme model object
#' @param upper.multiplier The multiplier for the upper bound of the confidence interval. Default is 10
#' Creturn A vector of the lower, point estimate, and upper bounds of the 95 percent confidence interva
#' @author Shinichi Nakaqawa - s.nakaqawa@unsw.edu.au
#' @author etc
coxme_icc_ci <- function(model, upper.multiplier = 10) {</pre>
  if(all(class(model) %in% c("coxme")) == FALSE)
    {stop("Sorry, you need to fit a coxme model of class coxme")}
  if(any(length(summary(model) random variance) > 1)) {stop("Sorry. At the moment, we can only have a m
  # Define a sequence of variance values``
  # the length of the response
  n <- nrow(model$y)</pre>
  cut = 100
  var_point <- summary(model)$random$variance</pre>
  # based on this pdf: https://cran.r-project.org/web/packages/coxme/vignettes/coxme.pdf
  # upper CI is limited to var_point*(10*log(n)) - so this could fail
  estvar1 <- seq(0.00000000000001, var_point, length = cut)</pre>
  estvar2 <- seq(var_point, var_point*upper.multiplier, length = cut+1)[-1]</pre>
  estvar <- c(estvar1, estvar2)</pre>
  # Initialize a vector to store the log-likelihood values
  loglik <- double(cut)</pre>
  # Loop over the variance values
  for (i in seq_len(cut*2)) {
    # Fit a coxme model with fixed variance
    tfit <- update(model, vfixed = estvar[i])</pre>
    # Compute the log-likelihood
    loglik[i] <- 2 * diff(tfit$loglik)[1]</pre>
  }
  # Compute the threshold for the 95% confidence interval
  temp <- as.numeric(2 * diff(model$loglik)[1]) - loglik</pre>
  # Find the variance values that correspond to the threshold
  # getting lower and upper CI using profile likelihood
  lower <- approx(temp[1:(cut)], sqrt(estvar[1:(cut)]), qchisq(.95, 1))$y</pre>
  upper \leftarrow approx(temp[(cut + 1):(2*cut)], sqrt(estvar[(cut + 1):(2*cut)]), qchisq(.95, 1))$y
```

```
# Return the 95% confidence interval
  ICC_lower <- lower^2 / (lower^2 + pi^2 / 6)</pre>
  if (is.na(ICC lower)) {
    ICC lower <- 0
  }
  ICC_point <- var_point / (var_point + pi^2 / 6)</pre>
  ICC_upper <- upper^2 / (upper^2 + pi^2 / 6)</pre>
  names(ICC lower) <- "lower"</pre>
  names(ICC_point) <- "ICC"</pre>
  names(ICC_upper) <- "upper"</pre>
  return(c(ICC_lower, ICC_point, ICC_upper))
}
coxme_pval(c1,fdata,boot=100) # as there were no fixed effects in our model, the likelihood ratio test
## liklihood_ratio_test bootstrapped_pval_1
coxme_icc_ci(c1, upper.multiplier = 10) # confidence interval and ICC estimate
##
        lower
                      ICC
                                upper
## 0.06143739 0.19488246 0.36584291
```

ICC from coxme with an exploded data set To check the equivalence in time-to-event ICC values between Cox proportional hazards models and binomial GLMM, we need a data set that includes discrete time intervals. This is referred to as an "exploded" data set (see Fig. 1 in the main text) and we can use the function survSplit in the survival package (Therneau, 2024) to easily transform the data. In a simulation study (see below), we found no difference in model estimates or ICC values when we used 2 or 4 time intervals for the exploded data sets.

```
# decide on the intervals
psych::describe(fdata$jump_s)
##
      vars
             n mean
                        sd median trimmed
                                             mad min max range skew kurtosis
## X1
         1 163 53.16 92.25
                               10
                                     29.18 10.38
                                                   1 300
                                                           299 2.02
                                                                          2.5 7.23
fdata_int = survSplit(Surv(jump_s,event) ~ repetition+ID, data = fdata,
                      cut = 150, start = "tstart", end = "tstop") # two time intervals
head(fdata_int,10)
```

```
##
      repetition
                         ID tstart tstop event
## 1
                1 i16-m-018
                                  0
                                       17
                                               1
## 2
               2 i16-m-018
                                  0
                                       150
                                               0
## 3
                                150
                                      300
                                               0
               2 i16-m-018
## 4
               3 i16-m-018
                                  0
                                        3
                                               1
## 5
               4 i16-m-018
                                  0
                                       49
                                               1
## 6
               1 i16-m-059
                                  0
                                         4
                                               1
```

```
## 7
                 1 i18-m-001
                                    0
                                          5
## 8
                 1 i18-m-009
                                    0
                                          5
                                                 1
## 9
                2 i18-m-009
                                    0
                                          4
                                                 1
## 10
                3 i18-m-009
                                    0
                                                 1
                                         11
```

In the exploded data set with two time intervals, for each individual and trial we have a time interval that ranges from 0 to 150 and 150 to 300 (the ceiling latency value for this data set). If the event occurs, the tstop value represents the time at which the frog approaches the speaker.

Now we can compare the Cox model output between the classic and the exploded data set. The response variable is again a "survival" object, but now combines the start and stop time of the interval and the event indicator.

```
c1_int = coxme(Surv(tstart,tstop,event)~1+(1|ID), data = fdata_int)
var <- VarCorr(c1_int)$ID[[1]]</pre>
```

You can see that the variance estimate for the random effect is the same in both Cox models.

In Equation 5 of the main text we show the calculation for the non-parametric version of the ICC, Kendall's tau, which assumes a Gamma distribution of the random effect. In addition to being non-parametric, another limitation is that it is not feasible to estimate the non-parametric ICC when there are more than one random effects. We wrote a function to obtain the non-parametric ICC numerically:

```
# function obtained from
# https://qithub.com/cran/parfm/blob/master/R/frailtydistros.R
g <- function(w, k, s, sigma2) {
  -k * w + exp(w) * s + w ^ 2 / (2 * sigma2)
}
g1 <- function(w, k, s, sigma2) {
  -k + exp(w) * s + w / sigma2
g2 <- function(w, k, s, sigma2) {</pre>
  exp(w) * s + 1 / sigma2
}
Lapl <- Vectorize(function(s, k, sigma2) {</pre>
  # Find wTilde = max(g(w)) so that g'(wTilde; k, s, theta) = 0
  WARN <- getOption("warn")</pre>
  options(warn = -1)
  wTilde <- optimize(f = g, c(-1e10, 1e10), maximum = FALSE,
                     k = k, s = s, sigma2 = sigma2)$minimum
  options(warn = WARN)
  # Approximate the integral via Laplacian method
  res <- (-1) ^ k *
    exp(-g(w = wTilde, k = k, s = s, sigma2 = sigma2)) /
    sqrt(sigma2 * g2(w = wTilde, k = k, s = s, sigma2 = sigma2))
  return(res)
}, 's')
```

```
intTau <- Vectorize(function(x, intTau.sigma2=sigma2) {</pre>
  res <- x *
    Lapl(s = x, k = 0, sigma2 = intTau.sigma2) *
    Lapl(s = x, k = 2, sigma2 = intTau.sigma2)
 return(res)
}, "x")
fr.lognormal <- function(k,</pre>
                          sigma2,
                          what = "logLT") {
  # if (!(is.numeric(sigma2) & (sigma2 > 0)))
  # stop("The parameter sigma2 is not a positive value.")
  if (what == "logLT") {
    # if (!(is.numeric(s) & (s > 0)))
          stop("The parameter s is not positive.")
    # Find wTilde = max(g(w)) so that g'(wTilde; k, s, theta) = 0
    WARN <- getOption("warn")</pre>
    options(warn = -1)
    wTilde \leftarrow nlm(f = g, p = 0, k = k, s = s, sigma2 = sigma2)$estimate
    options(warn = WARN)
    # Approximate the integral via Laplacian method
    res \leftarrow -g(w = wTilde, k = k, s = s, sigma2 = sigma2) -
      log(sigma2 * g2(w = wTilde, k = k, s = s, sigma2 = sigma2)
      ) / 2
    return(res)
  else if (what == "tau") {
    intTau <- Vectorize(function(x, intTau.sigma2=sigma2) {</pre>
      res <- x *
        Lapl(s = x, k = 0, sigma2 = intTau.sigma2) *
        Lapl(s = x, k = 2, sigma2 = intTau.sigma2)
      return(res)
    }, "x")
    tauRes <- 4 * integrate(</pre>
      f = intTau, lower = 0, upper = Inf,
      intTau.sigma2 = sigma2)$value - 1
    return(tauRes)
  }
}
c1_ph = coxph(Surv(jump_s, event)~1 + frailty(ID, distribution="gamma"), data=fdata)
var <- as.numeric(c1_ph$history$`frailty(ID, distribution = "gamma")`[1]) # Random effect variance is c
fr.lognormal(k,s,var,what = "tau") # non-parametric ICC estimate
```

[1] 0.1428707

The non-parametric ICC (ICC_{np}) estimate for these frog data is 0.14.

ICC from binomial GLMM with the exploded data set We now demonstrate the use of the binomial GLMM to analyze the exploded survival data. However, we also need to include a column as an identifier of the time interval to include as a fixed effect, which takes the place of the start and stop time from the Cox models.

The random effect variance from the binomial GLMM is 0.24, and the ICC value using Equation 7 is 0.13.

Example 2: Christmas tree worm re-emergence after a simulated predator attack

[1] 0.1291067

Our next example data come from Pezner et al. 2017 https://academic.oup.com/beheco/article/28/1/154/2453511. They tested the consistency of hiding time (latency to re-emerge from the hole) within and across days, and the impact of the social environment. They tested 30 worms, each received 4 trials within a day and 4 days of sampling for a total of 16 trials per worm. Censored data are not explicitly stated, but there are 5 NA values from 2 individuals that we here assume are censored data (~1% of data). In the original paper, the authors log-transformed the hiding time variable, used a linear mixed model to get variance estimates, then hand-calculated repeatability.

```
## Load the data
ctw = read.csv("https://raw.githubusercontent.com/kelseybmccune/Time-to-Event_Repeatability/main/data/C
# "HT" variable indicates hiding time, or the latency to emerge; "Whorls" is a visual indicator of age

# 2 individuals have NA values, but it is not explained why.
# For this worked example, we assume these are censored (the worm didn't emerge in the trial time) and
ctw$event = ifelse(is.na(ctw$HT),0,1) # create the event column
ctw$HT[which(is.na(ctw$HT))]<- 375 # identify ceiling values

## Coxme with classic data
ctw.cox = coxme(Surv(HT, event) ~ Whorls + (1|Worm_ID), data = ctw)
cox.zph(ctw.cox) # Whorls does change with time, violating the proportional hazards assumption

## chisq df p
## Whorls 19.5 1 1e-05
## GLOBAL 19.5 1 1e-05</pre>
```

```
# use functions for ICC, confidence interval and p-value
coxme_icc_ci(ctw.cox, upper.multiplier = 10)
       lower
                   ICC
                           upper
## 0.2189798 0.3286071 0.4758776
coxme_pval(ctw.cox,ctw,boot=100)
## liklihood_ratio_test bootstrapped_pval_1
## Coxme with exploded data
# decide on the intervals
psych::describe(ctw$HT)
##
                        sd median trimmed
                                            mad min max range skew kurtosis
             n mean
         1 475 37.75 57.78
                                     24.02 11.86
                                                   5 375
                                                           370 4.09
ctw_int = survSplit(Surv(HT,event) ~ Whorls + Trial_Total + Worm_ID, data = ctw,
                    cut = c(94,188,282), start = "tstart",end = "tstop") # 4 time intervals
ctw.cox.int = coxme(Surv(tstart,tstop, event)~Whorls + (1 | Worm_ID), data=ctw_int)
var <- VarCorr(ctw.cox.int)$Worm_ID[[1]]</pre>
var/(var + pi^2/6)
## [1] 0.3286071
# Original paper finds that the across-day hiding time repeatability was 0.42
## Binomial glmm model with exploded data
ctw_int$interval = NA
ctw_int$interval = ifelse(ctw_int$tstart == 0,1,ctw_int$interval)
ctw_int$interval = ifelse(ctw_int$tstart == 94,2,ctw_int$interval)
ctw_int$interval = ifelse(ctw_int$tstart == 188,3,ctw_int$interval)
ctw_int$interval[which(is.na(ctw_int$interval))]<-4</pre>
b2 <- glmer(event ~ as.factor(interval) + Whorls + (1 | Worm_ID), data=ctw_int,
                 family = binomial(link="cloglog"), nAGQ=7)
var <- b2@theta^2 # extract the random effect variance from the model output object
var/(var + pi^2/6)
```

Example 3: Repeatability of distance to cache a seed in smalll mammals

[1] 0.2633556

Brehm et al. 2019 (https://onlinelibrary.wiley.com/doi/full/10.1111/ele.13324) evaluated seed caching behavior in relation to multiple personality traits and habitat characteristics. They offered artificial seeds to

several species of small mammals that had reflective flags attached so that, if the individual cached the seed, it could be relocated and the distance from the food platform could be measured. The data include 17 short-tailed shrews, 47 red-backed voles and 46 unique deer mice that cached between 1-14 seeds (mean \pm se = 3.9 \pm 0.73). Understandably, it can be hard to relocate the cached seeds and so 52% of the caching distances were censored (seeds lost to follow up). For our worked example, we considered the repeatability of caching distance as a function of the fixed effect of silviculture (habitat) treatment, and random effects for individual ID and for species. The original paper did not run this particular analysis.

```
## Load the data
data <- read.csv("https://raw.githubusercontent.com/kelseybmccune/Time-to-Event_Repeatability/main/data/d
distmov<-data[-which(is.na(data$RECOVERED..Y.N.)),] # distance the seed is dispersed, only looking at s
distmov <- subset (distmov, CONS!=1 & REMOVE==1) # remove rows where the seed was consumed close by the fe
dist = distmov[-which(distmov$DIST..MOVED == 0),c(1,3,5,14,17)] #simplify data frame
# the Recovered column is analogous to the event variable in survival analysis. Modify it to be an inte
dist$event = ifelse(dist$RECOVERED..Y.N.== "Y",1,0)
dist$DIST..MOVED[which(dist$RECOVERED..Y.N. == "N")] <-1038 # give ceiling value to NAs
dist = dist[-which(is.na(dist$DIST..MOVED)),] # Remove rows where seeds are indicated as recovered, but
table(dist$RECOVERED..Y.N.) # how many cached seeds did they recover (i.e., how much censored data is t
##
##
             Y
## 130
         1 122
# 52% of data are censored
## Coxme with classic data set structure
dist.cox = coxme(Surv(DIST..MOVED, event) ~ TRT + (1 | ID) + (1 | SPP), data=dist)
cox.zph(dist.cox) # Fixed effect does not violate proportional hazard assumption
          chisq df
                      р
## TRT
          0.313 2 0.86
## GLOBAL 0.313 2 0.86
var <- VarCorr(dist.cox)$ID[[1]]</pre>
var/(var + pi^2/6)
## [1] 0.2796251
var2 <- VarCorr(dist.cox)$SPP[[1]]</pre>
var2/(var2 + pi^2/6)
## [1] 0.1838078
coxme_pval(dist.cox,dist,boot=100) # bootstrapping: ID p-value < 0.01; SPP p-value < 0.01
## liklihood_ratio_test bootstrapped_pval_1 bootstrapped_pval_2
            4.87077e-05
                                 0.00000e+00
                                                       0.00000e+00
```

```
# likelihood ratio test compares the increase in variance explained by the addition of the random effec
#coxme_icc_ci(dist.cox) # currently this function only works with one random effect
## Explode the data
# Create intervals
psych::describe(dist$DIST..MOVED)
##
                mean
                          sd median trimmed mad min max range skew kurtosis
## X1
       1 253 634.42 446.33
                               1038
                                      660.1 0
                                                  1 1038 1037 -0.3
dist_int = survSplit(Surv(DIST..MOVED, event) ~ TRT + ID + SPP, data = dist,
                     cut = c(346,692), start = "tstart",end = "tstop",zero=0)
## Cox model and ICC calculation
dist.cox.int = coxme(Surv(tstart,tstop, event) ~ TRT + (1 ID) + (1 SPP), data=dist_int)
var <- VarCorr(dist.cox.int)$ID[[1]]</pre>
var/(var + pi^2/6)
## [1] 0.2799994
var2 <- VarCorr(dist.cox.int)$SPP[[1]]</pre>
var2/(var2 + pi^2/6)
## [1] 0.1836764
## Binomial glmm model with exploded data
dist int$interval = NA
dist_int$interval = ifelse(dist_int$tstart == 0,1,dist_int$interval)
dist_int$interval = ifelse(dist_int$tstart == 346,2,dist_int$interval)
dist_int$interval = ifelse(dist_int$tstart == 692,3,dist_int$interval)
b3 <- glmer(event ~ as.factor(interval) + TRT + (1|ID) + (1|SPP), data=dist_int,
                 family = binomial(link="cloglog"))
var <- b3@theta[[1]]^2 # extract the random effect variance from the model output object
var/(var + pi^2/6) # ICC for ID
## [1] 0.1521422
var2 <- b3@theta[[2]]^2 # extract the random effect variance from the model output object
var2/(var2 + pi^2/6) # ICC for SPP
## [1] 0.126801
```

From McCune et al. 2019.

Data and code for Case Study in the main text

```
#Latency to solve a door on a puzzle box
jsolv = read.csv("https://raw.githubusercontent.com/kelseybmccune/Time-to-Event_Repeatability/main/data
jsolv = jsolv[,c(2:6,9)]
colnames(jsolv)[6] = "Time"
solv.su = coxme(Surv(Time, Solve)~Treatment + (1|ID), data=jsolv)
cox.zph(solv.su) # fixed effect does not violate the proportional hazards assumption
##
             chisq df
                   1 0.094
## Treatment
               2.8
## GLOBAL
               2.8 1 0.094
coxme_pval(solv.su,jsolv,boot = 100)
## liklihood_ratio_test bootstrapped_pval_1
##
           5.687028e-08
                                0.00000e+00
coxme_icc_ci(solv.su) # adjusted ICC
##
       lower
                   ICC
                           upper
## 0.3784428 0.6578112 0.8652368
solv.su2 = coxme(Surv(Time,Solve) ~ 1 + (1|ID), data = jsolv)
coxme_pval(solv.su2,jsolv,boot = 100)
## liklihood_ratio_test
                         bootstrapped_pval_1
##
coxme_icc_ci(solv.su2) # unadjusted ICC
##
       lower
                   TCC
                           upper
```

Simulations

0.4861004 0.7355558 0.8999777

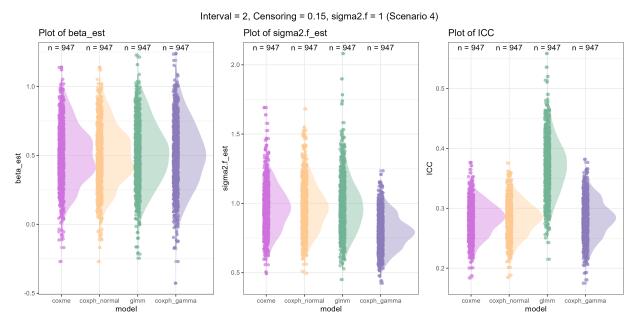
We conducted a small simulation study to examine whether model estimates are consistent across four different models and implementations: (1) a Cox proportional hazards model with a normal distribution of the random effect using the coxme function from the coxme package, (2) a Cox proportional hazards frailty model with a normal distribution of the random effect using coxph from the survival package, (3) a Cox proportional hazards frailty model with a gamma distribution of the random effect using coxph from the survival package, and (4) a binomial generalized linear mixed-effects model with a clog-log link using the glmer function from the lme4 package. The models (1), (2) and (3) are described in Equation 3 and model (4) is described in Equation 7 in the main text.

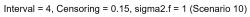
Time-to-event data were simulated with a single predictor variable (sex) and a single random effect (i.e. cluster or individual) α , as described in the main text. We considered n=1000 observations and $n_{cluster}=100$ clusters (i.e. individuals). To evaluate performance of the above models, we simulated different data sets where we varied the magnitude of the random effect variance (σ_{α}^2) from 1 to 3, the percent of censored data was either 0 or 0.15, and the number of time intervals was either 2 or 4. We ran each model on each data set

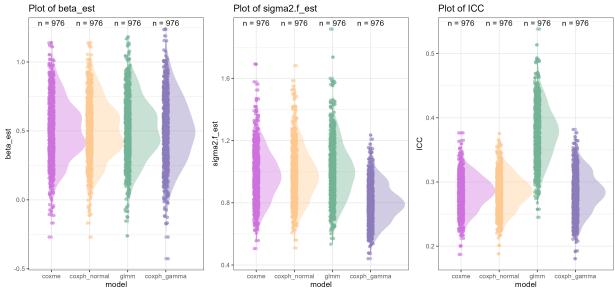
1000 times. We excluded results if at least one of the models did not converge or had a convergence warning message.

We present illustrative plots comparing model performance when the random effect variance is set to 1 or 2, censoring = 0.15 and time intervals = 2 or 4. These simulations had the least convergence problems or warnings out of the 1000 iterations.

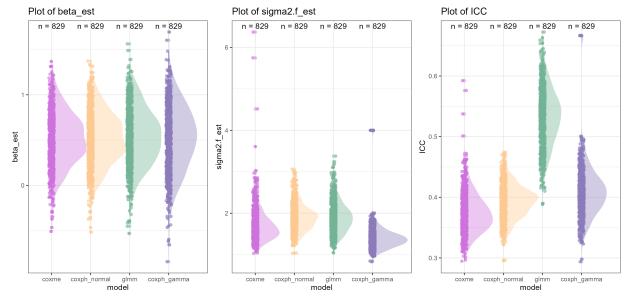
Examining the model performance within each plot, we found that the beta estimates of the predictor variable (β_{est}) are identical and relatively unbiased across all models. The variance estimates of the random effect $(\sigma_{f_{\text{est}}}^2)$ are very similar for the coxme, coxph, and glmm models, while the coxph_gamma model consistently underestimates this variance. Additionally, the intra-class correlation coefficient (ICC) is consistently higher in the glmm model compared to the coxme and coxph models. This aligns with Figure 2 in the main text, which shows that as the variance of the random effect increases, the ICC value from the GLMM model becomes proportionally larger than the non-parametric value (ICC_{np}) from the Cox models. Furthermore, comparing performance across plots demonstrates that the number of time intervals in the data (2 or 4) has no effect on the model estimates.



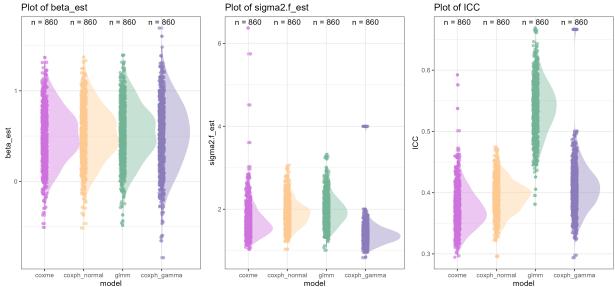




Interval = 2, Censoring = 0.15, sigma2.f = 2 (Scenario 5)







Note that 'sigma2.f' refers to the random effect variance.

The simulation study was conducted with R (R Core Team, 2024) and using the computational cluster Katana at UNSW Sydney (UNSW, 2024).

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